

Chemical Ecology of Vertebrates



This page intentionally left blank

Chemical Ecology of Vertebrates

Chemical Ecology of Vertebrates is the first book to focus exclusively on the chemically mediated interactions between vertebrates, including fish, amphibians, reptiles, birds, and mammals, and other animals, and plants. Reviewing the latest research in three core areas: pheromones (where the interactions are between members of the same species), interspecific interactions involving allomones (where the sender benefits) and kairomones (where the receiver benefits) This book draws information into a coherent whole from widely varying sources in many different disciplines. Chapters on the environment, properties of odour signals, and the production and release of chemosignals set the stage for discussion of more complex behavioral topics. While the main focus is ecological, dealing with behavior and interactions in the field, it also covers chemoreception, orientation and navigation, the development of behavior, and the practical applications of chemosignals.

Dietland Müller-Schwarze is Professor of Environmental Biology at the State University of New York.

Chemical Ecology of Vertebrates

DIETLAND MÜLLER-SCHWARZE

State University of New York



CAMBRIDGE UNIVERSITY PRESS

Cambridge, New York, Melbourne, Madrid, Cape Town, Singapore, São Paulo

Cambridge University Press

The Edinburgh Building, Cambridge CB2 2RU, UK

Published in the United States of America by Cambridge University Press, New York

www.cambridge.org

Information on this title: www.cambridge.org/9780521363778

© Cambridge University Press 2006

This publication is in copyright. Subject to statutory exception and to the provision of relevant collective licensing agreements, no reproduction of any part may take place without the written permission of Cambridge University Press.

First published in print format 2006

```
ISBN-13 978-0-511-24205-2 eBook (NetLibrary)
ISBN-10 0-511-24205-0 eBook (NetLibrary)
ISBN-13 978-0-521-36377-8 hardback
ISBN-10 0-521-36377-2 hardback
```

Cambridge University Press has no responsibility for the persistence or accuracy of URLs for external or third-party internet websites referred to in this publication, and does not guarantee that any content on such websites is, or will remain, accurate or appropriate.

CONTENTS

	Preface	page ix
	Acknowledgements	xii
1	The odorsphere: the environment for transmission of	
	chemical signals	1
1.1	Air	3
1.2	Water	15
1.3	Water-air interface	17
1.4	Influence of setting	17
2	Properties of vertebrate semiochemicals	20
2.1	Functional groups	20
2.2	Polarity	22
2.3	Solubility	22
2.4	Isomers and enantiomers	22
2.5	Volatiles	22
2.6	Non-volatiles	24
2.7	Multicomponent pheromones	26
2.8	Sex differences	31
2.9	Life expectancy of chemical signals	32
2.10	Spatial range of odor signals	33
2.11	Interaction of olfactory and visual signals	33
3	Odor production and release	36
3.1	Signals in excretions	36
3.2	Glandular secretions	38
3.3	Body odors and body region odors	48
3.4	Diet influences on odor production and venoms	49
3.5	Hormonal control of odors in urine and secretions	52
3.6	Microbial odors	53
3.7	Reservoirs	54

3.8	Pheromone transport	54
3.9	Environmental odors for communication	55
3.10	Supporting structures: osmetrichia, muscles	56
3.11	Special adaptations for broadcasting chemosignals	57
4	Chemical cues in orientation and navigation	60
4.1	Fish	61
4.2	Amphibia	68
4.3	Reptiles	69
4.4	Birds	71
4.5	Mammals	80
5	Chemoreception	82
5.1	Encounter and exposure: orientation, sniffing, licking, tasting	82
5.2	Receiving molecules: chemosensory organs	84
5.3	Structure–activity relationships	110
5.4	Neural pathways and decoding	112
5.5	Odor detection thresholds	113
5.6	Hormonal influences on chemoreception	119
5 · 7	Chemoreception and age	120
5.8	Interaction between chemical senses	121
6	Signaling pheromones I: discrimination and	
	recognition	124
6.1	Familiarization with environment and objects	125
6.2	Familiar versus non-familiar social odors	125
6.3	Recognizing kin	129
6.4	Individual odors	133
6.5	Odors in parental behavior	136
6.6	Species and population discrimination	142
6.7	Modulating behavior by status signals	144
6.8	Competition between conspecifics of the same sex	144
6.9	Liquid assets: marking territory and home range with urine and	
	secretions	151
6.10	Scent marking in mammals	153
7	Signaling pheromones II: sex and alarm pheromones	
	and evolutionary considerations	171
7.1	Sex pheromones: attracting and stimulating	171
7.2	Alarm and alert odors	191
7.3	Trail odors	196

7.4	Information about food	197
7·5	Evolutionary considerations	198
, •	•	
8	Intraspecific signals: priming pheromones	203
8.1	Fish reproduction	203
8.2	Amphibia	207
8.3	Reptiles	207
8.4	Mammals	207
8.5	Priming pheromones in humans?	224
9	Development of intra- and interspecific	
	chemical communication	227
9.1	Fish	227
9.2	Amphibia	228
9.3	Reptiles	228
9.4	Birds	231
9.5	Mammals	231
9.6	Learning	241
10	Allomones I: chemical defense by animals	246
10.1	Fish	246
10.2	Amphibia	250
10.3	Reptiles	256
10.4		259
10.5	Mammals	262
10.6	Pars pro toto: decoy odors	264
10.7	Invertebrate allomones that deter vertebrate predators	264
10.8	Recycled animal and plant materials	265
10.9	The question of coevolution between predator and prey	268
11	Allomones II: plant chemical defenses	
	against herbivores	270
11.1	Classes of plant defense compound	271
11.2	Physiological effects of secondary plant metabolites	284
11.3	Chemical defense strategies by plants	299
11.4	Feeding or avoiding? herbivores vis-à-vis plant	
•	defenses	303
11.5	Plant responses to herbivory	332
11.6	The question of coevolution of plants and herbivorous	
	mammals	334

CONTENTS vii

viii CONTENTS

12	Kairomones and synomones	338
12.1	Predator–prey interactions	338
12.2	Host odors used by parasites	371
12.3	Eavesdropping	374
12.4	The self-anointed: chemical mimicking	376
12.5	Evolutionary considerations	377
12.6	Plant chemicals used by vertebrates	378
12.7	Animal chemicals benefiting plants	383
12.8	Synomones	383
13	Practical applications of semiochemicals	391
13.1	Fish	391
13.2	Reptiles	393
13.3	Birds	394
13.4	Non-human mammals	397
13.5	Humans	418
	Glosssary	423
	References	428
	Index	530

PREFACE

Chemical ecology is developing by leaps and bounds. Thousands of growing points involve all organisms and offer ample opportunities for collaboration of chemists and biologists. The biological aspect draws on diverse fields, ranging from molecular genetics, anatomy, histology, and genetics to endocrinology, animal behavior, and systems ecology. Given this confluence of many strands of science, it is little wonder that there is not a textbook for chemical ecology courses.

While plant and insect studies dominate chemical ecology, the percentage of papers on vertebrates in the Journal of Chemical Ecology has held steady at 10-19% since its inception in 1975. Most papers on vertebrates deal with mammals, and birds have only recently attracted the attention of chemical ecologists (Müller-Schwarze, 2005). Chemical ecology is both a basic and an applied science. Fundamental questions include reproductive interactions in fish, olfactory imprinting, chemistry and functions of scent marking in mammals, olfactory foraging in seabirds, self-medication in animals, and protein chemistry. Practical applications consider, for example, challenges in fish migration, sea turtle conservation, pest control, and animal husbandry. To succeed in solving practical problems, we first have to establish the basic natural history, behavior, and ecology of a species: To lure brown tree snakes to scented traps, we need first to know what food odors or pheromones these animals attend to. In contrast to insects, much behavior of mammals is under multisensory control, and applications of repellents and attractants based on natural behavior are yet to be realized. Attractants are still in the art stage, much the same way as hunters and trappers have always used them. Sometimes we rediscover in the scientific literature what practitioners knew all along. After we published a paper on the existence and histology of the tail gland of reindeer (Müller-Schwarze et al., 1977), Swedish Saame told us how they avoid contaminating reindeer meat with this smelly gland, which was well known to them.

Our field has progressed from studying simple responses within a species to study of more complex ecological relationships. Research began with identifying the chemicals responsible for chemosensory communication: the classical pheromones. Now more complex ecological relationships are emerging for vertebrates. Investigations into predator-prey systems describe interactions between two species: predators cueing in on pheromones of their prey. Further, white-tailed deer presumably use wolf scent marks to orient and stay within the relative safe border areas of wolf territories. Increasingly more levels and larger pieces of the ecosystem are being studied: foraging seabirds cue in on dimethyl sulfide that is liberated when plankton feed on smaller organisms. Certain vultures spy on other vulture species that have detected carcass odors. Honeyguides and honey badgers form an effective partnership to exploit bee colonies. Such tripartite relationships are already well known for insects and other invertebrates.

The vast and diverse scope of the book precludes completeness. I had been advised to limit the book to pheromone communication, or just to mammals. However, interspecific interactions such as herbivory and plant and animal defenses have always been at the heart of chemical ecology, and the chemical ecology of most other vertebrates has just begun. As an animal behaviorist, I emphasize animals and their interactions with members of their own and other species. (By contrast, the Journal of Chemical Ecology, for example, "is devoted to an ecological understanding of the origin, function and significance of natural chemicals [italics mine] that mediate interactions within and between organisms.") This hybrid of textbook and review does not address methods. Good compendia of methods exist; that of Millar and Haynes (1998) is the best example. To avoid misleading conclusions from still fragmentary and unsettled research, I have refrained from textbook-style selectivity and generalizations. In this sourcebook, I present original data and consciously avoid premature generalization of studies still in flux. Birds' incorporation of fresh aromatic plants into their nests comes to mind as an example.

The chapters on pheromones, allomones, kairomones, orientation, and applications are the core of the book. The other chapters on environment, molecular properties, and chemoreception serve in supporting roles, always with animals in mind. I have deliberately included speculative ideas and open questions, both to encourage further research and to stimulate discussion in courses. For example, p. 365 suggests that mammals' wallowing, pawing, and urine soaking may be more adaptive in animals inhabiting cold countries. It is left to the reader to ponder why. Some ingenious stepping-stones show the path to today's knowledge: pioneer work on fish alarm pheromones, dog tracking, or palatability of birds. A certain planned redundancy keeps each chapter directly accessible; earlier ones need not be consulted.

I accumulated and honed the book's material during 14 years of teaching a course entitled *Chemical ecology of vertebrates* to graduate students and advanced undergraduates. Biology students have a complex relationship to chemistry.

I try to make chemistry interesting and relevant to a biologist. I emphasize ecology with a chemical twist, and not ecological chemistry. Because of its multidisciplinary nature, chemical ecology serves as a wonderful "capstone course" for graduate and advanced undergraduate students. They are challenged to remember disparate facts and principles from a variety of courses, such as the 12 cranial nerves, terpene classification, clay types, toxic plants, parasitism, carnivore ecology, fish migration, conditioned aversions, symbiosis, protein structure, bird behavior, nutrition, human health, livestock reproduction, behavior development, and much more. In examinations, students have listed numerous practical applications they have learned in this course. Responses included drinking tea with milk, eat dirt when poisoned, be wary of pufferfish, know estrogen mimics on livestock pasture, use chemical bird repellents, failure of chemical defenses vis-à-vis introduced species, and to make a car salesman take off his jacket (it could be artificially scented to boost sales). My heartfelt thanks go to the many friends and colleagues who helped by reviewing and providing information, suggestions, discussions, and constructive criticism over many years. I fondly remember countless discussions over decades with Drs. Robert M. Silverstein and the late John B. Simeone, both good friends. My wife Christine has participated in this project the longest, by searching literature and always looking out for new developments in the world of odors. I am especially indebted to Drs. Lee Drickamer, Donald E. Moore III, Tsutomu Nakatsugawa, Lixing Sun, Max M. Mozell, and Robert Mason for reviewing all or parts of the book. My colleagues Dr. José Giner, Dr. Neil Price, Christopher Sack, and Mangesh Goundalkar proofread chemical structures and names. Over the years, exchanges with former graduate students, Drs. Peter Houlihan, Bruce Schulte, Axel Engelhart, and Jan Herr, have enriched this work. I also benefited from the questions and comments by many students taking the course over the years. Even a single-author book depends on a team. I thank Cindi Gamage for essential keyboard help with references and both Cindi and Joyce Buczek for keeping track of the permissions for reproducing figures, and Dr. Jane Ward at Cambridge University Press for her tireless dedication and excellent editing. Jennifer Cheshire helped to track down numerous literature references.

ACKNOWLEDGEMENTS

Publisher and author thank the following publishers for permission to reproduce illustrations:

Alliance Communications Group/Allen Press:

Figs. 3.9 and 3.10. From: Quay, W.B. and D. Müller-Schwarze (1970). Functional histology of integumentary glandular regions in black-tailed deer *Odocoileus hemionus columbianus*). *Journal of Mammalogy* **51**, 675–694.

American Association for the Advancement of Science:

Fig. 4.1. From: Scholz, A. et al. (1976). Science 192, 1247–1249.

Figs. 4.9 and 4.10: From: Grassman, M. et al. (1984). Science 224, 83-84.

Fig. 5.7. From: Mori, K. et al. (1999). Science 286, 711-715.

Fig. 6.3. From: Desjardins, C. et al. (1973). Science 182, 939–941.

American Society for Ichthyology and Herpetology:

Fig. 3.7. From: Ehrenfeld, J.G. and Ehrenfeld, D.W. (1973). *Copeia* **1973**, 305–314.

American Society of Mammalogists:

Fig. 8.1. From: Batzli, G.O. et al. (1977). Journal of Mammalogy 58, 583-591.

Annual Reviews:

Fig. 5.6. From: Buck, L. (1996). *Annual Review of Neuroscience* **19**, 517–544.

Elsevier:

Fig. 3.13. From: Frey, R. & Hofmann, R.R. (1997). Skull, proboscis musculature and preorbital gland in the saiga antelope and Guenther's dikdik (Mammalia, Artiodactyla, Bovidae). *Zoologischer Anzeiger* **235**, 183–199.

Fig. 1.4. From: Wilson, E.O. (1970). Chemical Ecology, p. 138.

Fig. 4.11. From: Grubb, T.C. jr. (1974). Animal Behaviour 22, 192–202.

Fig. 3.14. Reprinted from Zeitschrift für Säugetierkunde, **21**, Ortmann, R. Über die Musterbildung von Duftdrüsen in her Sohlenhaut der weissen Hausmaus (Mus musculus albinus), 138–141, copyright 1956, with permission from Elsevier.

W.H. Freeman Publishing:

Fig. 1.5. From: Moen, A. (1973). Wildlife Ecology, p. 71.

The Herpetologists' League, Inc.:

Fig. 7.4. From: Greene, M.J. and Mason, R. T. (2000). Herpetologica 56, 166–175.

Journal of Herpetology:

Fig. 4.8. From: Chelazzi, G. and Delfino, G. (1986). *Journal of Herpetology* **20**, 451–455.

McGraw Hill Education:

Figs. 3.4 and 3.5. From: Duellmann, W. and Trueb, L. (1986). Biology of Amphibians.

Nature Publishing:

Fig. 2.3: From: Böcskei, Z. et al. (1992). Nature 360, 186-188.

Oxford University Press:

Fig. 5.9b: From: Rasmussen, L.E.L. and Hultgren, B. (1990). *Chemical Signals in Vertebrates*, vol. 5, p. 155.

Fig. 12.4. From: Müller-Schwarze, D. (1990). Chemical Signals in Vertebrates, vol. 5, p. 591.

Sigma Xi, The Scientific Research Society:

Fig. 6.8. From: Peters, R.P. and Mech, L.D. (1975). *American Scientist* **63**, 628–637.

Smithsonian Institution:

Fig. 3.3. From: Weitzman, S.H. and Fink (1985). *Smithsoniam Contributions to Zoology*, **421**, 1–121.

Springer (including Academic Press, Plenum):

Figs. 1.1 and 1.2. From: Regnier, F.E. and Goodwin, M. (1977). *Chemical Signals in Vertebrates*, vol. 1, 115–133.

Fig. 5.9a. From: Stoddart, M. (1983). The Chemical Ecology of Vertebrate Olfaction.

Fig. 4.7. From: Benvenuti, S. et al. (1992). in: Chemical Signals in Vertebrates, vol. 6, 429–434. New York: Plenum.

Fig. 4.13. From: Wiltschko, R. and Wiltschko, W. (1992). *Chemical Signals in Vertebrates*, vol. 6, p. 437.

Fig. 6.14. From: Müller-Schwarze, D. (1992). *Chemical Signals in Vertebrates*, vol. 6, p. 460.

Fig. 7.9. From: Frisch, K. v. (1941). Zeitschrift für vergleichende Physiologie **29**, 46–145.

Fig. 8.4. From: Bronson, F. H. and Coquelin, A. (1980). *Chemical Signals in Vertebrates*, vol. 2, p. 256.

Fig. 12.2. *Journal of Chemical Ecology* **21** (1995) p. 1357, Responses of beaver (Castor canadensis Kuhl) to predator chemicals, Engelhart, A. and D. Muller-Schwarze, fig 4. With kind permission of Springer Science and Business Media

Thomson Publishing Services:

Fig. 5.2. From: Bond, Carl E. (1979, 1996). Biology of Fishes.

John Wiley and Sons:

Fig. 3.8. From: Albone, E.S. (1984). Mammalian Semiochemistry, p. 43.

Fig. 5.5. From: Albone, E.S. (1984). Mammalian Semiochemistry, p. 245.

Fig 10.8. With kind permission of the Zoological Society of London.

The odorsphere: the environment for transmission of chemical signals

The scent of flowers does not go against the wind, not sandal, rosebay or jasmine, but the scent of the good goes against the wind; a good man is wafted to all quarters.

SUTTAPITAKA ("basket of discourse"), from Pali canon of Theravada Buddhists, ca. 500–250 BC

Land animals exploit the odorsphere, the world of vapors around them. In any given locale, they move in an odorscape, a landscape of volatiles. Even in fish we speak of odors because neurophysiologically the olfactory system is involved, even though water-soluble stimulants are not necessarily volatile. We expect vertebrates to have taken advantage evolutionarily of the physicochemical characteristics of their environment first to select and then to optimize chemical communication. The chemical communication system of a cold-water fish differs vastly from that of a tropical bat. Despite similar biological functions, each system has been shaped by, and is adapted to, a distinct set of environmental circumstances.

In air, temperature, relative humidity, barometric pressure, and air currents not only modulate the movement of molecules from the source but also affect odor reception once the molecules have arrived near the receptors. The evaporation of an odor from a surface such as animal skin, a scent mark, or vegetation is regulated by air temperature, relative humidity, the porosity of the surface, and other compounds present (Regnier and Goodwin, 1977; Figs. 1.1 and 1.2).

The evolution of chemical communication was probably influenced by such additional factors as adsorption of aerial pheromones to vegetation, or waterborne pheromones to suspended clay. The influence of these environmental features has very likely selected for both the choice of chemical constituents of the signals and the appropriate signal-emission behaviors (Gleeson, 1978).

Chemical signals have several advantages over cues in other sensory modalities. They work in darkness, around obstacles, and may last for a long time, ranging from seconds to months. This enables an animal to communicate with others in its absence, or even with

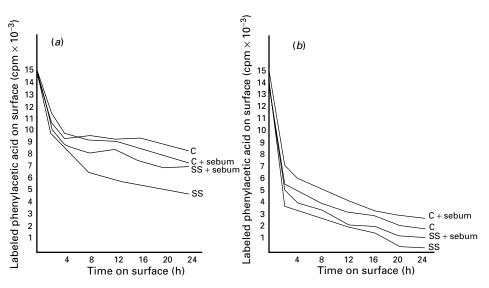


FIGURE 1.1 Effect of relative humidity of air on evaporation of phenylacetic acid from two surfaces in the presence and absence of sebum: stainless steel (SS) and cellulose (C). Relative humidity 0% (a) or 100% (b), both at 20°C. (Adapted from Regnier and Goodwin, 1977.)

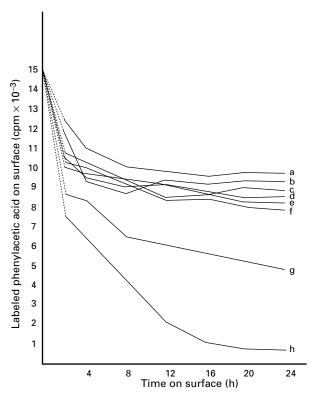


FIGURE 1.2 Effect of surface on evaporation of phenylacetic acid at 0% relative humidity and 20°C. a, montmorillonite; b, mylar plastic; c, glass; d, cellulose; e, kaolinite; f, balsa wood; g, stainless steel; h, platinum. (Adapted from Regnier and Goodwin, 1977.)

itself when returning to a previously scented site. A major disadvantage is a long "fade-out" time that would limit consecutive signals to a slow sequence. In the real world, air or water currents are nearly always present. They transport signals, and the role of diffusion is insignificant because it is too slow.

1.1 Air

For the medium air we can distinguish *close-range* from *long-distance* chemical signals. They differ in the behaviors that animals employ to take advantage of physical processes. Sniffing a scent mark, another animal, or potential food at close range is more than a passive receiving. Exhaling moist air onto the focus of attention moistens the surface and liberates more odorant. Thus, sniffing enables an animal to regulate the release of molecules from the substrate. At zero distance, the odor source can be licked to take up non-volatile compounds. An example of long-distance responses is the approach of seabirds to food odors such as dimethyl sulfide over the ocean. Here air currents and water–air interface transport of molecules are important.

Since virtually all communication by terrestrial vertebrates takes place in the air layer near the ground, the chemical ecologist has to understand the properties of that stratum. Volatiles in higher layers of air, however, may attract vultures to carcasses and could possibly serve as cues for migrating birds. Airborne odors are affected by temperature, relative humidity, barometric pressure, air currents, and vegetation, which, in turn, may influence temperature, humidity, and airflow. Complex interactions of these variables create countless unique environmental conditions for communication patterns of diverse species and for various specific purposes.

1.1.1 Temperature

The vapor pressure of volatile compounds, measured in atmospheres (or millimeters of mercury), varies with temperature. For example, the vapor pressure of acetone increases from 200 to 400 mmHg with a temperature rise from +20 to +40 °C, and that of *n*-heptane from 100 to 400 mmHg with a temperature change from +40 to +80 °C (e.g. Adams *et al.*, 1970). The half-lives of several acetates decreased by two- to fourfold when the temperature was raised from 20 to 30 °C (McDonough *et al.*, 1989). In temperate latitudes, temperatures can vary from about 40 to 0 °C within 24 hours. Therefore, it is important to know the vapor pressure of a given compound for the ambient temperatures under which a particular animal species operates. Diurnal and nocturnal animals may have selected different signal compounds (or mixtures). Do polar and tropical species differ in their choice of compounds for communication? Have cold-climate

pheromones higher vapor pressures? Do polar and montane animals use behavioral means such as sniffing at close range to optimize chemoreception? Are there environmental temperatures so low that pheromone communication is impractical, or even not feasible? The animal's ability to escape from ambient conditions by burrowing or seeking other shelter has also to be considered.

The sex pheromone of the male pig is adapted to the ambient temperature. The pheromone, a mixture of 5α -androstenol and 5α -androstenone, is bound to the protein "pheromaxein" in the saliva. At body temperature ($+37\,^{\circ}$ C) most binding is lost after 72 hours. In direct encounters with females, ample pheromone is given off. At $+4\,^{\circ}$ C, however, the binding of the pheromone to the protein is unchanged over 168 hours. Pigs deposit the frothy saliva in the environment during their breeding season in autumn and winter. At the prevailing low temperatures, the pheromone is released only slowly over a longer time (Booth, 1987).

Temperature may significantly affect *chemoreception*. For instance, electrical responses to amylacetate delivered to olfactory receptors of a tortoise, *Gopherus polyphemus*, were little affected by air temperatures between 20 and 30 °C at the nares but changed considerably above and below that range. Up to +35 °C and down to +10 °C, the olfactory response was a "monotonic slowly decreasing function of temperature" (Tucker, 1963; see also Grundvig *et al.*, 1967).

1.1.2 Humidity

The higher the humidity, the more odor molecules evaporate from a surface, because they compete with water molecules for surface sites. Dogs track better on moist ground and/or on humid days, and we are all familiar with the smell of a wet dog. This phenomenon has far-reaching consequences for the choice of compounds and communication patterns in humid versus arid climates. Some field studies have demonstrated behavior differences between wet and dry seasons. For example, the blackbuck, *Antilope cervicapra*, ceases to use dungpiles during the monsoon season (Prasad, 1989). Environments with high temperatures and humidity, such as tropical forests, call for range marks with active compounds that are either large molecules (Alberts, 1992a) or contain effective keeper substances that slowly release volatiles.

An animal can control release of odor molecules from body surfaces, especially skin gland areas, up to a point. But in moist air, volatiles from secretions on skin or hair will evaporate more easily. If only intermittent odor emission is desirable, humidity would interfere with the animal's odor release. Glands that produce such intermittent signals, such as alarm odors, appear to be more developed in species or subspecies in drier climates. An example is the metatarsal gland in

the North American deer genus *Odocoileus*. Within one species, the size of the gland, or even its presence or absence, varies with the climate. In populations of white-tailed deer, *Odocoileus virginianus*, the gland becomes smaller (or even absent) from eastern North America to more humid Central America and northern South America. In the western mule deer, *Odocoileus hemionus*, the gland is larger in more arid environments. Finally, comparing the two species, the mule deer of the more arid western North America has a larger metatarsal gland than the white-tailed deer of the more humid east (Müller-Schwarze, 1987). Similarly, of two species of the Indian gerbil genus *Tatera*, the one in a drier climate has a ventral gland, while it is smaller or absent from the species of a more humid environment (Prakash and Idris, 1982; Kumari and Prakash, 1983).

It is important to measure humidity exactly where an animal operates. In a meadow on a summer afternoon, the relative humidity can be over 90% at 5 cm above ground, but only 60% at 60 cm (Geiger, 1965). The implications of this would be different for a deer or a rodent. Furthermore, a species like a deer beds down on the moister ground but applies scent marks on branches in a higher, drier layer of air. The rodent, by comparison, experiences a higher and stable level of humidity in its burrow or cave. Humidity is high in stands of plants, and different plants offer different conditions. For instance, forest edges promote precipitation from fog. East-facing forest edges are in the wind and rain shadow and, therefore, experience less humidity.

Mammalian odor *reception* is modulated by relative humidity. For instance, neotropical bats (*Carollia perspicillata* and *Phyllostomus discolor*) are less able to approach an experimental banana odor correctly if the humidity is lowered from the normal 75% to under 60%. The sensory impairment results from drying out of the nasal mucosa in low humidity (Laska *et al.*, 1986).

It is well known that dogs track better in humid air. Rodents find buried seeds better in wet soil. This is important in arid climates. After rains, yellow pine chipmunks, *Tamias amoenus*, and deer mice, *Peromyscus maniculatus* found experimentally buried seeds of Jeffrey pine, *Pinus jeffreyi*, and antelope bitterbrush, *Purshia tridentata*, better than in dry soil. The recovered number of seeds increased 27- and 15-fold, respectively. In wet soil, seeds take up water rapidly and emanate volatile organic compounds that the rodents exploit. By extension, variations in humidity in arid environments may have profound effects on olfaction-dependent behaviors such as finding food, social interactions, preying, and predator avoidance (Vander Wall 1998).

Rodent species differ in their ability to smell buried seeds: those from arid climates perform better than species from mesic climates. Specifically, Panamint kangaroo rats, *Dipodomys panamintinus*, from arid and semiarid areas of the Great Basin Desert in North America were the only species that found deep caches

of seeds under dry conditions. By contrast, chipmunks from more humid eastern North America performed the poorest in finding buried sunflower seeds under dry conditions (Vander Wall *et al.*, 2003). Like the much-studied pesticides, volatiles adsorb to soil particles and desorb from theses particles when the moisture exceeds the thickness of a monomolecular layer, increasing the vapor pressure; this, in turn, facilitates finding buried seeds (Vander Wall, 2003). Furthermore, rodents, use memory as well as odors to find buried seed caches. As moisture favors searching by smell, pilfering occurs more often after rains (Vander Wall, 2000).

1.1.3 Barometric pressure

Hyperbaric pressure may intensify odors or render odoriferous some "odorless" gases such as methane. Professional divers, experimentally exposed to hyperbaric pressures, detected odors of krypton and methane when sniffing these during the decompression phase of a dive. The threshold for krypton was 2 ATA (atmosphere absolute), and 100% positive responses occurred at 6 ATA. For methane, the threshold was 3 ATA (100%: 13 ATA). The thresholds of individuals differed by as much as a factor of three (Laffort and Gortan, 1987).

1.1.4 Air currents

Odors travel in moving air. During their long evolutionary history, animals have adapted to detecting chemical signals from downwind. Many animals integrate chemotaxis with photo-, anemo-, or rheotaxis (Vickers, 2000). Several natural history accounts of large mammals report movements into the wind and presumably toward rain and fresh forage. According to French Camel Corps reports, dromedaries are said to detect water pools and fresh pasture from 40–60 km away. These animals turn into wind blowing from rain clouds and will head into that direction if permitted by the rider (Gauthier-Pilters, 1974). African water buffalo, elephant, and zebra are assumed to "smell rain" and migrate there (Daly, 1988). Arabian oryx, *Oryx leucoryx*, are reported to do the same over distances of about 50 km. One female was documented to have traveled 150 km to an area of fresh rain (Daly, 1988).

Factors affecting currents

Turbulent flow

The air flow in the microclimate of an animal's home range usually is more complicated than basic laminar flow (which does not even exist over large distances and open areas). Fluctuations in air velocity cause turbulence, and stationary objects may create such fluctuations. The air is almost always turbulent, and intact parcels of air laden with odor molecules travel in random fashion (Geiger, 1965). The multidirectional turbulence is superimposed on the horizontal wind flow pattern. Mechanical turbulence is caused by wind, and thermal or convective turbulence by heated air rising. The plume from a smokestack loops up and down on a hot day. Such a "looping plume" is shaped by thermal up- and downdrafts, which account for more vertical displacement than the small-scale mechanical turbulence (Thibodeaux, 1979). Clearings in dense forest experience eddies spinning off from the general flow over the treetops, while a sparse stand of trees has a more laminar air flow. Small eddies are typical for the air directly above the air-soil interface (Thibodeaux, 1979). Eddy diffusion varies in space and time: it is less intense and more variable near the ground, and the greatest mixing of air occurs during midday while at night the air is more stable. An exception is the thin layer of air that adheres to the ground, walls, or vegetation. Processes here follow the laws of molecular physics, and not those of eddy diffusion (Geiger, 1965). In most atmospheric odor movements, however, turbulence swamps molecular diffusion. Among insects, turbulent air flow around obstacles such as trees influences the response of a predatory beetle, *Rhizophagus grandis*, to the odor of their prey, bark beetles (Wyatt et al., 1993).

The more stable the air, the higher the concentrations of odor that can be carried over long distances. This, in turn, increases potential communication distances. However, unstable air conditions disperse the odor molecules more "sideways," with a wider "cone;" consequently more individuals can be reached, although the signal is attenuated over a shorter distance. "Parcels" of odorcontaining air travel straight, but subsequent parcels travel at different angles, as the wind direction changes over time (David *et al.*, 1982). Fluctuations in speed and direction ("meandering") expose an organism that is fixed in place to changes in odor concentration, including zero levels. This amounts to an on–off effect that, in turn, counteracts habituation of the animal's chemical senses.

Mountains

In the mountains, differential heating during the day causes upslope winds, which move up both sides of a valley, and upvalley winds that blow along the valley. During the night, the directions reverse to downslope and downvalley winds (Geiger, 1965; Fig. 1.3).

Large areas

Pressing problems of air pollution have spawned experiments and models of odor dispersion over larger areas. Strom (1976) and Beaman (1988)

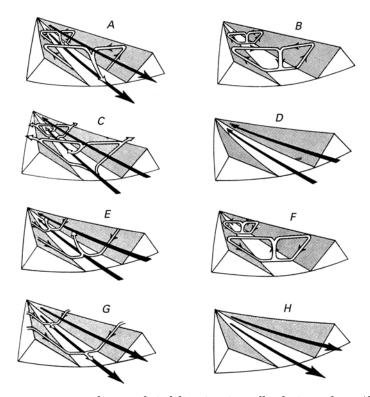


FIGURE 1.3 Changes of wind directions in a valley during 24 hours (from A [night] to H [night again]). Black arrows indicate valley winds and white arrows indicate slope winds. (From Geiger, 1965.)

analyzed transport of stack effluents. Beaman concluded that models of gas dispersion downstream from a source such as a factory predict the actual gas distribution better in rural than in urban areas.

Fluctuations in concentration in eddies

The changes in odor concentration during transport depend on eddy size. Large eddies carry small puffs of odor intact, with little concentration change over time (and distance). Eddies smaller than the puff let the puff grow slowly and concentration decreases, but slowly. Puffs of about the same size as eddies are torn apart, and concentration decreases rapidly. Mixtures of compounds of different molecular sizes will retain their relative concentrations in turbulent flows. Therefore, odor mixtures such as those from plants can contain compounds of high and low molecular weights, even though their separate diffusion rates would be different.

How have animals adapted their communication patterns to prevailing wind patterns? Air flows may be variable, as over slopes, valleys, wood slopes, and forest edges; continuous and strong, as on grasslands, tundra, or the Antarctic;

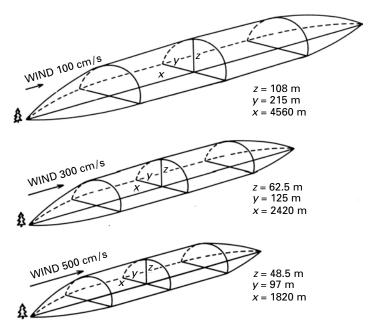


FIGURE 1.4 Active space for different wind speeds. A single female gypsy moth at the tree on the left emits sex attractant. The space where the concentration of pheromone is above threshold (for a searching male) is defined by the dimensions x, y, and z. The active space shrinks with increasing wind velocity. (From Wilson, 1970.)

minimal, as in woods; or absent, as in caves and burrows. In general, animals are expected to rely on wind only for longer-range attraction to food or conspecifics, avoidance, or warning of conspecifics or predators. Finer details, such as the exact outline of a territory, or the identity of an individual, can be assessed by sniffing scent marks or the animals themselves at close range. In some environments, air flow patterns can be very complex, and we have not even asked how vertebrates exploit these for effective communication. For instance, how do nocturnal downvalley winds and the water current along streams affect odor signals used by beaver? Active at night, beaver probably use these for signal propagation between neighbor colonies. They cannot easily exploit the daytime upvalley wind, because they stay in their lodge during the day.

Active space

The biologically significant *active space* where the odor concentration is above threshold is shaped like an overturned boat (Fig. 1.4). (If the molecules were able to spread in all directions, as from an elevated odor source, the active space would assume the shape of a cylinder with pointed ends.)

To calculate the active space, the investigator needs to measure odor emission rate, detection threshold of the animal, and wind speed.

Plume models

Pheromone propagation by wind depends on the release rate of the pheromone (or any other odor) and air movements (turbulent dispersion). In wind, the turbulent diffusivity overwhelms the diffusion properties of a volatile compound or mixture itself. Diffusion properties are now properties of wind structure and boundary surfaces, and preferably termed *dispersion coefficients*. Two models have dominated the discussion of insect pheromone propagation. These are the *time-average model* (Sutton, 1953) and the *Gaussian plume model*.

The time-average model considers the average concentration of airborne materials at sites downwind from a point source. The concentration (or density D) of a pheromone at any one point with the coordinates x (downwind direction), y (horizontal crosswind [transverse] dimension), and z (vertical dimension) can be estimated with the following formula.

$$D_{xyz} = \frac{2Q}{C_y C_z U x^{2-n}} \exp \left[-x^{n-2} \left(\frac{y^2}{C_y^2} + \frac{z^2}{C_z^2} \right) \right]$$

where Q is the release rate; U the mean wind speed; C_y and C_z are horizontal and vertical dispersion coefficients, respectively; and n a parameter ranging between 0 and 1. Wilson and Bossert (1963) have applied this model to pheromones. Dispersion coefficients are functions of atmospheric turbulence, terrain roughness, and vertical wind speed profile. According to Sutton (1953), with light winds of speeds between 100 and 500 cm/s, neutral atmospheric conditions, and level ground: $C_y = 0.4 \, \mathrm{cm}^{1/8}$, $C_z = 0.2 \, \mathrm{cm}^{1/8}$, and n = 0.25.

The release rate *Q* is doubled to 2*Q* because odor clouds released on or near the ground are "reflected" by this boundary layer. (For elevated odor sources, only *Q* is used because the molecules can disperse in all directions.)

Most pheromone biologists have used the Sutton formula. For elevated odor sources, a more complicated version of the equation exists. However, compared with the concentration differences at different distances from the source, those between elevated and ground-level sources are minuscule (Elkinton *et al.*, 1984).

Now we introduce the sensitivity (perception threshold) of the receiving animal. According to Bossert and Wilson (1963), the downwind maximum distance x_{max} (in cm) at which the concentration of an odor remains above threshold is

$$x_{\text{max}} = \left(\frac{2Q}{K\pi C_{y}C_{z}U}\right)^{\frac{1}{2}-n}$$

where Q is rate of odorant emission in molecules per second; K is the threshold (molecules/cm³); C_y and C_z are dispersion constants (0.4 cm^{1/8} and 0.2 cm^{1/8}, respectively; see above); U is the mean wind speed; and n is 0.25. For example, the emission rate Q for carcass odors that attract vultures is assumed to be up to 20/day or 7×10^{18} to 1.4×10^{20} molecules/s.

Again, 2Q is used instead of Q because the odor is "reflected" from the ground, and twice as many molecules fill the space above ground than would if the odor spread equally in all directions.

The *Gaussian plume model* estimates the average pheromone flux by multiplying the measured odor concentration by mean wind speed, using the following formula (Elkinton *et al.*, 1984). Everything is the same as in the Sutton model, except that σ_y and σ_z , respectively, replace the terms C_y and C_z of the Sutton model. Dispersion coefficients are determined for each experiment separately.

$$D_{xyz} = \frac{Q}{2\pi \sigma_y \sigma_z \, \overline{u}} \exp \left[-\left(\frac{y^2}{2\sigma_y^2} + \frac{z^2}{2\sigma_z^2} \right) \right]$$

Here σ_y and σ_z , the horizontal and vertical diffusion coefficients, are the standard deviations of the cloud dimensions in the horizontal and vertical directions, respectively. They are functions of the downwind distance x and, in this, differ from the constants C_y and C_z of the Sutton model:

$$\sigma_y = \frac{1}{2} C_y x^{(2-n)/2}$$
 and $\sigma_z = 2 \frac{1}{2} C_z x^{(2-n)/2}$

The rate of dispersion and the values of the dispersion coefficients depend also on terrain and atmospheric conditions. There are "prairie grass" coefficients and values for forests (Fares *et al.*, 1980). Both are discussed by Elkinton *et al.* (1984). (In one more complicated equation, the Gaussian model also considers absorption on the ground surface by introducing a factor α .)

Both the Sutton and the Gaussian models underestimate the width of the active space at each distance, as Elkinton $et\ al.$ (1984) showed. Currently, the Gaussian plume model is preferred over the Sutton model because the dispersion coefficients are measured anew in each experiment. In the Gaussian model, however, the dispersion coefficients are functions of the downwind distances x and correspond to the standard deviation of the vertical and horizontal distribution of the concentration along any axis perpendicular to the mean wind direction. Elkinton $et\ al.$ (1984) pointed out that the Gaussian plume model can be easily applied to a great range of atmospheric stabilities. The dispersion of the plume is affected by temperature changes and resulting turbulence. In the Gaussian model, the size of the active space shrinks as wind speed increases. One has to distinguish concentration (g/cm^3) from flux (g/s per cm^2). The flux increases in

direct proportion to wind speed. Flux is more pertinent for stimulation of receptors in animals. Both models consider average concentration over several (3 to 15) minutes. But insects and other animals experience meandering turbulent plumes; consequently most of the time the concentration is below threshold, interrupted by brief exposures to above-threshold concentrations. Therefore, the active space is actually larger than predicted by the models.

For triggering behavior, the concentration at one point in time is more important than the average concentration. Therefore, in the real world, considerable deviation from time-averaging models is observed. In addition to time-averaging models, *peak* concentrations of odors in turbulent systems have to be considered. Aylor (1976) estimated peak concentrations for air currents in forests. Average concentrations, as calculated by the Sutton formula, may be as low as only a few percent of maximum (peak) concentrations. It is often the latter, however, that would trigger an animal's response.

Webster and Weissburg (2001) visualized instantaneous versus time-averaged odor plumes by laser-induced fluorescence. The spatially varying plume at any particular point in time matters more to an animal than an average plume shape. The mean direction and speed of air flow may be relatively constant, but the animal may extract information from concentration differences on very small temporal and spatial scales.

1.1.5 Topography and vegetation

Topography and vegetation create air currents and modify existing wind. Vegetation affects air flow patterns and may adsorb and re-emit odor molecules (Perry and Wall, 1984). The flow over vegetation is "practically exclusively turbulent" (Geiger, 1965). Within vegetation there is little eddy diffusion. The scale of vegetation will affect communication patterns in terrestrial vertebrates: obviously, a vole in a grassy runway, deer in a mature forest, and soaring vultures separated from a carcass by a forest canopy all face different odor propagation problems.

In forests, mechanical turbulence is caused by trees, and temperature inversions by the forest canopy. Ventilation inside a forest is complex and not readily described by existing air flow models (Aylor, 1976).

Forest edges

In the ecologically important *forest edges*, during daytime a "forest breeze" blows from under the tree trunks into the open. However, because the trees act as obstacles, there is no reverse "field breeze" at night, as between land

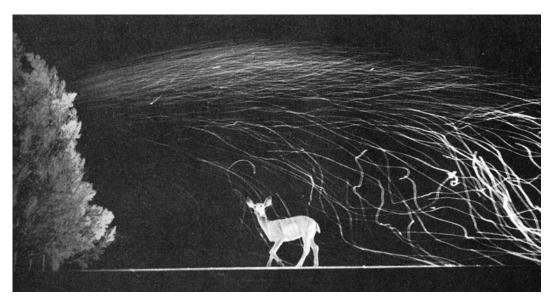


FIGURE 1.5 "Silent zone" in lee of a hedgerow. Wind-tunnel visualization of air movement over a windbreak, such as a hedgerow, with aid of a bubble generator. Wildlife seek such zones for protection from wind, but information flow through air can be reduced there. (From Moen, 1973.)

and sea. Only in mountainous wooded areas is there a night forest wind that blows into the open country. This downdraft from the radiating crown layer is a night-cooling phenomenon, and not a "forest wind" (Geiger, 1965). Uneven heating inside the forest drives night breezes of about 10 cm/s, but the heterogeneous canopy is more important than this differential heating.

A "quiet zone," about two to three tree heights long, forms in the lee of a forest (Fig. 1.5). It is much shorter behind a forest edge than behind a thin shelter belt. In the open, air currents reach again their maximum speed at a distance of 20 to 70 tree heights from the forest edge (American Meteorological Association, 2000). This will affect the behavior of animals such as deer bedding down or predators chemo-searching for prey; for thermoregulation, an animal should bed down in the "quiet zone," but to receive airborne chemical information on conspecifics, predators, or food it should stay in the wind beyond the "quiet zone."

Area odors

Model experiments have shown the rates of evaporation of volatiles from vegetated areas. The flux of ammonia and related amino compounds from

an Australian sheep pasture into the atmosphere was measured by collecting air samples in traps at different heights above ground level. The flux of nitrogen compounds was highest at midday and lowest during the night. Wind and humidity affected flux and gas composition. Pure ammonia was characteristic for cool, humid weather. The pasture lost on average 0.27 kg nitrogen/ha (Denmead *et al.*, 1974). Such "area odors" may serve as important cues for food-seeking herbivores, or migrating mammals or birds.

1.1.6 Chemical environment

In mixtures such as secretions and scent marks, the polarity of the non-pheromonal compounds may greatly affect the release of the active odor molecules (Regnier and Goodwin, 1977). The pH also affects volatility; with higher acidity, a volatile odoriferous base, such as an amine, will form larger amounts of non-volatile salts and, therefore, contribute less to perceived odor. Conversely, odorous acids such as volatile fatty acids will increasingly form salts as the alkalinity increases (Albone, 1984).

1.1.7 Environmental perturbances

More and more artificial changes of the modern environment interfere with communication behavior of animals and humans. Gases, radiation, and gravity all affect responses to odors.

Gases

As an example, low levels of carbon monoxide (CO), together with ingested alcohol, reduce sensitivity to guaiacol, which has a "smokey" or "burnt" odor (Engen, 1986). Smokers take up CO. If they also drink, they could be impaired in their ability to detect smoke from a fire.

Radiation

Patients who were irradiated for 5 weeks with photons of 8 MV of energy at 3 Gy/min, for nasopharyngeal carcinoma or pituitary adenoma, became less sensitive to odors. Their thresholds for amylacetate and eugenol were determined before and several times after radiotherapy. One week after irradiation, the thresholds had increased from 10 to 2 dilution steps. Sensitivity recovered over the subsequent weeks, reaching 6–8 dilution steps 6 months after treatment. The dose received by the olfactory area was estimated as 2 Gy/day (Ophir *et al.*, 1988).

Gravity

The gravitational field affects olfaction as it does vision, audition, or vestibular function. It could impair detection of dangerous fumes or burning electrical equipment in airplanes or space vehicles. Astronauts also report altered perception of food flavors under weightless conditions. Men and women tested with four "scratch-and-sniff" odor samples of the University of Pennsylvania Smell Identification Test identified odors more poorly when in an upside-down position (Mester *et al.*, 1988).

In another experiment, animals were accelerated. Puppies, aged 1–20 days, were rotated in a drum for 5 or 15 minutes and accelerated for a maximum of $3 \times g$. Their response to eucalyptus odor changed from neutral to negative, and that to maternal odor from positive to "uncertain." Odor aversion to eucalyptus lasted longer than that to maternal odor. When visual and auditory stimuli start to control behavior at the age of 12–13 days, the induced odor aversion is more easily suppressed (Kassil and Gulina, 1987).

1.2 Water

Aquatic animals use their chemical senses in all aspects of their lives, from reproductive behavior to feeding, habitat selection, and predator avoidance. The hydrodynamic properties determine the possibilities and limits of chemical communication in water. As a medium, water is as dynamic as air, so that convection and advection are far more important for odor transport than is diffusion. Distribution by currents is even more important in water because compounds of similar molecular weight diffuse four orders of magnitude more slowly than in air (Gleeson, 1978). Diffusion of odorants may be important only in the submillimeter range, while turbulence is typical for water masses above the centimeter range.

1.2.1 Boundary layers

In flowing water, the "boundary layer" is the water mass above the substrate. This boundary layer is defined as that part of the flow where the velocity ranges from 0 to 99% of the velocity of the stream. Most boundary layers of biological interest are turbulent (Webster and Weissburg, 2001).

The behavior of the animal in response to flow is important, not just the flows themselves. An animal searching for the source of an odor moves in the direction of increasing stimulus intensity and stays within the boundaries of the plume. If an animal needs to sample odor in turbulence frequently, it may have to reduce speed, to untenable levels in the case of moths or birds, or to an energetically

	Viscous environment	Turbulent environment
Reynolds number	Low	High
Signals	Change slowly	Chaotic structure
Flow	Low	Bulk flow, perceptible to animal
Gradient	Continuous	Intermittent
Animal	Searches in three dimensions for odor boundaries	Relies on flow
Sensory integration	Slower integration time	Rapid integration of brief odor pulses

Table 1.1 Properties of viscous versus turbulent environments

demanding level in the case of fish. This, in turn, means that it may not arrive in time or expose itself more to predation (Webster and Weissburg, 2001) (Table 1.1).

Depending on the organism, we have to consider water masses and movements ranging from the submillimeter "micropatches" all the way to the oceanographer's gyre on the 1000 km scale. Large bodies of water from freshwater lakes to oceans are stratified. Fish or other animals may sample several of these stable layers and select one that "smells right." Indeed, salmon (*Salmo salar*) with their olfactory sense experimentally obliterated did not sample certain water layers and were unable to select one as fish with intact sense of smell could do (Døving *et al.*, 1985). It has been suggested that the shear between two horizontal layers could be utilized to determine the lower or upper boundary of a particular layer (Westberg, 1984).

1.2.2 Sampling behavior and information currents

Most aquatic animals, notably vertebrates, have to sample *turbulent* water with patchy odor distribution. Two types of stimulus access ("sniffing") have been distinguished in fish: "cyclosmates" such as tuna or lobster, which sample a specific sniff volume in sniffs or flicks, and "isosmates," which sample a steady, ciliary-driven water flow. This latter type of stimulus access is found in slow-moving animals such as catfish, eels, dogfish, or mud snails (Atema, 1988). In turbulent water, patch boundaries will be the sharper the more recently the odor was released. When crossing an odor patch, the animal will learn about the distance to the odor source from the rate of concentration change.

In water, currents may be utilized or even created to communicate chemically. Beaver, river otter, mink, or various fish may simply release their odorants into a stream. Lobster, tail-waving newts, and possibly fanning fish produce *information currents* that propel chemical signals toward recipients. In the lobster, the female sends a current with pheromone into the male's shelter. The male, in turn, draws water toward himself and fans it out into the surroundings, signaling his mating status (Atema, 1986).

Water may be better than air for studying the dynamics of stimulus dispersal, because in this denser medium fluid dynamic processes are attenuated (Atema, 1988). As interindividual distances increase with increasing body size, pheromone communication becomes less and less practical in water, particularly because of signal delays, dilution, and cross-currents.

1.2.3 Water as solvent

The chemical properties of water determine what compounds will leach out of soil and vegetation, and hence what chemicals other plants and animals will be exposed to. For instance, acidic water will extract alkaloids from plant materials.

1.3 Water-air interface

Odor retention, release and propagation on water surfaces are important to many animals, but we know little of how animals exploit or actively manipulate these processes.

Freshwater mammals such as beaver may leave odors on the surface of their ponds and olfactorily sample the water or layer of air immediately above it. Lipids on water may form *micelles*, small blobs of molecules (from Latin *mica*, a grain, crumb, morsel) that enhance evaporation into the air layer by increased chemical potential. Some seabirds hunt by odor (e.g. Hutchison and Wenzel, 1980; Nevitt, 1999). They may respond to prey volatiles (from krill, squid, or fish) that rise to the water surface and evaporate into the air. The air—water equilibrium for dilute solutions can be expressed by using partition coefficients, relative volatility, or Henry's law (Thibodeaux, 1979).

1.4 Influence of setting

The circumstances of an odor experiment are important. For instance, male carp, *Cyprinus carpio*, spermiate in the presence of females that ovulate after having been injected with carp pituitary homogenate (as is already known for goldfish). This effect occurred in earthen ponds (200 m²) but not in bare tanks

 Table 1.2 Different behaviors in different settings

Species	Behavior	Field	Laboratory	Reference
Kin recognition				
Bufo americanus	Associate with kin	Yes	No	Waldman, 1981
Rana cascadae	Associate with kin	Yes	No	O'Hara and Blaustein, 1985
Reproductive behav	vior			
Carp Cyprinus carpio	♂spermiates in presence of ♀ injected with pituitary homogenate	In earthen ponds: yes	In bare tanks: no	Billard <i>et al.</i> , 1989
Domestic cattle	♂ mounts ♀ treated with estrus urine	Pasture: only treated ♀ mounted	Stalls: mount indiscrimi- nately	Sambraus and Waring, 1975
Feeding behavior				
Brown tree snake	Responds strongly	No	Yes	D. Chiszar,
Boiga irregularis	to fish attractants and trapper's lures			personal communication
Townsend's vole	Feeding in presence of repellent	Avoids repellent only in open, not in cover	Test without cover may be misleading	Merkens et al., 1991
Mongoose	Aversive conditioning to scented, toxic eggs	Avoid scented, toxic eggs from a distance	Will not avoid at distance, break shell, eat still	Nicolaus and Nellis, 1987
Responses to preda	tor chemicals			
	Avoiding area	Farmland, golf	Enclosure: avoid	Ward <i>et al.</i> , 1997
Hedgehog Erinaceus europaeus	tainted with badger (predator) feces	courses: avoid treated area for minutes or hours only	treated area for 2 days	
Chinook salmon	Conditioned to recognize predator (cutthroat trout)	Better survival in creek but only if raised in complex habitat	No survival benefit in hatchery raceway; and none if raised in simple habitat	Berejikian <i>et al</i> ., 1999
Gray-tailed voles Microtus canicaudus	Mink odor effect on reproductive rate, sexual maturation, juvenile recruitment	No effect	Reproductive behavior suppressed in M. agrestis and bank voles, Clethrionomys glareolus	Wolff and Davis-Born, 1997

(2m³) (Billard *et al.*, 1989). In this experiment, the effect may not be clearly pheromonal, as the fish could see each other.

The American toad, *Bufo americanus*, discriminates kin from non-kin (after having been raised in mixed groups of siblings and non-siblings) *only in the field*, but not in the laboratory. The same is true for the cascades frog, *Rana cascadae*. These results have been summarized by Blaustein and Waldman (1992). Conversely, the brown tree snake, *Boiga irregularis*, responds strongly to fish attractants and trappers' lures in the laboratory but hardly at all in the field (D. Chiszar, personal communication, 1994).

Finally, in cattle, bulls respond to female sexual odors on pastures but not in stalls (Sambraus and Waring, 1975). Table 1.2 summarizes these findings.

Properties of vertebrate semiochemicals

... the odors of ointments are more durable than those of flowers.

FRANCIS BACON (1561–1626): Essays, Of Praise.

On the banks of the Plata I perceived the air tainted with the odour of the male *Cervus campestris*, at half a mile to leeward of a herd; and a silk handkerchief, in which I carried home a skin, though often used and washed, retained, when first unfolded, traces of the odour for one year and seven months.

CHARLES DARWIN: The Descent of Man, p. 529.

The structures of vertebrate chemosignals reflect their functions and the environment they are used in. Temporal parameters, spatial range, localizability, intensity, detectability, and information content of the signal depend on both the chemical structure and the operating environment (Alberts, 1992a). The chemical properties involved include functional groups of the molecule; volatility; aromaticity; the number of compounds composing the entire signal; and when, where, and how it is emitted (Alberts, 1992b).

2.1 Functional groups

Functional groups can determine communicative activity of a molecule. For example, hypoxanthine 3*N*-oxide triggers alarm responses in Ostariophysan fish. The amineoxide (NO) functional group (Fig. 2.1) appears to be essential for the antipredator behavior of fathead minnows, *Pimephales promelas*, and finescale dace, *Phoxinus neogaeus* (Cyprinidae) to occur. Structurally similar molecules lacking the NO group did not release alarm responses in two tetra species (Charicidae). All four species belong to the Ostariophysi, and the response to the NO group is probably widespread in this order, also known as Cypriniformes (Brown *et al.*, 2000).

In another example for functional groups, Gower and Ruparelia (1993) noted that the odoriferous steroids that play a role in communication share certain

FIGURE 2.1 Hypoxanthine oxide, a fish alarm pheromone.

features. They are relatively volatile, lack any substituent at C-17, are unsaturated at C-16 and C-17, have only one oxygen function at C-3 and hence are very non-polar. The double bond at C-16–17 is not important for the odor to humans, while manipulations at C-3 change the odor quality (Gower *et al.*, 1989).

A third example is the role amino acids such as L-aspartate and L-glutamate have in deterring feeding by the oriental weatherfish, the loach *Misgurnus anguillicaudatus*, presumably signaling a predator. Monoalkyl and dialkyl esters of these amino acids are less repellent. This indicates that the carboxyl group is important for the repellent activity. In the same vein, acetic acid is repellent, while alkyl acetates are less so. Again, the carboxyl group appears to mediate the effect (Harada, 1989).

Further examples of the importance of functional groups for behavior are the responses of sunfish to steroids in beetles, their prey; reactions of birds and mammals to capsaicin-related compounds; and fear behavior of rats when exposed to sulfur compounds from fox urine and feces.

Of several related steroids in the defensive secretion of dytiscid beetles, deoxy-corticosterone was most effective. It deterred sunfish from feeding in 94% of the tests. Other steroids (pregnolones) that differed only by lacking a keto group at one carbon atom were either intermediate or not active at all (Gerhart *et al.*, 1991).

Capsaicin analogues that differ in only one functional group affect birds and mammals differently. A change from an acidic phenolic hydroxyl group in vanillyl acetamide to a methoxy group in veratryl acetamide reverses the effect on starlings and rats. The first was aversive to starlings, but attractive to rats, while the opposite was true for veratryl acetamide (Mason *et al.*, 1991).

Finally, fox urine and feces contain sulfur compounds that "stress" rats. In several mercaptoketones, the thiol (mercapto) group was essential for the effect, while the keto group was not (Vernet-Maury *et al.*, 1984).

In addition to their presence, the position of functional groups in a molecule can determine its smell. This is known as "regional selectivity."

Compound	Odor perceived by humans
Vanillin	Odor of vanilla
Iso-vanillin	Odorless
(S)-4-Carvone	Caraway
(R)-4-Carvone	Spearmint
(+)-Androsta-4,16-dien-3-one	Urinous, sweaty, musky, woody (only 10–20% can smell it)
(–)-Androsta-4,16-dien-3-one	Odorless

Table 2.1 Odor differences of isomers and enantiomers

2.2 Polarity

Most compounds in vertebrates and insects contain polar functional groups (Wheeler, 1977). An intriguing question is whether marking pheromones are less polar than water-soluble ones, such as those in urine. The polarity of non-pheromonal compounds in a mixture greatly affects pheromone release into the environment (Regnier and Goodwin, 1977).

2.3 Solubility

Chemical signals in urine, fresh-, and saltwater are water soluble. In water, many more compounds are pheromones candidates than in air, because molecules of a wide range of sizes are water soluble. Terrestrial scent marks have to survive humidity and precipitation and so here the active components are soluble in lipids.

2.4 Isomers and enantiomers

Humans and insects can discriminate enantiomers of the same compound (Friedman and Miller, 1971; Russell and Hills, 1971) (Table 2.1). Blacktailed deer (*Odocoileus hemionus columbianus*) discriminate between the two geometric isomers and somewhat between the enantiomers of the "deer lactone" (Fig. 2.2) (Brownlee *et al.*, 1969; Müller-Schwarze *et al.*, 1976, 1978a). Young male Asian elephants have more (+) than (-) enantiomer of frontalin in their temporal secretion. Their secretion and (+)-frontalin affect conspecifics little. However, the nearly recemic enantiomer mix from mature bulls attracts ovulating females and repels males (Greenwood *et al.*, 2005)

2.5 Volatiles

Secretions and excretions of terrestrial vertebrates contain compounds with a wide range of volatility. For convenience, the two ends of the continuum

$$CH_{2} CH_{2}$$

$$CH_{3}(CH_{2})_{4} - C CH_{2} - CH_{2}$$

FIGURE 2.2 The "deer lactone: (Z)-4-hydroxydodec-6-enoic acid lactone.

are arbitrarily termed volatiles and non-volatiles. Both can be integral parts of chemical signals between animals. The molecular weight of volatile compounds ranges from about 16 to 300 Dalton (Wilson and Bossert, 1963). Volatility can be increased by elevated temperature.

Long-distance communication via airborne odors requires volatiles, while non-volatiles can effectively be used in short-range communication that requires body contact, as in mutual licking by two animals or the nuzzling and licking of a female by a male in mating behavior. Indeed, mating pheromones are often of low volatility. Many communication systems employ both volatiles and non-volatiles, either simultaneously or in sequences, as when long-range attraction by volatiles is followed by close-range inspection of a conspecific – or of a plant in the case of herbivory – with sniffing, nibbling, biting, or chewing. Acceptance or recognition of young or group members may be the result of exchange of non-volatile signals at close range.

A mammal may emit many volatile compounds. Humans, for instance, give off hundreds of volatiles, many of them chemically identified (Ellin et al., 1974). The volatiles include many classes of compound such as acids (gerbil), ketones, lactones, sulfides (golden hamster), phenolics (beaver, elephant), acetates (mouse), terpenes (elephant), butyrate esters (tamarins), among others. The human samples mentioned before contained hydrocarbons, unsaturated hydrocarbons, alcohols, acids, ketones, aldehydes, esters, nitriles, aromatics, heterocyclics, sulfur compounds, ethers, and halogenated hydrocarbons. Sulfur compounds are found in carnivores, such as foxes, coyotes, or mustelids. The major volatile compound in urine of female coyotes, *Canis latrans*, is methyl 3-methylbut-3-enyl sulfide, which accounts for at least 50% of all urinary volatiles (Schultz et al., 1988).

In the secretions of the anal sacs of dogs (*Canis familiaris*) and coyotes (*C. latrans*) the most abundant volatile compounds are trimethylamine, short-chain (C_2-C_6 : acetic, propionic, isobutyric) acids, acetone, and 2-piperidone (Preti *et al.*, 1976).

Free fatty acids up to C_5 are found in urine and other mammalian secretions (Albone, 1984); C_6 and C_8 free fatty acids occur in the anal gland secretion of the aardwolf (Apps *et al.*, 1989), and C_7 , C_8 , C_9 , C_{10} , and C_{12} , "and their isoforms," in the marking fluid of the tiger (Poddar-Sarkar *et al.*, 1991).

The well-known defense secretion from the skunk's (*Mephitis mephitis*) anal glands contains thiols and disulfides (Andersen and Bernstein, 1975) and thioacetates (Wood, 1990). The seven major compounds are (*E*)-2-butene-1-thiol, 3-methyl-1-butanethiol, *S*-(*E*)-2-butenyl thioacetate, *S*-3-methylbutanyl thioacetate, 2-methylquinoline, 2-quinolinemethanethiol, and *S*-2-quinolinemethyl thioacetate (Wood, 1990). Table 13.3 (p. 403) and Fig. 10.9 (p. 403) list some of the sulfur compounds found in Mustelids. Thiols are responsible for the unpleasant smell of the skunk's spray. In the presence of water, they are formed from thioacetates, which also smell rather repulsive. However, the human nose is not necessarily a good judge of what is significant to animals. For instance, a krill extract fraction that humans perceived as fishy did not attract (fish-catching) petrels particularly strongly (Clark and Shah, 1992). At the other extreme, chin gland marks of rabbits appear odorless to humans (Mykytowycz, 1965).

Volatiles from fecal pellets of wild male house mice include ketones, alcohols, and carboxylic acids. Pellets aged over 24 hours had 20 major volatiles whereas fresh pellets had only 15 (Goodrich *et al.*, 1990).

The number of carbon atoms of saturated straight-chain compounds correlates with hydrophobicity, molecular volume, and molecular length. All three could affect the interaction of an odorant ligand with the olfactory receptors (Johnson and Leon, 2001).

The techniques for trapping, concentrating, isolating, and identifying volatiles have been developed by flavor chemists and insect pheromone researchers and are not detailed here. Some of the techniques useful for mammals are summarized in Albone (1984) and Millar and Haynes (1988).

2.6 Non-volatiles

2.6.1 Proteins

The molecular size of non-volatile compounds may be very high. The "mounting pheromone" or "aphrodisin" of the golden hamster, *Mesocricetus auratus*, is a protein with a molecular mass of 17000 daltons (Da) (Singer *et al.*, 1989). It belongs to the alpha- $\alpha_{2\mu}$ -globulin superfamily and is very similar to the major urinary proteins (MUPs) in mice (Fig. 2.3), the bovine β -lactoglobulin from cow's milk, and the pyrazine-binding protein in the nasal mucosa.

Mouse urinary proteins

The urine of male mice contains high levels of MUPs, which play an important role in scent communication. Male mice excrete higher levels (up to 20 mg/ml urine) in their urine than females, and secretion increases with sexual

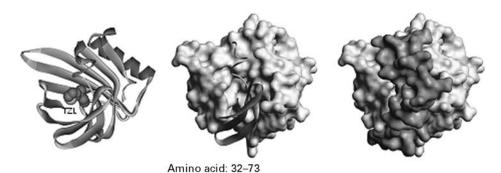


FIGURE 2.3 Major urinary protein (MUP) of mouse, showing region of polymorphic variation. Left: Ribbon model with a volatile ligand (TZL: thiazole) in binding pocket. Center: Ribbon in lower center shows location of variable amino acid sequence. Right: Space-filling model with variable polypeptide chain segment shaded darker, in middle region. (Courtesy Robert J. Beynon.)

maturation. After castration, levels fall to 2.5 to 3.5%, and androgen treatment restores higher levels. This protein type belongs to the lipocalin superfamily and is synthesized in the liver. The liver of males contains three to five times more mRNA for MUPs than that of females. The barrel-shaped (eight-stranded β -barrel), 18 kDa MUP has a hydrophobic cavity that serves as a "container" for volatile ligands (Beynon *et al.*, 1999). This pocket in the molecule binds small hydrophobic molecules and a wide range of odorants can be bound. Mouse urine contains 2-*sec*-butyl-4,5-dihydrothiazole and 2,3-dehydro-*exo*-brevicomin and these volatiles are assumed to signal the presence of a male, while the MUP protein itself seems to trigger puberty in females, acting as a priming pheromone. Volatile ligands attract the attention of male mice, while proteins and non-volatile protein–ligand complexes stimulate counter-marking (Humphries *et al.*, 1999).

The scent mark of the saddle-back tamarin, *Saguinus fuscicollis*, contains water-soluble proteins. The major protein (66kDa) of the scent mark occurs in the urine, while another major protein (18kDa) derives from gland secretions. If the proteins are removed by enzymatic degradation, tamarins still discriminate scent marks from different donor types. However, they are able to discriminate scents with intact proteins from those with the proteins digested. Consequently, the proteins are most likely an integral part of the scent image (Belcher *et al.*, 1990).

Proteins also occur in many excretions and glandular secretions, from turtles to mice and humans. Intensive studies are clarifying their role in signaling. In axillary secretion from human males, the main odoriferous acid, (E)-3-methyl-2-hexanoic acid, (E)-3M2H for short, is bound to two apocrine secretion-binding proteins (ASOB1 and 2) of molecular masses 45 and 26kDa, respectively. The

ASOB2 is apolipoprotein D (apoD) of the lipocalin or $\alpha_{2\mu}$ -microglobulin superfamily of carrier proteins (Preti *et al.*, 1992; Zeng *et al.*, 1996a,b).

The female elephant pheromone (Z)-7-dodecenyl acetate occurs bound to urinary proteins. When it is taken up by a male elephant, the acetate is bound to proteins of the trunk mucus. This might facilitate transport of the pheromone to the vicinity of the sensory epithelium of the vomeronasal organ (Rasmussen and Schulte, 1998).

Frontalin in the temporal gland secretion of Asian elephants is bound to elephant albumin (Schulte *et al.*, 2005). The bulk of the secretion of the chin gland of the rabbit is protein (Goodrich and Mykytowycz, 1972).

2.7 Multicomponent pheromones

2.7.1 Mixtures of volatiles

It was initially thought that, in insects, the major component in a pheromone blend attracted from the longest distance, while the minor components came into play at shorter distances from an odor source such as a "calling" female. However, the male Oriental fruit moth, *Grapholita molesta*, at least, responds from longer distances (100 m) more to the full female pheromone blend of three compounds than to the major component (Linn *et al.*, 1987). Similar tests have not been performed with vertebrates.

For mammals, if not vertebrates in general, multicomponent pheromones appear to be the rule. Such mixtures can comprise compounds of a wide range of volatility. They have been variously termed "odor profile," "pattern," "odor image" (Albone, 1984), "gestalt" (Evans *et al.*, 1978), or "mosaic" (Johnston, 2005). One of the best-investigated odor profiles is that of the scent mark of the saddle-back tamarin, *S. fuscicollis*, (Smith *et al.*, 1985). Here, not even the 16 butyrate esters are sufficient for subspecies recognition. Additional volatiles are also required.

A second example of a mammalian multicomponent pheromone is the puberty-delaying pheromone from female house mice, *Mus musculus*. Two acetate esters and a pyrazine are biologically active in various combinations, but the pyrazine is also active alone (Novotny *et al.*, 1985a).

The chinning response in tree shrews, *Tupaia belangeri*, is triggered by several lipophilic fractions of male urine. The combined fractions are more active than single fractions. Pyrazine compounds and a high concentration of several volatile monocarboxylic acids characterize male urine. Some pyrazine compounds and some monocarboxylic acids release the chinning response (Stralendorff, 1987).

The European badger, Meles meles, marks its territory with its subcaudal gland. This gland produces a mixture of many carboxylic acids and other unidentified compounds. In one recent analysis (Buesching et al., 2002a) 21 compounds, mostly fatty acids, were common to all 66 investigated samples, from both sexes. Analyses have indicated that the patterns of these compounds reflected specific characterists such as sex, individuals, group membership, and seasonality. Some or all of these compounds may signal species. The sexes differed in several respects. Females had more measurable peaks (average 36.1) in their chromatograms than males (30.7). (Peaks can represent one or several compounds.) Tetradecanoic acid was more abundant in males than in females. Three not fully identified compounds were more abundant in females than in males in all seasons, except winter. Individuals showed distinct patterns: each badger had a unique combination of 23 to 58 compunds. Group members' profiles were more similar to each other than to those of members of other groups. Consistent seasonal variation occurred in females. Their secretion was less complex in spring than in winter and summer. Certain patterns correlated with reproductive state: dodecanoic acid and two other compounds were more abundant in non-lactating females than in lactating ones. Males with descended testes (i.e. those reproductively active) had more dodecanoic acid than those with non-descended testes. With increasing age, the level of heptadecanoic acid increased (Buesching et al., 2002a). The levels of two monounsaturated C₁₆ acids decreased in the course of 2 days, potentially providing information about the age of a scent mark. Individual characteristics remained remarkably consistent from year to year. During the annual cycle, periods of stability of composition alternated with rapid changes (Buesching et al., 2002b). What exact combinations of compounds the badgers themselves use for the various levels of discrimination remains to be seen.

It is important to identify and measure the concentrations of a number of compounds in a mixture simultaneously for several reasons. First, among related compounds there may exist precursors of active ones, and pathways of pheromone synthesis may be elucidated. This is true for steroids in the human axilla. Nixon *et al.* (1988) determined the concentration of five steroids extracted from axillary hair of adult men aged 18 to 40 years. The relationships in concentrations between the two ketones 5α -androst-16-en-3-one and 4,16-androstadien-3-one suggest that axillary bacteria reduce the former to the latter with the aid of the enzyme 4-ene- 5α -reductase. Humans have a low olfactory threshold for several 16-androstenes, and the fact that some men have large quantities of 16-androstenes (Nixon *et al.*, 1988) is biologically suggestive.

For several possible functional reasons, pheromones are thought to be mixtures of compounds. In vertebrates, signal specificity is required more at the levels of individual, family, or clan, than the species (in many insects). This specificity arises from "fingerprints" of several compounds mixed in different ratios, and not unique compounds, as in insects. Why are odors so often complex mixtures of compounds? There are several possibilities

- to ensure that an odor is noticed in the first place in case some components vary in concentration over time or between individuals;
- to provide redundancy to counteract environmental obstacles;
- to provide enough variability for distinct specific signals to permit discrimination ("chemical fingerprints," especially in scent marks);
- to increase sensitivity, as a mixture can require fewer molecules of all of its compounds combined to reach an animal's threshold than the sum of the molecules needed to reach threshold for the same compounds tested singly (Laska and Hudson, 1991);
- to render an odor distinct against background odors;
- to be more resistant to sensory adaptation (consequently, mixtures would be better markers for signaling dangers to humans, as in gas leaks) (Commetto-Muñiz and Cain, 1993).

Other complex odors may be extremely variable among individuals and genotypes, and receiving animals have to learn their significance. For example, laboratory mice discriminate the urine odor of different genotypes that vary at the t-locus but no discrete chemosignal compounds have been found in these different genotypes. It is felt that "the variation of the overall pattern of general secondary metabolites" is used for discrimination, instead of specific compounds (Jemiolo et al., 1991). These authors stated that the "lack of discrete chemosignal compounds responsible for the genotype discrimination supports the concept of a non-specific influence of the t-locus (part of the major histocompatibility complex [MHC]) on olfactory cues." This overall pattern is possibly learned, as offspring are exposed to the odor of their parents at an early age. For urine odors determined by the MHC, it was also concluded that they result from multiple and redundant compounds (Tsuchiya et al., 1992). Volatiles alone convey information on MHC genotype, since mice still distinguish corresponding fractions of two odor types even after proteins have been depleted (Singer et al., 1993).

In addition to the simultaneous impact of an odor mixture, differential evaporation may alter the signal over time, indicating the age of a scent. As an example, the major component of the chest gland secretion of the thick-tailed galago, *Galago crassicaudatus*, is benzylcyanide. It evaporates rapidly and is present for only about 1 hour. The other two identified compounds last for several days. In a behavioral corollary, galagos retreat from a scent mark that is less than 1 hour old, while older marks have no such effect (Katsir and Crewe, 1980).

Two different types of scent mark from the same species can evaporate differentially. The brown hyena, *Hyaena brunnea*, pastes two different secretions on the same blade of grass: a brown mark with an odor that dissipates rapidly, and a white mark that – to the human nose – lasts for up to a month (Mills *et al.*, 1980). Similarly, the fade-out time for the anal gland secretion of the African *Helogale undulata rufula*, dwarf mongoose, used for range marking, is about 10 days, while cheek gland secretion, part of the threat display, lasts only 1 to 2 days (Rasa, 1973).

Interspecific chemical cues are also often mixtures. Mixtures of amino acids serve as feeding stimulants in fish. Among mammals, ferrets respond more to mixtures than to pure odors in their foraging responses. The mixtures are thought to contain more information (Apfelbach, 1973).

In some cases, animals do not seem to identify components of complex environmental chemical cues. The pool frog ($Rana\ lessonae$) provides an example. After being exposed to a mixture of morpholine and β -phenylethanol during development before metamorphosis, the froglets prefer the mixture but are not attracted to either of the two compounds (Ogurtsov and Bastakov, 2001).

2.7.2 Combinations of volatiles and non-volatiles

Volatiles and non-volatiles may be related in at least four ways.

- 1. Both kinds of compound may combine to form complex signals, sometimes releasing two-step responses. The volatiles attract from a longer distance, while the less-volatile compounds provide more information at close range. For example, volatile lipids from femoral glands are thought to alert iguanas (*Iguana iguana*) to the presence of a conspecific, but a non-volatile protein fraction, picked up from secretion deposits by tongue touches, helps to identify individuals (Alberts and Werner, 1993). Both volatiles and non-volatiles in bovine body fluids from glands at the mucocutaneous junction of the anogenital area stimulate sexual behavior in bulls (Rivard and Klemm, 1990). Cottontop tamarins, *Saguinus o. oedipus*, typically touch a scent mark to identify conspecifics but are able to rely only on volatiles when forced to do so by a wire screen over the mark (Belcher *et al.*, 1988).
- 2. The non-volatiles may not possess a signal value themselves but may act as release modulators, or "keeper substances." The MUPs in mice provide an example. Each individual expresses at least 4 to 15 different proteins and the combination differs between individuals in the population. Most of the variation between MUPs occurs on the surface of the molecule and is accessible to chemoreceptors. MUPs are stable over the life of the individual. On urine

FIGURE 2.4 Structure of isopentenyl methyl sulfide.

marks, they are also stable for weeks or months. Volatile metabolites that characterize MHC-different individuals appear to be bound to MHC peptides, to MUPs, or both. Sebum, ubiquitous in mammals, can also serve as a "keeper substance" for volatile signal compounds.

- 3. A volatile compound from a synthesis pathway such as that leading to the less-volatile steroid sex hormones may signal reproductive status. For example, Jorgenson *et al.* (1978) have suggested that in the red fox, *Vulpes vulpes*, the strong-smelling Δ^3 -isopentenylmethyl sulfide (Fig. 2.4) could signal an increase of steroid production, at the start of the mating season. This compound can derive from isopentenyl pyrophosphate, an intermediate in terpene biogenesis, leading to sterols and steroid hormones.
- 4. The non-volatiles may be precursors of signal compounds, breaking up into volatiles as needed. This has been suggested, for instance, for the esters in the subauricular gland secretion of the pronghorn, *Antilocapra americana* (Müller-Schwarze *et al.*, 1974), and (*E*)-*s*-2-butenyl thioacetate in the anal gland secretion of the hog-nosed skunk, *Conepatus mesoleucus*, which possibly breaks down to the smelly (*E*)-2-butene-1-thiol (Wood *et al.*, 1993). Finally, in brush-tailed possums, oleic acid breaks down into two molecules of nonanal.

2.7.3 Keeper substances

Mammalian secretions typically have large amounts of non-volatile compounds, while the actual chemosignals may represent a rather small part of the total weight. Squalene is one of the more common lipids in glandular secretions such as sebum. Squalene and sebum retard evaporation of volatiles. In laboratory experiments, the volatility of phenylacetic acid, a pheromone compound from the ventral gland of the Mongolian gerbil, *Meriones unguiculatus*, was affected by "keeper substances": being a polar compound, phenylacetic acid is the less volatile the more polar the keeper substance is. Volatility is lowered if a small amount of pheromone interacts with a large amount of solvent (dipole–dipole interaction). For example, polar diethylene glycol tightly retains phenylacetic acid, while non-polar mineral oil retains it only weakly. However, volatility is retarded more by mineral oil than by squalene. Humidity increases volatility as odor molecules compete with water molecules for surface sites

(Regnier and Goodwin, 1977). Less-polar odorants will be less affected by the polarity of their lipid substrate or by relative humidity. For these reasons, the chemical ecologist has to keep in mind that volatility results not only from the nature of the odorant but also from the characteristics of the substrate, and the environmental conditions.

Keeper substances in a scent secretion, and/or the nature of the substrate such as rock, wood or soil, impart longevity to a scent mark. In our own studies, scent marks of captive pronghorn, *A. americana*, were still noticeable to the human nose 4 months after all animals had been removed from a pen. Similarly, scent marks of the aardwolf, *Proteles cristatus*, last for up to 6 months (Apps *et al.*, 1989). 2-Phenoxyethanol in the secretions from the chin gland of the rabbit is typical in dominant animals, serving as a fixative to extend the life of the signal (Hayes *et al.*, 2001).

The urinary lipids in the urine marks of lions and tigers may serve in prolonging pheromone release from the mark into the air (Asa, 1993). Both sexes of tigers, *Panthera t. tigris*, spray a "marking fluid" onto vegetation. The fluid is released through the urinary tract. This lipid-rich fluid contains lipid fixatives at a level of 1–2 mg/ml. These lipids include cholesterol ester, wax esters, triacylglycerols, free fatty acids, diacylglycerols, monoacylglycerols, free sterol, and phospholipid (Poddar-Sarkar, 1996). Wax esters occur in anal gland secretion of beaver, *Castor canadensis* (Grønneberg and Lee, 1984).

2.8 Sex differences

Some male-specific compounds have been identified in mammalian secretions that may have a role in communication (Table 2.2)

The anal gland secretion of beaver, *C. canadensis*, differs between the sexes (Grønneberg, 1978–79). The male grey duiker, *Sylvicapra grimmia*, has more 2-heptanone and 2-nonadecanone in its preorbital secretion than the female. Two thiazoles (2-isobutyl-1,3-thiazole and 2-isobutyl-4,5-dihydro-1,3-thiazole) and an epoxy ketone (3,4-epoxy-2-dodecanone) are also more abundant in the male's secretion. Correlated with these chemical differences is the fact that only males scent mark with the preorbital gland (Burger *et al.*, 1990).

In European moles, *Talpa europaea*, a series of carboxylic acids dominates the anal gland secretion of both adult males and anestrous females. These acids disappear in proestrous and estrous females but are present again in pregnant and lactating females. The acids are also absent in juveniles. The anal scent appears to constitute a "keep out" signal, and its absence in estrous females permits mating (Khazenehdari *et al.*, 1996).

Species Compound		Source	Reference	
Boar Sus scrofa	16-Androstenes	Saliva	Melrose et al., 1971	
Goat Capra hircus	4-Ethyloctanoic acid	Cornual organ	Sugiyama, 1983	
Asian elephant Elephas maximus	Frontalin	Temporal gland	Rasmussen and Greenwood, 2003	
Humans Homo sapiens	(E)-3-Methyl-2-hexenoic acid	Axilla	Zeng et al., 1991	
Spotted skunk Spilogale putorius	(E)-2-Butene-1-thiol	Anal glands	Wood <i>et al.</i> , 1991	
Striped skunk Mephitis mephitis	Thiols, thioacetates	Anal glands	Wood, 1990	
Egyptian mongoose Herpestes ichneumon	2,4,6,10-Tetramethyl 1-undecanoic acid	Anal glands	Hefetz et al., 1984	
Wolf Canis lupus Isopentyl sulfide and 3,5-dimethyl-2-octanone		Urine	Raymer et al., 1984	

Table 2.2 Some male-specific compounds in mammalian secretions with a role in communication

The mixtures of volatiles in the urine of tree shrews, *T. belangeri*, are sex specific but no single sex-specific compounds have been found (Stralendorff, 1987).

Axillary odors of male and female humans contain the same compounds (C_6-C_{11} acids and E-3-methyl-2-hexenoic acid), but in different ratios. The characteristic odor resides in acidic compounds (Zeng *et al.*, 1996b).

Other chemical studies did not find sex or seasonal differences in the composition of mammalian scents. No sex differences in the composition of mixtures of volatile compounds from glands have been found in the brushtailed possum, *Trichosurus vulpecula*, for example. The same profiles of low-molecular-weight branched carboxylic acids were found in paracloacal gland secretions of males and females (Woolhouse *et al.*, 1994). Branched carboxylic acids also occurred in the preorbital gland secretion of a female sika deer (*Cervus nippon*) (Wood, 2004). Comparisons between the compositions of secretions in different, related species permit assumptions about functional adaptations and possible evolutionary pathways. Such comparisons are available for five *Mustela* species (Brinck *et al.*, 1983), and three species of hyenas (Buglass *et al.*, 1990).

2.9 Life expectancy of chemical signals

Volatiles evaporate, and signals become weaker over time. In mixtures, evaporation of the more-volatile compounds can change the quality of the odor and the range over which it can be detected. This means that decaying and

changing signals potentially provide information about the age of the scent mark and, in turn, the recent behavioral history of the odor donor.

Ferkin and Johnston (1995a) aged anogenital area scent and odor from the posterolateral region in meadow voles (*Microtus pennsylvanicus*) for 15 minutes to 30 days. Males "preferred" female to male anogenital odor if it was 10 days old or less. Females preferred male to female anogenital odor if its age was 25 days or less. Both sexes preferred posterolateral odor of males to that of females if it was up to 1 day of age. In a second experiment, both sexes preferred fresh odors from either source over the same scents that were older. In conclusion, information about sex may get lost with the age of the scent mark.

Pheromones in urine will suffer degradation, hydrolysis, oxidation, and ultraviolet radiation effects. For example, the (*Z*)-7–12-acetyl derivative in elephant urine will gradually hydrolyze (Rasmussen, 1988). In this case, the lipoprotein carriers of the elephant acetate may also determine the life time of the signal besides serving to filter and select odorants, confer specificity, and play a critical role in the transport and transfer of an active ligand to the vomeronasal organ (Rasmussen and Schulte, 1998).

Keeper substances and commonly used marking substrates such as clay and wood can extend the "fade-out time" of the signal. Licking or exhaling on scent marks or the body surface of another individual liberates "on-demand odors". Alberts (1992a) characterized such signals as having multiple rise times.

2.10 Spatial range of odor signals

Animals boost the range of a signal in various ingenious ways. Multiple scent marks, distributed over territory or home range, reach more receivers and assure reception by one individual as it moves about and misses certain places. Species using multiple marks are commonplace. Examples are hyenas, wolves, antelopes, and beavers. Placing a scent mark as high as possible maximizes its range (active space).

2.11 Interaction of olfactory and visual signals

Rocks, stumps, trees, scraped soil, scarred plant stems, or otherwise disturbed vegetation attract the attention of animals who then examine the spot more closely for olfactory information left by conspecifics. The "urine balls" made by sandrats of the Algerian Sahara provide an elaborate example (Daly and Daly, 1975). Sandrats dig up sand, urinate on it and work it into a ball that other sandrats examine. Urine balls from estrous females strongly attract males.



FIGURE 2.5 Visual signal is combined with odor release when an alarmed pronghorn (*Antilocapra americana*) flares his "rump patch" of long hair covering the ischiadic gland. (Photograph: D. Müller-Schwarze.)

Crepuscular and nocturnal animals can use visual signals to attract attention to their scent glands. Hair tufts or brightly colored hair that stands out against the background of the animal's fur highlight many odor-producing skin glands, especially when hair is erected during a display. Black-tailed and white-tailed deer spread their tarsal hair tuft, located on the tarsal gland. Pronghorn spread

white hair on the ischiadic glands ("rump patch;" Fig. 2.5) when alarmed. White, orange, and black hairs draw attention to the dorsal gland of the rock hyrax. Guinea pigs and cuis swivel their rump toward a conspecific and flash the pink inner skin of their perineal pouches as a threat (Rood, 1972).

Visual anomalies in the environment, such as those created by thrashing, pawing, or tearing of bark, render olfactory marks more detectable (Roberts and Gosling, 2001). Finally, the height of marks on tree trunks, saplings, or rocks may signal the size of the marking animal.

Odor production and release

The odour emitted must be of considerable importance to the male, inasmuch as large and complex glands, furnished with muscles for everting the sack, and for closing or opening the orifice, have in some cases been developed. The development of these organs is intelligible through sexual selection, if the most odoriferous males are the most successful in winning the females, and in leaving offspring to inherit their gradually-perfected glands and odours.

CHARLES DARWIN: The Descent of Man, p. 530

Among mammals, which mostly "think through their noses," it is not surprising that *olfactory* marking of their territory plays a large role. The most diverse methods have been chosen, various scent glands evolved, and the most remarkable displays of depositing urine and feces developed; of these, the leg-lifting of the domestic dog is most familiar to everyone.

KONRAD LORENZ: On Aggression (1963), p. 53 of original German version, translated by author.

Chemosignals in vertebrates come from a great variety of sources. These sources include excretions, secretions, material recycled from other organisms, and even from the environment.

3.1 Signals in excretions

Metabolites in urine or feces provide the energetically least expensive, and evolutionarily probably the original, chemical signals in vertebrates. "Much of history of evolution has concerned the development by living things of responses to metabolites, sometimes their own and sometimes produced by others. Those organisms which developed 'satisfactory' responses succeeded, and those which did not, failed." (Lucas, 1944). Interested parties, such as members of the opposite sex, can then "spy" and read pertinent information about sexual and dominance status, health and body condition, quality of diet, and more. For instance, female goldfish release sex pheromones in their urine that

$$\begin{array}{c|c} H_3C \\ \\ H_3C \\ \hline \\ S \\ CH_3 \\ \hline \\ Trimethylthiazoline \\ \end{array} \begin{array}{c} N \\ N \\ H \\ \hline \\ Indole \\ \end{array}$$

FIGURE 3.1 Structures of trimethylthiazoline, found in red fox feces, and indole, found in dog feces (also in coal tar, orange blossoms).

are precursors of androgens and estrogens. In terrestrial animals, potentially many volatile, but also less-volatile, components of urine or feces may serve as chemical cues. For instance, experiments suggested that in the shrew, *Crocidura russula*, males locate females by markers in urine or fecal pellets (Cantoni and Rivier, 1992).

3.1.1 Urine

The volatiles in urine of mice, rats, coyotes, white-tailed deer, and many other species carry chemical cues. Male–male aggression in mice is triggered by a urinary mixture of 2-sec-butyl-4,5-dihydrothiazole and 3,4-dehydro-exobrevicomin (Novotny et al., 1985b; Harvey et al., 1989). Dominant males emit (E,E)- α -farnesene and (E)- β -farnesone (Harvey et al., 1989). Adrenal-dependent urinary volatiles, found in female mice, delay puberty in other females. The time of the first vaginal estrus in delayed by appoximately 2 days by n-pentylacetate, cis-2-penten-1-yl acetate, and 2,5-dimethylpyrazine, Odocoileus hemionus columbianus, (Novotny et al., 1985a). The "deer lactone" occurs in urine of blacktailed deer and is applied to the hair tuft of the tarsal gland on the deer's hock. There it forms part of a recognition odor (Müller-Schwarze, 1971). Beavers use castoreum in their castor sacs for scent marking. This secretion is, in large part, concentrated and perhaps slightly modified urine.

3.1.2 Feces

Red fox, *Vulpes vulpes*, feces contain trimethylthiazoline (2,5-dihydro-2,4,5-trimethyl thiazole; Fig. 3.1) which alarms a prey species, the Norway rat (Vernet-Maury, 1980).

Indole (Fig. 3.1), in dog feces a smelly matabolite has no effect on sheep (Arnould *et al.*, 1998), although its odor is strong and repulsive to humans. However, mixtures of fatty acids and neutral compounds, as well as sulfur compounds, from dog feces inhibit feeding in sheep (Arnould *et al.*, 1998).

Often animals add glandular secretions to their excretions to produce a complex specific signal. Urine marks of lions and tigers are examples. A whitish material in urine marks, first thought to represent anal gland secretion, tested positive for urinary lipids. When dyed with green food dye, anal gland secretion failed to show up in urine marks. These lipids may both prolong pheromone release from the urine mark and indicate the general nutritional condition of the animal, as urinary lipids correlate with kidney fat (Asa, 1993).

Some pheromones in urine may originate in the reproductive tract, rather than being metabolites. In white-tailed deer, *Odocoileus virginianus*, estrus urine elicits some chases and associating by bucks, but vaginal secretion is more active. It is assumed that the sexual signal originates in glands of the reproductive tract and is transported by urine to the outside (Murphy *et al.*, 1994).

3.2 Glandular secretions

Vertebrates, especially mammals, have evolved a bewildering variety of specialized glands that produce secretions which in turn carry chemical signals, independently of excretions.

3.2.1 Fish

In fish, proven or suggested sources of pheromones are, in addition to urine and feces (including bile), the reproductive organs, skin, and secondary sex structures. Several glands are sources of reproductive pheromones in males: the blenny, *Blennius pavo*, has specialized anal fin appendages that enlarge at maturity and produce a factor that attracts females (Lauman *et al.*, 1974). A second type of gland, the caudal gland in males of the glandulocaudine fishes, is suspected to be its source of sex pheromones. It is located at the base of the tail fin and covered by a large, highly modified scale, or several scales (Fig. 3.2). The glands' secretory cells shrink when males are kept isolated, and enlarge with courtship (Nelson, 1964). Courtship behavior includes tail fanning. It is thought that such movements provide a passive pumping mechanism that releases pheromones from the gland and propels them toward the female (Weitzman and Fink, 1985). In fish, female pheromones typically originate in the ovaries.

3.2.2 Amphibia

Amphibians have mucus glands over their whole body to provide the mucus needed to keep their skin moist, and fields of granular glands that secrete alkaloid toxins. In addition to these general glands, salamanders possess glands

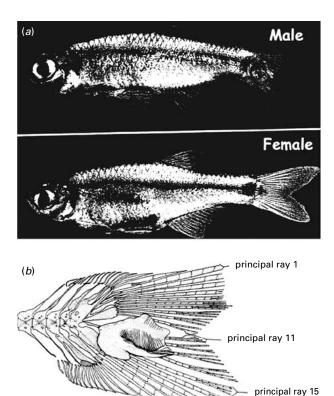


FIGURE 3.2 Glands in fish. (a) Caudal gland at the tail base of *Scopeocharax atopodus* (Characidae). Note the difference between male and female. (b) The pouch scale is a large, modified scale that covers the caudal gland in a glandulocaudine fish (*Iotabrycon praecox*). (From Weitzman and Fink, 1985.)

principal ray 18

of three different types in the cloacal area: cloacal, pelvic, and abdominal glands. These are generally more developed in males. Pelvic and cloacal glands function in spermatophore formation. A communication function of these glands is starting to emerge. Male red-bellied newts, *Cynops pyrrhogaster*, attract females with their abdominal gland secretion. The active ingredient was shown to be a decapeptide, named sodefrin (Kikuyama *et al.*, 1995).

Some salamanders, such as *Plethodon*, have "hedonic" glands on their chin; glands occur also around the eyelids, temporal region, dorsal tail base or entire dorsal region in various species (Fig. 3.3). Finally, the plethodontid salamanders have nasolabial glands on their lips. The glands hypertrophy seasonally in males, and *Plethodon cinereus* show "nose-tapping" and "chin-touching" of the substrate, suggesting a communication function (Tristram, 1997). A "unique cloacal vent gland" has been described for male *Rhyacotriton olympicus* (Dicamptodontidae).

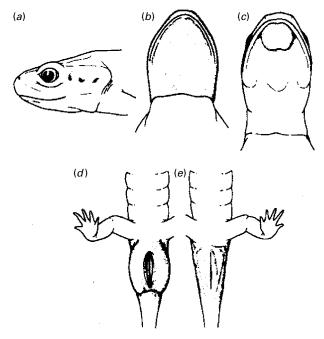


FIGURE 3.3 Glands in salamanders. (a) Genial glands on the side of the head of *Notophthalmus viridescens*; (b) diffuse submandibular glands of *Taricha torosa*; (c) mental gland of *Pseudoeurycea smithii*; (d, e) *Ambystoma jeffersonianum*: in the male (d) the glandular area around the vent and cloacal papillae is swollen, which is not the case for the cloacal area in the female (e). (From Duellman and Trueb, 1986.)

This species may release pheromones during "tail curling" (Sever, 1988). Little is known about glands for communication in anurans, other than the general mucus and granular glands (Fig. 3.4). In the Australian tree frog *Litoria splendida*, the pheromone *splendipherin* is produced in rostral and parotoidal glands on the head of the male (Wabnitz *et al.*, 1999).

3.2.3 Reptiles

In lizards, femoral glands (Fig. 3.5) are arranged in rows on the ventral surface of the hindleg. They are most active in the breeding season and are larger in males than in females. Femoral secretion stimulates tongue flicking and may serve in species recognition and home-range marking. Chemically, more lipids can be extracted from femoral gland secretions during, than outside, the breeding season. Saturated C_{14} – C_{26} fatty acids, unsaturated C_{16} – C_{24} fatty acids, and eight sterols were found, including cholesterol (Alberts *et al.*, 1992).

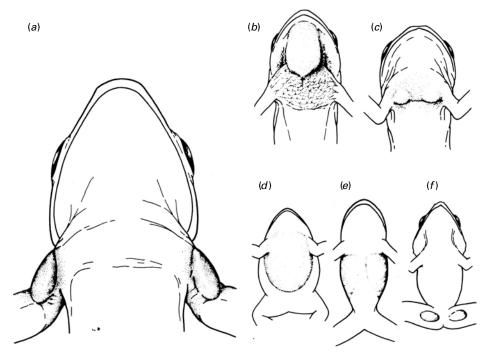


FIGURE 3.4 Glands in male frogs. (a) Mental gland of Kassina senegalensis; (b) pectoral glands of Leptopelis karissimbensis; (c) humeral glands on the foreleg of Hylarana albolabris; (d) abdominal gland of Kaloula verrucosa; (e) ventrolateral glands of Ptychohyla schmidtorum; (f) femoral glands of Mantidactylus pseudoasper. (From Duellman and Trueb, 1986.)

In desert iguanas, *Dipsosaurus dorsalis*, the femoral glands are more developed in males than in females. The percentage of females with active glands varied with the population density of three populations studied. In the one with the highest density, 12.3% of the females had active glands. At the intermediate site, 31.7% of the females had active glands, with 59.2% at the low-density site (Alberts, 1992b).

Snakes have paired "scent glands" that open into the cloaca. Some snakes such as *Natrix* and *Macropisthodon* spp., have "nucho-dorsal" glands under the skin of the dorsal neck area. Their secretions are assumed to serve in defense and/or intraspecific communication (Madison, 1977). Turtles have "mental glands" on their chins, inguinal, and axillary ("Rathke's glands;" Fig. 3.6), lateral, and cloacal glands. Their role in producing chemical signals is little explored. Crocodiles have three main types of "musk glands": mandibular (or throat) musk glands, cloacal, and dorsal glands. The mandibular glands are most active during the breeding season and are discharged during courtship behavior.

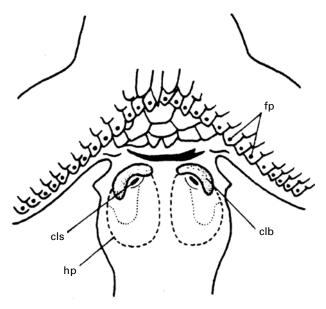


FIGURE 3.5 Glands in lizards are illustrated by the femoral pores and cloacal sacs in a gecko, *Gymnodactylus pulchellus*. clb cloacal bone; cls, cloacal sac opening; fp, femoral pore; hp, hemipenis. (From Bellairs, 1970.)

Skin glands are thought to have evolved from scraped-off corneous material of the skin. Such "uncontrolled semiochemical release" may have given way to "facultative, controlled semiochemical release" of desquamated, loosely joined plugs of keratinous material when the germinal portion of the integumentary gland moved "below" the body surface. Invagination then created a "gland" (Maderson, 1986).

3.2.4 Birds

Birds are not exactly known for scent communication or scent-producing organs. The preen (uropygial) gland is usually considered the only, most developed, and ubiquitous skin gland in birds. It serves to waterproof the plumage and is larger in aquatic than in land birds. However, there are also sex differences in the composition of waxes in the uropygial gland of domestic ducks, possibly for communication (Jacob *et al.*, 1979). In the hoopoe, *Upupa epops*, the uropygial gland is largest at 12 days of age. Surrounded with a ring of specialized feathers, it is said to have a repellent effect on intruders. A predator is squirted with liquid from the large intestine, accompanied by a hissing sound (Sutter, 1946). The roles of cloacal and anal glands in birds are poorly understood (Quay, 1977).

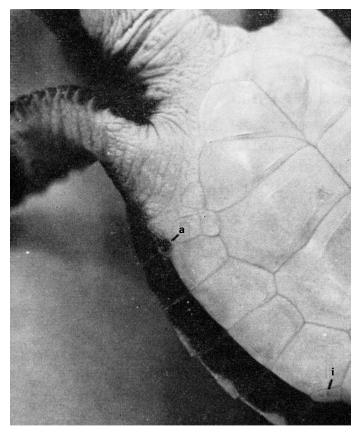


FIGURE 3.6 Rathke's glands in turtles and tortoises. Axillary (a) and inguinal (i) pores (gland openings) are shown in *Chelonia mydas*. (From Ehrenfeld and Ehrenfeld, 1973.)

3.2.5 Mammals

Skin glands in mammals are widespread and diverse. We know so much about their behavioral roles and secretion chemistry that even a brief survey would fill several volumes. Skin glands can occur on all body parts but are found more often on the head, the extremities, and the anogenital region. Animals grow scent glands where vehicles carry lights: front and rear. But there are also neck, dorsal, ventral, and lateral glands. The two basic types of gland component are sebaceous and apocrine (modified sweat) glands (Table 3.1 and Fig. 3.7). Sebaceous gland produce a lipid secretion, while apocrine glands secrete a milk-like aqueous fluid. Both types of gland are associated with hair follicles and secrete into the pilosebaceous canal. By contrast, true sweat (eccrine) glands are not associated with hairs and secrete directly to the skin surface. Sebaceous and apocrine

Table 3.1 Examples of specialized mammalian skin glands

Dominating gland type	Skin gland organ	Species
Holocrine (sebaceous)	Ventral gland	Mongolian gerbil, Meriones unguiculatus
	Tarsal gland	Black-tailed deer, Odocoileus hemionus columbianus
	Subauricular gland (jaw patch)	Pronghorn, Antilocapra americana
Apocrine (modified sweat	Metatarsal gland	Black-tailed deer
glands)	Axillary gland	Humans

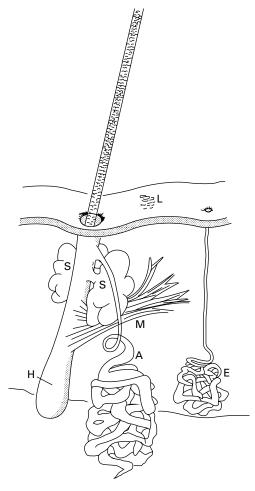


FIGURE 3.7 Elements of mammalian skin glands. S, sebaceous gland; A, apocrine gland; E, eccrine gland; M, errector pili muscle; H, hair follicle. (From Albone, 1984.)



FIGURE 3.8 Subauricular gland (SG: the "jaw patch", a black area below the ear) in male pronghorn, *Antilocapra americana*. (Photograph: D. Müller-Schwarze.)

glands can occur in concentrations of a single type, or combined, depending on the function of a particular gland and its secretion: for scent marking on the substrate, a lipid secretion from the sebaceous glands is called for. An example is the subauricular gland of the pronghorn, *Antilocapra americana* (Fig. 3.8). Another example of a skin gland dominated by sebaceous elements is the tarsal organ (Fig. 3.9) in deer of the genus *Odocoileus* (white-tailed, black-tailed, and mule deer). The odors that are supposed to volatilize fast, as alarm pheromones, typically arise in aqueous secretions from modified sweat glands. Examples are the axillary gland in humans and the metatarsal gland on the hindlegs of deer of the genus *Odocoileus* (Fig. 3.10). In the female brown hyena (*Hyaena brunnea*) two separate glands, one sebaceous, one apocrine, produce a double scent mark (Mills *et al.*, 1980).

In most mammal species the skin glands typically occur only in, or are larger or more numerous in, males. For example, only the male pronghorn has a pair of

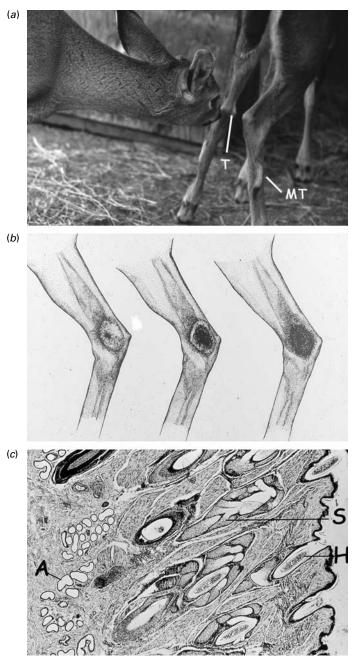


FIGURE 3.9 The tarsal gland of black-tailed deer, dominated by sebaceous glands. (a) Location of the gland on the hock (T); a young male sniffs the gland. MT, metatarsal gland. (b) Increasing opening of the secretion- and urine-covered tuft of modified hair is seen left to right. (c) A section of skin (surface on the right) showing the tarsal gland with its apocrine (A) and sebaceous (S) gland elements and a hair (H). (From Quay and Müller-Schwarze, 1970.)

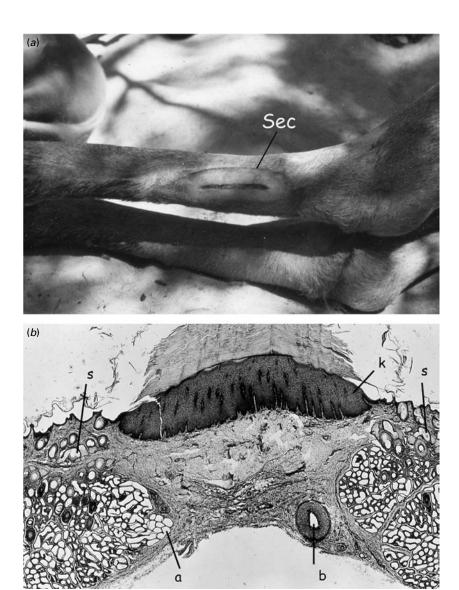


FIGURE 3.10 The metatarsal gland of black-tailed deer, dominated by apocrine glands. (*a*) The location on the outside of the hindleg; there is a dark keratinized ridge in the center with glandular tissue either side. The droplets are secretions (Sec), stimulated by epinephrine injections. (*b*) Histology of a 4-year-old male shows the metatarsal gland. a, apocrine gland portion; b, blood vessel; k, keratinized ridge; s, sebaceous gland. (Photograph (*a*): D. Müller-Schwarze; (*b*) from Quay and Müller-Schwarze, 1970.)

subauricular glands and a single dorsal gland. Male wolves, especially the alphamale, deposit anal gland secretion on feces more often than do females or juveniles (Asa *et al.*, 1985). However, skin glands can be larger in females, as in the cotton-top tamarin, *Saguinus o. oedipus* (Epple *et al.*, 1988).

The Harderian gland, wrapped behind the eyeball, is especially prominent in rodents. It produces a variable, primarily lipid secretion and drains through the Harderian-lacrymal tract and the external nares. In Mongolian gerbils, the secretion is spread over the body during grooming (Thiessen *et al.*, 1976). A pheromonal function has not been clearly established.

The composition of a scent mark of glandular origin may be quite different from that of the original glandular secretion. For example, the scent marks that rabbits have applied to poles by "chinning" behavior are impoverished compared with the secretion on the chin or the head (Hayes *et al.*, 2002). However many scent marks comprise several different secretions. Two examples are the beaver, which marks the same spot with castoreum and anal gland secretion, and the female giant panda, whose scent mark contains compounds from glands, urine, and the vagina (San Diego Zoo, 2002).

Saliva may also contain specific chemical information for individual recognition (Blass and Teicher, 1980) or kin recognition (Block *et al.*, 1981; Smith and Block, 1990). The best-known example of a salivary pheromone is the mixture of androstenone and androstenol in the submaxillary glands of the boar.

Blood can be a source of pheromones. The chemical cues in secretion from the mucocutaneous junction in domestic cows that stimulate sexual behavior in bulls have been traced back to the blood (Rivard and Klemm, 1989).

3.3 Body odors and body region odors

"Body odor is the sum of all perceivable compounds in excreta and secreta" (Bryant and Atema, 1987). Body odors can change with diet but are nevertheless important in communication: bullhead catfish (*Ictalurus nebulosus*) use body odors in dominance and territorial relationships.

Odors from different body parts of the same individual can differ in their biological effects. In reptiles, dorsal skin odors of colubrid snakes release body bridging in crotaline snakes, but ventral and anal gland secretion do not (Bogert, 1941). Also, the urine of wolf, coyote, and red fox inhibits feeding in herbivorous mammals, while the feces of these species do not (Sullivan *et al.*, 1985a). Hamsters (*Mesocricetus auratus*) did not respond to polecat (*Mustela putorius*) urine but showed an extreme negative reaction to bedding (possibly fur odor) of polecats and other carnivores (Dieterlen, 1959).

Multiple sources can make for complex signals. For example, odors emanating from the anogenital area of the cow can arise from feces, bladder urine, secretions from the upper reproductive tract, vagina, vestibular glands, enlarged sweat and sebaceous glands of the vulval skin, and the activity of microorganisms (Albone *et al.*, 1986). Other examples of complex odor sources are the anal glands of the brown hyena, urine–tarsal gland interactions in black-tailed deer, and the diverse glands in the anal area of the dog (Schaffer, 1940).

Different secretions and body regions can provide redundant signals. Meadow voles, *Microtus pennsylvaticus*, investigate odors from urine, feces, or anogenital area of the opposite sex more than those of the same sex. Males sniffed scents from the mouth of females more than other regions, but females did not discriminate. Finally, males and females investigated odors from the posterolateral region of males more than those of females (Ferkin and Johnston, 1995a).

3.4 Diet influences on odor production and venoms

3.4.1 Fish

Changing the diet of a fish may change the behavior of conspecifics it interacts with subsequently. For instance, if one of a pair of male brown bullhead, *I. nebulosus* (a catfish), is removed from the tank and fed beef liver instead of the usual trout chow and then returned to his partner in their original tank, the resident will behave differently than if the same male is reintroduced without a diet change. The former tank mate is now a "chemical stranger." The behavior changes include loss of territory and more activity by the smaller, manipulated fish and more aggression and activity by the resident fish. These dietdependent odors are not specialized pheromones, and yet they are probably important social chemical cues in the natural territorial and dominance behavior of bullhead catfish. "Body odor" is the more appropriate term (Bryant and Atema, 1987).

3.4.2 Amphibians

The toxic alkaloids of poison dart frogs appear to depend on diet (Daly *et al.*, 1994, 2000; Caldwell, 1996). Details are described in Section 10.2.2.

3.4.3 Reptiles

Snake venom composition can reflect diet. The Southeast Asian pitviper, Calloselasma rhodostoma (Viperidae), preys on different animals in different

regions of its range. In some areas, only reptiles are taken; in others endotherms predominate in their diet, and in still others the diet includes a large proportion of amphibians. Accordingly, venom composition varies with diet. Other possible explanations for this variation, such as geographic proximity, or patristic (phylogenetic) distance, were ruled out statistically (Daltry *et al.*, 1996).

3.4.4 Birds

Similar to amphibians, the pitohui, a toxic bird in New Guinea, appears to derive its toxic alkaloids from its invertebrate diet (Dumbacher *et al.*, 2000).

3.4.5 Mammals

Several rodent species produce social odor signals that vary with diet. Lactating laboratory rats produce an odor in their caecal contents that attracts their own young. However, the pups are also attracted to the caecal odor of a different female if she lived on the same diet as their mother. The kind and numbers of caecal bacteria also change with diet. Such a variable odor implies that the young have to learn their mother's cues (Leon, 1975). Male guinea pigs (Cavia sp.) investigate female urine for a longer time if the donor females were eating standard guinea pig diet than if they were fed a commercial rat chow. It is possible that chemical signals in the urine provide information on available food sources (Beauchamp, 1976). In addition to fecal and urinary odors, scent gland secretions can also be altered by diet. Young of the Mongolian gerbil, Meriones unguiculatus, prefer odors of adults who share their and their parents' diet. The odor of whole animals and that of soiled sawdust or sebum from the ventral scent gland carry the diet-dependent signals (Skeen and Thiessen, 1977). Many compounds that are found in castoreum of beaver, Castor canadensis, are known as metabolites of compounds in trees the beaver feeds on (Fig. 3.11).

Diet is only one of the factors responsible for odor variability. For example, genotype and diet contribute additively to the complex odor signatures that permit kin recognition in spiny mice, *Acomys cahirinus* (Porter *et al.*, 1989), and the levels of 5-methyl-2-furoic acid and homogentisic acid in the sternal gland secretion of brushtail possums, *Trichosurus vulpecula*, rise after experimental feeding on *Eucalyptus* for 2 days (Salamon, 1995).

To discriminate diet-dependent odors can be vital in the context of reproductive behavior. Supporting the hypothesis that animals discriminate and prefer potential mates that are in good nutritional condition, Ferkin *et al.* (1997) showed that meadow voles preferred odors of members of their own species that are on a high-protein diet (Table 3.2).

Table 3.2 Some examples of diet-dependent composition or effect of mammalian secretion

Species	Secretion	Dietary factor	Affected compound(s) or response
Brushtail possum Trichosurus vulpecula	Sternal gland	Eucalyptus leaves	5-Methyl-2-furoic acid, homogentisic acid
Guinea pig	Urine	Commercial guinea pig food versus rat food	Preference by males
Mongolian gerbil	Ventral gland	Type of laboratory chow	Phenylacetic acid; pups prefer odor of gerbils on same diet as their mother
Meadow vole	Anal gland, urine, feces	Protein, 9, 15 or 25%	Mate choice: voles with high-protein diet preferred

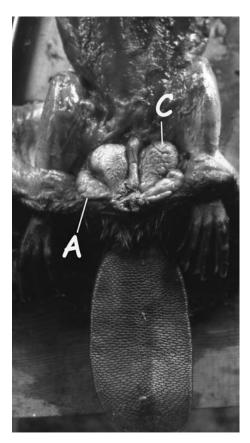


FIGURE 3.11 Castor sacs (C) and anal glands (A) in the beaver. (Photograph, D. Müller-Schwarze; dissection, B. Stagge.)

3.5 Hormonal control of odors in urine and secretions

Numerous studies have shown that reproductive hormones regulate skin gland activity.

3.5.1 Reptiles

The protein-rich secretion of the femoral gland of the agamid lizard, *Amphibolurus ornatus*, depends on hormonal control (Fergusson *et al.*, 1985). In male iguanas, *Iguana iguana*, femoral gland activity and androgen levels are correlated with dominance status (Alberts, 1993). In advanced snakes, the levels of non-volatile integumentary lipids that serve in species identification vary with hormonal state, skin-shed state, and season (Mason, 1992).

3.5.2 Mammals

Testosterone stimulates secretion in sebaceous and apocrine glands in rabbits, rats, guinea pigs, and hamsters, while estrogen in most cases inhibits secretion. Progesterone tends to be neutral (reviewed by Ebling, 1977).

Androgens affect many skin glands. An example is the flank gland of the golden hamster, M. auratus. The size and pigmentation, combined in an index, correlate with the relative levels of circulating androgens (Vandenbergh, 1973). The perineal and neck glands of cattle calves are affected by estradiol-17- β , but in different ways. The volume of the sebaceous glands in the perineal region increases, while that of the neck glands decreases. The sweat glands in the perineal region also increase in size while those in the neck do not change (Blasquez $et\ al.$, 1987). Elephants have high concentrations of testosterone and dihydrotestosterone in their temporal glands (Rasmussen $et\ al.$, 1984). The size of the snout scent gland (morilla) of the male capybara ($Hydrochaeris\ hydrochaeris\ hydrochaeris\$

As expected, castration affects gland size, activity, and composition of the secretion profoundly. For example, castration changes the levels of porphyrins, indoles, and proteins in the Harderian gland secretion of the Syrian hamster (Buzzell *et al.*, 1991).

The secretion of the sebaceous glands on the haunch of male and estrous and diestrous female rats varies considerably between individuals. No one single compound is characteristic of sex or reproductive condition. Gas liquid chromatography with principal component analysis of the 22 common peaks showed sex-specific patterns and 79.5% of variation was linked to sexual status (Natynczuk and MacDonald, 1992).

Hormone implants can be used for long-term stimulation or suppression of odor production. Scent marking is stimulated by testosterone in gerbils and wolves (Asa *et al.*, 1990).

Urine composition depends on testosterone, as in male mice (Schwende *et al.*, 1986) and the wolf, *Canis lupus* (Raymer *et al.*, 1986). In the wolf, these volatiles signal both sex and sexual maturity (Raymer *et al.*, 1986). Similarly, composition of female urine changes with the estrus cycle (Schwende *et al.*, 1986).

3.6 Microbial odors

Microbial activity can be critical for odor production in glandular secretions. Frequent body contact in social animals ensures sharing of microorganisms, which, in turn, results in a shared group odor.

In the anal sac contents of the red fox, *V. vulpes*, six genera of bacteria have been identified. The most common is *Clostridium*, with nine species. Also found were *Eubacterium*, *Bacteroides*, *Peptostreptococcus*, *Bifidobacterium*, and *Fusobacterium* (Ware and Gosden, 1980). These bacteria produce fatty acids. Aerobic microorganisms in the anal gland of the red fox include *Streptococcus* spp., *Proteus* spp., coliform bacteria, *Staphylococcus* spp., *Pseudomonas* spp., *Neisseria* spp., and *Bacillus* spp. (Gosden and Ware, 1976).

Svendsen and Jollick (1978) studied the microbes in the castor sacs and anal glands of the beaver, *C. canadensis*. No bacteria cultured from the castor sacs, while the anal sacs contained the aerobe *Escherichia coli* and the anaerobe *Bacteroides fragilis*. Species and numbers of bacteria did not differ between the sexes, age classes, or beaver colonies.

Female laboratory rats seem to depend upon an intact vaginal bacterial flora to be olfactorily attractive to males. When given a choice in a four-arm maze, adult males spend more time with an untreated estrous female than with one whose vaginal bacteria had been killed by daily injections of an antibiotic (Merkx *et al.*, 1988). Generally speaking, individual odors of rats have microbial, genetic, and dietary components (Schellink and Brown, 2000).

Staphylococcus aureus and the yeast Candida kruzei dominate the inguinal gland pouches of rabbits, with Bacillus subtilis, E. coli, and Streptococcus faecalis also occurring (Merritt et al., 1982).

Saddleback tamarin monkeys, *Saguinis fuscicollis*, harbor a complex microflora in the secretions of their circumgenital glands. Coagulase-negative staphylococci, Gram-negative bacteria, *Streptococcus* spp., and coryneform bacteria were

most abundant, followed by S. aureus and Bacillus spp. (Nordstrom et al., 1989).

In the human axilla, bacteria produce odoriferous steroids. The concentration of 5α -androst-16-en-one in the axillae of adult men reduced after treating the "superior" (more-producing) axilla with the germicidal agent Povidone-iodine (Bird and Gower, 1982). Faint or acid odors are associated with micrococcaceae, while the more pungent axillary odor that resembles that of androstene and other $C_{19}\Delta^{16}$ -androgen steroids correlates with a strong population of lipophilic diphteroids (Leyden *et al.*, 1981). The typical axillary odor is produced by incubation with coryneform bacteria, but not Micrococcaceae, propionibacteria, or Gram-negative organisms. Less-odorous steroids are transformed to the more odorous 5α -androst-16-en-3-one and 5α -androst-16-en-3 α -ol (Gower, 1989). The suggested pathway to $3\alpha(\beta)$ -androstenol is from 5,16-androstenol via the 4,16- and $5\alpha(\beta)$ -derivatives, and finally $3\alpha(\beta)$ -androstenol (Gower *et al.*, 1989).

Besides steroids, axillary odor also contains normal, branched, unsaturated aliphatic acids with 6 to 11 carbon members. Most abundant is (*E*)-3-methyl-2-hexenoic acid. Both this compound and its (*Z*)-isomer are found in the aqueous phase hydrolysate and the aqueous phase incubated with bacteria. It is assumed that precursors are water soluble and converted to odoriferous compounds by the axillary flora (Zeng *et al.*, 1992).

Finally, preputial secretion of male mice probably assumes pheromone activity as attractant for females only after having been metabolized by microorganisms (Ninomiya and Kimura, 1988).

3.7 Reservoirs

After a secretion has been produced, it can be stored for later use or even accumulated for massive or repeated signals. Anal sacs of canids, mustelids, and felids, and the castor sacs of beaver are examples. The skunk provides the most dramatic example for such reservoirs with large amounts of often very potent secretions. We do not understand well if and how the various compounds are transformed in these reservoirs.

3.8 Pheromone transport

Many pheromones travel from the tissue of their synthesis via the bloodstream to the surface of the body. They are also transported by larger molecules, notably proteins, both when being emitted by an odor donor ("outgoing") and when being received in the olfactory mucosa of an addressee ("incoming"). In the saliva of the male pig, the pheromone-binding protein

pheromaxein binds outgoing 16-androstene steroids. Two compounds serve as pheromone: 5α -androst-16-en- 3α -ol (3α -androstenol) and 5α -androst-16-en-3-one (5α -androstenone). Both are produced in the testes, transported by the blood, concentrated in the submaxillary salivary glands, and bound to pheromaxein there. Pheromaxein is formed in the submaxillary salivary glands (reviewed by Booth, 1989). The binding is strongly temperature dependent. At 4 °C the binding of the pheromone to the protein is unchanged over 168 hours. But at 21 and 37 °C (body temperature) most binding is lost after 72 hours. This is ecologically significant: pigs breed in the cold season (i.e. late autumn and winter). The salivary foam is present around the mouth of the boar when courting a sow, but it also deposited in the environment. On the body with its higher temperature, odor release is facilitated, while away from the animal retention of pheromone in saliva is optimized because the degradation of pheromaxein is retarded at lower temperatures (Booth, 1987).

The androst-16-enes in humans are also produced in the testes but spread widely by transport through the body in blood, saliva, seminal fluid, and axillary secretion of males (Gower, 1989).

3.9 Environmental odors for communication

Some chemosignals are not produced by the sending animal itself, but rather appropriated from other individuals, species, or the environment. "Scent rolling" in manure or on carcasses by canids or hyaenids (Drea *et al.*, 2002) and self-anointing by hedgehogs (Poduschka and Firbas, 1968) belong here. Rats use food odors lingering about their snout for communicating information about food sources (Galef and Kaner, 1980; Galef and Stein, 1985).

Mammals often damage plants or the substrate before or during marking, such as when deer thrash a sapling or tree branch with the head or antlers, or scrape the ground before urinating on it. Odors from the plant or soil may become part of the active signal.

Environmental odors can also play a role in interspecific interactions such as preying. The domestic dog uses cues from disturbed soil and crushed plants when tracking and they can be trained to follow tracks that lack scent. When following a scent, such as a human track, they can be diverted by another scent crossing the first pathway. The choice made by the dog is influenced by the odors and cues available at any point (Most and Brückner, 1936; see p. 414).

The environmental odors used for homing in fish, amphibia, and birds, or possibly for long-range navigation, are not yet understood and will be discussed in Chapter 4.

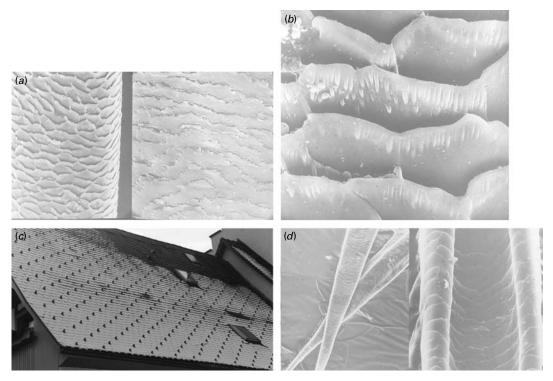


FIGURE 3.12 Osmetrichia (scent hairs). (a) Specialized hair from the tarsal gland of the black-tailed deer (left) and human axillary hair (right); (b) cuticular scales of tarsal scent hair; (c) surface of a snow-retaining roof (Germany); (d) modified U-shaped scent hair from the ventral gland of the Mongolian gerbil. (a, b, d, from Müller-Schwarze et al., 1977; c, photograph by D. Müller-Schwarze.)

3.10 Supporting structures: osmetrichia, muscles

Scent glands often have accessory structures that store secretion, enlarge the surface for scent dispersal, serve as "applicators" during scent marking, or provide opportunities for bacterial action or interaction of compounds that may come from different sources.

Special modified hair in the region of a scent gland can enhance its function. Such "scent hairs" have been termed *osmetrichia* (Müller-Schwarze *et al.*, 1977; Fig. 3.12). They may be stiff bristles with surface chambers formed by their cuticular scales, as in the tarsal tuft of black-tailed deer (Fig. 3.12a), spoon or spatula like, as in the ventral gland of the Mongolian gerbil *Meriories unguiculatus* Fig. 3.12d), or a "wick" formed by a hollow medulla and vacuolated cortical

region, as in the African crested rat, *Lophiomus imhausi* (Stoddart, 1979). Hairs from scent glands in bats of the families Pteropodidae and Molossidae differ from ordinary body hair by being much thicker, and in some species also have a differently sculptured scale pattern on their surface (Hickey and Fenton, 1987). The red fox has heavily sculpted bristles on its supracaudal gland (Brown and MacDonald, 1985).

3.11 Special adaptations for broadcasting chemosignals

The output of signals can increase only within limits because of the cost of producing signal compounds. In addition, as signal strength rises, the effect becomes progressively more inefficient: the perceived odor magnitude increases as a function of signal strength raised to a power of much less than one (i.e. logarithmically). A better way to increase odor detectability is for the receiver to become more sensitive, rather than the sender to emit more material.

Yet there are many ways the producer can improve detectability. Animals increase the spatial range of chemical signals by ingenious means. Many increase the active space, the three-dimensional portion of the animal's home range where the stimulus is above threshold, by broadcasting from an elevated place, as by the mudpiles beavers build for their scent marks. The evaporative surface can be enlarged, as in red deer (wapiti) or goats, which spray urine onto their mane. The saiga antelope (Saiga tatarica) wears a large hair tuft below its eye when in its winter coat. It is thought that secretion from the preorbital gland accumulates there on the large surface (Frey and Hofmann, 1997; Fig. 3.13). Male tomb bats saturate their brushlike beard with glandular secretion (Quay, 1970). Hairs on the anal pockets of the male capybara are saturated with secretion that is deposited on vegetation and the ground (MacDonald et al., 1984).

A scent can be propelled into the environment. The most spectacular example is the "manure spreader" of the hippopotamus: the rotating tail sprays about urine and feces (Olivier and Laurie, 1974).

Some mammals have evolved elaborate behaviors to aim odors at conspecifics during critical encounters. Black-tailed deer *O. h. columbianus*, spray urine as a threat. Facing an opponent, a male, female, or fawn rubs the hocks together and urinates over them, soaking the brush-like hair patch on the inner sides of the hocks in the process. Urine odor in the air is noticeable to a human observer farther away than the distance between the interacting animals, and also at a greater distance than during regular urinating. Such rub-urination usually results in spacing of adult deer, while fawns spray urine when suddenly separated from their mother or – in captivity – a caretaker. It is important to know that the hocks

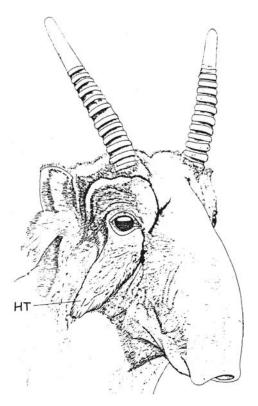


FIGURE 3.13 The hair tuft (HT) of the Saiga antelope is possibly a scent distributor. (From Frey and Hofmann, 1997.)

carry the tarsal gland, dominated by sebaceous glands (Müller-Schwarze, 1971). Gas chromatographic analysis of urine before and after it passes the tarsal tufts showed that during rub-urination material is picked up from the gland and hair surface (D. Müller-Schwarze, unpublished data).

Male bats have pouches on the leading edge of their wings, termed ante-brachial sacs. During courtship, males hover in front of, and waft odor toward, roosting females (Bradbury and Vehrencamp, 1977). The sacs themselves do not contain scent glands. Rather, the bat transfers the perfume from other glandular areas on its body. First, a resting male bends his head down toward his genital region and picks up urine with his mouth. Then he licks his wing sac. This is seen as cleaning the pouch. Finally, the male reaches down to the genital region again. This time, he presses his throat with the gular glands to the penis and transfers a droplet of secretion from the penis to the wing sac (Voigt and von Helversen, 1999; Voigt, 2002).

The arrangement of skin glands on the foot of a mouse, for instance, might convey information. As the mouse travels, the glands stamp a pattern on the



FIGURE 3.14 The pattern of skin glands on the sole of the foot of a mouse may provide information on the direction of travel. (From Ortmann, 1956.)

substrate that may signal size of the mouse and the direction it has been traveling (Ortmann, 1956; Fig. 3.14).

Short-range transfer of chemical factors may require body contact. Male Appalachian woodland salamanders press or slap their mental gland on the nares of the female (Arnold, 1966). The pygmy salamander even pierces the skin of the female's head with his modified teeth and "vaccinates" it with the secretion from his mental gland.

Chemical cues in orientation and navigation

Where will I find a patch of fresh grass?
The answer, my friend, is blowing in the wind.
With apologies to Bob Dylan

Nec vident terras, sed in odorem earum natant. (They don't see the land, but swim following its smell.)

(Red deer, *Cervus elaphus*, when supposedly swimming from Cyprus to the mainland of Asia Minor.)

C. PLINIUS SECUNDUS (Pliny the Older): *Naturali Historiae*, Book 8: *Zoology; Land Animals*, 115: *De cervis*, p. 89. Translated into German. Tusculum Library, Heimeran Verlag, 1976.

Humans depending on fishing and hunting have learned at their peril about animal migrations very early on. Today we still marvel at how precisely migrating organisms time their departures and arrivals and almost never fail to find their destinations, even passing on this vital information to their offspring. Young wandering albatrosses circumnavigate the Southern Ocean many times in the course of 8 or more years, before they return to breed at exactly the same place they were born. What cues guide these migrants in their epic journey? We know how insects, fish, and birds use the sun, the stars, and the magnetic field of the earth. But what role do chemical cues play? The physical and biological environments abound with volatile chemicals that vertebrates might use to orient themselves in space. In air, point sources such as a volcano, a small pond, or a tree in bloom are available, as are landscape odors such as those from pinewoods or the ocean. Even ship-borne humans can smell large seabird rookeries on ocean islands from afar. Arriving from sterile Antarctica, an air passenger landing in New Zealand is overwhelmed by its steamy, musty, moldy greenhouse odor. If humans notice such landscape odors, we can expect animals to be much more sensitive and attentive to them. Indeed, early explorers reported striking cases of animals smelling land. Alexander von Humboldt noticed that pigs on board his ship were excited and sniffed toward land when still 30-40 miles from the

Mexican coast with its aromatic vegetation (Faak, 1990). There is every reason to expect that wild animals exploit such landscape odors for their needs.

Vertebrates use many different cues for orientation in space and during navigation when traveling long distances. *Orientation* is defined as moving in relation to an external stimulus, for example light or a feature such as a more complex landmark. A landmark can be a prominent tree, a river, or a coastline. In *navigation*, a goal is approached by means other than the use of landmarks. Navigation is "the self-controlled movement toward an unperceived goal" (Dusenberry, 1992). Navigation is thought to require knowledge of one's geographical position (map component) and a compass. The chemical senses have been implicated in fish migration for a long time, while we are only starting to appreciate this sensory channel for other classes of vertebrate.

4.1 Fish

Among vertebrates, the homing of migrating fish of the anadromous type (undertaking upstream migration) is the classical example of olfactory orientation in space. Such a spawning mechanism was proposed over 100 years ago. Hasler (1954) formulated four testable hypotheses: (1) each stream has a characteristic odor; (2) migrating fish can differentiate among them; (3) the odor of the home stream is learned (imprinted) by young fish; (4) this chemical information is retained in memory and used by adults for orientation during their spawning trips upstream.

4.1.1 Salmon

Numerous experiments with salmon have revealed several facets of their orientation mechanism: Fish can be trained to distinguish odors of different streams; homing behavior is impaired in anosmic experimental fish; the odor is learned early in life, as transplanted presmolt salmon will return to their adopted stream and not their natal stream; and the odor is remembered after only a few days in a particular stream. Atlantic salmon, *Salmo salar*, acquire lasting information about home-stream odor during the period of smoltification (i.e. the transformation from *parr*, which lives in freshwater, to *smolt*, which is salt-water adapted, silvery, and swimming downstream). Conditioning different age groups to L-cysteine and subsequent measuring of their heart rate in response to this compound demonstrated this "imprinting" during a sensitive period (Morin *et al.*, 1987).

In an experiment, hatchery-reared Coho salmon have been imprinted to the artificial odors of morpholine, a heterocyclic amine (C_4H_9NO ; 5 × 10⁻⁵ mg/l);

HN
$$\beta$$
—Phenylethyl alcohol

FIGURE 4.1 Structures of morpholine and phenylethyl alcohol.

and phenylethyl alcohol ($C_8H_{10}O$ at 1×10^{-5} mg/l) (Fig. 4.1) as smolts at the age of about 18 months. (Unconditioned salmon detect morpholine at 1×10^{-6} mg/l water). The fish experienced these odors for 6 weeks between 1 April and 13 May during their presmolt and smolt stage (up to 16 months of age). (Presmolt fish start migrating downstream from their home stream.) In May, the two groups, plus a control group, were tagged and released into Lake Michigan midway between the mouths of two rivers, 4.7 km apart (Fig. 4.2). Eighteen months later, during the upstream spawning migration in autumn, morpholine (final dilution in stream 5×10^{-5} mg/l stream water) was added to one river, and phenylethyl alcohol (5 \times 10⁻³ mg/l) to the other. At 19 monitoring stations along the shore, fish were counted by electrofishing, gill nets, and creel censuses (information from fishermen). The imprinted fish were trapped in both rivers. They had been successfully decoyed by the artificial odors: of the total number of morpholine-exposed salmon that were recovered, 94.1% were found in the morpholine-treated stream in 1 year of the experiment, and 97.6 in the second. Almost 93 % of the alcohol-imprinted fish recovered were caught in the alcoholscented river (Scholz et al., 1976; Hasler et al., 1978). Hasler and Scholz (1983) have summarized these studies.

4.1.2 Mechanisms

Three mechanisms of home-stream recognition have been proposed: the original *fish stream imprinting* hypothesis by Hasler and Wisby (1951) assumed the memory of one single odor, that of the home stream. The odor is thought to emanate from rocks, soil, and vegetation. The *sequential imprinting* hypothesis by Harden-Jones (1968) postulated successive learning of several water odors along the migration route. Finally, the *pheromone hypothesis* of fish migration sees a role for conspecific odor in the home stream as a critical part of the environmental odor (Nordeng, 1971).

There is evidence for all three hypotheses.

1. The cited experiments by Hasler and coworkers demonstrated fish stream imprinting (Scholz *et al.*, 1976; Hasler *et al.*, 1978).

63

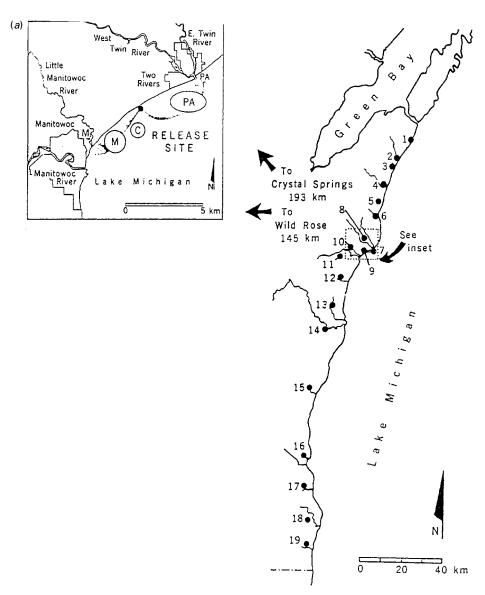


FIGURE 4.2 The classic field experiment to demonstrate chemical stream imprinting in salmon. (*a*) Lake Michigan is on the right in the main picture and the land area of Wisconsin is on the left. Three groups of fish were released between rivers No. 8 (twin Rivers) and 10 (Little Manitowoc River). (This are is shown enlarged in the inset.) One group was imprinted on morpholine (M) and another on phenylethyl alcohol (PA). The third group served as an untreated control (C). To test their chemical orientation, river No. 8 was scented with PA, and river No. 10 with M. Other rivers, numbered 1 to 19, were also monitored for entering fish. They served as control rivers. (*b*) Most M-imprinted salmon showed up in the M-scented river, and most PA-imprinted fish were caught in the PA-scented river. Most untreated control fish swam into unscented control rivers. Almost no imprinted fish appeared in the "wrong" rivers. (From data in Scholz *et al.*, 1976.)

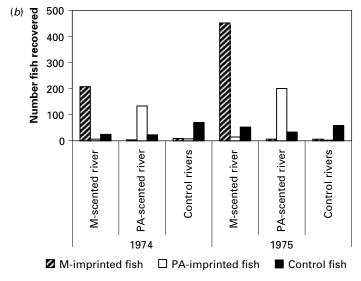


FIGURE 4.2 (cont.)

- 2. When transported 4 or 90km from their Norwegian home stream into the sea, both intact and anosmic adult Atlantic Salmon, *S. salar*, did not return to their home stream but entered many streams along the coast. These transported fish lacked the opportunity to gather experiences during the outward smolt migration. These experiments support the sequential imprinting hypothesis (Hansen *et al.*, 1987).
- 3. Support for the pheromone hypothesis has come from preference tests in Atlantic salmon parr and experiments with sea lampreys migrating upstream to spawn.

The bottom-dwelling juveniles (parr) of salmon mark their substrate, gravel, probably with fecal material. The source of the material is the gastrointestinal tract. It originates probably in the liver but is not found in skin mucus. When given a choice, parr prefer the odor of their own strain but are also attracted by that of another strain if only that is available. This suggests a hierarchical order of chemical recognition, ranging from sibling to strain to species. Outside the migrating season, the salmon do not respond to these odors, suggesting a role in orientation (Stabell, 1987). A different experiment also supports Nordeng's pheromone hypothesis. If a basket with smolt of Atlantic salmon is placed in a freshwater pool, salmon will enter the pool (Johannesson, 1987).

Migrating male sea lampreys, *Petromyzon marinus*, are attracted to water from conspecific larvae (Teeter, 1980). This jawless fish originates in the Atlantic Ocean but has invaded the Great Lakes, where it parasitizes fish populations,

posing a serious environmental danger. Adults migrate up streams to build nests and spawn in spring. The blind larvae filter feed in stream sediments. After 3 to 18 years, they metamorphose into the parasitic form, migrate downstream, and feed on other fish in lakes or the ocean. After this parasitic life, during which they gain 100-fold in body mass over 15 months, the lampreys stop feeding, start maturing sexually and migrate upstream to spawn. At the end of this 4–8 week migration phase, the fish spawn and die. Since sea lamprey larvae live in less than 10% of 6000 tributaries draining into the Great Lakes, although many more are suited as habitat, these populations may be limited because lampreys attract each other by pheromones (Bjerselius *et al.*, 2000).

In the laboratory as well as in the field, larval waterborne stimuli attracted sexually immature males and females. (The field tests were run from 22:00 to 04:00 hours, the activity time of migrating lampreys.) Certain conditions have to be met: the fish responded only early in the night, and before being fully sexually mature.

Some experiments do not support the pheromone hypothesis. Atlantic salmon grilse (salmon a year older than smolt) do not need a conspecific (smolt) cue for their return from the sea to a saltwater bay where they had been released as smolt. They return to the release site on their own accord but will not proceed from there and enter freshwater pools (Johannesson, 1987). This behavior is not in accordance with the pheromone hypothesis. Similarly, Arctic charr (Salvelinus alpinus) could not be lured into a non-natal stream that contained adult conspecifics (Black and Dempson, 1986). Pacific salmon (Oncorhynchus kisutch) returned to their natal hatchery, even though on their way from the ocean to their natal area they had to pass a major hatchery where their full-sibling relatives discharged their odor into the water (Brannon and Quinn, 1990).

Barging salmon down the Columbia and Snake rivers impaired their homing. The fish returned but they did not return upstream to spawn, for unclear reasons. It is possible that they failed to imprint on the home stream when barged. To avoid this problem, spillways are recommended (Lorraine Bodo, American Rivers/Columbia River Alliance). This is reminiscent of trucked reindeer in Sweden (see Section 4.5).

Brook trout (*Salvelinus fontinalis*) have both migratory (anadromous) and non-anadromous populations. Fish from each type were tested for their responses to home-stream water. Both preferred home stream over control water but were equally attracted to water from their home-stream and from an unfamiliar stream. Therefore, specific local odors may not be critical cues. Furthermore, anosmic fish made the same choices, indicating that taste may be involved. In the non-migratory population, responses to home-stream water were noted only in summer, not in winter (Keefe and Winn, 1991).

4.1.3 Chemical nature of odor cues

Salmonid fishes such as charr (*S. alpinus*) and grayling (*Thymallus thymallus*) are assumed to use *bile acids* for several functions, most notably homing. Bile acids are prime candidates for this function because they are diverse, specific, potent, and stable. Bile acids elicit electrical responses in the medial part of the olfactory bulb. By contrast, amino acids elicit responses in the lateral part. Taurine-conjugated bile acids are up to 1000 times more potent than methionine. There may be two different types of receptor: one for bile acids and one for amino acids. Because of their "great potency as odorants," their evolutionary history and variability, and their "renowned adherent properties," bile acids are "interesting candidates for specific signals in the aquatic environment" (Døving *et al.*, 1980).

Sea lamprey larvae (*P. marinus*) produce and release petromyzonol sulfate (PS) via their feces, a bile acid not found in any other vertebrate, and allocholic acid (ACA) (Fig. 4.3), which increased swimming activity by adult female sea lampreys in their migratory phase but did not attract them within the confines of the laboratory tanks (Bjerselius *et al.*, 2000). These authors conclude: "our studies lend support to the hypothesis that unique bile acids released by larval sea lamprey attract migratory adults into rivers to spawn." Sea lampreys are very sensitive to P and ACA, and only to these compounds, as revealed by electro-olfactograms, recorded from their olfactory lamellae. The fish detected these two compounds at a concentration of 10^{-12} mol/l (Li *et al.*, 2002). In lamprey streams, the concentrations of PS and ACA are high enough to be detected by adult sea lampreys (Polkinghorne *et al.*, 2001). The migratory pheromone contains at least three compounds (Fine and Sorensen, 2004) and attracts more than one species (Fine *et al.*, 2004).

Eels are catadromous fish that travel downstream to spawn in saltwater. Glass eels are the eels' juvenile stage. They migrate only once from their natal area in the ocean to freshwater bodies, as far as 1000km inland, growing and developing during that year-long journey. Therefore, eels cannot imprint on a home-stream odor. Yet they are the attracted to bile salts and taurine dissolved in freshwater (Sola and Tosi, 1993). Since taurine occurs in fish mucus, and bile acids in urine and intestinal waste, these stimuli are candidates for pheromones (Fig. 4.3). Geosmin, (E)-1,10-dimethyl-(E)-9-decalol (Fig. 4.3), produced by actinomycetes, is the most abundant vegetation odor in surface freshwater. Glass eels swim toward this "surface water odor" in the laboratory (Tosi and Sola, 1993) when presented in freshwater. In saltwater, however, the fish avoid the geosmin odor. Salinity affects the response: a very weak geosmin concentration (10⁻¹³mg/l) attracted the eels at a salinity of 15°/oo, while a concentration of

Allocholic acid

$$C - N - CH_2 - CH_2 - SO_3$$

Taurine-conjugated bile acid

Geosmin

FIGURE 4.3 Structures of allocholic acid (petromyzonol sulfate is identical apart from replacement of the carboxyl group with sulfate), taurine-conjugated bile acid, and geosmin.

 10^{-10} mg/l had this effect only at a salinity of 10° /oo (Tosi and Sola, 1993). In summary, it is assumed that glass eels heading for a stream follow decreasing salinity and increasing geosmin odor. Near the coast, geosmin odor may become more important and guide them to a particular stream.

Upstream migrating glass eels also swim toward compounds with earthy and "green" (vegetable) odors. These are pyrazines, thiazoles, and alcohols of the cyclohexanol type (Sola, 1995). One of these compounds, 2-isobutyl-3-methoxypyrazine, is common in rivers and lakes (Hwang *et al.*, 1984).

4.2 Amphibia

Anurans and urodeles are known to home to their ponds. The chemical senses as well as vision and other senses are required for this ability.

4.2.1 Anurans

Among anurans, Fowler's toad, *Bufo woodhousei fowleri*, the Mexican toad, *Bufo valliceps*, and the chorus frogs *Pseudacris clarki*, and *Pseudacris streckeri* prefer the water from their own breeding pond to that of another pond (Grubb, 1973a,b, 1976). Artificial odors can be learned: Mexican toads, tested in a complex maze used odor cues such as anise oil, benzaldehyde, cedarwood oil, citral, and others to orient themselves in a complex environment (Grubb, 1976). The toad *Bufo boreas* uses olfaction for distance orientation, and vision when near the pond (Tracy and Dole, 1969), while the reverse is true for *Salamandra* sp. The green frog (*Rana clamitans*) locates its breeding site with more difficulty when the olfactory tracts are experimentally ablated, but it does not lose the homing ability.

After metamorphosis, first-year individuals of the common European frog *Rana esculenta* stay in their home pond, even if it dries out. Experiments in a two-way choice apparatus showed that these animals prefer to head toward the odor of mud from their home pond as opposed to some other mud sample (Bastakov, 1986). Newly metamorphized (at Gosner stages 43–46) pool frogs (*Rana lessonae*) caught in the wild preferred water from their pond to tap water, and also to water from an unfamiliar pond. When tadpoles were raised in water from their home pond, or water with boiled nettle as food, the froglets later preferred water with these respective stimuli. They had developed a preference during larval development (Ogurtsov and Bastakov, 2001).

The signal for homing appears to be a complex pond odor, originating in vegetation and soil. The frog *Pseudacris triseriata*, tested in a T-maze, chooses the odor of lowland muck and filamentous algae from its breeding habitat over the odor of soil and decaying vegetation from non-breeding upland habitat suspended in distilled water (Martof, 1962).

The dart-poison frog *Dendrobates pumilio* uses odors for homing. This species lives in the understory of lowland tropical forests in Central America. The eggs are laid on land, and the female carries the newly hatched tadpoles on her back to water-filled leaf axils of bromeliads. She feeds the tadpoles with unfertilized eggs, while the male defends the territory. Captive dart-poison frogs tested in a Y-olfactometer chose the odor from their own communal tank over odors from tanks planted with different plants. However, they did not distinguish between

the odor of their own tank and that of a tank with bromeliad genetically identical to that in their home aquarium. In addition to plant odors, passive marking possibly takes place, as the frogs had been in their aquarium for 3 weeks before the test (Forester and Wisnieski, 1991).

4.2.2 Urodeles

Among the Salamandridae, displaced Western newts, *Taricha rivularis*, home to the same section of a stream year after year. Blinded newts still homed successfully, and anosmic specimens were reduced in their homing ability but still did not move in random fashion. They return to their home pond from up to 12km (Twitty, 1966). Another salamander, *Ambystoma maculatum*, tested in the laboratory, preferred paper towels soaked with mud or pond water from their home pond over towels treated with materials from a distant pond (McGregor and Teska, 1989).

4.3 Reptiles

4.3.1 Snakes

Among snakes, over 20 species in five families follow scent trails of conspecifics. The trails are probably used for homing and locating dens and, therefore, serve more purposes than strictly social behavior. Odor trails may be more important for younger snakes and in northern populations. The garter snake *Thamnophis sirtalis* undertakes autumn migrations of up to 17.7km (Costanzo, 1989). Corn snakes, *Elaphe guttata*, and garter snakes, *Thamnophis radix*, use chemical cues when aggregating and selecting their shelters. The response depends on an intact vomeronasal organ. In corn snakes, a social odor is involved: individual snakes prefer shelters that had been used by groups previously (Eyck and Halpern, 1988). Eastern garter snakes do not depend on their vomeronasal organ for orientation in their home range: severing the vomeronasal nerve does not prevent these snakes from finding previously used sites. However, the distance travelled each day by these avomic snakes (7.7 m) was only about one third of that of controls (24.6 m) (Graves *et al.*, 1993).

4.3.2 Tortoises and turtles

Land tortoises (*Testudo hermanni*), especially males, caught in the wild and displaced 500–1000 m returned to the point of their capture. However, if their olfaction was impaired by washing the nasal cavity with zinc sulfate, their

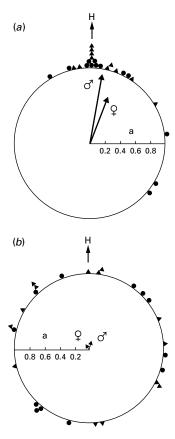


FIGURE 4.4 Homing in intact (*a*) and anosmic (*b*) land tortoises. Triangles indicate males and dots females. While intact tortoises head in the home direction (H), animals rendered anosmic by zinc sulfate treatment scattered in all directions. The inner arrows are resultant vectors for all animals of one sex and represent directedness. a, vector length, with 1.0 being perfect directedness. (From Chelazzi and Delfino, 1986.)

homing was poor (Fig. 4.4). These treated animals did move in many directions and, furthermore, did cover only very short distances during the 14 days of the experiment (Chelazzi and Delfino, 1986).

Freshwater turtles home to their pond when experimentally displaced. Painted turtles, *Chrysemys picta*, homed from 100 m, but not from 1.6 km (Emlen, 1969). They use chemical cues: painted turtles discriminate chemical cues from their home ponds and other ponds. Males and females prefer water from their home pond to that from other ponds (Quinn and Graves, 1998). The Eastern long-necked turtle, *Chelodina longicollis*, of southestern Australia uses solar cues during migration between a permanent lake and an ephemeral swamp, as

orientation is impaired on overcast days. Olfactory cues also seem to play a role: in a Y-maze, the turtles headed toward swamp mud and debris in preference to plain water (Graham *et al.*, 1996).

The sea turtles Kemp's ridley, Lepidochelis kempi, have been "head-started" by imprinting them to the odor of the water they hatched in. Eggs from the threatened beach at Rancho Nuevo, Mexico were placed in sand and water from Padre Island, Texas and transported to Padre Island. When 4 months old, these turtles were tested in a multiple choice of different kinds of water (Fig. 4.5). They preferred water from Padre Island, indicating that some form of olfactory imprinting had taken place (Grassman et al., 1984). Starting in 1978, 2000 eggs from Rancho Nuevo were hatched annually for 10 years. Later, the hatchlings were released at Padre Island National Seashore (see also Ch.12). In 1996, the first two turtles came ashore at Padre Island and laid a total of 176 eggs. These females were 10 and 13 years old, respectively. The artificial colonization of the Padre Island beaches has been an unqualified success. However, at this time it is not clear what cues the turtles use to return to Padre Island from the ocean. As the young turtles stay in shallow water for a while, they may imprint on any of many factors other than the beach chemistry. These possible other factors include solar cues, star patterns, the Earth's magnetic field, or salinity gradients in the ocean; they may follow other turtles or use still other cues.

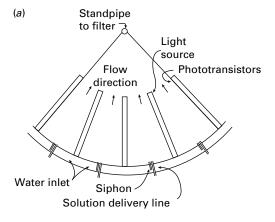
4.4 Birds

Homing by olfactory cues has been the most researched topic in bird navigation research for many years. Homing pigeons and petrels use airborne cues for orientation under certain conditions. However, many experimental results do not clearly show that *odors* are the crucial stimuli.

4.4.1 Procellariiforms

Birds' homing to their burrow

Many petrels and shearwaters approach their nest burrow, often located under forest cover, at night. When Leach's petrel (*Oceanodroma leucorrhoa*) return to their nest, they first hover over the spruce-fir canopy near their burrow. Then they plummet to the ground several meters downwind from their nest site and walk upwind to their burrows (Fig. 4.6). In still air, they landed closer to the burrow and followed a more roundabout route than in wind. With external nares plugged or olfactory nerves transected, displaced birds did not return to their burrows for 1 week. In a laboratory two-choice apparatus, breeding petrels



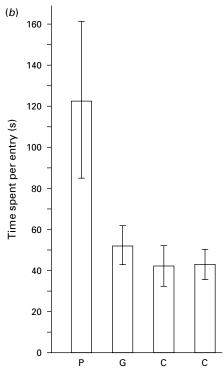


FIGURE 4.5 Sea turtle orientation. (a) Bird eye's view of apparatus to test whether sea turtles discriminate sea water samples from different locations. Sea water flows from water inlet toward the standpipe (arrows). The turtle is placed near the standpipe. Phototransistors record when the animals move into different compartments. The water washes of sand come from Padre Island (P) and Galveston (G), and unscented water in two compartments serves as control (C). (b) Turtles spent more time in water extract of sand from Padre Island than in that from Galveston. (From Grassman et al., 1984.)

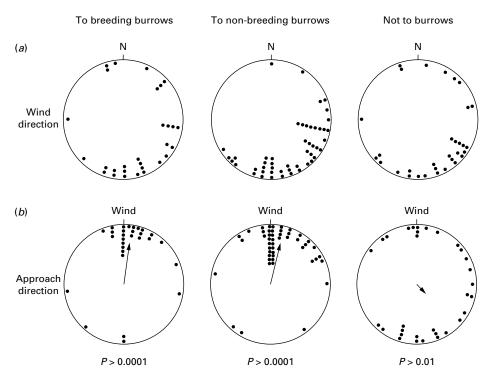


FIGURE 4.6 Orientation of Leach's petrels toward their burrows in relation to wind direction. (a) Wind directions measured during observations. (b) Approach to burrows on foot on the forest floor related to wind directions. Regardless of its absolute compass direction, wind direction during each observation is depicted as "north" in the diagram. Birds orient into the wind when heading for their burrows, whether breeding or non-breeding, but walk randomly when they are not going to burrows. (From Grubb, 1974.)

preferred an air stream from their own nest material to other forest floor material. These findings suggest that this species uses olfactory cues to locate the nest (Grubb, 1974).

Materials from the birds themselves most likely provide the chemical cues that are implied in this behavior. Petrels use odor in finding their nests early in life. Chicks of the British storm petrel, *Hydrobates pelagicus*, orient toward their nest from 30 cm, and they fail to do so when their nares are blocked. In a T-maze with one arm inserted into a nest burrow, the chicks were able to orient toward a nest crevice (Mínguez, 1997). Other birds may also use odor to orient toward their nest: domestic chicks, *Gallus gallus domesticus*, prefer odor of their own bedding (Jones and Gentle, 1985). Experiments show that possibly only nocturnal burrowing petrels use olfaction to find their nest, while surface nesters and dayactive species are visually orientated (Bonadonna and Bretagnolle, 2002).

The snow petrel, *Pagodroma nivea*, of the Antarctic accumulates stomach oil around its nest. These deposits can grow up to 0.5 m thick and have been radiocarbon-dated as 4000 and 9000 years old in the Shackleton range and the Wohlthat Massif, respectively, in Queen Maud Land (Hiller and Wand, 1984). The birds might use odors from these accretions for orientation around their nests. However, a small-scale field experiment in Antarctica's Mühlig-Hofmann mountains suggested that snow petrels do not use odors to find their nest for distances over 1–2 km. The same percentage of birds returned to the breeding colony from those with nasal plugs and those intact (Haftorn *et al.*, 1988). Manx' shearwaters, *Puffinus puffinus*, however, do not seem to use odors for finding their burrows (James, 1986). Blue petrel (*Halobaena caerulea*) find their burrow by smell, as field experiments in the Kerguelen showed (Bonadonna *et al.*, 2004).

Finding food

On the high seas, petrels and albatrosses unfailingly locate patches of krill, their main food. Such patches are as small as a few cubic meters, and the larger ones extend over an area the size of several football fields. On their feeding trips of thousands of kilometers, birds such as black-browed albatrosses or white-chinned petrels find the "needle in a haystack" by celestial cues, the earth magnetic field, and by detecting high odor concentrations in an "olfactory landscape" over the ocean. On such a large scale, they possibly use an odor that emanates from the water when zooplankton such as krill feed on phytoplankton. The relief of the ocean floor is ultimately responsible for this "odor landscape": undersea ridges create upwelling that transports nutrients to the upper water layers, benefiting phytoplankton, the basis of the food chain. We do not know how these seabirds accomplish long-range orientation. Once close to a krill patch, they can use chemical cues blown downwind, or visual cues such as other birds feeding (Nevitt, 1999). The close-range responses of these tube-nosed birds to chemicals released into the air when krill is feeding on phytoplankton are discussed on p. 351. The best known of these food-signaling compounds is dimethyl sulfide.

4.4.2 Homing pigeons

During the last 30 years, several laboratories have examined homing pigeons for their ability to use odors for orientation. Early experiments suggested that olfaction is important in orientation: homing pigeons with their olfactory nerves cut were "generally found to be unable to home from short

distances" (Benvenuti *et al.*, 1973). According to Papi's (1976) hypothesis, pigeons learn their loft odor and foreign odors blown their way by the winds. These odors provide the map component, while the sun or magnetic compass is used to deduce flight direction. Papi based his hypothesis on four phenomena. (1) Pigeons prevented from breathing through their nostrils were incapable of correct initial orientation when released at a distance from their loft. (2) If wind deflectors at the loft had shifted air currents, the initial orientation of the birds was also deflected when released at a distance. (3) When an odor was applied to their beaks at the time of release, displaced pigeons flew in the opposite direction of that from which the same experimental odor came on the wind at the loft. (4) Birds transported to the release site by different overland routes differed in their initial flight direction. This effect was abolished if they were transported with their nostrils plugged (reviewed by Papi, 1976).

In a typical early experiment, pigeons were exposed to artificial odorous winds from the time of fledging. One group experienced south wind with olive oil odor, and another group north wind with a mixture of solvent odors (toluene and terpene hydrocarbons). The birds were tested later by applying olive oil or the turpentine mixture to their nostrils while being displaced from their loft. They flew in the direction opposite from that which they had experienced the "training" odor: that is, they "homed". The olfactory information used for navigation seems to be acquired during the first months of life. If the nostrils of young pigeons are covered with masks for 3 days after fledging so that they cannot, or only barely, breathe through their nostrils, they cannot associate odors with wind directions and subsequently home poorly after displacement (Papi *et al.*, 1973). Numerous experiments, however, have raised many questions to which there are no answers at present.

How can a pigeon use olfaction to navigate? The two competing hypotheses are the *mosaic map* (Papi *et al.*, 1972) and the *gradient-map* (Wallraff, 1989) models. In the mosaic map model the pigeon is assumed to qualitatively discriminate different environmental odors that occur in a patchwork over an area of up to 200 km around the home site (loft). The gradient map model postulates quantitative discrimination of long-range environmental odor gradients that extend over thousands of kilometers. So far, neither is convincingly supported (Schmidt-Koenig, 1987).

Airborne stimuli are probably necessary, but not sufficient, for homing. If air samples from future release sites are brought to the loft and pigeons exposed to them, only birds that had been exposed to the odor of their actual release site oriented toward home. In this experiment, the birds were rendered anosmic for the displacement so that they could not pick up odors during the trip to the release site (Kiepenheuer, 1985, 1986).

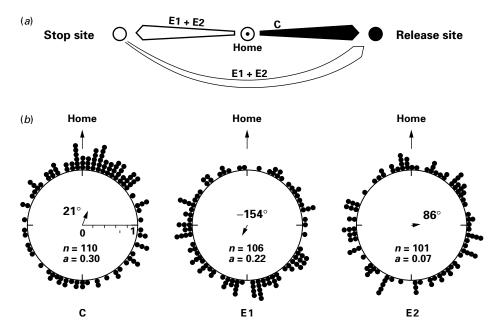


FIGURE 4.7 Pigeons can be "fooled" about where they are in relation to their loft if they are allowed to smell the air at a "distraction site." (a) Three groups of pigeons were tested for their homing ability. Experimental groups 1 and 2 (E1 and E2) were brought to a distraction site (stop site) and then transported to the actual release site in the opposite direction from the loft (white arrows). E1 were allowed to smell the local air at the distraction site but not at the release site. E2 were never allowed to smell local air anywhere. Controls (C) were directly transported to the release site (black arrow) and allowed to smell the air there. (b) The dots represent individual pigeons' bearings. The controls homed correctly. The E1 group traveled in a direction opposite of home. The E2 group lacked meaningful information and randomly headed in various directions. (From Benvenuti et al., 1992.)

Pigeons can be fooled into using a cue from a "false release site;" before their release, the birds are first brought from their home loft to a site located in the opposite direction from the release site and permitted to smell the air there (Fig. 4.7). In this site simulation test, the released birds flew in the direction opposite from home (Wallraff, 1990; Benvenuti *et al.*, 1992). It is clear that pigeons gain information about the spatial relationship between the strange release site and their home loft by odors they perceive at the unfamiliar site (Benvenuti *et al.*, 1992).

Inexperienced young pigeons, when rendered anosmic before release, have oriented normally in one experiment (Wiltschko and Wiltschko, 1987) but were impaired in another (Gagliardo *et al.*, 1988). In the latter experiment, the birds

peculiarly were still directed but flew in the wrong direction (i.e. away from their home loft).

Other experiments, however, have failed to confirm that homing pigeons used odor cues. For instance, pigeons were made anosmic by carrying nasal plugs and being sprayed in their nostrils with the local anesthetic tetracaine. Displaced shortly after treatment and released, they were not impaired in their homing. Their vector length (a measure of "directedness," regardless of where the birds flew), homeward component, and vanishing intervals between time of release and directed departure did not differ consistently or significantly from controls (Wiltschko and Wiltschko, 1987; Wiltschko *et al.*, 1987a). In one experiment, pigeons were transported to the release site unimpaired and allowed to smell the release site for 15 minutes. Then the nostrils of one half of the group were sprayed with tetracaine, and the birds were set free. The control birds were sprayed with propellant only. Birds that were sprayed with tetracaine for the first time were most affected. It is suggested that a non-specific, traumatic effect is responsible for impaired navigation, and not altered olfactory input (Wiltschko *et al.*, 1989).

Strangely, Papi's homing pigeons in Italy appeared to use odors, while the results of experiments in New York were negative (Papi et al., 1978). Later experiments confirmed that the Italian pigeons relied more on odors for homing than pigeons in Germany or the USA. During transfer to the release site, pigeons were rendered anosmic by cotton nasal plugs soaked with citrus oil 30-40km from their home loft, and the local anaesthetic tetracaine was applied shortly before the release which rendered the birds anosmic for 4-6 hours. The experiments were performed in Pisa, Italy, at Cornell University in Ithaca, New York, and in Frankfurt, Germany (Fig. 4.8). The variables measured were the direction and length of mean vector (i.e. the average travel direction), the deviation of this mean vector from the home direction, the homeward component (derived from mean vector and deviation from home), the median vanishing interval, the median homing speed, and the return rate as a percentage (Wiltschko et al., 1987b). Only the Italian pigeons were impaired, while pigeons in New York and Germany homed normally (Wiltschko and Wiltschko, 1992).

Pigeons appear to use two maps, one olfactory in nature, the other a mosaic of familiar sites. These maps are redundant for intact birds in familiar terrain, but if impaired sensorily, the pigeon can find home without one or the other of these mechanisms. This was shown in experiments with clock-shifted and anosmic pigeons (Luschi and Dall'Antonia, 1993).

Pigeons kept in "deflector lofts" (with panels that change the wind direction that they experienced) were affected in some experiments (Papi, 1990). This may

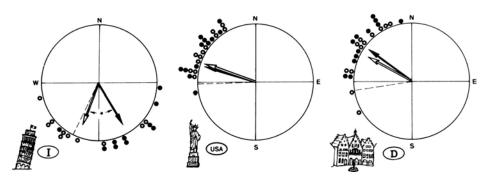


FIGURE 4.8 Pigeons in Italy misorient when deprived of olfaction, while those in New York and Germany were unaffected. In Italy (left), anosmic birds headed into a direction different from home, while no such difference between anosmic and control birds was found in New York and Germany. (Anosmic birds had their nostrils plugged with citrus oil-soaked cotton until release, and an anesthetic was sprayed into their nostrils at the time of release. This renders the birds anosmic for 4–6 hours.) Dashed radius, home direction; solid dots, individual anosmic birds; open circles, control birds; arrows mean directions for anosmic and control birds. (From Wiltschko and Wiltschko, 1992.)

actually have been a consequence of changes in the reflection pattern of polarized light, and not odors (Waldvogel *et al.*, 1988).

The methodology of odor deprivation has been problematical. It is difficult to isolate pigeons from airborne odors. Occlusion of the nostrils reduces the stimulus intensity to 20-30%, but not to zero. Bilateral section of the olfactory nerve irreversibly eliminates responses to weaker stimuli, but responses to higher concentrations remain, probably via trigeminal stimulation. Spraying the nasal cavity with anaesthetic abolishes sensitivity to odors, but the effect is variable and depends on the way the drug is applied (Wallraff, 1988a). Some of the experimental results can possibly be explained by the fact that zinc sulfate produces anosmia that lasts several days, much longer than that induced by xylocaine (Wallraff, 1988a; Benvenuti et al., 1992). Furthermore, using unconditioned responses to olfactory stimuli has its disadvantages. (a) The stimulus intensity must be high; no response does not mean that the animal did not receive the stimulus. (b) Without reward or punishment, the response will not be stable over many repetitions. Therefore, more naive birds are needed for testing. (c) This introduces individual-specific amplitudes and response curves to even the same stimulus (Wallraff, 1988a).

What do we know about the olfactory apparatus and performance of homing pigeons? The size of the olfactory bulb lies in the middle range of 151 bird species examined (Bang, 1971). What odors do pigeons use? The home site air, the air

that surrounds the pigeon during displacement, and the air at the release site have all been tested. The birds home normally if fiberglass filters remove airborne particles, ranging in size from 0.04 to 2.0 μm from the air around the birds during displacement. However, if charcoal filters are used, they home more poorly over 24–155 km than pigeons in unfiltered air (Wallraff and Foà, 1981). Odor gradients of mixtures in the atmosphere near the ground have been proposed as important for pigeon homing (Wallraff, 1988b).

The concentrations of environmental odors have been calculated to test whether they suffice to serve as odor cues for birds. Each year, there are 17 imes 10^6 metric tons of sulfur released from natural sources between 30° and $50^\circ N$ alone. Less than 10kg sulfur and ammonium are emitted daily from 1 ha. Pineoak forest in the United States gives off 1-5 kg/m² α -pinene daily. Air sampling at midday from a scaffold tower above the canopy of a loblolly pine forest in North Carolina yielded α -pinene concentrations as high as 2.5 μ g/m³ air (Arnts et al., 1982). Do these amounts provide concentrations above the olfactory threshold of the pigeons? There are 1-5 parts per billion (ppb) sulfur and ammonium in the atmosphere, and 10–100 ppb α -pinene. These concentrations are below the pigeon's detection thresholds (0.1 to 40 parts per million; Henton et al., 1966). The differences between the performances of the New York and Italian pigeons may be a consequence of atmospheric differences. The Italian experiments were carried out near the coast, which runs in a north-south direction. Indeed, east and west-displaced pigeons oriented better than north and southdisplaced birds. Seabreezes from the west may provide odor cues and these will extend for approximately 45 to 85 km inland (Waldvogel, 1987).

Other odor fields possibly available to birds are ammonia from pastures and, over the ocean, dimethyl sulfide released by herbivorous action of plankton on marine plants (Nevitt *et al.*, 1995).

Serious obstacles to olfactory navigation have been pointed out. There are drastic differences between actually observed air currents and *mean* wind directions for a given locality. Also, the odor sources are unstable temporally and spatially. Finally, there is no evidence for long-lived, strong, and stable chemical gradients in the lower atmosphere (Becker and van Raden, 1986). The Cross-Appalachian Tracer Experiment (CAPTEX 1983) showed that an aerosol released in Ohio or Ontario could be found in various concentrations over 300 to 1000 km downwind, with irregular distribution that would provide little directional information to birds for homing (Waldvogel, 1987).

Chemical cues and the earth's magnetic field may interact in pigeon navigation. Pigeons kept in an oscillating magnetic field were affected in their initial orientation only if they could smell atmospheric odors (Wallraff *et al.*, 1986). Olfactory restriction and magnetic deprivation have different effects on homing

in pigeons. In one set of experiments, pigeons were exposed to an artificial random magnetic field by one pair of horizontal and two pairs of vertical Helmholtz coils on their heads. Olfactory deprivation was accomplished by plugging the nostrils with cotton and sealing with adhesive tape. Only intact birds (no experimental magnetic field, nostrils not plugged) oriented toward home. Birds prevented from smelling were disoriented; they scattered widely in their departure directions. Magnetically impaired, but olfactorily intact, pigeons were directed, but in their preferred compass direction for that particular year and not towards home (Papi and Ioalè, 1988).

The most important research areas for the future are the central mechanisms in the bird and the nature and distribution of odorous substances in the atmosphere (Benvenuti *et al.*, 1992). Reviewing the current state of olfactory navigation in pigeons, Wallraff (2004) has presented maps of the distribution of different concentrations of six hydrocarbons in a 300×300 km area in Germany. Pigeons may well be able to use these atmospheric gas gradients for navigation.

4.5 Mammals

Small mammals are well oriented within their familiar home range but also when exploring beyond that home range, and most likely when dispersing. They home well when experimentally displaced from their home site. Finding one's way within the familiar home range is termed *topographical navigation*, in contrast to operating beyond the home range, where *geographical navigation* takes over. Topographical navigation involves three mechanisms:

- guided orientation (also: known as guidance): a direct response to objects;
- path integration: continuous monitoring and computation of the twists and turns while moving about so that the path back to the starting point ("home") can be found;
- *landmark navigation*: "movement by means of distal visual cues toward a goal not directly perceived" (Alyan and Jander, 1997).

Spatial orientation in the home range has been studied in house mice, domestic rats, golden hamsters, and Mongolian gerbils. All these species use distant (visual) landmarks as reference points. However, most of the laboratory studies focused on very short distances in the 1m range (e.g. Alyan and Jander, 1994). While most studies in mammals examined how the animals find their way after they have committed their surroundings to memory, we know less on how these animals learn to orient themselves in their home range in the first place. The brain structure instrumental in such spatial learning and memory is the hippocampus.

The "two blind mice" experiment of Alyan and Jander (1994) used the pupretrieving behavior of female house mice to determine the *visual* cues they use for orientation in an arena of 1 m diameter. A strain of blind mice did not orient as well as intact mice. In different experiments, neither sighted nor blind mice appeared to use olfactory cues such as scent trails in sand, or the odors of wooden blocks in the arena, to find their nest after their arena had been rotated by 90 degrees. Such rotation misled them on their way home, despite landmarks in their arena with odor that should have been familiar to them. We know little on the role of olfactory cues in such short-range orientation.

Given their acute sense of smell, it is reasonable to assume that mammals use environmental odors for orientation, but we lack experimental evidence. Particularly suggestive are observations of desert animals that head into the direction of the wind that blows from bodies of water, areas with rains, or fresh vegetation suited for grazing. Such so-called "smelling of rain" has been reported for camels, antelopes, water buffaloes, and others. Migrating mammals such as caribou, reindeer, or wildebeest may also use conspecific odors on or near trails. These may originate in dung, urine, or interdigital glands. Whitefooted mice (*Peromyscus leucopus noveboracensis*) homed over distances of about 200 m whether or not they had been made anosmic by treatment with zinc sulfide (Parsons and Terman, 1978).

Even marine mammals, especially baleen whales, may use odors from land or sea for navigation. They may also locate their food, krill, near the surface by sampling the air for "krill odor" (Cave, 1988).

Chemoreception

We might fairly gauge the future of biological sciences, centuries ahead, by estimating the time it will take to reach a complete, comprehensive understanding of odor.

LEWIS THOMAS, in: Late Night Thoughts on Listening to Mahler's Ninth Symphony

To appreciate the possibilities and limitations of exchange of chemical information, the chemical ecologist needs to understand how the chemical senses work. There are a number of outstanding reviews and books on chemoreception *per se.* In addition to the general principles of chemoreception, the chemical ecologist is particularly interested in adaptations of anatomy and performance to specific abiotic and biotic conditions that optimize signal detection and maximize information gain. Here odor reception will be treated from an ecological viewpoint and only to the extent that it is an essential step in the behavior of an animal moving about in its environment and attending to vital cues. While sensory studies in the laboratory employ simple and clean stimuli, *noise* and the need for *filtering* are important factors in sensory ecology (Ali, 1978).

5.1 Encounter and exposure: orientation, sniffing, licking, tasting

The ecologist is interested in behaviors that animals employ before odor molecules even reach their chemical sense organs. Animals can manipulate the amount of odorants reaching the chemoreceptors by manipulating airflow toward the receptors, body movements, or locomotion. In its natural environment, an animal may seek or avoid stimulation by odors. Such behaviors cannot be satisfactorily addressed in the confines of laboratory studies. A foraging carnivore or herbivore exposes itself continuously to new chemical signals. Alerted by a sound or visual stimulus, an antelope or deer may turn into the wind and sniff. In many contexts, such orienting movements are the first step in a sequence of behaviors whose later steps are primarily guided by chemical cues.

As any dog owner knows, sniffing, licking, and various body movements in the service of detecting biologically significant chemical signals may be elicited by visual, tactile, auditory, or chemical stimuli, not just by chemical ones. The Russian reflexologists have formally worked out these stimulus-response relationships. Chiszar (1986) has termed such investigatory behavior "dedicated motor patterns" because they are dedicated to bringing sensory receptors in contact with semiochemicals. Such dedicated motor patterns may occur in the total absence of chemical cues. Chiszar has pointed out that many experiments in chemical ecology that measure orienting or investigatory behaviors as dependent variables failed to demonstrate odor effects. A control stimulus may be sniffed as much as the putative semiochemical. As examples, Chiszar discussed the tongue flicking by rattlesnakes after striking a prey animal (strike-induced chemosensory searching), the male guinea pig's investigation of female urine, and the male golden hamster's investigation of vaginal secretion. In all these cases, rates of investigation of various samples do not depend on a particular chemical cue. That is why measuring the intensity of sniffing and/or licking in a bioassay is often a poor indicator of bioactivity of a stimulus. However, complex chemical signals may include volatile "alerting compounds" that attract the attention of conspecifics, while other components in the mixture provide more specific information or trigger typical responses (Müller-Schwarze, 1998).

5.1.1 Fish

Fish manipulate stimulus access to the olfactory organ in different ways. *Isosmates* move water by beating cilia on the olfactory lamellae and thus receive a continuous laminar flow. By contrast, *cyclosmate* species draw in water in pulses by muscles associated with the gills (sniffs or flicks). The commonly observed jaw movements termed "coughing" are a form of "sniffing" in flounders. The pressure in the olfactory sac drops rapidly during such coughing, drawing in water. The jaw movement rotates the lachrymal bone, which in turn, pushes or pulls the olfactory sac. Food odors trigger such coughing (Nevitt, 1991).

5.1.2 Reptiles

Among reptiles, desert iguanas (*Dipsosaurus dorsalis*) may first visually spot conspecific scent marks that absorb long-wave ultraviolet light and then bring their chemoreceptors close. Their femoral gland secretion is non-volatile and active only at close range. In the laboratory, iguanas detected samples of femoral secretion on tiles better in ultraviolet light than under incandescent

light. Once close to the scent mark, the iguanas' response is measured as the number of tongue-touches by blindfolded animals. The femoral gland secretion is a pheromone candidate because the gland is larger in males, is activated by testosterone, and there is a cycle of glandular activity that coincides with the breeding season. The secretion contains proteins of 10–30 kDa daltons (Alberts, 1989).

5.1.3 Mammals

Mice are able actively to seek or avoid *priming pheromones* that modulate their ovarian cycle and onset of puberty. Peripubertal female mice avoid the urine odor of adult males, known to accelerate puberty in females, and are more attracted to the odor of grouped adult females. This behavior is particularly effective because the active space of the (almost) non-volatile male pheromone is small, and prolonged exposure is required for the effect to occur (Coppola and O'Connell, 1988). Likewise, prepubertal female mice do not urinate near urine marks of adult males, while grouped, estrous, and diestrous *adult* females do. Such behavior may help young females to avoid exposure to male odors until they reach puberty. This way they would be protected from mating too early, and their eventual reproductive success would be enhanced (Drickamer, 1989a).

Various specialized structures may aid in sampling air or improving directional smelling. The elephant's trunk can be seen as an olfactory periscope; tube-nosed bats (*Murina* and *Harpiocephalus* spp., Vespertilionidae) and hammerhead sharks (Sphyrnidae) represent bizarre examples of specialized olfactory anatomy. Nostrils of lower vertebrates tend to be farther apart than those of higher vertebrates that have more flexible heads and necks (Stoddart, 1983).

5.2 Receiving molecules: chemosensory organs

Apart from taste, vertebrates have five different chemoreceptor systems for airborne chemosignals: the main olfactory system, the vomeronasal organ (VNO), the trigeminal nerve, the septal organ of Masera, and the nervus terminalis. They each will be discussed in turn. All five are fully functional in most mammals (Fig. 5.1).

5.2.1 Main olfactory system

This chapter will only deal with a few facts pertinent to the ecological operations of animals. Excellent reviews and monographs deal with the olfactory system.

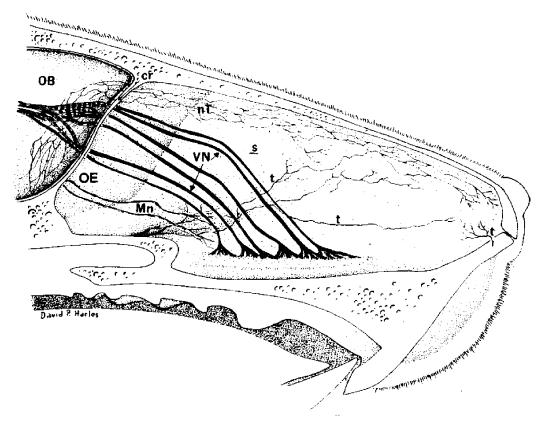


FIGURE 5.1 The five neural components that serve in olfactory reception, shown in an rat; nostrils at upper right, incisors at lower right. The main olfactory bulb (OB) receives axons from the olfactory epithelium (OE) through the cribriform plate (cr). The lines (VN) represent the vomeronasal nerve, conducting signals from the vomeronasal organ above the roof of the mouth. NT, nervus terminalis; t, branches of the trigeminal nerve; Mn, septal organ of Masera; s, septum. (Modified from Graziadei, 1977.).

Overall structure

Fish

Elasmobranchs have their paired olfactory organs on the ventral side near the mouth. As the fish takes the respiratory water current into the mouth, water passes through the olfactory sacs. Thus, elasmobranches use the respiratory water current for supplying the olfactory organ with waterborne stimuli.

By contrast, bony fish have their olfactory organs on the dorsal side of the snout at some distance from the mouth. The olfactory system in fish involves the first (olfactory) cranial nerve, while the ninth (glossopharyngeal) and other nerves serve the sense of taste.

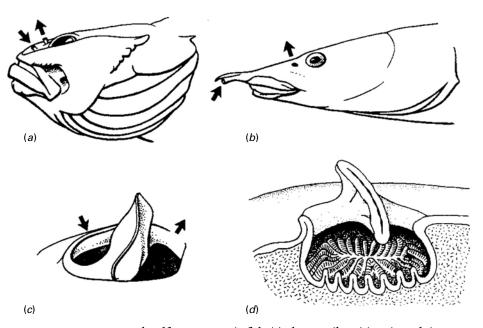


FIGURE 5.2 The olfactory organ in fish. (a) The nostril positions in sculpin (Cottidae); (b) nostril position in spiny eel (Mastacembelidae); (c) a skin flap separates in- and outflow, an arrangement typical for bony fish (here: Catastomidae); (d) the olfactory lamellae are located in the floor of the olfactory capsule (here: minnow, *Phoxinus*). (From: C. E. Bond: *Biology of Fishes*.)

The olfactory receptors of fish are located on lamellae, arranged in an oval or round olfactory rosette; this in turn, is housed in the olfactory capsule, a chamber at the front of the head (Fig. 5.2). The olfactory capsule is totally separated from the respiration system. Inlet and outlet openings permit a stream of water to pass through the olfactory capsule. Fish with round rosettes have the fewest chemoreceptors and rely less on olfaction, while elongated rosettes are characteristic for species with a keen sense of smell. Cycloosmatic fish possess accessory sacs, branching off from the main olfactory sacs. The accessory sacs act as pumps drawing water through the olfactory organ, activated by muscles of the jaw and gill regions. With the exception of lungfish, the olfactory sacs in fish are not connected to the mouth. Therefore, the lungfish resemble terrestrial vertebrates in this regard. The olfactory sensory cells are unevenly distributed over the epithelium, with 50 000 to 100 000 cells/mm², on average (Jobling, 1995). The olfactory epithelium sends its information to the olfactory bulb, which is organized into glomeruli, clusters of neurons. The olfactory tract transmits the chemosensory information from the bulb to the brain. The olfactory tract in the Atlantic cod has four nerve bundles that specialize in conveying specific information, leading to four different behaviors: flight, courtship, snapping with open mouth, and food seeking (Jobling, 1995).

In many deep-sea fish, the olfactory capsule and olfactory lobes are much larger in males than in females. The olfactory lobe of the brain may even change in size in the same individual: ripe males of *Gonostoma bathyphilus* (Gonostomatidae) have extremely large olfactory lobes. With sex reversal, their olfactory lobes regress (Badcock, 1986).

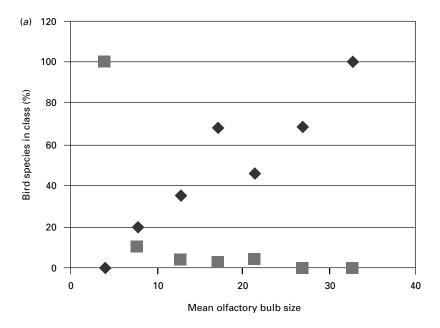
In fish, certain odorants activate specific glomeruli in the olfactory bulb. Amino acids stimulate glomeruli in the lateral region of the bulb, while glomeruli in the medial region of the bulb process information on bile acids (Yoshihara *et al.*, 2001).

Reptiles

The forked tongue of squamate reptiles, together with the VNO, has received considerable more attention than the sense of smell *per se*. Recent strong evidence suggests that the forked tongue serves in odor trail location by simultaneously comparing two odor concentrations ("directional smelling"). Ecologically, reptiles that follow prey trails have forked tongues, while ambush hunters do not (Schwenk, 1994). Comparing dinosaur skulls with living reptiles and birds strongly suggests that even in those extinct reptiles the fleshy nostril was located forward (rostral) in the enormous bony nasal aperture – contrary to the traditional assumption of a location farther back on the skull. This would have permitted the dinosaurs to pass the inhaled air over their olfactory epithelium (Witmer, 2001).

Birds

The sense of smell in many birds is better developed than previously thought. The relative size of the olfactory bulb varies from largest in procellariforms to smallest in songbirds (Bang and Cobb, 1968; Bang, 1971). Olfactory bulb size is correlated with life habits: carnivorous and piscivorous, colonial, burowing and/or sexually monomorph bird species have the largest, seedeaters the smallest (Fig. 5.3). In some species, the numbers of olfactory receptor cells and glomerular mitral cells approach those of mammals with a keen sense of smell, such as the rabbit (Wenzel, 1986). Accordingly, olfactory performance is acute in some bird species. Using Pavlovian techniques and measuring cardiac acceleration, the importance of the olfactory system in the reception of four odorants has been examined in pigeons (Henton *et al.*, 1966). The birds were most sensitive to *n*-butyl acetate (10^{-4} of vapor saturation), followed by butanol ($10^{-4.3}$), *n*-amyl acetate (10^{-4}), and benzaldehyde ($10^{-3.3}$).



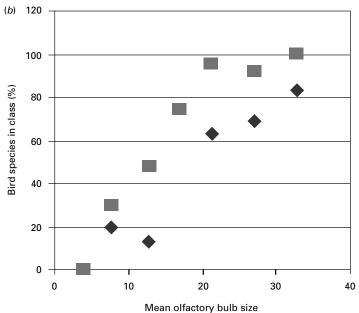


FIGURE 5.3 Food habits and relative size of olfactory bulb in birds. Species were devided into seven classes by mean olfactory bulb size calculated as the largest diameter of the bulb as a percentage of the largest diameter of the whole brain. (a) Relationship with food habits; birds with large olfactory bulbs are more likely to be carnivorous or piscivorous (), while all bird species in the smallest class are seedeaters (). (b) The larger the olfactory bulb, the more species are colonial breeders () and sexually monomorphic (). (After data in Bang and Cobb (1968) and Bang (1971.)

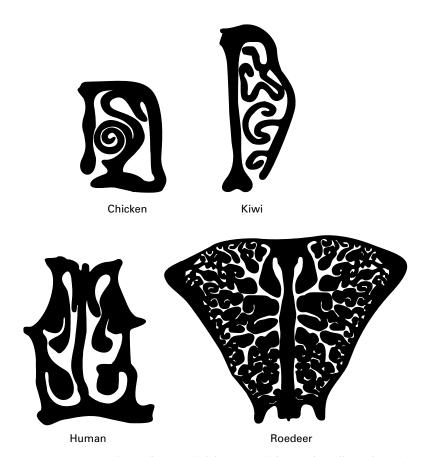


FIGURE 5.4 Different degrees of elaboration of the nasal scrolls (turbinates) in some birds and mammals. Only part of the scrolls contain olfactory epithelium. Note the difference between humans and deer. (Redrawn from various sources.)

Birds smell food not necessarily from a distance. They may do so via the choanae while the food is in the mouth. Waxwings discriminate berries treated with methyl anthranilate from controls only after picking them up with their bills (Avery *et al.*, 1992).

Mammals

In mammals, the main olfactory system is the "work horse" in the perception of odors. Excellent detailed reviews of the mammalian olfactory system are available elsewhere. In brief, the olfactory epithelium is located on a portion of the scroll bones (endoturbinales and posterior part of nasoturbinales; Fig. 5.4), in humans it is located about 1 cm beneath the bridge of the nose. Olfactory reception is affected by several factors, such as the size, shape, or wetness of the nasal passages. In the dog, the olfactory membrane extends over 75–150 cm² depending on body size, while in humans it is only 2–4 cm². It consists of three

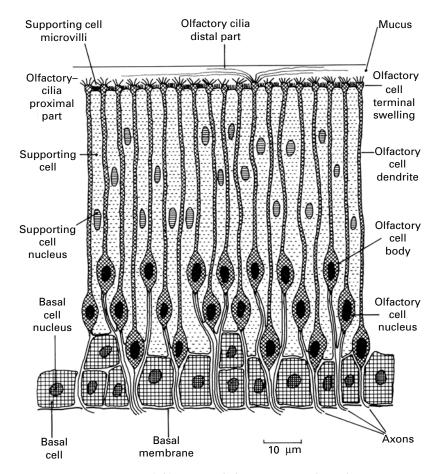


FIGURE 5.5 Structure of olfactory epithelium in mammals, a schematic view. Only one receptor cell is shown with its entire cilia floating in the olfactory mucus layer. (From Albone, 1984.)

cell types: receptor cells are bipolar and connected to the olfactory bulb; supporting cells space the dendrites out, and basal cells are precursors of both receptor and supporting cells (Fig. 5.5). There can be as many as 9.0×10^6 receptors/cm². The olfactory epithelium of the mouse contains over 2 million sensory neurons.

The dendrites on the receptor cells have swellings, the olfactory knobs. These knobs have smooth vesicles that may be open to the mucus covering the epithelium. From each knob extend 1 to 150 cilia, floating in the mucus. The precise number of cilia varies with the species. The rabbit, for example, has 10–12 cilia on each knob. The surrounding mucus consists of mucopolysaccharides, lipids, and phosphatides. The axons of the olfactory receptor cells extend through the cribriform plate and terminate in the olfactory bulb (Fig. 5.1).

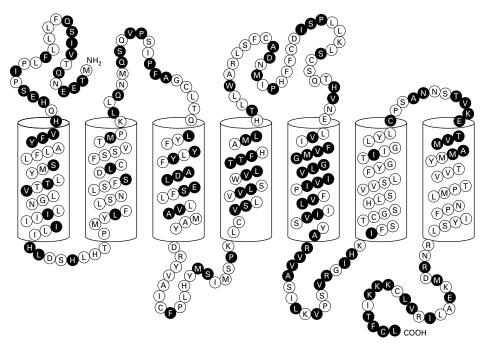
How olfactory receptors work

The olfactory epithelium contains a large number of different receptors. Buck and Axel (1991) found large families of genes for proteins that appear to be receptors linked to G-proteins and occur only in the olfactory epithelium. These are the largest gene families known to date for any animal genome. Mouse and rat have over 1000 olfactory receptor genes (OR), humans 500-750, and zebrafish and catfish around 100 (Mombaerts, 1999). The genes code for a range of receptors that recognize diverse odorants. Does olfaction use "labeled lines" or "diffuse signals" ("broad tuning")? A study of olfactory receptor neurons in vivo in anesthetized rats supports the notion of broad tuning. As many as 10 different odors can excite the same olfactory receptor neuron as measured by the electro-olfactogram; 32% responded to six different odors, and only 12% to just one single odor. The study employed odors from the terpene, camphor, aromatic and straight-chain ketone groups, previously used in frogs (Duchamp-Viret et al., 1999). The several compounds that one olfactory receptor type responds to share certain molecular features. The rat neurons responded more broadly but with less sensitivity to odors than did those in the frog. One single olfactory receptor neuron is thought to express only one or only few of the 1000 OR genes. Some consider the "one neuron, one OR" hypothesis far from proven (Mombaerts, 1999); however, recent work has shown how feedback from an expressed OR gene inhibits the activation of other OR genes, ensuring the "one receptor - one olfactory neuron rule" (Serizawa et al., 2003).

Molecular events at the olfactory receptors

The mucus of the olfactory epithelium contains odorant-binding proteins that are specific to various odorants, such as anisole, camphor, benzaldehyde (cherry-almond odor), 2-isobutyl-3-methoxypyrazine (green bell pepper odor), and 5α -androst-16-en-3-one (urine odor) (summarized by Leffingwell, 2001).

Odorants are thought to bind to integral membrane receptors on the cilia of the olfactory sensory neurons. The receptors are thought to be specific; different olfactory neuron types recognize different odorants that share certain characteristics (Buck, 1993). The odorant receptors transduce signals via interactions with G-proteins (so-called because guanosine trisphosphate is involved in their activation). These G-protein-coupled exhibit seven hydrophobic domains (Fig. 5.6). Variation in the amino acid sequence of the transmembrane domain may account for specificity and selectivity of odor reception.



cylinders reside in the cell wall. These are the seven transmembrane hydrophobic domains. Loops on top (and the N-terminus) are outside the cell, those at the bottom (and the C-terminus) are inside the cell. For clarity, the "cylinders" are spread out but, in the cell, they are bundled into a ligand-binding pocket. The residues shown as black dots are especially diverse among known odorant receptors. This diversity enables the receptor family to interact with many different odor molecules. (From Buck, 1996.)

The receptors start a second messenger cascade that is initiated by activation of G-proteins in the cell. These, in turn, interact with membrane-bound adenylyl cyclase, which catalyzes the formation of cyclic adenine monophosphate (cAMP) and opening of cAMP-gated cation channels. Depolarization then brings about an action potential, which travels along the axon of the olfactory sensory neuron. Many of the molecular components of this cascade are olfactory specific.

To be ready for the next odor stimulus, β -adrenoceptor kinase (Bark) 2 inactivates a receptor only a tenth of a second after the first stimulation occurred. This kinase phosphorylates the activated receptor, which allows another protein, B-arrestin, to bind to the receptor and inactivate it. This is a specific example of a group of enzymes that deactivate hormone or neuroreceptors (Dawson *et al.*, 1993).

The first olfactory neuron-specific protein, termed olfactory marker protein was isolated by Frank Margolis in 1972 and was cloned in the 1980s. This 19 kDa cytoplasmatic protein is unique to the mature olfactory system and is found in vertebrates from salamanders to humans (reviewed by Margolis *et al.*, 1993).

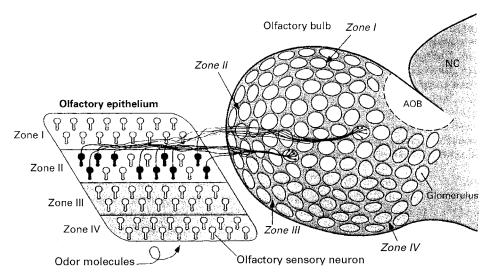
In the olfactory epithelium, different *OR* genes are expressed in different regions so olfactory neurons of one type are concentrated in a particular zone. For instance, the K4 and K7 subfamilies are always expressed together in the same (middle) region, while the K18 subfamily is always located in a different region dorsal and medial from K4 and K7 (Ressler *et al.*, 1993). This spatial segregation is preserved in the projections of the olfactory receptor neurons to the olfactory bulb (Ressler *et al.*, 1994). However, in each of the four zones, many different types of olfactory receptor neuron occur in a mosaic. Such zonal arrangements exist in mouse, rat, and zebrafish and may represent an efficient wiring system conducive to easy decoding (Mombaerts, 1999).

Processing in the olfactory bulb

The olfactory bulb contains glomeruli where the dendrites of mitral cells and tufted cells concentrate. The mouse has about 1800 glomeruli in its olfactory bulb. In the rabbit, the input from 5×10^7 receptor cells converges on 1900 glomeruli (Fig. 5.7). In some bats, 900 receptor cells converge on each secondary olfactory neuron.

Associated with the glomerulus are two additional cell types, the periglomerular and granular cells. Both establish lateral connections between tufted and mitral cells. With 1000 OR genes estimated in mouse and rat, each olfactory receptor type may correspond to two glomeruli (Mombaerts, 1999). Thus, a glomerulus is the site where the axons of olfactory sensory neurons expressing a specific OR type converge, an important stage in integrating olfactory information (Mombaerts, 1999).

With about 1000 OR genes, there are about two glomeruli for each. The connection of olfactory neurons to the olfactory bulb follows two principles: zone-to-zone projection and glomerular convergence (Mori *et al.*, 1999). An optical imaging technique, termed intrinsic signal imaging, identifies how specific glomeruli respond to particular odors in living animals. A light illuminates the glomeruli through an opening or thinned spot in the skull above the olfactory bulb. Changes in neural activity affect levels of blood oxygenation and light-scattering properties of neural membranes. (The hemoglobin of the blood absorbs light, and the membranes scatter light in varying ways.) This causes



glomeruli in the main olfactory bulb in mice. The epithelium is organized into four zones defined by expression of odorant receptors. Olfactory neurons of a particular zone project to a corresponding zone in the bulb. Axons of these olfactory neurons that express the same odorant receptor (such as those shown in black) converge to a small number of glomeruli. AOB, accessory olfactory bulbs, NC, nucleus coeruleus. (From Mori *et al.*, 1999.)

changes in the light bouncing back that can be visualized as differently colored small areas of the olfactory bulb (Rubin and Katz, 1999).

Processing at higher centers

After only one synapse – in the glomerulus – connections with the cortex are made. Approximately 24 output cells emerge from each glomerulus. We are now starting to understand how the olfactory bulb connects functionally to the cortex. Linda Buck and her colleagues have labeled receptor cells by inserting the gene for a marker protein called barley lectin next to the gene for a mouse olfactory receptor protein. The lectin marker could be demonstrated in both the neuron carrying that receptor and in connecting neurons. In this way, the investigators traced the connections from receptor neuron through the olfactory bulb to the olfactory cortex. Each of two *OR* genes thus marked resulted in one or two stained glomeruli on each side of the main olfactory bulb. The olfactory cortex showed several clusters of stained neurons. Each mouse exhibited the same pathways, showing for the first time a clear organization in the projection from the bulb to the cortex. Moreover, projections from different receptor

types converge in the cortex, permitting comparison and integration of information. While in each half of the bulb, each receptor is represented in 1 out of 1000 glomeruli (0.1% of the bulb's area), a receptor's projections cover about 5% of the area in the cortex. This suggests that a single cortical neuron receives input from as many as 50 different olfactory receptors. Leading smell researchers see this as a means for the brain to analyze and distinguish the often complex smells.

Beyond the olfactory bulbs, complex signal processing takes place. Connections between the olfactory bulbs permit summation of input from the two olfactory mucosae, thus enhancing sensitivity (Bennett, 1968). The central olfactory apparatus involves many brain areas, including the mediodorsal thalamic nucleus and the prefrontal insular cortex (cited in Ferreira *et al.*, 1987), the hippocampus, fornix, gyrus supracallosus and dentalus, septum lucidum, and uncus.

Other developmental and functional aspects

A number of pertinent observations have been made.

- Odor exposure during early ontogeny (e.g. 2-week-old rats) alters the mitral cells in specific regions of the olfactory bulb (Døving and Pinching, 1973; review by Reasner, 1987).
- Domestication can change the olfactory system. In the domestic pig, most olfactory structures in the forebrain are 30% smaller than in wild boar (Kruska and Rohrs, 1974).
- The olfactory receptors can be stimulated by bloodborne odors. Garlic odor, carried in the blood, can be perceived olfactorily (Maruniak *et al.*, 1983).
- The main olfactory system is designed with considerable redundancy. Rats that have been bulbectomized on one side still have the same absolute threshold and intensity difference threshold as intact rats (Slotnick and Schoonover, 1984).
- Centrifugal neural pathways affect the processing of meaningful biological information in the olfactory bulb; rates of habituation particularly appear to be modulated this way. (Responses to food and sex odors habituate little.) This centrifugal innervation also has long-lasting effects on olfactory learning. In short, the olfactory bulb is part of a neural network (rather than a strict one-way street for incoming stimuli) that supports learning and memory for biologically significant odors (Gervais *et al.*, 1988).

The main olfactory system mediates numerous behaviors that will be discussed in the section on signaling and priming pheromones, and also

interspecific responses. For any given mammalian species, the role of this system in particular behaviors cannot be easily predicted. For instance, in prairie voles, *Microtus ochrogaster*, females associate strongly with familiar males. This response is lost after removal of one or both olfactory bulbs. Bulbectomy reduces mating behavior but does not eliminate it, while – unlike in mice – maternal behavior is not affected (Williams *et al.*, 1992a).

5.2.2 Vomeronasal system

The vomeronasal system, also known as the accessory olfactory system, consists of chemoreceptors, organized into the VNO, the vomeronasal nerve, its terminal, the accessory olfactory bulb, and more central pathways. First described by Jacobson in 1811, the VNO has been studied intensely. We now know how stimuli reach it and what behaviors it mediates. The VNO occurs in amphibians, reptiles, and mammals. Among mammals, it is best developed in marsupials and monotremes. In birds it only appears during embryogenesis. The VNO and its function are best known for squamate reptiles, particularly snakes, and rodents and ungulates among the mammals.

Amphibia

The best developed vomeronasal system among amphibia is found in plethodontids, a family of lungless salamanders that originated in the Appalachian mountains of North America, where diverse forms still abound. This family enjoys special chemosensory adaptations and the VNO is situated anteriorly in the nasal cavity (Fig. 5.8). The genus *Plethodon* (woodland salamanders) has been well investigated. In the red-backed salamander, *Plethodon cinereus*, nasolabial grooves lead to the VNO; the behavior of nose tapping draws material up the nasolabial grooves, which probably facilitates stimulus detection (Dawley, 1987, 1992). Nose tapping is the functional equivalent of tongue flicking in snakes. Both behaviors pick up material for chemical evaluation. The red-backed salamander uses its VNO in finding prey such as *Drosophila*, although other senses are also important (Placyk and Graves, 2002).

There are sex differences: in woodland salamanders (*Plethodon*) and brook salamanders (genus *Eurycea*) of the lungless salamander family Plethodontidae, females have smaller VNOs than males (Dawley, 1992). There are also species differences: the VNO in *Eurycea wilderea* is larger than that in *Plethodon cinereus*, and the size of the hypertrophied nasal grooves, called cirri, correlate with size of VNO (Dawley, 1992).

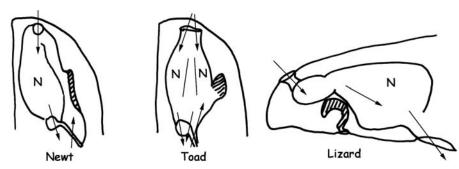


FIGURE 5.8 Location of the vomeronasal organ (VNO, cross-hatched) in amphibians and reptiles. Arrows show air entering through the external nares at top and exiting through the choanae at bottom. From there, VNO receives air with odors. N' nasal cavity. In lizard, VNO receives stimuli from mouth cavity below, with help from tongue. (Redrawn after Romer, 1959.)

In the frog *Rana temporaria*, the VNO shares the nasal compartments with the main olfactory organ. The VNO has three cavities, and water enters from the external nares via two fissures. The VNO is used to sample water while the frog is submerged, while above water air is inhaled and the olfactory system stimulated (Døving *et al.*, 1993).

Reptiles

The reptilian VNO develops from the nasal cavity but is separate from it in the adult stage. The best-known behavior involving the VNO in reptiles is tongue flicking. The forked or simple tongue samples air and is assumed to transport odorants into the openings of the VNO in the roof of the mouth, although this view has been challenged (Schwenk, 1994). The VNO processes volatiles and non-volatiles, while the nasal olfactory system is limited to volatile stimuli. Odorants can be directly moved into the VNO by pressing the back of the tongue against the vomeronasal duct or indirectly after first wiping the tongue over the sublingual plicae (folds) (Schwenk, 1994). The VNO is highly developed in snakes and lizards, though turtles possess some VNO tissue. The VNO of garter snakes projects to the accessory olfactory bulb in a zone-to-zone fashion, as in several mammalian species.

The VNO plays an important role in feeding behavior in garter snakes (Halpern and Frumin, 1979). Lizards also appear to use their VNO for food detection. Examples are the scincid lizard *Chalcides ocellatus* (Graves and Halpern, 1990) and the desert iguana *D. dorsalis* (Cooper and Alberts, 1991). The latter

Class and taxon	Context	Behavior	Reference
Amphibia			
Plethodontids (lungless salamanders): woodland salamanders (<i>Plethodon</i>)	Chemical evaluation	Nose tapping	Dawley 1987, 1992
Frog (Rana temporaria)	When submerged	Sample water	Døving et al., 1993
Reptiles			
Garter snakes (<i>Thamnophis</i> spp.),	Feeding	Response to earthworm wash	Halpern et al., 1985
Lizard Dipsosaurus dorsalis	Food finding	Detection of carrot extract	Cooper and Alberts, 1991
Red-eared turtle Pseudemys scripta	Chemoreception in water	Accessory bulb activity to salt solutions and soluble vapors	Hatanaka and Hanada, 1987

Table 5.1 Role of vomeronasal organ in behavior of amphibians and reptiles

were unable to detect carrot extract odor or cologne on cotton swabs if their vomeronasal ducts had been sealed experimentally (Cooper and Alberts, 1991).

In garter snakes (*Thamnophis sirtalis*) a VNO-mediated stimulus can reinforce behavior: dried earthworm wash or earthworm bits can be used to reward correct performance in a conditioned response to an arbitrary stimulus, such as dots versus stripes in a Y-maze (Halpern *et al.*, 1985). In red-eared turtles the VNO "is considered to involve aqueous chemoreception in water." Salt solutions and soluble vapor substances generated activity in the accessory olfactory bulb (Hatanaka and Hanada, 1987).

Table 5.1 lists some behavioral roles of the VNO in amphibians and reptiles.

Mammals

Structure of the vomeronasal organ

The VNO of mammals consists of a pair of parallel tubes located above the palate on either side of the nasal septum (Fig. 5.9). The organ communicates with the outside by the nasopalatine duct, which, depending on the taxon, may open (a) to the mouth cavity via the incisive duct and incisive papilla (Fig. 5.10); (b) to the nasal cavity, as in alcelaphine antelopes, such as Topi, *Damaliscus korrigum*, and Coke's hartebeest, *Alcelaphus cokii* (Hart *et al.*, 1988), and the horse (here the VNO does not "get it from the horse's mouth, but rather is paid through the nose"); or (c) both (e.g. the ring-tailed lemur, *Lemur catta*; Bailey,

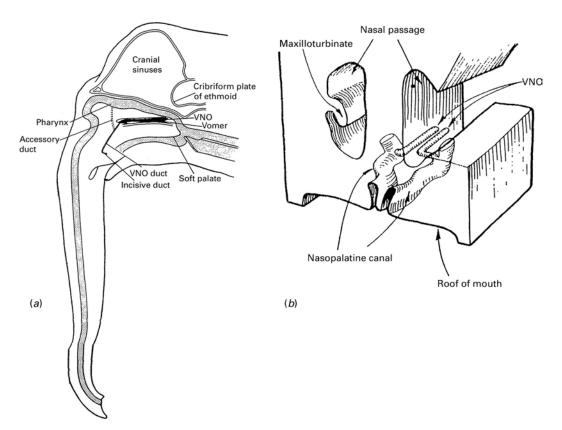


FIGURE 5.9 Mammalian vomeronasal organ (VNO). (a) Location of the VNO in a female Asian elephant, *Elephas maximus*. (b) Location of the VNO in the European hedgehog, *Erinaceus europaeus*. ([b] From Rasmussen and Hultgren, 1990; [a] from Stoddart, 1983, after Poduschka and Firbas 1968).

1978). There may be intermediate stages: the common wildebeest (*Connochaetes taurinus*) lacks incisive papillae but has small incisive ducts.

Blood vessels, cavernous tissue, and muscles surround the VNO, and all in turn are enclosed by the vomer bone capsule. This arrangement permits the lumen of the VNO to be compressed by swelling of surrounding tissue and opened by deflation, permitting stimuli to move out or in. This mechanism is known as the vomeronasal pump (Meredith and O'Connell, 1979).

Receptor cells

The VNO is lined with bipolar receptor cells (Fig. 5.11). The receptor cells possess microvilli, in contrast to the cilia on the receptor cells of the main olfactory epithelium. The VNO of male and female mice have sensory cells with receptors that respond to male urine, and others that respond to female



FIGURE 5.10 Openings of incisive ducts (leading to the vomeronasal organ) in the roof of the mouth of a male black-tailed deer, *Odocoileus hemionus columbianus*. The two openings are the small pores in the left and right corner of the rhomboid groove close to the upper lip (indicated by thin lines). (Photograph: D. Müller-Schwarze.)

urine (Holy *et al.*, 2000). The VNO can be very sensitive. VNO neurons, exposed to six pheromonal compounds from mice, responded to concentrations near 10^{-10} mol/l (Leinders-Zufall *et al.*, 2000).

$Me chanisms\ of\ stimulus\ access$

Stimuli gain access to the organ via the incisive foramen in the roof of the mouth or the nasal cavity. The vomeronasal pump, described for the hamster by Meredith and O'Connell (1979), aids in moving molecules into the organ. This way the animal can control stimulus access and its intensity. Experiments

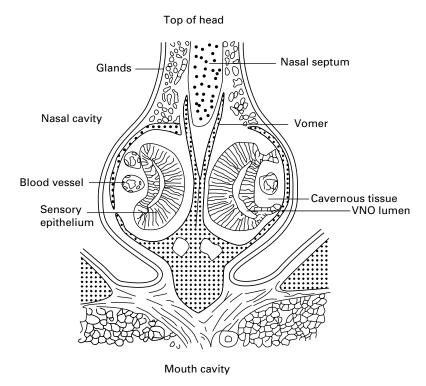


FIGURE 5.11 Structure and location of the vomeronasal organ in the mouse, seen in a transverse cross-section through the middle of the vomeronasal organ. (Redrawn after Døving and Tritier, 1998.)

with dyed materials have shown that non-volatiles reach the VNO in a variety of behavior contexts. For instance, the VNO of male pine voles (*Microtus pinetorum*) and meadow voles (*Microtus pennsylvanicus*) takes up female urine during investigation, and also materials from the fur during grooming, and even dyes applied to food (Wysocki *et al.*, 1980). The VNO plays an important role in mediating reproductive behavior.

Even in primates, the vomeronasal system can be functional. If the *liquid* phase of urine from several species of the same genus is micropipetted into the oral opening of the nasopalatine duct of mouse lemurs (*Microcebus murinus*, Prosimia), they respond with sniffing, licking and "VNO testing." By contrast, urine *volatiles* stimulate only the main olfactory system in this species (Schilling *et al.*, 1989).

Flehmen

Flehmen (or "lipcurl"; Fig. 5.12), a behavior typical for males when examining female urine, occurs in several mammalian groups. After the buck



FIGURE 5.12 Flehmen in a male pronghorn, *Antilocapra americana*. (Photograph: D. Müller-Schwarze.)

has brought urine to his upper palate by licking or mouthing movements, he raises his head, curls up the upper lip, and breathes deeply and rhythmically. Various species differ in details of flehmen. Estes (1972) postulated that the VNO receives material during flehmen. Studies of male goats with dyed urine showed that indeed urine enters the VNO during flehmen (Hart, 1983). In several better-investigated species such as black-tailed deer, *Odocoileus hemionus columbianus*, (Müller-Schwarze, 1979) or domestic sheep (Bland and Jubilan, 1987) Flehmen occurs *least* often during estrus. This indicates the analytical role of flehmen but does not exlude a stimulatory function. Wildebeest males do flehmen to female urine, but lick their nostrils during flehmen. This licking may deliver stimulus material to the VNO via the nasal cavity, perhaps to compensate for

the reduced oral access to the VNO in this group of mammals (Hart et al., 1988).

Central projections from the vomeronasal organ

The VNO axons form the vomeronasal nerve, which connects with the accessory olfactory bulb. In contrast to the main olfactory system, axons from neurons expressing a particular vomeronasal receptor project to glomeruli of the accessory olfactory bulb (Mombaerts, 1999). The apical VNO region, with mostly type 1 (V_1) receptors, projects to the anterior (rostral) part of the accessory olfactory bulb, while the basal region (with V₂ receptors) projects to the posterior part. The representation of sensory neurons of the VNO in the glomeruli of the accessory bulb differs from that of the main olfactory system: to make up for the smaller number of receptor types, the VNO achieves odor decoding by a more complicated spatial representation. "Neurons expressing the same receptor gene project to many different glomeruli, while a single glomerulus may receive input from more than one receptor type" (Keverne, 1999). Beyond the accessory bulb, more complexity occurs: one mitral cell can gather information from more than one glomerulus. It is thought that these arrangements ultimately serve as a "difference detector" (Keverne, 1999), most other analysis being done by the main olfactory system.

From the accessory bulb, projections lead to the medial nuclei and posteriomedial portion of the cortical nuclei of the amygdala. The amygdala in turn is connected to higher centers via the stria terminalis, and hypothalamic structures. Thus, the accessory olfactory system represents a neural pathway separate from the main olfactory system. Both project into absolutely different parts of the amygdaloids (Powers and Winans, 1975; Powers *et al.*, 1979).

Functional magnetic resonance imaging permits the central processing of urinary pheromones to be followed. The VNO and accessory olfactory bulb are involved in processing pheromonal stimuli. In an Australian marsupial, the brown antechinus (*Antechinus stuartii*), some projections from the accessory olfactory bulb respond indiscriminately to male and female urine odors while others are activated only by male urine cues. These findings show that urinary pheromones may act on the hypothalamic–pituitary–adrenocortical axis via the paraventricular nucleus of the hypothalamus (Toftegaard *et al.*, 2002).

$Role\ of\ the\ vomeron as al\ organ\ in\ behavior$

In mice, extirpation of the VNO has many behavioral effects. It reduces aggression in male mice (Bean, 1982; Wysocki *et al.*, 1986), reduces urine marking and aggressive behavior in sexually naive male mice (Maruniak *et al.*, 1986); lowers marking to some extent also in sexually experienced males (Labov and

Table 5.2 Chemosensory systems required for behaviors in golden hamsters

Behavior	Chemosensory system required
Species recognition	
Responses to vaginal secretion marks	Main olfactory system
Responses to flank gland secretion	Main olfactory system
Reproduction	
Androgen responses of male to vaginal secretion	Vomeronasal organ
Ultrasonic calling	Vomeronasal organ and main olfactory system

After Johnston, 1992.

Wysocki, 1989); reduces ultrasonic vocalizations by male mice when a female is present (Wysocki *et al.*, 1982); and abolishes maternal aggression in primiparous female mice (Lepri *et al.*, 1985). Deafferentiation of the VNO in male mice impairs sexual behavior. The effect is greatest if the male was deafferentiated before experience with any adult female (Clancy *et al.*, 1984). Aggressive behavior in male mice is less reduced after deafferentiation of the VNO if they had had fighting experience (Daranzo *et al.*, 1983).

Wysocki *et al.* (1986) have reviewed the relationships between learning and the VNO. The VNO plays a role in responses to infant conspecifics. Intact virgin male rats kill newborn pups. If their VNO is removed surgically, the rate of this infanticide decreases (Mennela and Moltz, 1988). A virgin female rat will accept foster pups if her VNO has been deafferentiated (Fleming *et al.*, 1979). The same is true for male rats. They will show maternal behavior toward pups after removal of the VNO (Saito, 1986a). Lactating rats are impaired in their retrieving of pups if their VNO has been extirpated (Saito, 1986b).

In the golden hamster, the VNO is essential for certain reproductive behaviors, while the main olfactory system mediates responses that involve species recognition (Johnston, 1992; Table 5.2). As in mice, removal of the VNO impairs sexual behavior in male golden hamsters, but only if carried out before the animal had had sexual experience (Meredith, 1986). The same is true for ultrasonic vocalizations.

Virgin male prairie voles (*M. ochrogaster*) are less likely to sire offspring if their VNO is removed (Wekesa and Lepri, 1992). In the guinea pig, stimulation of the VNO is assumed to be "inherently reinforcing." Male guinea pigs cease to head-bob to female urine after their VNO is surgically removed (Beauchamp *et al.*, 1985).

The role of the vomeronasal organ in priming

The VNO is extremely important in mediating endocrine responses to primer pheromones. Puberty acceleration in female rats by male urine odors can be prevented by electrolytic damage to the vomeronasal nerve. Also, effects of male urine odor such as shortening of the estrus cycle (see Ch. 8) can be eliminated by section of the vomeronasal nerve, or bilateral electrocoagulation of the accessory olfactory bulb (Sánchez-Criado, 1982). In rats, the odor of males stimulates ovulation in females, an effect that is lost if the VNO is extirpated (Johns *et al.*, 1978). Female prairie voles, *M. ochrogaster*, respond to odors from males with reproductive activation. Surgical removal of the VNO from adult females impedes this reproductive activation by the stud male. The weights of the uterus and the ovaries of these females were lower than those of normal or sham-operated individuals. However, the females without a VNO were still able to locate food by chemical cues (Lepri and Wysocki, 1987).

At the endocrinological level, the VNO mediates the surge of luteinizing hormone and testosterone in males after exposure to females. This surge does not occur if a male with deafferentiated VNO is exposed to an anesthetized female or her urine (Wysocki *et al.*, 1983). But VNO-deafferentiated males will show a surge in luteinizing hormone in response to *awake* females (Coquelin *et al.*, 1984). In female mice, stimulation of the VNO by male urinary cues activates the limbic system (discussed in Ch. 8). The roles of the VNO in the behavior of some rodents are listed in Table 5.3.

Genetic basis of function in vomeronasal organs

Some sensory neurons of the VNO express two gene superfamilies, termed *V1r* and *V2r*, that encode over 240 proteins of the seven-transmembrane type (Matsunami and Buck, 1997). These G-protein-linked putative pheromone receptors are distantly related to the main olfactory system's receptors. Receptors of the VNO are linked to different G-proteins, and their extracellular N-terminal domains are longer than those of the receptors in the main olfactory system. (V₁ receptors are linked to G_i-proteins and V₂ receptors to G_o-proteins). The intracellular excitation mechanism in VNO sensory neurons also differs from that in the main olfactory systems; instead of linking to adenylyl cyclase, the VNO receptors activate the phosphoinositol second messenger system. This has been demonstrated in several mammalian species. In hamsters, aphrodisin increases inositol 1,4,5-trisphosphate (IP₃) levels in VNO membranes. Boar seminal fluid and urine stimulate increases of IP₃ in the VNO of the female pig. (However, in the pig, the VNO is not necessarily essential for responses to pheromones [Dorries *et al.*, 1997]).

T 11 = a	n 1 C.1	1 • 1	. 1 1 .
Table 5.3	Role of the vomeronasa	i organ in roder	it behavior
10010000	TOTO OF CITE TOTAL OFFI	1 015411 111 10401	it believiel

Species	Sex	Context	Behavior	Reference
Mouse Mus domesticus	M	Aggression	Aggressive behavior; urine marking	Bean, 1982; Wysocki <i>et al.</i> , 1986;
	M	Sexual behavior	Ultrasonic sounds	Wysocki et al., 1982
	F	Maternal behavior	Maternal aggression	Lepri <i>et al.</i> , 1985
Rat Rattus norvegicus	M	Infanticide	Kills newborn pups	Mennela and Moltz, 1988
	F	Maternal behavior	Neophobia to pups by virgin F	Fleming et al., 1979
	F	Maternal behavior	Retrieving pups	Saito, 1986b
Prairie vole Microtus ochrogaster	M	Reproduction	Siring offspring	Wekesa and Lepri, 1992
Golden hamster Mesocricetus auratus	M	Mating	Vaginal secretion elicits mounting	Clancy et al., 1984
	M	Sexually native male	Sexual behavior; ultrasonic sounds	Meredith, 1986
Guinea pig Cavia porcellus	M	Female urine	Head bobbing	Beauchamp et al., 1985
Meadow vole Microtus pennsylvanicus	M, F	Feeding	Food dye found in VNO	Wysocki et al., 1985
Pine vole Microtus pinetorum	M, F	Feeding	Food dye found in VNO	Wysocki <i>et al.</i> , 1985

M, male; F, Female; VNO, vomeronasal organ.

In rats, both male and female urine activate both subtypes of G-protein. (Lipophilic volatile odorants act on G_i , and a urinary lipocalin protein on G_0). There are specific receptors for urinary lipocalins in the VNO (Krieger *et al.*, 1999). In summary, the two subtypes of receptor may be activated by distinct ligands; V_2 receptors are particularly receptive to non-volatile proteins (summarized in Keverne, 1999).

Deleting a cluster of 16 *V1r* genes (2 of the 12 *V1r* gene families; representing about 12% of the *V1r* repertoire) impairs sexual and maternal behavior. Gene-deficient male mice initiate fewer sexual encounters, and lactating females attack intruders less. Furthermore, both mutant male and female mice showed "specific anosmia" in that VNO did not respond to three of eight tested pheromonal compounds (6-hydroxy-6-methyl-3-heptanone, *n*-pentyl acetate, and isobutylamine) (Del Punta *et al.*, 2002).

Peptides that serve as ligands for major histocompatibility complex (MHC) class I molecules can activate vomeronasal sensory neurons. These peptides contain nine amino acid residues and activate sensory neurons from the V_2 receptor

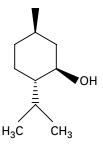


FIGURE 5.13 Structure of menthol, a trigeminal irritant.

family, which are located in the basal (deep) layer of the vomeronasal epithelium. This finding establishes a connection between the MHC molecules that signal individuality (genotypic diversity) and receptor mechanisms that read information about individuality (individual recognition), thus influencing social behavior (Leinders-Zufall *et al.*, 2004).

Why do animals need a VNO? Meredith (1983) suggested three possible reasons: (a) VNO receptors may be more sensitive to certain chemicals, but also more vulnerable to damage; (b) reception of certain chemicals may require special glandular secretions over the receptors, which is only possible in the VNO lumen; and (c) the information reaching the brain via the VNO may be so potent in triggering behavioral or physiological processes that inadvertent triggering should be avoided.

5.2.3 Trigeminal nerve

The trigeminal nerve, or fifth cranial nerve, innervates the face and eyes. It responds not only to touch, painful stimuli, and temperature but also to irritating and noxious chemical stimuli. Pungent, tickling, tingling, stinging, burning, cool, warm, and painful sensations such as those of ammonia, menthol, peppermint, eucalyptol, or carbon dioxide are mediated by the trigeminal nerve. Menthol (Fig. 5.13) is perceived in the nasal cavity as cold at lower concentrations and hot at high concentrations. Mustard oil (allyl isothiocyanate), onion (containing diallyl sulfide), hot Chile powder (capsaicin; Fig. 13.1), and mace spray (capsaicin) also stimulate the trigeminal receptors. In humans, the trigeminal nerves plays an important part in chemoreception.

In birds, the trigeminal nerve appears to play a role in food selection. Starlings more easily accepted commercial feed treated with otherwise avoided coniferyl benzoate after bilateral section of the ophthalmic branch of the trigeminal nerve. Therefore, the trigeminal nerve may help to protect the animal by detecting plant defense compounds. Many of these compounds are astringent or irritating (Jakubas and Mason, 1991).

Blocking trigeminal input by subcutaneous injections of capsaicin does not critically affect perception of olfactory or taste stimuli in starlings (Mason *et al.*, 1987a). Yet, the trigeminal nerve may interact with the olfactory system in perceiving stimuli: patients with unilateral destruction of the trigeminal nerve report a lower intensity of the odors of propanol and butanol for the impaired nostril than for the intact one (Cain, 1978). Trigeminal signals may add a painful or irritating note to food. At low levels, substances stimulating the trigeminal nerve may actually enhance the flavor of foods, as do carbon dioxide in soft drinks, capsaicin in hot peppers and sauces, acetic acid in vinegar, and allylisothiocyanate in mustard (Rozin and Schiller, 1980).

5.2.4 Septal organ of Masera

The septal organ is a small patch of sensory epithelium on the wall of the septum, in the anterior part of the nasal cavity, and ventral to the olfactory epithelium. It is found primarily in rodents, has chemical receptors similar to olfactory receptors, and is sensitive to volatile odorants. It projects into the main olfactory bulb, but not into the accessory olfactory bulb (Pedersen and Benson, 1986). Because of its "forward" location, the septal organ may serve as an "early-warning system" that arouses resting or sleeping animals when volatiles are present (Wysocki, 1989).

5.2.5 Nervus terminalis

Lesioning of the terminal nerve system in hamsters disrupts mating behavior (Demski and Northcutt, 1983; Wirsig and Leonard, 1985). The terminal nerve is probably the sole source of all brain neurons containing gonadotropin-releasing hormone (Schwanzel-Fukuda and Pfaff, 1989).

5.2.6 Taste

In fish, both taste and olfactory stimuli are waterborne. However, taste involves the seventh, ninth or tenth cranial nerves, in contrast to the first cranial nerve for smell. Elasmobranchs have their taste buds in the mouth and pharynx, but in bony fish they occur around the gills, on barbels and pectoral fins, and also scattered over the rest of the body surface. They crowd particularly in the roof of the mouth, forming the palatal organ. The taste receptor cells are arranged as a bundle to form a taste bud. Like other vertebrates, fish have receptors for sweet, sour, salty, and bitter. For instance, goldfish reject quinine-treated food pellets (Jobling, 1995). Many fish species are particularly sensitive to acidic taste characteristics. The responses of fish to amino acids will be discussed in Chapter 12.

In addition to taste buds, many fish have solitary chemosensory cells, which resemble taste sensory cells and are distributed over the whole surface of the fish body. Some play a role in food detection, while in other species they appear to help in detecting predator chemicals (Jobling, 1995).

Among the rest of the vertebrates, many social, sexual, or feeding stimuli are non-volatile. Contact with the material is necessary, and animals typically sample by licking. Licking often is one step in a sequence of responses, as in male ungulates' sampling of female urine. Here the animal first smells, then approaches and licks the urine before it exhibits flehmen, thought to serve in moving an odor sample to stimulate yet another chemical sense organ, the VNO. Some authors assume that taste contributes little to the perception of pheromones (Wysocki, 1989).

Structure

Taste receptors on the mammalian tongue occur in three different papillal types: fungiform, foliate, and circumvallate. The pinkish fungiform papillae are located around the edge of the tongue and can be visualized with milk or food color. There are approximately 12 circumvallate papillae arranged in a V shape at the back of the tongue. Finally, the foliate papillae lie in small grooves on the side of the rear of the tongue. One papilla will contain as few as two to five taste buds or as many as 200. Each onion-shaped taste bud in turn contains 50-100 receptor cells. Taste receptors have microvilli, like the vomeronasal receptors. The five types of cell mediate sweet, sour, salty, bitter, and umami (meaty, savory) sensations. (Umami is the taste of glutamate, one of the 20 amino acids making up proteins in meat, fish and legumes. Monosodium glutamate is a food additive.) Each taste bud has an opening, the taste pore, through which the microvilli of the taste cells reach out into the coat of saliva. Food chemicals, called tastants, are dissolved in saliva. The often-published "map" of the tongue with a neat geography of receptors segregated by taste quality is considered outdated. Taste buds in all areas of the tongue can respond to all taste qualities (Smith and Margolskee, 2001).

Function

Molecular events at taste receptors

After the saliva has carried the tastants into the taste bud, they interact with the taste receptors on the surface of the cells, or with ion channels, which are pore-like proteins. Salty and sour tastants act through ion channels, and sweet and bitter sensations are mediated by surface receptors. The different taste submodalities rely on specific mechanisms: Na⁺ flux through Na⁺

channels accounts for salty taste; sour taste results from H⁺ blockade of K⁺ or Na⁺ channels; and bitter and sweet rely on G-protein mechanisms, involving the G-protein subunit gustducin, which resembles transducin, known to transduce light signals into electrical impulses in the retina (McLaughlin *et al.*, 1993). Genetically engineered mice with defunct gustducin do not distinguish between sweet and bitter (Wong and Gannon, 1996).

The sweet, sour, salty, and bitter primary tastes are thought to serve nutrition, pH levels, ion balance, and protection from toxins, respectively. Bitter receptors deserve a closer look. Far from mediating only a general "bitter" sensation, taste cells on the rat's tongue can distinguish different bitter compounds. The family of bitter receptors consists of 50 to 100 related proteins. Individual taste bud cells express the genes for most of these receptors. This, at first, suggested that a cell would fire when stimulated by any of a great number of bitter compounds. However, 65% of the cells responded to only one of five different bitter compounds tested. About 25% responded to two compounds, and only 7% to three or more. (The cells' activity was monitored visually by fluorescence, which detected the release of Ca²⁺ inside the receptor cell [Caicedo and Roper, 2001]). In short, different taste cells appear to be tuned to different bitter compounds. They may be specialist cells rather than generalists. This specificity, though, seems to be at odds with the fact that one cell has so many different receptor types. As soon as many more than the five bitter compounds have been tested, we will know whether the taste cells are more generalist than appears now.

Central processing of responses

Researchers have oscillated between emphasizing specificity of neurons ("labeled lines") and responses to a spectrum of tastants by one cell. More recently, patterns of activation of a number of sensory cells are favored for coding specific taste sensations (Smith and Margolskee, 2001). Neural distinction of different tastes requires simultaneous activation of different cell types. The brain receives a single channel of information, simply "bitter" for a number of different compounds.

5.3 Structure-activity relationships

5.3.1 Receptor cells: generalists and specialists

Fish have receptor cells specific for certain compounds. Examples are the reception of the fish toxins tetrodotoxin (TTX) and saxitoxin (STX) by rainbow trout, *Salmo gairdneri*, and Arctic charr, *Salvelinus alpinus*. Both toxins are extremely potent taste stimuli. Not only are the receptors extremely sensitive

to these compounds (Section 5.5), but specialized receptor cells appear to exist. Cross-adaptation experiments showed that the receptor(s) for TTX are probably distinct from those that detect amino acids and bile salts. Further, TTX and STX do not share the same receptor populations. The extreme sensitivity of these fish probably protects them from ill effects by poisonous prey (Yamamori *et al.*, 1988).

Since prostaglandins serve as fish pheromones, specific receptors for these compounds have been searched for. Of 12 prostaglandins tested with Arctic charr, six gave clear electro-olfactogram responses at a concentration of under 10^{-8} mol/l. The threshold for the two most potent compounds, prostaglandin $F_{2\alpha}$ and its synthetic analogue dimethyl prostaglandin $F_{2\alpha}$, was 10^{-11} mol/l. The concentration–response curve on a semilog graph gave a typical sigmoidal shape, saturating at 5×10^{-8} mol/l. This suggests a single receptor type. The receptors are highly stereospecific. If the stereochemistry of the chemical bond of the hydroxyl group at C-9, C-11, or C-15 is changed, the receptor affinity for the compound is reduced by more than two log units. High species specificity was observed: lake char (*Salvelinus namaycush*) are also sensitive and show specificity, but none of the prostaglandins tested were active in rainbow trout (*O. salvelinus mykiss*) and brook char (*S. fontinalis*) at the concentration of 10^{-8} mol/l (Sveinsson and Hara, 1990).

5.3.2 Stimulus generalization and generalist receptors

How specific are the responses to certain compounds? Will slight changes in the size, shape, or functional groups of the molecule render it unrecognizable for a certain response?

The methods to investigate the specificity of behaviorally active compounds include *spontaneous responses* of untrained animals and *discrimination tests*, where a discriminated stimulus is substituted by another closely related compound to detect the degree of *generalization* from one stimulus to another. In field studies, the first is the method of choice.

An example for stimulus generalization are responses of rats to stress-inducing odors. Laboratory rats of the Wistar strain respond to predator odors, specifically mercapto compounds in fox droppings, with stress reactions, for example avoidance behavior such as "freezing" and increased plasma cortico-sterone concentrations (Vernet-Maury *et al.*, 1984). The rats were trained to avoid water scented with a mercapto odorant that contained both a keto- and a sulfhydryl group (4-mercapto-4-methyl-2-pentanone). As the animals licked a waterspout, a mild electric shock was applied to their tongue. When different compounds were tested thereafter, the rats avoided compounds with similar

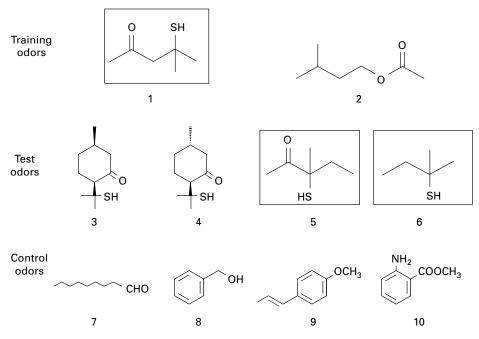


FIGURE 5.14 Stimulus generalization. Rats that experienced electric shocks when drinking water laced with compound 1, later also avoided compounds 5 and 6, which are structurally similar. The keto group was not necessary for the effect. See text. (After Fombon and Polak 1987.)

structure containing the sulfhydryl group, but the keto group was not essential for the effect (Fombon and Polak, 1987; Fig. 5.14).

Other examples of stimulus generalization are the alarm chemicals in salamanders (Mason and Stevens, 1981a) and behaviorally irrelevant pure odors (Braun and Marcus, 1969).

5.4 Neural pathways and decoding

The long-held dichotomy of "macrosmatic" and "microsmatic" vertebrates is no longer tenable. Neuroanatomists had assumed that taxa with relatively few olfactory receptor cells and small olfactory bulbs would also be inferior in olfactory performance (threshold and number of compounds detected) to those with more receptors and larger bulbs. However, we now know from single-cell recordings that a particular receptor cell type can respond to a wide range of odor compounds that share certain features. Keverne (1983) pointed out that the olfactory bulbs act as a filter, while more complex pattern analysis takes place in the neocortex. The more evolved the neocortex, as in primates, the

greater the potential for decoding olfactory messages. We now know that birds, cetaceans, and primates, formerly considered "microsmates," command impressive chemoreception.

Many vertebrate odors consist of several to many components. How mammals in particular process and use information coded in such mixtures of odorants is still not clear. One experiment posed the question "do rats discriminate complex odors by recognizing unshared components, or do they treat such odors as gestalten, i.e. in a 'unitary fashion'?" Rats were trained to distinguish threecomponent odors (ABC versus ABD) by rewarding them at one of the odors with water. They did not recognize compounds C or D when each was presented alone. Also, the rats treated the mixtures AC or AD (two-thirds of original mixtures) as novel odors, with no evidence that they remembered these from the ABC and ABD combinations. This indicates that the animals treated complex odors as units, or "gestalts." However, inclusion of a previously learned one-compound odor into a mixture changed learning of that odor: In a different experiment, rats were preexposed on day 1 to compounds X and Y. X was rewarded (X+), Y was not (Y-). The following day, this group and a not preexposed group were tested with mixtures ABC+ (rewarded) and ABX- (not rewarded). The preexposed animals were slightly slower in learning this reversal from the originally rewarded compound X to one to be avoided (as part of a mixture). This shows that the single compound was remembered in some form (Staubli et al., 1987).

5.5 Odor detection thresholds

The odor sensitivity of an experimental animal needs to be known to appreciate communication distances and performance, but it is not easily measured. The *detection threshold* has to be distinguished from the *recognition threshold*. The former is the concentration at which an odor is noticed, and the latter – typically much higher – the concentration at which behavior tests in animals or verbal responses in humans show that the odor has been recognized as a specific signal, compound, or mixture.

5.5.1 Fish

Eels, Anguilla anguilla, are extremely sensitive to odors. Their detection threshold for β -phenylethyl alcohol lies at the unbelievable concentration of 3×10^{-18} mol/l. This corresponds to 1 ml of this alcohol diluted in 58 times the volume of Lake Constance (80km long) in southern Germany where these experiments were performed. It has been calculated that only three molecules are in the olfactory sac at any one time (Teichmann, 1957, 1959). Coho salmon

detect morpholine in water at a concentration of 1×10^{-7} mg/l (Hassler and Kucas, 1988).

Rainbow trout (*S. gairdneri*) and Arctic char (*S. alpinus*) are extremely sensitive to TTX and STX. Electrophysiological recordings from the palatine (seventh cranial) nerve showed that the rainbow trout's threshold concentration of TTX is 2×10^{-7} mol/l. At 10^{-5} mol/l the response is still four times as strong as that to 10^{-3} mol/l of L-proline, the most potent amino acid for this species. The threshold for STX is lower (10^{-8} mol/l), but the response reached its maximum at 10^{-6} mol/l. Arctic char also had a lower threshold for TTX (10^{-8} mol/l) than STX (10^{-7} mol/l). The response magnitude never exceeded that of 10^{-3} mol/l for L-proline. Such a highly sensitive gustatory receptor system is probably effective in protecting predatory fish from the effects of toxins of their prey (Yamamori *et al.*, 1988).

Young (2–3 months old, 80–120 mm long) Russian, Siberian, and stellate sturgeons (*Accipenser gueldenstaedtii*, *A. baeri*, and *A. stellatus*) had a threshold of 0.001 mmol/l for 19 amino acids (Kasumyan, 1994).

Thresholds of the olfactory receptors of male goldfish are 35 pg/ml water for the prostaglandin $F_{2\alpha}$, and 100 times less for its 15-keto-derivative. The males' receptor threshold for 17,20-progesterone (from females) is "a tiny fraction of 1 pg/ml water." Three grams (one teaspoonful) would provide an above-threshold stimulus when diluted in $500 \times 500 \times 500$ m water (Bjerselius and Olsen, 1993). In lampreys, testosterone from males attracts females at a concentration of 29 pg/ml water but urine with a testosterone concentration of 29×10^{-4} pg/ml is active! (Adams *et al.*, 1987).

The threshold for amino acids can vary with the hunger level. Hungry sablefish, *Anoplopoma fimbria*, have a lower threshold $(1.4 \times 10^{-11} \,\text{mol/l})$ than satiated fish $(4.4 \times 10^{-8} \,\text{mol/l})$ (Løkkeborg *et al.*, 1995).

5.5.2 Amphibians and reptiles

Behavioral thresholds for n-butyl acetate and n-butyl alcohol in the tiger salamander ($Ambystoma\ tigrinum$) were 8.9×10^{-5} and 6.7×10^{-5} mol/l, respectively (Mason and Stevens, 1981b). The threshold for amyl acetate has been measured as 10^{-4} mol/l in frogs and 10^{-7} mol/l in turtles (Enomoto $et\ al.$, 1992).

5.5.3 Birds

Since we now know that birds rely on odors in a variety of contexts, we are interested in how sensitive they are to odorants and how their olfactory performance compares with mammals. Birds' sensitivity has been measured in different ways: behavioral thresholds differ from electrophysiological

thresholds and can be either spontaneous or experimentally conditioned. The latter pushes an animal more to its limits.

Foraging turkey vultures, Cathartes aura, find carcasses by smell. Behaviorally measured, turkey vultures are about as sensitive to ecologically relevant odorants of carcasses as pigeons are to heptane, hexane, pentane, and amyl acetate, commonly used odorants (Smith and Paselk, 1986). (According to Henton et al., [1966] the behavioral thresholds of pigeons ranged from 0.1 to 39.7 ppm). The carrion odorants tested were butanoic acid (BA), ethanethiol (ET), and trimethylamine (TMA): BA has the odor of rancid fat and is a byproduct of decomposition of fat, carbohydrates, and protein; thiols, including ethanethiol, are formed from the breakdown of sulfur-containing amino acids (they are also used to give natural gas an odor); and TMA has a fishy odor and is released during decomposition of muscle tissue. Both BA and ET elicited olfactory responses in turkey vultures (Stager, 1964). The thresholds measured by Smith and Paselk (1986) were 1×10^{-6} mol/l for BA and ET, and 1×10^{-5} mol/l for TMA. However, a turkey vulture should smell it only from 0.17 m above a carcass, as estimates of concentrations near carrion have shown. The threshold needed to detect a carcass from 3 to 60 m altitude would lie between 1×10^{-10} and 1×10^{-12} mol/l orders of magnitude lower than actually observed in vultures. The detection rate would be 1 to 20 moles per day. For comparison, humans detect BA and ET at a concentration of $1 \times 10^{-11} \, \text{mol/l}$.

Brown-headed cowbirds (*Molothrus ater*) can discriminate ethyl butyrate and s-limonene, as evidenced by cardiac conditioning. Furthermore, they discriminate among concentrations of ethyl butyrate. To discriminate the two odorants, vapor saturation must be at least 0.6%. This suggests that for ethyl butyrate the discrimination sensitivity is at least 1.9×10^{13} molecules/ml, or 0.76 ppm (Clark and Mason, 1989).

Electrophysiologically measured thresholds for butanoic acid and ethanethiol in Manx shearwater (*Puffinus puffinus*) and black-footed albatross (*Diomedea nigripes*) are as low as 0.01 ppm (Wenzel and Sieck, 1966). More electrophysiological thresholds for some compounds in tree swallows and cedar waxwings (Clark, 1991), starlings (Clark and Smeraski, 1990), and brown-headed cowbirds (Clark and Mason, 1989) are listed in Table 5.4.

5.5.4 Mammals

The most celebrated mammalian olfactory detector is the dog's nose. As a predator, the dog locates its prey by "air scenting" (following a gradient of airborne odors) and tracking. Practitioners are familiar with the extreme olfactory sensitivity of the dog. The detection threshold for butyric acid has been determined as 9×10^3 molecules/cm³ air (Neuhaus, 1953). Considering that

Table 5.4 Thresholds for odorants in birds, obtained by behavioral, cardiac, and electrophysiological techniques

Species	Compound	Threshold (ppm)	Reference
Manx shearwater Puffinus puffinus	Amyl acetate, trimethylpentane	0.01	Wenzel and Sieck, 1966
Black-footed albatross Diomedea nigripes	Amyl acetate, trimethylpentane	0.01	Wenzel and Sieck, 1966
Turkey vulture Cathartes aura	Butanoic acid Ethanethiol Trimethylamine	$\begin{aligned} &1.0 \left(1.0 \ 1 \times 10^{-6} \text{mol/l}\right) \\ &1.0 \left(1.0 \ 1 \times 10^{-6} \text{mol/l}\right) \\ &1.0 \left(1.0 \ 1 \times 10^{-5} \text{mol/l}\right) \end{aligned}$	Smith and Paselk, 1986
Domestic fowl Gallus gallus	Heptane Hexane Pentane	0.31-0.57 0.64-1.00 1.58-2.22	Stattelman et al., 1975
Pigeon Columba livia	Heptane Hexane Pentane Amyl acetate Ethanethiol Butanethiol	0.29-0.38 1.53-2.98 16.45-20.76 0.31-29.8 10080 13825	Stattelman <i>et al.</i> , 1975 Henton, <i>et al.</i> , 1966 Snyder and Peterson, 1979
Northern bobwhite Colinus virginianus	Heptane Hexane Pentane	2.14–3.49 3.15–4.02 7.18–10.92	Stattelman et al., 1975
Black-billed magpie Pica pica Starling Sturnus vulgaris	Ethanethiol Butanethiol Cyclohexanone	8400 13 416 Varies seasonally (0.3% vapor sat. [3.778 × 10 ¹⁴ molecules])	Snyder and Peterson, 1979 Clark and Smeraski, 1990
Tree swallow Tachycineta bicolor	Cyclohexanone	73.4–317.8	Clark, 1991
Cedar waxwing Bombycilla cedrorum	Cyclohexanone	6.8–86.46	Clark, 1991
Brown-headed cowbird Molothrus ater	Ethyl butyrate	0.76	Clark and Mason, 1989

the number of olfactory receptor cells in dogs ranges from 125 to 225 million, there would be one molecule of odorant for every 82 sensory receptors, on average, if all molecules reached the olfactory epithelium. In reality much fewer molecules reach the epithelium and it was concluded that olfactory receptor cells may respond to only one molecule (Neuhaus and Müller, 1954). Of six fatty acids tested, ranging from acetic acid (C_2) to caprylic acid (C_8) , dogs

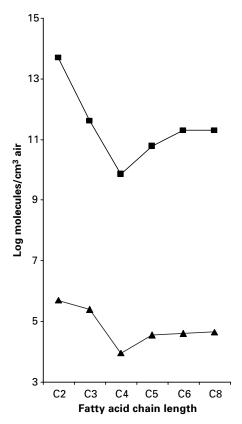


FIGURE 5.15 Detection thresholds for fatty acids in dogs (\triangle) and humans (\blacksquare). Both species are most sensitive to butyric acid (C_4). (From data in Neuhaus, 1953.)

were most sensitive to butyric acid (C_4) (Fig. 5.15). Thresholds were higher for fatty acids with lower or higher chain lengths. Humans showed the same V-shaped threshold curve (Neuhaus 1953). However, compared with humans, dogs (beagles) were found to be 300 times more sensitive to amyl acetate (Krestel *et al.*, 1984), and 1000 to 10000 times more sensitive to α -ionone (Marshall and Moulton, 1981). Laboratory rats proved more sensitive than humans to n-propanol (8-fold), benzaldehyde (10-fold), cyclohexanone (10-fold), isobutyl n-butyrate (30-fold), and n-heptanol (50-fold) (Laing, 1975).

Thresholds can vary between and within individuals, with the estrus cycle, and with the chemical background. Some benchmarks will be given here. In laboratory (Wistar) rats, the olfactory detection threshold for ethyl acetate was measured as 7.3×10^{-5} vol% in subadults, while adult rats were more sensitive (1.4 × 10⁻⁵ vol%) (Apfelbach *et al.*, 1990). (By comparison, among insects the gypsy moth is 10^{13} times as sensitive. The threshold for bombykol, defined as

the concentration at which 50% of the males fan their wings, was approximately 1×10^{-8} g/cm³ air).

Neotropical bats (*Desmodus rotundus*, *Artibeus literatus*, and *Phyllostomus discolor*) are very sensitive to butanoic acid: their detection threshold lies between 1.5×10^{-10} and 1.5×10^{-11} mol/l (Schmidt, 1975). The short-tailed fruit bat (*Carollia perspicillata*) has detection thresholds for 18 odorants ranging from 3.6×10^{13} to 2.7×10^{10} molecules/cm³ air. The animals were most sensitive to fruit-typical compounds such as ethyl butyrate (5.4×10^{10} molecules/cm³), *n*-pentyl acetate (2.8×10^{10} molecules/cm³), and linalool (1.8×10^{11} molecules/cm³), suggesting nutritional specialization of chemoreception (Laska, 1990). The bats increased their respiration rate from a basal rate of 2-4 Hz to as much as 12 Hz when confronted with an odor of high concentration.

A complex biologically relevant odor, such as that of a fruit for fruit bats, may require fewer molecules overall for detection than each of its single components alone (Laska *et al.*, 1990; Laska and Hudson, 1991). Vampire bats (*Plasmodus rotundus*) have a threshold of 1.5×10^{-10} mol/l to 1.5×10^{-11} mol/l for butanoic acid (Schmidt, 1975). Similarly, subthreshold amounts of two or more single compounds may become noticeable when mixed together. Dogs responded to mixtures of subthreshold concentrations of butyric plus isovaleric acids and of butyric plus *n*-caproic acids (Neuhaus 1956a).

Some biologically relevant smells may have two "functional thresholds," the higher one resulting in a qualitatively different odor. For instance, geosmin in high concentrations smells like "musty basement" or "soil," in low concentration like "beets."

In some mammals, the sexes differ in their odor thresholds. The sow detects the boar pheromone androstenone (5α -androsten-16-en-3-one) easily at 3.67×10^{-3} mol/l, while this concentration is near or below the threshold for boars. Boars initially could not detect the odor at all but became more sensitive after repeated exposure (Dorries *et al.*, 1991).

Humans may be programmed to be more sensitive to natural food contaminants. For instance, (–)geosmin (trans-1,10-dimethyl-trans-9-decalol) occurs in earth, natural surface water, and in foods in contact with soil or water, such as beets, clams, or fish. Geosmin is a microbial, fungal, or algal metabolite. It is a water pollutant and off-flavor compound. The naturally occurring (–)enantiomer has a threshold 11 times lower than the (+) enantiomer (Polak and Provasi, 1992).

Humans are extremely sensitive to hydrogen sulfide, most likely as a danger signal. The recognition threshold lies at 0.00047 ppm (vol) (cited in Cain, 1978). Only 40 molecules of methyl mercaptan, distributed over several receptors, sufficed for detection (cited in Harborne, 1993).

The thresholds of human subjects can vary over minutes, hours, or days as much as between individuals. Therefore, measured interindividual thresholds may reflect nothing more than having been recorded at different levels of these sensitivity fluctuations (Stevens *et al.*, 1988).

Considering the biologically important 16-androstenes, humans are most sensitive to 5α -androst-16-en-3-one (threshold: 0.2 ppb). The detection threshold for 4,16-androstadien-3-one is 1.0 ppb and that for 3α -androstenol is 6.2 ppb (Amoore *et al.*, 1977).

Odor thresholds in humans appear to be more influenced by the environment than genetics. The detection thresholds for acetic acid, isobutyric acid, and cyclohexanone varied as much in monozygotic twins as in fraternal twins. Instead, smoking and diabetes were related to lower olfactory sensitivity, and body fatness and alcohol consumption to greater sensitivity to the latter two compounds (Hubert *et al.*, 1980).

Thresholds are affected by the complexity of an odor. In tests with mixtures of 3, 6, and 12 compounds, human subjects varied less from one another in their thresholds the more complex the odor was. The same trend was observed within individuals (Laska and Hudson, 1991).

In general, perceived odor intensity is a power function of the odor concentration. The exponent is below 1, meaning that boosting the perceived intensity requires increasingly more odor production. As odor production becomes more and more costly, fine-tuning of chemoreception (instead of odor production) becomes the more attractive alternative evolutionarily.

5.6 Hormonal influences on chemoreception

Sensory performance often varies between the sexes and with different stages of the reproductive cycle. In humans, females outperform males at all ages (Doty, 1981; Doty *et al.*, 1984). Even infants show this difference: at 2 days old, girls could discriminate cherry from ginger odor, but boys could not (Balogh and Porter, 1986).

Olfactory thresholds also vary with the ovarian cycle. Women are most sensitive to odors around the time of ovulation, when estrogen levels are highest, and less sensitive to odors during menstruation. This may have sensory—physiological reasons. During menstruation the mucus layer on the olfactory epithelium is thicker and more likely to trap molecules, while the thin mucus layer at the time of ovulation renders the receptors more accessible. The thickness of the mucus layer, in turn, is controlled by testosterone and estrogen (Mair *et al.*, 1978).

Sex differences in olfactory performance have been described for many mammals, implying hormonal differences. In domestic cats, as in other species, males typically perform flehmen in response to conspecific urine. But spayed females can be stimulated to show flehmen by administering testosterone. If paired with estrogen-treated females, they frequently inspect the genital area of the female partner and subsequently exhibit flehmen. Males flehmen in 80% of the time to either female or male urine that is applied to the naso-oral surface. Testosterone-treated females flehmen to 90% of the male urine samples, and to 70% of those from females (Hart and Leedy, 1987).

Males also depend on testosterone for olfactory performance. If in hamsters testosterone is converted to estrogen by subcutaneous silastic implants of the aromatase inhibitor 1,4,6-androstatriene-3,17-dione, their sexual sniffing decreases. They sniff less toward novel females and no longer discriminate between males and females (Steel and Hutchinson, 1987). Castration affects odor detection performance in male rats (Doty and Ferguson-Segall 1989).

Central norepinephrine modulates systems that regulate the attraction to conspecific odors in the rat. A noradrenergic neurotoxin (DPS4), lowers the level of norepinephrine in the olfactory bulb, the olfactory cortex, and frontal cortex. When given to male rats, their response to odors from females is impaired: while sexually experienced, intact males are attracted to pine shavings from the nest of a female and her litter, males treated with DSP4 are not attracted. Furthermore, while normal males enter a chamber with pup-scented shavings more often if odor from anesthetized females is added, males treated with DSP4 do not respond in this way (Cornwell-Jones *et al.*, 1988).

Hormonal events around parturition affect olfactory interactions between mother and offspring. Expulsion of the fetus is followed by liberation of oxytocin in the brain. The oxytocin level rises also in the olfactory bulb. This suggests that oxytocin modulates olfactory processing at the level of the olfactory bulbs (Kendrick *et al.*, 1988). Vaginal stimulation during birth increases noradrenergic activity, especially in the olfactory bulb (Kendrick *et al.*, 1988), possibly resulting in olfactory memory (Rosser and Keverne, 1985). Evidence for a noradrenergic influence on bulbar neural networks during parturition comes from studies with rats. If a female rat's central noradrenergic projection to the olfactory bulb is experimentally lesioned prior to parturition, cannibalism will result, but not general anosmia or gross impairment of maternal behavior (Dickinson and Keverne, 1988).

5.7 Chemoreception and age

Humans perform at their olfactory best during the third to fifth decade of life. A marked decline occurs after the seventh decade. Over 50% of people

aged from 65 to 80 years have major impairments. In one study, 75% of subjects 80 years of age or older suffered such loss. The gradual loss of sensitivity to odors is responsible for frequent complaints by the elderly that food lacks flavor. The elderly are also disproportionally often poisoned by household gas (Doty *et al.*, 1984).

Aging does not necessarily affect the process by which overall intensity in different sensory modalities is determined. Young people (18 to 21 years) and older people (61 to 94 years, average 74.5 years) differed in their magnitude estimations of almond odor and almond taste. However, they did not differ when presented with the taste of sucrose or the visual task of estimating the length of a line (Enns and Hornung, 1988). Likewise, young and old did not differ in their magnitude estimations for odor of lime (Stevens and Cain, 1985).

5.8 Interaction between chemical senses

5.8.1 Olfaction-vomeronasal organ interaction

Male mice learn about female cues. In the presence of females, a male emits ultrasonic vocalizations. These vocalizations become less frequent after extirpation of the VNO. The more experience a male has had with females, the smaller the deficit he suffers. The learning is mediated by interaction between the main olfactory system and the accessory olfactory system: after vomeronasal deprivation, olfactory cues maintain the behavior (Wysocki *et al.*, 1986).

5.8.2 Odor-taste interactions

Odorants and tastants in foods interact in complicated ways. For instance, strawberry odor enhances the sweetness of whipped cream, while peanut butter odor does not, and strawberry odor did not enhance the saltiness of sodium chloride (Frank and Byram, 1988).

The best investigated odor-taste interactions occur in conditioned flavor aversions. Tastes that precede a delayed food-related illness are often avoided after only one experience. Odors are not avoided under similar conditions. However, if taste and odor are presented together before the "malaise," animals will avoid odor when encountered later by itself. Taste affects odor, but not vice versa. If only the taste intensity is increased, both taste and odor aversion increase. Conversely, if only the odor stimulus is increased, only the odor aversion increases (Garcia *et al.*, 1986).

5.8.3 Olfaction–trigeminus interactions

Interactions of trigeminal fibers and olfactory chemoreception have been extensively studied. In frogs, trigeminal fibers lead into the olfactory mucosa. The trigeminal system could thus modulate olfaction peripherally as well as centrally. Indeed, antidromic electrical stimulation of the ophthalmic branch of the trigeminal nerve evokes slow potentials in the olfactory mucosa, modifies the activity of receptor cells, and modulates responses to odors. Trigeminal stimulation by odorants might trigger local axon reflexes that induce the sensory neurotransmitter substance P, which, in turn, elicits electrical responses in the mucosa (Bouvet *et al.*, 1989). Blocking trigeminal input by subcutaneous injections of capsaicin does not critically affect perception of olfactory or taste stimuli in starlings (Mason *et al.*, 1987a).

An interaction between "main olfaction" and the trigeminal somatosensory system has been proposed to facilitate "directional smelling." For carbon dioxide and menthol, human subjects were able to tell the odor direction in 96% of cases, while the results for hydrogen sulfide and vanillin were random (Kobal *et al.*, 1989). Rats can discriminate odor direction in one sniff (Rajan *et al.*, 2006).

5.8.4 Chemoreception and other sensory modalities: mammals

Chemical cues often have to be accompanied, preceded, or succeeded by visual, auditory, or tactile stimuli for appropriate behavior to occur. Bats, for example, use both olfactory and acoustic information for individual recognition (Kunz, 1982).

Visual cues can help in the detection of a scent mark in the first place. This may be accomplished in different ways. An animal may use a visually prominent natural landmark for scent marking, such as a rock, stump, or root along a frequently traveled trail. Other species produce their own visually conspicuous support for a scent mark. Carnivores, such as otters (*Lutra* sp.), use feces, called spraint, as carrier for secretion from the anal gland (Chanin, 1985). Beaver, *Castor* sp., have carried this behavior one step further. They build a platform for their scent mark by dredging up mud from their pond and depositing it on the bank. This scent mound is marked with secretion from the castor sacs and anal glands. Finally, the scent mark itself may be visually conspicuous, as that of the brown hyena, *Hyaena brunnea*. During scent marking, the two lobes of its anal glands produce white and black secretion in succession. This results in a bicolored scent mark on a blade of grass that has the black part above the white one and separated from it by 1 cm (Kruuk, 1972; Fig. 6.7).

Relatively non-volatile materials such as the proteins (10–30 kDa) and lipids in the femoral gland secretion of desert iguanas, *D. dorsalis*, may strongly absorb long-wave ultraviolet (300–500 nm). Desert iguanas actually locate scent marks better under ultraviolet light (Alberts, 1989).

Numerous studies have addressed the relative importance of different senses in modulating behavior. Mating behavior of domestic sheep is an example. Rams use mostly vision for sexual activity but smell for seeking the partner. Odors are important for distinguishing estrous from non-estrous ewes. In ewes, blindfolding interfered with seeking rams (Fletcher and Lindsay, 1968).

One sense can be substituted for another in feeding behavior in sheep. Even if smell, taste, and touch are impaired, sheep still show food preferences but undesirable plants become more acceptable. However, the senses contribute specific information: flowering stems are not taken if smell only is impaired. It is assumed that flowers have an attractive smell to sheep. Taste appears to be important in discriminating fine- and broad-leave *Dactylis glomerata* (from different geographic areas). Finally, it is thought that the sense of touch is responsible for the fact that brome grasses are always highly acceptable (Arnold, 1966). Merino weaners learned from experienced adult sheep to eat wheat, a novel food. They learned rapidly, even if one or two senses were impaired. Only weaners with impairment of olfaction, vision, and hearing failed to eat wheat for all 5 days of the experiment; weaners with no "teachers" also did not eat the wheat (Chapple and Wodzicka-Tomaszewska, 1987).

Spiny mice, *Acomys cahirinus*, use all senses interchangeably in locating prey (Langley, 1988). These are cases of "adaptive redundancy" at the level of the sensory modalities. As is well known, deaf-blind humans use smell and taste more.

Signaling pheromones I: discrimination and recognition

My dear friend, the last time you were so good as to come and see me – for nobody comes any more to see the wretched invalid I am – I was obliged to take the chair you sat in and keep it out in the courtyard for three days: it was impregnated with scent.

MARCEL PROUST, according to Léon Pierre-Quint, 1925

Optimus odor in corpore est nullus [The best body odor is none]. SENECA, Epistulae ad Lucilium, *Epis* CVIII, 16.

Signaling pheromones are animal-produced, interindividual chemicals that modulate behavior in conspecifics. Like visual and auditory signals, they have comparatively rapid effects: exchange of signals takes seconds or minutes. (Priming pheromones [Ch. 8], by comparison, trigger slower endocrine or developmental processes.) The pheromone concept, originally based on insects (Karlson and Lüscher, 1959), has been debated for vertebrates, notably mammals (e.g. Beauchamp *et al.*, 1976; Johnston, 2001). Often it is better to use the term "body odors" to avoid particular assumptions. Now the term pheromones is widely used for vertebrates, without any particularly narrow definition implied.

A pheromone is functionally defined as a conspecific compound(s) that affects a receiver. Sources such as urine or gland secretions typically contain many compounds of which only some are pheromonally active. So in most cases, a pheromone is more than a single compound and less than a secretion ("scent"), it is rather a group of active compounds in a secretion or excretion that supply information to, or change behavior in, another conspecific.

The following text discusses first the ability of animals to distinguish and recognize other animals by odors without necessarily exhibiting specific behaviors and then the behaviors that are modulated by status signals. Chapter 7 discusses the sexual and evolutionary implications of signaling pheromones.

6.1 Familiarization with environment and objects

Strange as it sounds, the absence of odor often serves as a powerful stimulis; unfamiliar and still unmarked objects or areas prompt vigorous scent marking. This applies to many mammals, including ungulates, such as pronghorn, *Antilocapra americana* (Müller-Schwarze *et al.*, 1972) rodents, such as house mice, *Mus musculus* (Hurst, 1987), and carnivores (Kleiman, 1966). For example, pine martens, *Martes martes*, mark most consistently unmarked objects and do not mark objects that already carry their own odor. It is concluded that marking primarily serves familiarization.

In isolated male mice, own odor regulates the amount of urine deposited in marking. If it is present, they mark less, while clean surfaces and also other males' urine trigger more frequent marking (Daumae and Kimura, 1986). In our laboratory experiments, students are impressed by how a mouse stops at a clean tile in the middle of a soiled open field. A "scent-the-habitat" function for odors from both sexes has been assumed for the gland secretions in the brushtail possum, *Trichosurus vulpecula*, since no sex differences in chemical composition were found (Woolhouse *et al.*, 1994).

The "security blankets" of children belong in this context. Apart from tactile stimuli, they also provide a familiar odor, particularly in strange surroundings. Many children will not easily accept a familiar blanket, piece of cloth, or tattered teddy bear that has been washed. A particularly inventive 3-year old boy held a cloth toy close to his nose when tired or stressed. Asked why he did so, he answered "to get the smell right" (Russell, 1983).

6.2 Familiar versus non-familiar social odors

Discrimination of familiar from unfamiliar social odors is widespread among vertebrates. It serves for recognition and interaction with group (or colony) members. Being familiar with other individuals not only reduces aggression but also may facilitate sexual behavior. Group members can be discriminated on the basis of familiarization with individuals; sharing common environmental odors, such as those of nest or diet; "labeling" of one animal by another; and sharing individual odors that result in a colony odor, as in sugar gliders, *Petaurus breviceps* (Schultze-Westrum, 1965). These can combine into rather complex body odors, determined by diet, maternal labeling, and genetic factors, as for example in spiny mice, *Acomys* sp. (Porter *et al.*, 1989).

 $^{^1}$ Animal names follow the original literature. Thus, the house mouse appears here variously as Mus musculus, Mus musculus domesticus, or Mus domesticus. Retaining the original version may facilitate searching the literature. "Charr" is Canadian for "Char."

6.2.1 Discrimination of own odor versus that of other conspecifics *Reptiles*

Lizards have femoral glands that are more developed in males. The secretion is proteinaceous, and its function is largely unknown. Desert iguanas (*Diposaurus dorsalis*) probably use femoral secretion for advertising their home range. Iguanas experimentally exposed to own secretion and that of a strange male tongue flicked oriented toward their own tails in response to own secretion. This may be "scent matching" of scent with "scent owners." They marked primarily their own secretion. This may indicate self-recognition (Alberts, 1992c). Bluetongued skinks (*Tilique scincoides*) also discriminate their own odor from that of other conspecifics (Graves and Halpern, 1991).

Mammals

In male house mice, urine marks from other males trigger intensified marking, while grouped male mice do not distinguish between own odor and that of other males (Daumae and Kimura, 1986). The aboriginal house mouse Mus spretus carries or pushes away fresh fecal pellets. Experiments demonstrated that they recognize their own feces. Compared with fecal pellets of other conspecifics of both sexes, these mice are more likely to remove their own and are less likely to investigate them (Hurst and Smith, 1995). Mice (Mus sp.) avoid the odor of mice from other groups. Here it is not necessary to be familiar with the odor of the other group. Odors of mice from discontinuous, completely strange demes are also avoided (Cox, 1989). In male mice, volatiles from feces of strange males depress the heart rate more than volatiles from own feces (Goodrich et al., 1990). In pine marten, M. martes, own scent marks reduced marking behavior, while all other types of scent mark were marked in turn (deMonte and Roeder, 1990). Pine martens did not respond differentially to familiar/unfamiliar martens, males versus females, or other species such as stone marten or genets (deMonte and Roeder, 1990).

6.2.2 Familiar individuals

Exposure to the odor of *particular* conspecifics can affect behavior toward these individuals when encountered.

Birds

Antarctic prions (*Pachyptila desolata*) have a "musky" smell and appear to use conspecific odors to recognize their burrow. These birds chose the odor of

their breeding partner over that of another conspecific in Y-maze experiments on the Kerguelen archipelago. They also preferred a conspecific's odor to their own, and the latter to "no odor" (Bonadonna and Nevitt, 2004).

Rodents

In meadow voles, *Microtus pennsylvanicus*, males are more aggressive toward familiar males than unfamiliar ones, while females behave in the opposite way: encounters between familiar females entail less agonistic behavior and more "amicable acts" than those between unfamiliar females. This is true for voles kept together or apart in the laboratory and also for voles caught in the wild, whether they had been neighbors or lived at greater distances from one another. In contrast to same-sex encounters, familiarity has no effect on male-female interactions (Ferkin, 1988). Spiny mice (*Acomys* sp.) prefer to huddle with familiar cagemates. They do not recognize each other if rendered anosmic by treatment with zinc sulfate (Matochik, 1988).

6.2.3 The "dear enemy" phenomenon

Ecologically, female meadow voles are territorial, know their neighbors, and are more tolerant of each other. They exemplify the "dear enemy" concept: familiar neighbors reduce aggression toward one another because they pose less threat to each other than newcomers without a territory, who might compete for territory, mates, or resources. Males are dispersal prone, and neighbor relations are more ephemeral. Each male's home range overlaps with those of several females (Ferkin, 1988).

Eurasian beavers (*Castor fiber*) show the dear enemy phenomenon: they respond more strongly (by destroying scent mounds and over-marking them) to experimental scent marks from strangers than from neighbors. This is true for marks containing either of the two beaver secretions castoreum or anal glands secretion (Rosell and Bjørkøyli, 2002). The dear enemy phenomenon is thought to be particularly adaptive in species that maintain multipurpose territories containing the breeding site, mate, and food supply (Temeles, 1994).

Males of the Columbian ground squirrel (*Spermophilus columbianus*) sniff scent marks from oral glands of non-residents longer than marks from residents. However, they marked their own, neighbor's, and strangers' scent equally often (Harris and Murie, 1982). Similarly, male woodchucks, *Marmota monax*, spent more time investigating scent marks of strangers than those of neighbors (Meier, 1991). (In woodchucks, adults and juveniles of both sexes mark objects near their burrows with their oral angle gland.)

Preexposing rodents to the odor of a conspecific can alter the response to this individual when it is again encountered later, compared with responses to conspecifics whose odor is unfamiliar. For example, female brown lemmings, *Lemmus trimocrunatus*, were experimentally exposed to the odor of a male. These females engaged in more contact social behavior with that now "familiar" male than females who had experienced odor from a different male or none at all. The males whose odor the females had experienced ejaculated more frequently than males under the other two conditions (Coopersmith and Banks, 1983).

Female golden hamsters can distinguish individual males by the odors of their flank gland secretion, urine, feces, and soiled bedding. Females were less aggressive and showed more sexual behavior toward males whose odor they had been exposed to before than to unfamiliar males. The preexposed females also had larger litter sizes. Thus, familiarity with the odor of a particular male appears to promote mating and enhance mating success (Tang-Martinez *et al.*, 1993).

Carnivores

Urban feral cats of both sexes sniff marks of sprayed urine more if the donor is a strange cat (from a different town) and respond least to the urine of a familiar cat. Since one male roams over the territories of several females, it is assumed that male–male competition has selected males that spray mark more and respond more strongly to urine marks (Natoli, 1985).

Another example from carnivores is the odor of anal sac secretion in the ferret, *Mustela furo*. Ferrets discriminate strange from familiar individuals by this odor. (They also use anal sac secretion to distinguish males from females, a familiar individual's from own odor, and fresh from 1-day old odor, but not between fresh and odor only 2 hours old nor anestrous from estrous females [Clapperton *et al.*, 1988]).

Bats

Female pipistrelle bats (*Pipistellus* pipistrellus) recognize familiar individuals by scent. They preferred the odor of females from the same colony to that of females from a different colony (deFanis and Jones, 1995).

Primates

Lemurs (*Lemur fulvus* and *Lemur macacao*) showed more interest in the scent marks of familiar than unfamiliar individuals (Fornasieri and Roeder, 1992).

Table 6.1 Kin recognition by chemical cues in vertebrates

Species	Behavior	Reference
Fish		
Coho salmon Oncorhynchus kisutch	Phenotype matching	Quinn and Busack, 1985
Rainbow trout Oncorhynchus mykiss	Staying in odorized water current	Brown et al., 1993
Amphibia		
Polyphenic salamanders	Eating parasitized prey	Pfennig, 1997
Toad tadpoles Bufo americanus	Negative response to non-siblings in Y-test	Waldman, 1981
Cascade frog tadpoles Rana cascadae	Naive tadpoles group with siblings; waterborne cues	O'Hara and Blaustein, 1985 Blaustein and O'Hara, 1981
Mammals		
White-footed mouse <i>Peromyscus leucopus</i>	Possibly by odor	Grau, 1982
Spiny mice Acomys cahirinus	Associate with familiar siblings anosmics do not discriminate	Porter, 1986
13-lined ground squirrel Spermophilus tridecemlineatus	Reduced social exploration when raised together	Holmes, 1984
Beldings ground squirrel Spermophilus beldingi	Phenotype matching	Holmes, 1986
Beaver Castor canadensis	Differential response to scent marks of relatives and strangers, phenotype matching	Sun and Müller-Schwarze, 1997

6.2.4 Indirect familiarization

Mammals may become familiarized with one another by body odors, without having direct contact. For instance, Columbian ground squirrels, *S. columbianus*, show more-cohesive and less-agonistic behaviors toward unfamiliar non-colony members if they had experienced the odor of these individuals in traps that had been used in both colonies (Hare, 1994).

6.3 Recognizing kin

Many questions in inclusive fitness theory turn on whether and how animals distinguish genetic relatives from non-kin. Vertebrates, from fish to humans, use odors, in addition to cues in other sensory modalities, to recognize kin (Table 6.1). This recognition, in turn, may enable relatives to cooperate,

resulting in kin selection, or to adjust reproduction, as in avoiding incest. Generally, there are four kin recognition mechanisms that involve various sensory modalities, including chemoreception (Fletcher and Michener, 1987).

- 1. *Spatial cues*: individuals from one common place are considered kin.
- 2. Familiarity: cues are learned during earlier association.
- 3. *Phenotype matching*: to recognize an individual as kin, it is not necessary to have met it before. Instead, it can be recognized by cues shared with oneself or a known related individual (also known as the "armpit effect").
- 4. *Recognition alleles*: both expression of the identifying cue and the recognition process do not rely on experience.

Tang-Martinez (2001) suggested that kin recognition possibly relies on only one mechanism: "learning, particular associative learning and habituation" (i.e. point 2 in this list).

6.3.1 Fish

Juvenile coho salmon, *Oncorhynchus kisutch*, recognize kin. This is considered to be advantageous for schooling, predator avoidance, and avoiding inbreeding (Quinn and Busack, 1985). Whether fish recognize kin may depend on how they were reared. In Arctic charr, *Salvelinus alpinus*, only fish that were raised with siblings later preferred water scented by unknown siblings over water from non-siblings. Charr raised in isolation do not discriminate the two odors. Even when the isolated fish, at 15 months of age, are reared with siblings for 50–62 days, they still do not discriminate sibling odor. Therefore, it appears that in Arctic charr social experience during the first 15 months of life is necessary for sibling preference (Winberg and Olsén, 1992; Olsén and Winberg, 1996).

The major histocompatibility complex (MHC) plays a role in kin recognition. Arctic charr preferred water scented by a sibling of the same MHC (class II) type over water from a sibling that differed in MHC type from themselves. The charr did not discriminate in this way if they had been living in isolation since fertilization. However, a sibling of a different MHC type was not distinguished from a non-sibling that shared the MHC type of the responding fish. As the earlier experiments, this confirmed the role of early learning in sibling recognition. Arctic charr probably use MHC plus other odors, possibly based on the MHC class I region, for kin recognition (Olsén *et al.*, 2002a,b).

Striped bass, *Morone saxatilis*, juveniles develop in estuaries, notably Hudson River and Chesapeake Bay. Juveniles school during the day but disperse and feed at night. Schools provide the advantages of improved prey location, predator avoidance, and swimming hydrodynamics. How are the fish attracted to each other? In Y-maze tests in the laboratory, juveniles are attracted to both

familiar siblings and unfamiliar unrelated juveniles but they can distinguish between familiar siblings and unfamiliar, unrelated juveniles, and prefer the former. This recognition is probably important as a simple mechanism to re-form schools. Lost or disoriented individuals probably use a species-specific response (Kelley, 1988).

Juvenile Atlantic salmon (*Salmo salar*) and juvenile rainbow trout (*Oncorhynchus mykiss*) that live in kin groups fight less, thus saving energy and reducing the risk of injury (Brown and Brown, 1993). Rainbow trout discriminate unfamiliar kin from non-kin, but not familiar from unfamiliar kin (Brown *et al.*, 1993). This appears to be an example of phenotype matching: kin have matching odors, while kin and non-kin have not.

6.3.2 Amphibia

In tadpoles of the American toad, *Bufo americanus*, the prehatching environment influences postnatal preferences. Tadpoles reared isolated from any conspecifics, with siblings only, or exposed to both siblings and non-siblings differed in their later association with siblings in a test pool. The first and second group preferred to associate with siblings, while the third did not prefer to associate with siblings, unless they had been reared in sibling groups during early development. The tadpoles reared in isolation from conspecifics discriminated paternal, but not maternal half-siblings from full siblings (Waldmann, 1981).

6.3.3 Reptiles

Hatchling green iguanas, *Iguana iguana*, recognize kin by the odor of their feces, but also by their body odor (Werner *et al.*, 1987).

6.3.4 Non-human mammals

It has been known for a long time that certain rodents avoid mating with siblings. This, for example, is true for European common voles, *Microtus arvalis* (Frank, 1954). After living in isolation, adult common voles sniffed nonsiblings of the opposite sex more than siblings (Bolhuis *et al.*, 1988). Similarly, female house mice, *M. musculus*, prefer the smell of non-sibling males to that of siblings or males of other strains. This has been interpreted as a sign of *optimal outbreeding* (Gilder and Slater, 1978). Female prairie voles (*Microtus ochrogaster*) spend more time investigating the anogenital and mouth region odors of nonsiblings than siblings. They also discriminate saliva and urine of non-siblings from those of siblings. It is thought that examining one male more than another may help in mate choice (Smale *et al.*, 1990). In the prairie vole (Gavish *et al.*, 1984)

and the gray-tailed vole, *Microtus canicaudus* (Boyd and Blaustein, 1985), social experience before weaning is necessary for incest avoidance to occur. Laboratory mice choose their mates based on both relatedness and familiarity (Barnard and Fitzsimmons, 1988). By cross-fostering, it is experimentally possible to "create" "non-genetic" or phenotypic "kin" in laboratory mice (Kareem and Barnard, 1982) and deer mice, *Peromyscus leucopus* (Grau, 1982). Female laboratory mice appear to avoid mating with close relatives by preferring genetically dissimilar mates. Specifically, in one experiment, females chose males that were dissimilar at the MHC (Egid and Brown, 1989). This does not apply to members of the same sex: female house mice prefer to nest with other females with whom they share MHC genes (Manning *et al.*, 1992).

In rats, nulliparous females kill unrelated young. Such infanticide is reduced if these females are exposed to bedding soiled by a pregnant rat (Menella and Moltz, 1989), an odor that also reduces infanticide by male rats (Menella and Moltz, 1988).

How does kin recognition develop? Rats learn some kinship characteristics (Hepper, 1983). Rat pups whose mother and siblings were rubbed with different odors prefer the odor of their siblings when tested in a Y-maze (Hepper, 1987). But genetic factors also play a role. Rats can distinguish chemosignals in the urine of other rats that differ only in the MHC. This may be the basis for kin recognition. In the mouse, the MHC is located on chromosome 17. Proteins encoded by the MHC are found on all cell surfaces, are antigen receptors, but also contribute to urinary odors specific to individuals (Hurst *et al.*, 2001). The binding groove on the protein molecule, a binding site for antigenic peptides, may also serve as binding site for volatile signal compounds (Beynon *et al.*, 2002). There are 50 genes in this complex, and they are highly variable, with about 50 alleles. The gene *H-2Kb* is the most mutable. This variability lends itself to signaling individual signatures.

6.3.5 Humans

The odors of T-shirts worn by mothers and their children appear similar even to strangers (Porter *et al.*, 1986). Adults can recognize the odor on shirts worn by their full siblings after several months of separation (Porter *et al.*, 1986).

Mothers recognize their newborn babies by odor (Russell *et al.*, 1983; Schaal *et al.*, 1980) even if there had been little contact after a Cesarean delivery (Porter *et al.*, 1983). Mothers distinguished the odor of their own infant from those of two other infants on a non-soiled undershirt that the infant had worn for at least 13 hours. If the mothers had been exposed to their own infant for less than 10 minutes after birth, only 20% were successful at recognition. After

10–60 minutes, 90% succeeded. All mothers exposed to their infant longer than 1 hour identified their odors. Maternal analgesia or anesthesia had no influence on the discrimination (Kaitz *et al.*, 1987). These results show that the infant's odor is most likely very important for identification during the immediate postpartum period (Eidelman *et al.*, 1987). Even fathers, grandmothers, and aunts, after only 0 to 8 hours of exposure to a newborn, can correctly identify an infant 1–2 days old by the odor on its garment (Porter *et al.*, 1986). Human infants actually mark their mothers with saliva. Sleeping infants aged 6–8 weeks responded to the mother's breast odor or to the odor of their own saliva on gauze pads, but not to cow's milk (Russell *et al.*, 1989).

Breast-fed neonates themselves distinguish breast (Macfarlane, 1975; Russell, 1976; Schaal *et al.*, 1980) and axillary (Cernoch and Porter, 1985) odors of their mother from the same body region odors of other lactating women.

6.4 Individual odors

Every individual of a species can have its own olfactory signature or "fingerprint" owing to the many odor sources that make up the body odor and the many compounds in each secretion or excretion. The relative concentrations of all these compounds may vary, providing for endless variety. These individual signatures can potentially be monitored and exploited by conspecifics. "True individual recognition" is a more selective and complex process than "social recognition" (categorization of conspecifics) (Gheusi et al., 1997). Animals are able to discriminate between the odors of two individuals other than self, such as its mate and another individual of the same sex and status. Such individual recognition or, better, discrimination (Halpin, 1986) is thought to increase genetic fitness. Examples are recognition of one's mate in species with strong and/or lasting pair bonds, recognition of own offspring, recognition of relatives where favoring of kin results in increased chances of propagation of one's own genes, and recognition of a potential rival such as a territory owner. True recognition of the odor of an individual, as opposed to discrimination of odors of age, sex, or physiological status, has rarely been demonstrated.

6.4.1 Amphibia

Male red-backed salamanders (*Plethodon cinereus*) in Petri dishes were given a choice between own feces and those of another male. They preferred their own. The same was true for "washes" from the cloacal glands (Simon and Madison, 1984). While this may represent discrimination of "own" and "other," rather than true *individual* recognition, another experiment showed

discrimination of individual odors in *Plethodon* spp. In a Y-maze, *Plethodon* salamanders discriminated individual odors in addition to sex and species odors (Dawley, 1984).

6.4.2 Non-human mammals

Among rodents, chemosensory discrimination of individuals occurs in mice (Bowers and Alexander, 1967), rats (Krames, 1970), Mongolian gerbils (Halpin, 1974), eastern chipmunks, *Tamias striatus* (Keevin *et al.*, 1981), cavies, *Cavia aperea* (Martin and Beauchamp, 1982), golden hamsters, *Mesocricetus auratus* (Johnston, 1993; Johnston and Jernigan 1994), and prairie voles, *M. ochrogaster* (Newman and Halpin, 1988). Individual odors are also discriminated by African dwarf mongoose, *Helogale undulata rufula* (Rasa, 1973) and the Indian mongoose, *Herpestes auropunctatus* (Gorman 1976), badgers, *M. meles* (Kruuk *et al.*, 1984), and ferrets (Clapperton *et al.*, 1988).

Rats form an *olfactory image* about individuals that they retain in their *olfactory memory*. Gheusi *et al.* (1997) trained rats by operant conditioning to distinguish two other rats. After successful training, the reward and non-reward stimulus rats were switched (reversal test). The responding rat's error rate increased drastically after this reversal. This test avoids novelty effects that confound habituation–dishabituation experiments. In a second type of experiment, rats were first trained to discriminate pairs of rats, and then given the odor (bedding) of these rats. They were able to transfer the discrimination of whole animals to their odor signature alone (Gheusi *et al.*, 1997).

In primates, lemurs distinguish and remember individual odors. In *L. ful-ves* the anogenital scent mark of an evicted individual was still recognized (i.e., scent-marked) 10 months after the eviction (Fornasieri and Roeder, 1992).

Individual recognition can be important for selecting and correctly recognizing mates, reducing aggression, incest avoidance, and sexual re-arousal, in the Coolidge effect (named after an anecdote about a US President and refers to increased sexual arousal with a new mating partner) (Dewsbury, 1981).

Mate recognition

Male and female Mongolian gerbils, *Meriones unguiculatus*, respond to the urine of their respective mates by investigating it more and uttering more ultrasonic calls than to unfamiliar gerbils of the same sex. Members of same-sex pairs do not respond differently to the odors of the familiar and an unfamiliar individual of the same sex. To recognize the odor of one's mate may be important for pair maintenance by promoting affiliative behavior and reducing aggression (Brown *et al.*, 1988). Similarly, males and females of the prairie vole,

M. ochrogaster, recognize and prefer the odor of their mate to that of a non-mate. This was established with three techniques. First, after habituation to the odor of soiled bedding or urine from one male or female, material from another individual of the same sex was substituted. The voles discriminated this new odor from the previous one. Second, in a Y-maze, females preferred the odors of their mates over those from other mated or unmated males; males preferred the odors of their mates over those from other mated females but they did not discriminate between their mate and a virgin female. Third, when allowed to stay for over 10 hours in a scented Y-maze and provided with nest material, females stayed and built a nest on the side of their mate's odor. Likewise, males build a nest on the side with their mate's odor, when juxtaposed with the odor of a virgin female. It is not difficult to see that to recognize and prefer the odor of one's mate can play a vital role in monogamy (Newman and Halpin, 1988). Females of the shrew, Crocidura russula, distinguish flank gland secretion of their mate from that of another male (Cantoni and Rivier, 1992).

Sex differences

Discrimination of individual odors may depend on the sex of the odor donor. In the Virginia opossum (*D. virginiana*), females distinguish individual female, but not male odors (Holmes, 1992).

Genetic basis and chemical variation of odors

In the rat and other rodents, individual odors probably reflect genetic differences. Laboratory rats can distinguish individuals. They discriminate between two intact males, two castrated males, two estrous/proestrous females, two diestrous/metestrous females, or two ovariectomized females. Urine odors differ individually despite differences in the levels of gonadal hormones. Individual recognition may be independent of reproductive state or social status, even though hormone-influenced body odors may be used for individual recognition (Brown, 1988).

In the rat, the major histocompatibility complex (MHC) is responsible for individual odors (Brown *et al.*, 1987a,b). Genetic differences in the *IA* region actually result in different urine odors (Singh *et al.*, 1987).

The relative concentrations of constituents of the anal sac secretion in male stoats, *Mustela erminea*, are distinct for different individuals, but consistent over time for each individual. This possibly permits individual recognition of territory owners (Erlinge *et al.*, 1982). The chemical composition of the anal gland of the otter, *Lutra lutra* (Gorman *et al.*, 1978) and the subcaudal gland of the badger, *Meles meles* (Kruuk *et al.*, 1984) also differ with the individuals.

Body regions

Odors of different body regions can convey different types of information. In the golden hamster, *M. auratus*, five scents provide individual information. These are the flank glands, ear glands, urine, feces, and vaginal secretion. However, hamsters did not discriminate individuals by the odors of saliva, feet, chest fur, back fur, area behind the ear, and flanks from males with flank glands surgically removed (Johnston *et al.*, 1993). The golden hamster recognizes flank gland and vaginal secretions as coming from the same individual (Johnston and Jernigan, 1994).

6.4.3 Humans

Mothers and siblings can distinguish the odor of young children in their family from those of unrelated children of the same age. Parents could identify T-shirts worn by boys or girls as those of siblings or offspring and could correctly identify the T-shirt worn by each of two of their children (aged 35–58 months and 64–94 months (Porter and Moore, 1981). This shows that the identifying odors were individual signatures and not household odors common to several siblings (Porter and Moore, 1981). The odors of identical twins are so similar that humans cannot easily distinguish these (Wallace, 1977). Even dogs confuse the odors of identical twins, especially if not available simultaneously (Kalmus, 1955).

6.5 Odors in parental behavior

6.5.1 Chemosignals from parents in mammals

Offspring of many different vertebrate groups cue in on chemosignals from the mother, father, or both, and vice versa. Here we focus on mammals. From birth on, odors play a central role in discrimination of individuals. Odors from the mother fall into two categories: general odors, usually signifying a food source, and individual odors that the young use to recognize their mother as a specific individual.

Nipples, milk, and suckling

Nipple pheromone in altricial mammals

Newborn rabbits, *Oryctolagus cuniculus*, nurse only once per day for 5–7 minutes. During this short bout, pups can drink up to 25% of their body weight. They respond first to vibration and tactile cues from the approaching mother and then find the nipples with the aid of chemical cues. If one covers the nipples with

FIGURE 6.1 Structure of 2-methylbut-2-enal, the rabbit mammary pheromone.

plastic, there is little or no sucking. Earlier studies postulated that the "highly stereotyped and reliable response of rabbit pups to the odour presented by the nursing doe seems to qualify for consideration as a true pheromone" (Hudson and Distel, 1983). Newborn rabbits prefer the odor of females in early, rather than late, lactation; in the prenursing rather than postnursing phase; and nipple areas over other abdominal, or back, areas (Coureaud *et al.*, 2001). The mammary pheromone, a compound in rabbit milk that triggers searching by head movements and grasping the nipple, is 2-methylbut-2-enal (Fig. 6.1) (Schaal *et al.*, 2003). The pheromone is specific to rabbits and not produced by, or effective in, hares, rats, or mice. The nipple is the source of the mammary pheromone. Milk before it passes the nipple is not active as pheromone. It remains to be seen whether the pheromone comes from skin glands, symbiotic microorganisms, or compounds complexed to proteins or lipids that are secreted in the mammary glands (Moncomble *et al.*, 2005).

Likewise, odors on the domestic sow's ventrum regulate attachment of piglets to her nipples. Piglets take longer to attach to the nipples after washing the ventrum of the mother with organic solvent or impairing piglet olfaction either by blocking their nares or by flushing their olfactory system with lidocaine, a local anaesthetic. Attachment is still possible, since the response is controlled by tactile and taste stimuli as well as olfaction (Morrow and McGlone, 1988).

Kittens rely little on olfactory stimuli to attach to the nipples of their mother. Instead, tactile cues are important. The mouth area and the trigeminal projection field mediate these tactile sensations (Blass *et al.*, 1988).

Milk

During the days following birth, the odor of milk may activate young mammals. In rats at least, 3- to 9-day-old pups respond to the odor of milk by probing. This behavior wanes at 12 days of age. It is assumed that the odor of milk acquires its activating effect during the first hours and days by the suckling experience. If the dam's diet contains an artificial odor such as eucalyptol, this odor elicits as much activity, mouthing, and probing as milk odor Terry and Johanson, 1987). In rat pups 8 days old or older, suckling is critically dependent on olfaction. If 7-day-old rats are bulbectomized and then tested for

nipple attachment 24 hours later, they will not attach to the nipples, lose weight, and become moribund. Cortical lesions have no such effects (Risser and Slotnick, 1987).

Precocial Mammals

Lambs of domestic sheep use different senses for different stages of attachment (bonding, or imprinting) to their mothers. During their first hours, lambs stand up, nose the ewe, and suck. Blindfolded lambs did not stand. When the blindfold was removed, they approached the udder and sucked. This shows the importance of vision for this first step. After spraying the nasal passage with xylocaine to impair olfaction, the lambs did not suck. Finally, lambs with the upper lip anaesthetized with xylocaine needed longer to touch the udder and suck. In summary, visual stimuli guide the initial approach to the mother (Vince *et al.*, 1987). Smell may not be as important for the lamb at that age as it is for the ewe. Initially, "teat seeking" is directed at any ewe, even though the lamb is able to discriminate the odor of its mother from that of other ewes (Vince and Ward, 1984). The smell of a ewe's *inguinal wax* stimulates the lamb's nosing, munching, sucking, and bunting (Vince and Ward, 1984; Vince and Billing, 1986). Calves of free-ranging Labrador Caribou are attracted to the fresh urine of their mothers (Müller-Schwarze and Müller-Schwarze, 1985).

Fecal odors

Feces may also be a source of maternal odors. Domestic piglets prefer maternal feces odor to water, but they do not respond differently to fecal odors of their mother and other lactating or non-lactating females (Morrow and McGlone, 1987). In the laboratory rat, the odor of caecal material from the mother attracts the young once they are mobile. The diet of the mother affects the odor. It is also diet specific: pups will even be attracted to the caecal odor of another lactating female as long as she is on the same diet as their mother. Stimulated by prolactin, lactating females consume more food and water and thus produce more caecal material. This is released to the environment, as lactating rats do not eat caecal material. At weaning, young rats prefer the diet of their mother even if they had been exposed to maternal diet signals only before the time of weaning (Leon, 1975).

Amniotic fluid

Chemical stimulation of a young mammal starts already *in utero*. Amniotic fluid is highly attractive for rat pups. This response is not necessarily

acquired during perinatal exposure to this fluid. Rat pups delivered by Cesarean section with no *ex utero* experience did prefer their own mother's amniotic fluid over that of other females (Hepper, 1987). Amniotic fluid is considered important in initial bonding between mother and offspring.

Stress and fear in rodent pups

Laboratory mouse pups show signs of stress, such as higher rates of ultrasonic calls, when the odor of the mother is removed by exchanging the mother's litter with clean bedding (D'Amato and Cabib, 1987).

Some experiments have failed to demonstrate an effect of maternal odors on stressed offspring. The heart rate rose in rat pups removed from their home cage and placed in an unfamiliar environment; this was taken as an index of "fear." Tests with mothers and soiled bedding from mothers or other rat pups showed that tactile and thermal stimuli reduced "fear" in 16-day old pups, but olfactory cues from the mother or odors from the home cage did not (Siegel *et al.*, 1988).

6.5.2 Filial odors: chemosignals from offspring

Olfactory stimuli emanating from the young guide initial bonding, subsequent recognition, and acceptance for nursing in a variety of mammals.

Genital stimulation during parturition stimulates many aspects of maternal behavior, including attraction to amniotic fluid and forming a selective bond with an alien lamb. This is possibly mediated by stimulation of oxytocin release, and/or activation of afferent noradrenergic pathways in the olfactory bulbs (Poindron *et al.*, 1988).

In domestic sheep, the *amniotic fluid* on the coat of the neonate is important for establishing maternal behavior, but only for inexperienced ewes. Washing of the coat of newborn lambs reduces licking and acceptance at the udder by primiparous ewes and increases aggressive behavior by the mother. Multiparous females, however, did not need amniotic fluid for acceptance or for less-aggressive behavior. They merely licked the lamb less (Levy and Poindron, 1987). In dogs (Dunbar *et al.*, 1981) and rats (Kristal *et al.*, 1981), among other species, the amniotic fluid is also important for establishing the first contact between mother and newborn.

The mother's responses to her offspring's odors may have far-reaching consequences for the young later in life. For instance, if a male rat pup is perfumed on his anogenital area, the mother will lick his anogenital area less. Such males

later show diminished "masculine" behavior such as mounting and intromission. Licking of the anogenital area of pups by the mother is assumed to produce greater genital sensitivity to peripheral feedback when adult (Moore, 1984; Birke and Sadler, 1987a).

Cross-fostering

Since time immemorial, animal breeders have had to cross-foster motherless lambs or calves, or had to attach newborn mammals to a mother of a different species. They have been aware of odor barriers and developed methods to overcome them. A ewe will accept a non-related lamb if it has been rubbed with the hide or amniotic fluid of her own, perhaps stillborn, lamb. A classical case of successful cross-fostering between species is a technique employed by Peruvian livestock breeders to produce hybrids between alpacas and vicuñas. The cross is called *paco-vicuña* and combines the large quantity of wool of the alpaca with the fine quality of vicuña hair. To breed an alpaca female with a vicuña male, first a male has to be imprinted on alpacas. A newborn male vicuña is covered with the hide of a newborn alpaca and presented to a lactating female alpaca without young. The young vicuña is accepted and nursed on account of his alpaca odor. Successfully raised by his alpaca mother, he will imprint on, and breed with, alpacas when adult.

Recent experiments have confirmed these practitioners' experiences and revealed the precise mechanisms better. A ewe accepts an alien lamb if it wears a stockinette that had been worn before by her own lamb. The salient cue for acceptance is the odor of her own lamb rather than absence of alien odor (Price et al., 1984). A ewe will even accept an additional lamb if it smells like her own lamb. Such "add-on experiments" work better with primiparous ewes than with multiparous ones. To collect the odor, the own lamb wore a stockinette for 20 hours after birth. The stockinette was then pulled over the alien lamb. It should be noted, however, that visual cues such as the color of the lamb's head also influence the ewe's choice (Martin et al., 1987).

In some species, bonding depends solely on the *mother's* ability to recognize the odor of her pup, as for instance in the Mexican bat, *Tadarida brasiliensis mexicana*. In this species, the pups do not discriminate odors of their (presumed) mother and other lactating females but mothers discriminate well between the pup they nurse and other pups. They also distinguish the odor of their own muzzle glands from that of randomly selected lactating females. It is possible that in this bat species mothers mark their young with secretion from their muzzle glands (Gustin and McCracken, 1987).

FIGURE 6.2 Structure of dodecyl propionate, found in the preputial gland of rat pups.

Other olfactory effects on maternal behavior

During lactation, female rats eat more, are aggressive toward adult conspecifics, and are less fearful than usual. Lesions of the peripheral and central olfactory system interfere with these behaviors. If the olfactory epithelium is ablated, lactating females eat less, weigh less, and maternal aggression decreases. However, "fear" (i.e. freezing in response to a sound) is not affected by this treatment. Centrally, the olfactory system involves brain structures such as the mediodorsal thalamic nucleus and the prefrontal insular cortex. Thalamic and cortical lesions in these regions lower the frequency of attacking male intruders, but eating behavior and fear responses are left intact (Ferreira *et al.*, 1987).

Behaviorally active chemical compounds

Dodecyl propionate (Fig. 6.2) from the preputial gland of rat pups attracts the mother. She responds by sniffing and licking the secretion, resulting in grooming and stimulating of her young (Brouette-Lahlou *et al.*, 1991).

Paternal behavior

Chemosignals from mate

In the California mouse, *Peromyscus californicus*, both sexes participate in taking care of the young. A chemosignal in the mother's urine maintains parental behavior in the father. This paternal behavior consists of licking the pups and crouching over them in a "nursing position." The active principle resides in the volatile fraction of the female's urine. The paternal behavior was maintained by an experimental dose of 100 μ l urine given on the male's nares twice per day (Gubernick, 1990).

In Mongolian gerbils (M. unguiculatus; a "biparental" species), males avoid their newborn sons until they are 3 days old. The pups' surge of testosterone around birth is seen as the proximate cause for this behavior. The males appear to respond to an odor from the neonates, because if rendered anosmic by zinc sulfate infusion they do not avoid neonatal pups. The male faces conflicting

demands: either attending to the pups or mate-guarding a female during her postpartum estrus. A male engaged in fathering will miss the estrus and mating will be delayed by at least 30 days. It is not clear why only sons have this effect (Clark *et al.*, 2003).

6.6 Species and population discrimination

Discrimination of the own species from other closely related and sympatric species is essential not only for reproductive behavior but also in the contexts of competition for resources and antipredator behavior.

6.6.1 Fish

Juveniles (elvers) of the European eel (*Anguilla anguilla*) migrate upstream in rivers. To test whether they might follow conspecific odors, elvers were taken from the Arno river in Italy and exposed to water in which other juvenile eels had lived for 8 months. They were attracted to this stimulus much more than to uncontaminated water or water from mosquito fish (Pesaro *et al.*, 1981).

Young (i.e. 2-summers old) Arctic charr, *S. alpinus*, are attracted to odors of conspecifics. These odors include those of intestinal contents, fed and starved fish, and urine (Olsén, 1987). In a T-maze fluvarium, fry of Arctic charr of one population was attracted to water from a tank with conspecific fry, while those of another population were not (Olsén, 1990). Juvenile coho salmon, *O. kisutch*, from British Columbia, Canada discriminate between chemical cues from similarly aged conspecifics from their own and a different population. Some experimental results suggest that the odor emanates from the feces. This discrimination of population odors may play a role in imprinting for homing, sibling recognition, or mate choice (Courtnay and Masel, 1997).

Blind cave fish are attracted to water in which conspecifics had lived. This is true for the Congo blind barb, *Caecobarbus geertsi*, and the Somalian cave fish, *Phreatichthys andruzzi* (Berti *et al.*, 1982). The population density can be critical for the strength of a produced chemosignal. The blind cave fish *Astyanax mexicanus* (formerly *Anoptichthys antrobius*; Characidae) is attracted to water in which conspecifics had been. Tested with water from groups of 4, 8, 16, and 32 fish, the response was the stronger the more fish served as stimulus. All other factors, such as familiar versus alien group, did not matter (De Fraipont, 1987). However, test fish may prefer the odor of known individuals, but only for 2 minutes. After that time, the movements of the test fish become random (De Fraipont and Thines, 1986). The Mexican blind cave fish *Astyanax jordani* spends more time in the

section of a tank that had been occupied by a conspecific. In the presence of conspecific odor, locomotor activity is reduced in this species (Quinn, 1980).

6.6.2 Amphibians

Courtship pheromones are not necessarily species specific. Pairs of the woodland salamander, *Plethodon shermani*, courted for an equally long time (about 35 to 50 minutes) whether male pheromone from the mental gland of conspecifics or the allopatric species *P. montanus* or *P. yonahlossee* was present, even though the composition of the proteinacous pheromones (plethodontid receptivity factor of these three species differ considerably (Rollmann *et al.*, 2003).

6.6.3 Reptiles

A scincid lizard species, *Eumeces laticeps*, can distinguish the cloacal odors of males of conspecifics from those of *Eumeces fasciatus*. When the odor is presented on cotton-tipped applicators, the lizards flick their tongue more often to their own species' odor. The adaptive advantage of interspecific discrimination may be to recognize sexual rivals or to avoid injury in interspecific fighting (Cooper and Garstka, 1987).

Garter snakes identify their species by tongue flicking at non-volatile, integumentary lipids which they also use in courtship and following conspecific trails. The levels of these lipids fluctuate with hormonal state, skin-shed state, and season (Mason, 1992).

Aggregation pheromones belong in this context. Garter snakes produce an aggregation pheromone in their skin that chemically is a cholesterol ester (Devine, 1977). These snakes can find and follow each other by depositing *trailing* pheromones on vegetation or soil. These have been considered functionally different from the "sexual attractiveness pheromone" (Ford, 1978, 1981, 1982; Garstka and Crews, 1986).

6.6.4 Birds

Social odors have rarely been reported in birds. The respiration rate of wedge-tailed shearwaters (*Puffinus pacificus*) increases in response to the odor of an unfamiliar conspecific (Shallenberger, 1975). Domestic ducks altered social and sexual behavior after bilateral section of the olfactory nerve, or after treatment with amyl acetate or ethyl acrylate (Balthazart and Schoffeniels, 1979). Some bird species are known for their strong characteristic odors. For instance,

the hoatzin, *Opisthocomus hoazin*, smells like cow manure (Grimmer, 1962). Oilbirds (*Steatornis* sp.) and hoopoes (*Upupa epops*) are further examples of "smelly" birds. The functions of these odors are still unknown.

6.6.5 Mammals

Mammals easily discriminate the odor of conspecifics from odors of other species.

Investigators have searched for specific compounds that signal the species, while groups of other constituents form the "fingerprint" for individual recognition. The Eurasian otter, *L. lutra*, may provide an example. Its anal sacs contain a secretion with many compounds. One of these, less volatile and hence long lasting, occurred in nearly all individuals, making it a candidate for the signal "otter." The proportions between other compounds varied between individuals and remained constant for 25 days for each individual (Trowbridge, 1983).

Among primates, females of the saddleback tamarin, *Saguinus fuscicollis*, discriminate conspecific scent marks from those of other species or subspecies (Epple *et al.*, 1988), based on a complex mixture of compounds (p. 168). Likewise, *L. fulvus* and *L. macaco* distinguish scent marks by species and "show more interest" in the other species' odor than their own (Fornasieri and Roeder, 1992).

Mole rats of the superspecies *Spalax ehrenbergi* occur in four main chromosome forms: 2n = 52, 54, 58, and 60. Females of two of these forms (52 and 58) were given choices between soiled bedding (or urine) from males of a homochromosomal or a heterochromosomal form. The females were estrous or diestrous. Only estrous females preferred soiled bedding and urine of homochromosomal males, measured in time spent near the odor samples. Diestrous females showed no preference (Nevo *et al.*, 1976).

6.7 Modulating behavior by status signals

In the second part of this chapter, we discuss communication by pheromones and pheromone-like chemicals in the context of competition, aggression, dominance, and territorial behavior.

6.8 Competition between conspecifics of the same sex

Competition between conspecifics of the same sex may either lead to spacing, often as territorial behavior, or dominance orders among animals that stay together, at least temporarily.

6.8.1 Amphibians

Some salamanders show signs of odor-mediated spacing mechanisms. In Western red-backed (lungless) salamanders, *Plethodon vehiculum*, both sexes avoid the odor of males in a choice test. Ovaska (1988) suggested that the "males use pheromonal markers to space themselves out for mating purposes." Some evidence suggests that male red-spotted newts, *Notophthalmus viridescens*, produce a repellent pheromone that inhibits other males from approaching a cluster of several males courting a female (Park and Propper, 2001).

6.8.2 Mammals

In mammals, dominant individuals typically scent mark more – or in a more effective manner – than subordinate ones.

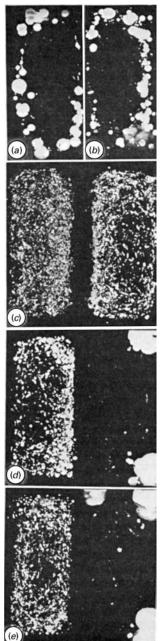
Marsupials

Dominance status information, coded in whole-body odor, can travel between animals in an air stream. When exposed to the odor of a familiar, dominant male, the sugar glider, *P. breviceps*, increases cardiac and respiration rates within 10 minutes, and levels of glucose and catecholamine in the plasma rise after 30 minutes (Stoddart and Bradley, 1991).

Rodents

In caged laboratory mice, dominant males mark over their entire area, while subordinate neighbors, separated behind a partition, urinate only in the corners (Desjardins *et al.*, 1973; Fig. 6.3). In wild house mice, *M. musculus*, territorial males advertise their "aggressive dominance over other resident and intruder males" by vigorous marking, while subordinate males investigate the dominants' marks. Both use the marks of the dominant males as beacons to orient to their own territory and to avoid areas marked by other dominant males (Hurst, 1990a). Subordinate males also contribute to the communal odor of a group's substrate. A subordinate that is experimentally prevented from contributing his odor, will be attacked more by the dominant and the other subordinate males of the group (Hurst *et al.*, 1993).

One of a pair of fighting mice will be more "confident" if his feces are present: if two male house mice (M. musculus domesticus) are placed together in a container along with fecal pellets from one of them, the donor of the fecal pellets shows more and longer aggressive contacts and succeeds more often in



dominant and subordinate adult male mice. (*a*, *b*) Two individually isolated males (without immediate neighbors) urinate along the periphery. (*c*) The same males, but in one cage, only separated by a wire mesh partition. They saturate the entire area with small marks. (*d*) The same animals after an aggressive encounter established the male on the left as dominant. (*e*) The same animals, after five aggressive encounters. The dominant mouse on the left continued to distribute urine over the entire cage floor, while his subordinate neighbor voided urine in corners only. (From Desjardins *et al.*, 1973.)

aggressive encounters (Goodrich *et al.*, 1990). Male mice advertise their dominance by urine marks. In staged encounters between familiar, unfamiliar, dominant, and subordinate males, a urine mark was provided. The dominance status of the urine donor influenced the outcome of the encounter (Hurst, 1993). Male mice also over-mark female urine marks heavily (Hurst, 1989), as is common in ungulates (see below).

Female mice spend more time with the odors of winning male mice than with odors of losers. Even more remarkably, they also spend more time with the odors of the sons of winners than with the odors of sons of losers (Drickamer, 1992).

In other rodents, subordinate males also smell scent marks quite often and so keep informed on the presence, status and activities of higher-ranking group members. For instance, dominant males of the hispid cotton rat, *Sigmodon hispidus*, urine mark more than subordinates. The social status of the male urine donor affects the response of other males to the odor. The response of a reproductive female to feces of either sex depends on her dominance status (Gregory and Cameron, 1989).

Dominant male bank voles (*Clethrionomys glareolus*) over-mark urine and fecal marks from strange dominant males, hierarchically naive males, and sexually unstimulated virgin females. They sniff urine but handle and lick (possibly mask the odor of) feces. Dominant males also scratch their flanks and drag their genitals near conspecific marks, and more so at marks of unknown dominant than unknown naive males. Consequently, in this species, chemical signals provide information on the hierarchical background of an individual in addition to species and sex. Furthermore, bank voles may mask conspecific odors with saliva and specific skin gland secretions (Rozenfeld and Rasmont, 1991).

In the golden hamster, *M. auratus*, both dominant and subordinate males use the flank glands to *communicate* their social status to inhibit overt aggression during encounters (Ferris *et al.*, 1987). However, they do not need their flank glands to *develop* dominant/subordinate relationships.

The dominance status of an individual can be predicted by its scent-marking rate before social interactions take place: Woodchucks (*Marmota monax*) scent mark with their oral glands. When presented with isolated secretion of the oral gland of other woodchucks, future subordinates marked the scent of future dominant animals more often than vice versa (Hébert and Barette, 1989).

Lagomorphs

Rabbits, *O. cuniculus*, "chin-mark" (Fig. 6.4) near their warren entrances and at boundaries to neighboring groups. Only the dominant male marks. This has been demonstrated by comparing gas chromatograms of the chin gland



FIGURE 6.4 A domestic rabbit scent marks by "chinning". (Photograph: R. Mykytowycz.)

secretion of males with those of "environmental" scent marks in the territory of these males. All scent marks contained 2-phenoxyethanol. This compound is typical for dominant males. Furthermore, the scent from the chin gland of the dominant male can be chromatographically found on the forehead of subordinate males and of females, as the dominant buck marks these group members (Hayes *et al.*, 2002). Such "allomarking" is thought to produce a group odor, an olfactory membership badge. However, specific individual relationships between the dominant male and other individuals may also be "recorded" and broadcast this way.

Ungulates

The scent marking of male feral boar, *Sus scrofa*, with metacarpal and preputial glands varies with rank and the social environment. The dominant boar marked more often than the subordinate one when paired, but did not mark at all when alone (Mayer and Brisbin, 1986).

Males of equids urinate and defecate over female urine. In this way, males possibly advertise themselves to other males as a dominance display (Klingel, 1974), while Trumler (1958) proposed that the stallions conceal estrus odors of females from other males. Pronghorn bucks, *A. americana*, cover spots of female urine on the ground in a stereotyped sequence of sniffing, pawing, urination, and defecation, but only when no other bucks are present. This supports the concealment hypothesis (Moodie and Byers, 1989). Males of several other species also cover female urine with their own urine or feces. This is true for blackbuck,

Antilope cervicapra (Dubost and Feer, 1981), dik-dik, Madoqua kirki (Hendrichs and Hendrichs, 1971), muntjacs, Muntiacus reevesi (Barrette, 1977), and ponies, Equus caballus (Feist and McCullough, 1976).

During the rut, a dominant muskox (*Ovibos moschatus*) performs a "superiority display": he tilts his head and walks slowly past his rival in a stiff-legged gait, with the prepuce everted as a 12cm long pendulous tube. The tube swings about and urine dribbles from the opening. The long belly hair is soaked, emitting a typical strong rutting odor. Washings of the preputial gland contain large amounts of benzoic acid and p-cresol. By contrast, the secretion of the muskox infraorbital glands has a "light sweetish, ethereal smell." It contains cholesterol, benzaldehyde, and a homologous series of saturated γ -lactones ranging from 8 to 12 carbons. These lactones smell like the natural secretion (Flood et al., 1989).

6.8.3 Active chemical compounds in mammals

Two compounds in the urine of male mice trigger attacks by other males: 3,4-dehydro-*exo*-brevicomin and 2-*sec*-butyl-4,5-dihydrothiazole (Novotny *et al.*, 1985b). The behavior of male mice depends on the context in which they encounter these compounds. When smeared on castrates or females, they release aggressive behavior, but applied to pups, they inhibit infanticide. Therefore, it appears to be more appropriate to consider the these as signals of "maleness" rather than "aggression pheromones" (Mucignat-Caretta *et al.*, 2004).

The concentrations of 16 constituents of male mouse urine vary with the male's dominance status. Dihydrofurans, ketones, and acetates decreased in subordinates. Two sesquiterpene compounds, α - and β -farnesene, are elevated in dominants' urine 1 week after establishing dominance. The bladder or voided urine of dominants contains more 2-sec-butyl-4,5-dihydrothiazole. Four compounds depend on hormones: α - and β -farnesene, dehydro-exo-brevicomin, and 2-sec-butyl-4,5-dihydrothiazole. The latter two are absent in urine of immature or castrated males, and testosterone treatment restores their presence. In addition, α - and β -farnesene do not occur in urine of immature males and are merely reduced in urine of castrates. They are not found in bladder urine and originate in the preputial glands (Harvey et al., 1989). While subordinate male mice have reduced levels of farnesenes, levels of their major urinary proteins remain high (Malone et al., 2001).

Dominant males of the European rabbit, *O. cuniculus*, have 2-phenoxyethanol in their chin gland secretion. Behaviorally subordinate males lack this compound. When a subordinate becomes dominant after removal of the originally dominant male, 2-phenoxyethanol starts showing up in his secretion. The perfume industry uses this compound as a fixative. Rabbits perhaps also employ this

alcohol as a fixative to keep alive the dominant's odor in the environment (Hayes *et al.*, 2001).

6.8.4 Females

Rodents

Female wild house mice mark frequently to advertise their dominant breeding status to other females. Resident breeding females overmark the urine of other breeding females (Hurst, 1990b).

Female mice "prefer" odors of dominant males over those of submissive males. Such preferences disappear if the dominant male is preputialectomized. A male mouse needs rivals to be aggressive, which, in turn, maintains his preputial gland and hence his attractiveness to females (Hayashi, 1987). Social dominance can be influenced by strange male odor (Hayashi, 1989). In bank voles (*C. glareolus*) the effect of urine odor of a familiar female depends on the status of the receiving female. Before the receiver's pregnancy, the odor inhibits aggression; later in pregnancy and early in lactation it releases aggressive behavior and scent marking. This is thought to aid in spacing (Rozenfeld and Denoël, 1994).

Ungulates

In female cattle, olfactory stimuli are more important than visual cues for dominance relationships. In one experiment, the 10 most dominant of 30 Holstein cows were sprayed with anise oil, or painted. They were kept apart from the herd for 2 hours and then reintroduced. Visual alteration had no effect. However, olfactory alteration resulted in less interaction with other cows in the group. Specifically, other cows investigated the altered individuals *less* and reduced their submissive behavior toward them (Cummins and Myers, 1988).

Here's the rub: "dominance areas" as transition between dominance and territory

White-tailed deer, *Odocoileus virginianus*, bucks compete with one another by three behaviors: they rub the bark off tree saplings with the base of their antlers, scrape the ground at various places in their home range during the rutting season (Fig. 6.5), and rub their muzzle and forehead on overhanging dry twigs while the antorbital gland opens, and its secretion is transferred to the substrate (Fig. 6.6 shows this for a male black-tailed deer). The first two, rubs and scrapes, can occur together or separately. Bucks also defend the area



FIGURE 6.5 A "rub" on a sapling and "scrape" on the ground made by a white-tailed deer buck during the rutting season. Such sites attract females and are thought to constitute a "dominance area." (Photograph: D. Müller-Schwarze.)

around estrous females (Moore and Marchinton, 1974). Only males older than 2.5 years regularly maintain the scrapes and rubs. One male may distribute scrapes and rubs over an area as large as 4 miles long and 1.5 miles wide (Moore and Marchinton, 1974).

6.9 Liquid assets: marking territory and home range with urine and secretions

6.9.1 Fish

Even fish may mark their substrate. Juvenile Atlantic salmon (parr) are attracted to extracts from gravel over which salmon parr had been reared. They

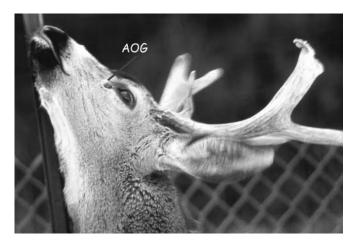


FIGURE 6.6 Secretion marking in the black-tailed deer. A male scent marks by rubbing the side of his head against a post while opening the pouch of the antorbital (AOG) (or preorbital) gland. (Photograph: D. Müller-Schwarze.)

preferred the odor of their own strain. It is assumed that fecal material serves as "scent marks" in this species (Stabell, 1987).

6.9.2 Amphibia

Amphibians appear to use territorial pheromones. The best-investigated species is the red-backed salamander, *P. cinereus*. Females deposit scent on fecal pellets. They are more aggressive to fecal pellets of other females than males are to fecal pellets of other males. These pellets may advertise territories (Horne and Jaeger, 1988). Territory owners of this species use pheromones as the first line of defense, followed by aggressive and submissive signals to intruders, and finally by biting and expelling intruders (Jaeger, 1986).

Salamanders of the ambystomatid family may also use territorial marks. In contrast to red-backed salamanders, which are *repelled* by conspecific odors or fecal pellets (Jaeger and Gergits, 1979; Jaeger, 1986), spotted salamanders, *Ambystoma maculatum*, are *attracted* to paper towels on which conspecifics had lived for 4 days, whether the latter were familiar or unfamiliar (Ducey and Ritsema, 1988).

6.9.3 Reptiles

Reptiles advertise their territories mostly by visual displays, but some olfactory marking may occur. For instance, western fence lizards, *Sceleporis* sp., of North America deposit fecal boli in prime basking sites, which are important

for thermoregulation. These fecal boli provide chemical as well as visual cues. Lizards are still attracted to boli that are covered with translucent plastic. The boli possibly signal that the area is occupied, the sex of the marker, breeding or non-breeding status, and the individual's resource-holding power. It is an energetically inexpensive way to maintain a territory (Duvall *et al.*, 1987).

6.9.4 Birds

Birds do not usually rely on scent for territorial marking. Some species, however, such as burrowing owls, apply odoriferous substances to their nest cavities or burrows. The functions are not well understood. Non-avian predators are thought to be the addressees, placing such marking outside the intraspecific behavior of this chapter.

While birds or nests in a number of species are "malodorous," the question is: does any bird "mark" in an area outside its nest, and do conspecifics respond to this chemical cue? Candidates are the "mimuyi," deposits of stomach oil near the nest of Antarctic and snow petrels that have been dated as 9000 and 4000 years old (Hiller and Wand, 1984). Petrels spray stomach oil at intruders as far as 2 m. However, most investigators see bird odors as antipredator, rather than territorial.

6.10 Scent marking in mammals

Scent marking for demarcation of territory or home range of individuals or groups is the hallmark of mammals and this will be discussed in some detail below. Depending on the social organization, scent marking in mammals can take many different forms. Solitary species, such as musk deer, *Moschus* sp., aardvark, *Orycteropus afer*, or aardwolf, *Proteles cristatus*, for example, simply cover their feces with soil or leaves. Pangolins, *Pangolinus arboricoles*, maintain individual distances by means of scent marks (Pages, 1972). Many social species, by contrast, mark in elaborate behaviors with complex or specific spatial and temporal patterns. Examples are the well-studied hyenas and canids such as wolves and coyotes.

6.10.1 Territorial marking in carnivores

Hyenas: a case study in territorial marking

The four species of hyena all scent mark plants with anal gland secretion ("pasting"). Different species concentrate their scent marks in different parts of their territories. In the resource-poor Kalahari, both brown (*Hyaena brunnea*) and

spotted (Crocuta crocuta) hyenas maintain large group territories of up to 500 and 1000 km², respectively. Given their average group sizes of 1 to 9 and 3 to 15 individuals, respectively, each individual would have to mark between 7 and 37 km border on average, were they to concentrate their scent marks at the periphery. This would strain their time and energy budget. Instead, these two species increase their marking from the edge toward the center of their territory where they are active anyway. The striped hyena of the Serengeti, a solitary species, also practices such hinterland marking. With a territory size of 40-70 km², the single animal would have to renew scent marks along a 22-30km circumference. By contrast, border marking is feasible in smaller territories. The spotted hyenas of the Ngorongoro crater live in much larger groups (30–80) than the same species at Kalahari. In addition, their territory is rather small (about 30 km²). They have only a quarter to two thirds of a kilometer circumference to mark per group member, on average. Indeed, they mark mostly at the border. Finally, the aardwolf, P. cristatus, lives alone or in pairs in a small territory (1.5 km²). This requires only 1.7 to 3.4 km periphery to mark per individual. Consistent with the assumption, the aardwolf follows a border-marking strategy (Gorman and Mills, 1984).

The brown hyena has been studied in detail. Its scent mark, typically on a grass stalk, consists of two parts: an upper black watery portion that loses its odor quickly, and a lower, lipid-rich white part (Fig. 6.7) the odor of which humans still detect after 30 days. The mark is placed at nose level of hyenas, ensuring that newcomers will encounter them easily. Hyenas travel on average 30–40 km per night and paste at a rate of 2.64 marks/km. In the course of a year, an individual may produce 29 000 paste marks, a considerable energetic cost, indicating an important function (Mills *et al.*, 1980).

Hyenas also maintain latrine sites that accumulate 5–50 feces. While the latrine sites are concentrated near, but not on, the border, the paste marks become denser toward the center of the territory, the main area of hyena activity. The driving force for the pasting pattern is not primarily the need to signal at the periphery, which is met by the latrine sites, but the impossibility to mark regularly the long border of a huge territory, as mentioned above (Gorman and Mills, 1984). Would mammals in such situations develop chemical marks that stay fresh longer?

Canids: a second case study in territorial marking

Urine marking by canids has received much attention. Wolves, *Canis lupus*, mark by raised-leg urination (RLU), squat urination (SQU), defecation, and scratching. In Minnesota's Superior National Forest, wolf packs maintain

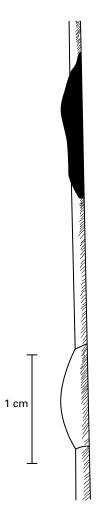


FIGURE 6.7 The double scent mark of the brown hyena. The black mark on top originates from apocrine glands and the white mark from separately located sebaceous glands. (Redrawn from Mills *et al.*, 1980.)

territories that range in size from 125 to 310 km². Wolves RLU-mark conspicuous objects in their territory such as blocks of snow, trees, rocks, or snow banks. They re-marked the same spots even though their own odor had not faded completely, nor had other wolves marked there. Fresh marks (2–6 days old) stimulate more marking than those 8 or more days old. Located along trails (Fig. 6.8), scent marks will be encountered often by resident wolves, informing them if they are still in their own territory or reaching the boundary zone with another pack's territory. From the age of a scent mark, wolves can tell how recently they have been in an area. Temporarily single pack members may also read from urination

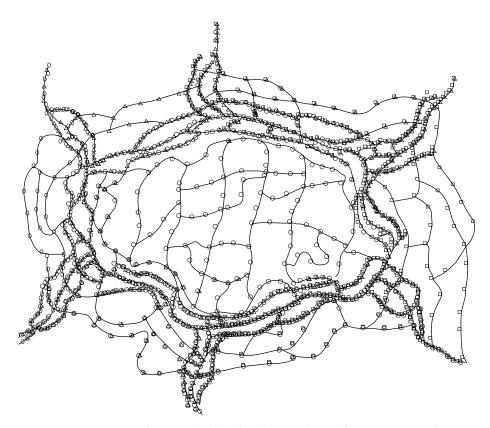


FIGURE 6.8 Schematic sketch of distribution of raised-leg-urination marks in a 20 km wide territory of a wolf pack. Dots are urine marks, lines are trails. Higher marking density along the periphery creates a bowl-shaped pattern. Six neighboring pack territories are indicated by dots in different colors. (From Peters and Mech, 1975.)

and defecation marks whether other pack members have hunted an area recently, if a member is nearby, or which animals are traveling together. Wolves appear to avoid unfamiliar marks as an integral part of unfamiliar terrain (Peters and Mech, 1975).

Urine marking in coyotes, *Canis latrans*, resembles that of wolves. Males also mark with RLU (Fig. 6.9). In a study at Grand Teton National Park, Wyoming, RLU was associated with courtship and mating, traveling, and aggression. Urine marking becomes more frequent during the breeding season (December to February), peaks in March, and reduces in April when the pups are born. Females squat urinate (SQU) year round and this is associated with acquisition and possession of food; it is particularly frequent during the denning season. The marking rate is higher in groups of two or more coyotes. Marking with RLU is higher



FIGURE 6.9 A signature urine mark (left center) on a snow bank made by a coyote. To the right, the animal has defecated, pawed, and partially covered the feces with snow. (Photograph: D. Müller-Schwarze.)

in areas of high rates of intrusion than near the denning area or in areas with less-frequent intrusions. Marking by SQU was most frequent in both denning areas and high-intrusion areas. It was concluded that "scent odours are important in orienting individuals in space but do not represent in and of themselves barriers to movement" (Wells and Bekoff, 1981).

Specialized site creation for marking: beavers

Unlike most terrestrial mammals, beavers maintain territories along a line following watercourses. Both species, the Eurasian beaver, *C. fiber*, and the North American beaver, *C. canadensis*, dredge up mud from the bottom of their home pond, carry it on land, deposit it, and apply a scent mark on top of this mud

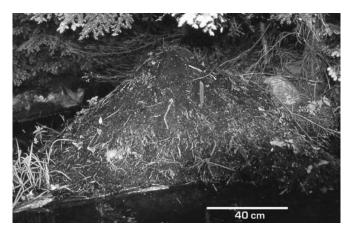


FIGURE 6.10 A giant beaver scent mound. (Vertical kitchen knife, center, for scale). (Photograph: D. Müller-Schwarze.)

pile. The mark on these scent mounds consists of castoreum, a urine-derived liquid from the castor sacs, and secretion from the anal glands, also known to trappers as oil sacs (Fig. 6.10).

Scent mounds of North American beavers are concentrated at strategic points in a beaver territory, such as shoreside trailheads of paths leading to feeding grounds. This suggests that beavers might use these scent mounds as "odor beacons" during the night to find their currently active feeding trails. Eurasian beavers make smaller mudpiles or mark directly on rocks or wood. Their scent marks are often concentrated at common boundaries between territories.

It is less likely that beavers will colonize unoccupied beaver sites if these sites are artificially scented with castoreum and anal scent secretion (Welsh and Müller-Schwarze, 1989).

6.10.2 Spatial distribution of scent marks in territory or home range

As we have seen in the carnivore examples, the spatial pattern of scent marking varies between and within species. These comparative studies hint at the function(s) of scent marking, even though we are far from understanding its precise mechanisms. Some species mark throughout their area; some mark more at or near the periphery, while others concentrate their marks around centers of daily activity, or at areas of contact (or conflict) with neighbors or transients, and still others combine some of these patterns (Table 6.2). The scent-marking pattern can vary within a species according to ecological differences, as in the spotted hyena.

Table 6.2 Distribution of scent marks in mammalian territories

Territory area marked by species	Behavior, secretion	Reference
More at periphery		
Wolf Canis lupus	Urine marks: RLU	Peters and Mech, 1975
Coyote Canis latrans	Urine marks: RLU	Bowen, 1978
Red fox Vulpes vulpes	RLU	White et al., 1989
Spotted hyena Crocuta crocuta	Feces and AGS	Kruuk, 1972
Brown hyena Hyaena brunnea	Pasting	Gorman and Mills, 1984
Aardwolf Proteles cristatus	Latrines	Richardson, 1990
Water deer Hydropotes sp.	Pasting, feces, urine	Sun et al., 1994
Beaver Castor fiber	Feces, urine, castoreum, AGS	Rosell and Nolet, 1997
More at contact zones		
Tiger Panthera tigris	Urine, AGS	Smith et al., 1989
Wild European Rabbit	Chin gland secretion	Hayes et al., 2002
Oryctolagus cuniculus		
More in center		
Brown hyena H. brunnea	Pasting	Gorman and Mills, 1984
Woodchuck Marmota monax	Oral angle glands	Ouellet and Ferron, 1988
Gerenuk Litocranius walleri	Antorbital secretion	Gosling, 1981
Honey badger Mellivora capensis	Token urination, latrines	Begg et al., 2003
More at "activity centers"		
Beaver Castor canadensis, C. fiber	Castoreum, AGS	Müller-Schwarze and
		Heckman, 1980; Rosell
	al. 1 1 .	and Nolet, 1997
Wild European rabbit O. cuniculus	Chin gland secretion	Hayes <i>et al.</i> , 2002
Entire area		
Wolf C. lupus	RLU	Paquet and Fuller, 1989
River otter L. lutra	Feces, urine, AGS	Trowbridge, 1983,
Thomson's gazelle Gazella thomsoni	Antorbital secretion	Walther, 1978
Vicuña V. vicugna	Dung piles: feces and urine	Franklin, 1983

RLU, raised leg urination; AGS, anal gland secretion.

Woodchucks, *M. monax*, like other sciurids, possess oral angle glands and rub their muzzle on objects such as fences, woodpiles, shrubs, rocks, or burrow mounds. The vast majority (95%) of the scent marks are within 6 m of the burrow. Males and females mark equally often. While social interactions are not immediately associated with marking, sighting of a conspecific, with or without interaction, "may release scent marking." Woodchucks also mark when moving into a

different burrow. These observations suggest that scent marking in woodchucks advertises that a burrow is occupied. It is unclear whether marking is directed at self or conspecifics (Ouellet and Ferron, 1988).

Free-ranging Mongolian gerbils, *M. unguiculatus*, mark more at territorial boundaries, plus at their burrow entrances (Ågren *et al.*, 1989). Similarly, rabbits, *O. cuniculus*, mark throughout their territory but concentrate their scent marks at the center of a group's area, and at boundaries with other groups' territories (Hayes *et al.*, 2002).

Tigers of both sexes mark more at contact zones (Smith *et al.*, 1989), and wolf packs mark more frequently by RLU at the edge of their territory than in the center (Peters and Mech, 1975). Captive red foxes, *Vulpes vulpes*, deposited fecal pellets with anal gland secretion more frequently in the perimeter strip of a 4 ha pen (White *et al.*, 1989).

Thomson gazelles, *Gazella thomsoni*, mark the entire territory. An 8000 m² large territory had 18 dung piles and 110 preorbital marks. Dung piles were most frequent in border sections where there were frequent agonistic encounters with neighboring territory owners. In the center of the territory was only one dung pile, near a bedding site. The preorbital marks were arranged in a "broad belt" around the territory. A central area remained unmarked. Walther (1978) concluded that the scent marks were more for the owner's own orientation than for territorial defense.

Another antelope, the gerenuk (*Litocranius walleri*) marks an oval-shaped line in only the core area of its territory, with lines of scent marks extending from it radially like spokes in a wheel. This ensures that intruders will encounter some scent marks, no matter from what angle they approach (Gosling, 1981). Similarly, Eurasian otters, *L. lutra*, in coastal Scotland place feces, urine, and anal gland secretion at so-called spraint stations along trails near the ocean at an average rate of 266 marks/km. This insures that strange otters entering the marked area from the sea will soon encounter a spraint site. Inland trails along streams have only 20 marks/km. Here otters are funneled along trails and will easily encounter scent marks. Otters mark throughout their territories, ensuring that other otters will find a spraint station soon (Trowbridge, 1983).

Linearly arranged territories, as those of beaver, *C. canadensis* and *C. fiber*, along a stream require yet another marking pattern. Trespassing conspecifics are expected to arrive along the watercourse, not usually overland. Therefore, if directed at potential invaders, marking is expected to be particularly intense at the up- and downstream ends of the territory, and less along the banks. Eurasian beavers mark indeed more heavily at territorial borders, and especially at the upstream limit of their territory (Rosell *et al.*, 1998).

Since resources are never completely evenly distributed within a territory, resource-related marking in many species will be concentrated in certain areas. Herbivores with vegetation covering most of their territories will mark the periphery and a ring farther "inland," or a core area ("hinterland marking"), if the territory is very large (Roberts and Gosling, 2001). After all, marking costs the animal in terms of producing chemicals, spending time marking (instead of foraging), and risk of predation. Pronghorn, *A. americana*, males undertake regular, exclusive "marking trips" along the periphery of their territory to refresh their subauricular scent marks (Gilbert, 1973) (Fig. 6.11).

6.10.3 Marking and population density

In some rodents, scent marking intensity reflects population density: the more scent mounds a beaver colony has, the more neighboring colonies exist. This is true for both beaver species (Houlihan, 1989; Rosell and Nolet, 1997; Fig. 6.12). The scent marking syndrome (marking behavior plus gland structure, size, and activity) can vary according to population density within a species in one geographical area. The Indian gerbil, *Tatera indica*, one of the predominant rodents in the Rajasthan desert of northwest India, is solitary in scrub grassland but develops dense urban populations. The urban populations have up to 70 individuals in one burrow. This gerbil makes scent marks by a "perineal drag," depositing urine and sebum from the ventral gland. These glands are larger in males than in females. Low-density populations mark more often than the urban ones. Moreover, the gland is present in more animals of the solitary type (91.4% of males, 38.5% of females) than those of the urban, gregarious type (85.6 and 3.2%). Experimental groups of 1, 3, 6, or 12 wild-caught males or females from solitary grassland populations were kept in one cage. The greater the density, the more they fought and chased each other. However, the frequencies of sebum marking, urine marking, and urination declined with density (Idris and Prakash, 1987).

6.10.4 Marking territory with family odors

In mice, urine is the main source of social odors. Wild house mice (*M. domesticus*) families over time build bizarre small posts of solidified urine by repeated marking (Hurst, 1987; Fig. 6.13). A mouse family is habituated to its own background odor, which permeates its living area. The ubiquitous family odor is dominated by the odor of the dominant male and identifies the home area to residents as well as non-residents. (When the author trapped 26 deer mice over





FIGURE 6.11 Marking by a pronghorn buck with his subauricular gland of a thistle in his natural environment (top; Yellowstone National Park) and an experimental Teflon rod (bottom). (Photograph: D. Müller-Schwarze.)

2 weeks in his cabin in the woods, the strong mouse smell disappeared once a big male had been caught.) Any unusual change is detected, and re-marking will restore the familiar background odor. Any novel odor is investigated. However, the urine marks of an unfamiliar family do *not* deter adult males or other mice from invading a new area. Adult males do not investigate, and marked little, the

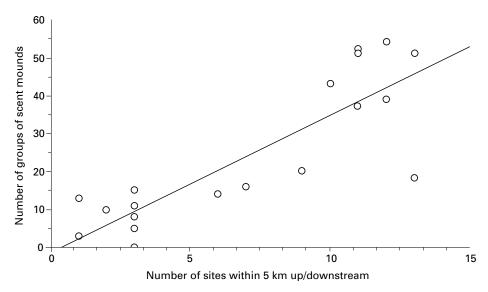


FIGURE 6.12 Correlation between population density and scent marking intensity in the beaver, *Castor canadensis*. (From Houlihan, 1989.)

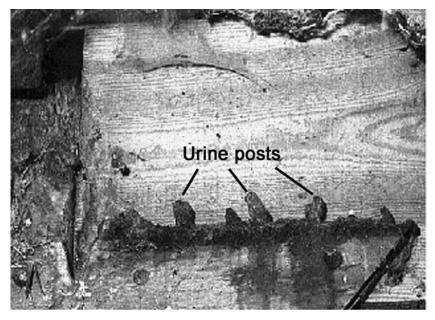


FIGURE 6.13 Scent turrets resulting from repeated scent marking by mice. (Courtesy Jane Hurst.)

urine marks of unfamiliar males. Isolated adult males are attracted to the urine of adult females. Females investigate and mark clean areas and also investigate the marks of adult unfamiliar males. Even a family member's odor can be novel if it is encountered at an unusual concentration, at an unexpected place, or against the background of a habituated odor. A bout of marking by a resident or a familiar neighbor is also treated as "novel" stimulus and can be used to communicate with familiar mice. In this paradox, a *familiar* odor can become a *novel* odor. This happens by virtue of contrast. As signals decay, they become part of the familiar background. Only fresh signals are detected. Immature females maximize their exposure to reproduction-suppressing odor cues: they investigate the urine of resident *females* but *not* the urine of the resident adult *male* (which might accelerate their puberty; see Section 8.4.2). But subadult females investigate urine of *unfamiliar* adult males (Hurst, 1989).

6.10.5 Latrine sites

Many mammals use latrine sites. They often are distributed over the entire home range, but not particularly dense at its periphery. This is true for civets and spotted hyenas (Bearder and Randall, 1978; see also above) and raccoon dogs, Nyctereutes procyonoides (Ikeda, 1984). The latrine sites of European badgers, M. meles, are large $(2-4 \text{ m}^2)$, contain feces, urine and secretions from the anal sac and subcaudal gland, and are bigger and more visible at territory borders with neighbor contact (Kruuk, 1978). Badgers locate their latrines under trees, particularly conifers, which are relatively rare in the English study area. Conifers may protect the latrines from rain and render them more conspicuous than they would be near the more common deciduous trees. The badgers also often place the latrines near linear features such as trails, ditches, or fence lines. Different behaviors point to specific functions: all age and sex classes participated in squat marking with the subcaudal gland, indicating a possible general territorial significance. By contrast, digging with forepaws and scratching with hindfeet occurred in the context of mating and may signal breeding condition. The latrines may communicate a variety of types of information (Stewart et al., 2002).

6.10.6 Effects of scent marks

The precise *effects* of territorial scent marks on conspecifics are still a matter of debate. They may warn of a possible agonistic encounter and/or induce a state of increased probability to loose a fight. Gosling (1982) proposed that an intruder matches the scent of a competitor, or of a mark it is seen to have made, with that of other marks nearby. If the odors match, the competitor is probably

the territory owner. This mechanism of *scent matching* helps the owner to reduce the costs of territory defense.

Territorial scent marking may greatly decrease the energetic costs of maintaining a resource-defense territory. In fact, a male mammal may only be able to maintain such a territory if he marks it. Otherwise the costs, such as walking, defending, predation risk, staying out in the heat instead of seeking shade, etc., outweigh the reproductive benefits. Indeed, all species with resource territories also scent mark (Gosling, 1986). Contests with other males establish the high social status of a territory owner even before he becomes territorial. It is to the advantage of the territory holder to be recognized as such. This saves energetically expensive and risky fights with intruders. The scent marks are located where encounters with the owner are likely. Odors of marks and the owner can be compared. Such scent matching (Gosling, 1982) is essential for the signal to work; information merely intrinsic to the scent mark, as assumed before, would not suffice. In other herbivores, such as beaver, scent marks may serve in population regulation before food becomes a limiting factor (Aleksiuk, 1968).

6.10.7 Countermarking: a visitors' register?

Over-marking is common. The best-known example is urine marking in dogs. How complex a visiting register can dogs read? Does only the top odor count or can they read the complex history of odor layers? Johnston and his group (1995) have experimentally examined the function of over-marking in golden hamsters, *M. auratus*, and Hurst and coworkers (Hurst and Rich, 1999) in wild house mice. In general, male hamsters remember and mark a female's vaginal secretion that is on top of an earlier mark (Johnston, *et al.*, 1995). Female house mice prefer a urine mark of a male mouse that is on top of that of another male. They also prefer a male (or its odor) who had exclusively marked or over-marked other urine marks in a territory that the female had encountered earlier. Male mice do not completely over-mark each other's marks. This provides information about relative competitive ability not otherwise available (Hurst and Rich, 1999).

Neighbors and newcomers, including potential mates, can "read a story": unchallenged scent marks inform of exclusive territory ownership and hence a high-quality individual, thought to be preferred in mate choice. Owners typically will destroy and/or over-mark a challenger's scent mark found in their own territory. This is the basis for the bioassays of beaver pheromones. An experimental scent mound is placed in an occupied territory. Resident beavers obliterate it by pawing and over-marking (e.g. Müller-Schwarze, 1992). In house mice, a single drop of competitor's urine placed in a male's territory not only

increases the owner's aggression against competitors but also reduces evasion of this owner by other males and increases challenges against him (Hurst, 1993).

Female mice given a choice between territorial marks of exclusive (one odor) and invaded (mixed odor) territories (in the absence of the owner) "chose" the first odor. Therefore, females use not only the odor of the territory owner but also the absence or presence of marks by competitors (challengers). The absence of any interfering mark identifies the marking male as one of high quality (Hurst and Rich, 1999). In short, individual A can read the story of interactions between individuals B and C and act accordingly to maximize its own fitness.

6.10.8 Factors stimulating scent marking

Scent marking can be stimulated by internal and external factors. Internal factors include most importantly gonadal steroid hormones, as in Mongolian gerbils, *M. unguiculatus* (Yahr, 1977), and the hormone-dependent high dominance status, as in rabbits, *O. cuniculus* (Mykytowycz, 1968). External factors include closeness to neighbors (population density), as in beaver, *C. canadensis* (Müller-Schwarze and Heckman, 1980; Houlihan, 1989), or encountering the mark of another individual, as in canids. Beaver also increased their scent marking drastically on days when transient conspecifics were observed. Even 1 day later the scent marking rate was still elevated (Bollinger, 1980).

Domestic female rabbits mark more when in estrus and prefer to over-mark scent marks by males. They also prefer to mark over chin marks, as opposed to urine and control odors, and prefer to over-mark chin marks from donors living in a long daylight cycle over those from short-day animals (Hudson and Vodermayer, 1992).

6.10.9 Feeding and scent marking

Feeding and scent marking are often connected: gerenuk, *Litocranius walleri*, mark on frequently eaten plants. These would most likely be encountered by an arriving stranger (Gosling, 1981). Tree shrews (*Tupaia glis*) also mark fig trees where they have been feeding (Kawamichi and Kawamichi, 1979). Beavers, *C. canadensis*, frequently place scent marks or scent mounds at feeding sites on the shore of their pond, sometimes where they have felled a tree (author's own observations). Marmosets (*Callithrix* and *Cebuella* spp.) place circumgenital secretion at holes on branches that they have gouged and extracted sap from (Lacher *et al.*, 1981). This marking of favored food trees may communicate information on food resources and avoid aggression, since several

groups feed and mark in the same area but avoid each other (Epple *et al.*, 1986; Roberts and Gosling, 2001).

In coyotes, *C. latrans*, SQU by females is associated with acquisition and possession of food, and the denning season (Wells and Bekoff, 1981). Red foxes, *V. vulpes*, urine mark a buried food cache at each visit; the more depleted it is, the more urine odor has accumulated, amounting to "book-keeping" (Henry, 1980). Foxes urine mark inedible food remains on repeated visits. This "no-food-left" signal, in turn, decreases the foxes' interest, and they investigate the site very little (Henry, 1980). Here, an animal may chemo-communicate with *itself* about food. Wolves, *C. lupus*, also urine mark their food caches (Harrington, 1981).

The latrine sites of European badgers, *M. meles*, also relate to food resources. In a 2-year study of a Mediterranean coastal population, the badgers produced more latrines in a year with few fruit because of a drought. This suggests increased defense of a scarce resource (Pigozzi, 1990).

Otters, *L. lutra*, scent mark with feces, called spraints in Britain. One study noted 12 times more frequent marking in winter when food is scarce than in summer. Spraints perhaps advertise depletion of patchy, recurrent resources to other otters and space out the animals. This non-competitive resource partitioning does not require agonistic reinforcement (Kruuk, 1992). Carnivores in general may heavily mark stable resource patches (Macdonald, 1980).

6.10.10 Sex differences in scent marking

Some glands and secretions are unique to one sex, such as preputial glands in males or vaginal secretion in females. Examples of skin glands typical for one sex are the subauricular and dorsal glands in the male pronghorn. Other glands occur in both sexes, such as the castor sacs and anal glands in beavers, and flank glands in golden hamsters. Even when present in both sexes, secretions may differ in composition, visible by differing color or viscosity, as in the anal gland secretion of both species of beavers (Grønneberg, 1978–79; Schulte *et al.*, 1995; Rosell and Sun, 1999). Although urine, feces, and saliva are common to both sexes, sex-specific accessory gland secretions may be added. Also urine marking can be ritualized by one sex, as RLU in canids shows.

Who scent marks in breeding pairs, families or extended kinship groups? In river otter, *L. lutra*, both sexes mark equally often. Tree shrews, *T. glis*, mark by rubbing chin, chest, and anogenital regions against trees. Adult males mark more than adult females and also have more secretion with a stronger odor. Juvenile males and females mark about 10 times less often (Kawamichi and Kawamichi, 1979). In captive red foxes, *V. vulpes*, males produced 84% of fecal pellets (with anal gland secretion) that served as scent marks (White *et al.*, 1989).

As in other sensory modalities, sex differences of anatomical structures such as scent glands, chemical composition of secretions, and behaviors associated with scent communication are more pronounced in species with polygamous mating systems than those with monogamy.

6.10.11 Chemical composition of scent marks

In a few cases we understand the chemical composition of territorial scent marks well enough that synthetic scents have succeeded in triggering typical scent marking in free-ranging mammals. Red foxes, *V. vulpes*, responded during their courtship season (January/February) to an aqueous solution of ethanol, polyethylene glycol, and eight volatile fox urine components on artificial mounds of fresh snow. These artificial scent marks were placed 20–40 m apart along trails traveled by foxes. The foxes marked the "fox urine" mixture more often than a control sample consisting of water, ethanol, and polyethylene glycol (Whitten *et al.*, 1980).

If artificial castoreum scent marks are placed on the banks of a pond, beavers, *C. canadensis*, are more likely to visit, destroy, and re-mark the sites as the complexity of the artificial odor composition increases (Fig. 6.14). While some single phenolics from castoreum such as 4-ethylphenol trigger marking (Müller-Schwarze and Houlihan, 1991), the response increases as the mixture grows to 4, 6, 10, 13, and finally 15 compounds. A mixture of 14 phenolics and 12 neutrals (mostly oxygenated monoterpenes) released responses almost as strong as whole castoreum (Schulte *et al.*, 1995).

The male pronghorn, *A. americana*, scent marks his territory with the sub-auricular gland. Among the constituents of the subauricular secretion, isovaleric acid (Fig. 6.15) released the strongest marking responses by pronghorn bucks (Müller-Schwarze *et al.*, 1974).

Non-volatile, hydrophilic compounds of high molecular weight in the polypeptide fraction of female (but not male) urine stimulate scent marking in male Mongolian gerbils, *M. unguiculatus*. The major volatile constituents of scent marks (suprapubic/perineal gland secretion) of the saddle back tamarin, *S. fuscicollis*, are butyrate esters with 20 to 28 carbon atoms (Smith *et al.*, 1985).

6.10.12 Time course of scent marking

The seasonal distribution of scent marking often suggests its function. The latrines of European badgers (*M. meles*) are largest in April and October, coinciding more with breeding than foraging, while the smaller, temporary

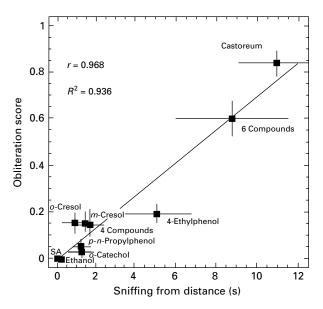


FIGURE 6.14 Free-ranging beavers respond more to mixtures than single compounds from castoreum. Sniffing from a distance was sniffing from water toward experimental scent mark on shore. The obliteration score measured pawing and re-marking of the experimental mark. The control was ethanol. Single castoreum compounds are shown and a four compound mixture groups with these while a six compounds mixture releases a stronger response, and complete castoreum the strongest. Obliteration score-0.014 \pm 0.07 (sniffing distance). (From Müller-Schwarze, 1992.)



FIGURE 6.15 Structure of isovaleric acid, which stimulates scent marking by male pronghorn.

defecation sites are most numerous in December and January and correlate with food supply and foraging (Roper *et al.*, 1986). Beaver, *C. canadensis*, scent mark most often from April through June (Svendsen, 1980) when the young disperse from their home colonies and search for new sites to settle. Beavers breed much earlier, in February when in the north their ponds still carry ice. Scent marks most likely signal occupancy to trespassers, who compete for colony space. In Norway, Eurasian beavers, *C. fiber*, also marked most heavily during the time of the subadults' dispersal in May (Rosell *et al.*, 1998).

6.10.13 Age of scent marks

The age of a scent mark may provide important information. Male domestic cats sniff fresh urine marks that are 0.5 to 4 hours old more (and perform flehmen more often) than urine that is over 1 day old. The length of examination may be an attempt to identify the donor and the time of emission (de Boer, 1977).

Urine of estrous females is attractive to males for only 24 hours in mice (Lydell and Doty, 1972) and 48 hours in guinea pigs (Beauchamp and Berüter, 1973). Puberty-delaying cues in urine of grouped female mice (Coppola and Vandenbergh, 1985) and the puberty-accelerating factor in urine of pregnant and lactating female mice are active for 5–7 days. Hamster flank gland and vaginal marks are active for 45 and 100 days, respectively (Johnston and Schmidt, 1979).

The number of measurable volatiles from fecal pellets of house mice, *M. musculus*, increased from 15 to 20 over a period of 24 hours (Goodrich *et al.*, 1990). Urine of female house mice contains a potent but ephemeral pheromone that elicits ultrasonic (70 kHz) mating calls from males, and a longer-lasting, weak pheromone. The first disappears within 15–18 hours, while the latter remains for at least 30 days (Sipos *et al.*, 1995). (For additional information on the life time of chemical signals, see p. 32.)

6.10.14 Complex scent marks

A species may scent mark in two different ways that convey different messages. In the stoat, *M. erminea*, body rubbing is correlated with threat behavior, while the anal drag permeates an area with an individual odor (Erlinge *et al.*, 1982).

Signaling pheromones II: sex and alarm pheromones and evolutionary considerations

And all your courtly civet cats can vent, Perfume to you, to me is excrement.

POPE, Epilogue to the Satires, Dial. 2,1, 183

Chapter 6 discussed signaling pheromones that allow discrimination, recognition, and broadcasting dominance and territorial status. This chapter explores the role of pheromones and other odors in reproduction, alarm, trail following, and in connection with food. Some evolutionary considerations conclude the discussion of signaling pheromones.

7.1 Sex pheromones: attracting and stimulating

Chemical cues are important to *advertise* one's sex and to *attract* the opposite sex as the first step in sexual behavior. Other functions of sexual signals are to signal current sexual status and to alter the behavior of the potential partner(s) via courtship or scent marking to facilitate mating. Typically, the odor of the opposite sex is attractive, at least in the breeding season. (Priming pheromones are covered separately in Chapter 8.)

7.1.1 Jawless fish (Agnatha)

Traditionally, French fishermen have used the male sex attractant of the sea lamprey, *Petromyzon marinus*. They bait a trap with an adult male and catch numerous females at night. Land-locked sea lamprey such as those in the Great Lakes of North America spawn in streams. Males arrive before the females and build nests. In laboratory choice experiments, ovulating females selected water from spermiating males and increased their searching behavior there. In a natural stream, females responded to male odors from as far as 65 m. The active

compound has been identified as a bile acid, specifically 7α , 12α , 24-trihydroxy- 5α -cholan-3-one 24-sulfate. The similar petromyzonol sulfate [Fig. 4.3] has a 3-keto group instead of an a hydroxyl group. It plays a role in migrating of non-reproductive adults. Males release the pheromone at a rate of about $250 \,\mu\text{g/h}$. The pheromone is probably synthesized in the liver and carried in the blood-stream to the gills where, in spermiating males, glandular cells are thought to secrete the pheromone actively. This would make male lampreys "active signalers," instead of females acting as "chemical spies" who cue in on their males' metabolites. Bile acids have advantages: they are more soluble in water and can be produced on a larger scale than steroids or prostaglandins (Li *et al.*, 2002; Yun *et al.*, 2003). To find spawning streams, adult sea lampreys use a migratory pheromone: petromyzonol sulfate plus allocholic acid, which larval sea lampreys release into the water. Larvae of 10 lamprey species produce and release petromyzonol sulfate in large amounts, very little prostaglandin, and only two species had allocholic acid (Fine *et al.* 2004; see also Ch.4).

7.1.2 Bony fish (teleosts)

Sex pheromones are widespread among fish. Fish in murky water or in caves benefit particularly from using chemical cues. For instance, the blind goby *Typhlogobius californiensis* (formerly *Othonops eos*) lives in burrows of the ghost shrimp *Callianassa affinis* and pairs for life. These gobies recognize the sex of conspecifics by odor in water (MacGintie, 1939). Chemically, hormones and their metabolites are common as fish pheromones. Tables 7.1 and 7.2 list some fish pheromones.

Male Pheromones in fish

All male fish pheromones are steroidal (Sorensen and Stacey, 1990). Many male fish release chemicals that attract females, stimulate them to spawn, and inhibit their aggression. As early as 1982, Liley compiled a long list of examples: testes, as in goldfish (Stacey and Hourston, 1982), urogenital fluid, glands on the caudal peduncle (an anal fin appendage), mucus and urine, can be sources of male pheromones.

Waterborne chemical stimuli from males in breeding condition attract female fathead minnows (*Pimephales promelas*). Females distinguish these cues from those of other females or sexually regressed males (Cole and Smith, 1992).

Pacific herring (*Clupeus harengus pallasi*) spawn synchronously by the millions near shore, their suspended milt discoloring the water. Small clusters of males start, then activity spreads to an entire school. Spawning lasts several hours in

Species	Compound(s)	Effect	Reference
Goldfish Carassius auratus	17α , 20β -Dihydroxypregnenone (preovulatory pheromone)	Stimulates milt production in male	Sorensen and Stacey, 1990
Goldfish	Prostaglandin $F_{2\alpha}$ and its metabolites (postovulatory pheromone)	Sexual arousal of male	Sorensen and Stacey, 1990
Zebra danio Brachydanio rerio	Testosterone glucuronide and estradiol glucuronide	Attracts males	van den Hurk <i>et al.</i> , 1987

Table 7.1 Some pheromones in female fish

Table 7.2 Some pheromones in male fish

Species	Compound(s)	Effect	Reference
Sea lamprey	Bile acid	Induces preference and searching in ovulating females	Li et al., 2002
Black goby Gobius jozo	Conjugated reduced steroid	Attracts ovulated females	Colombo et al., 1980
African catfish Glarias gariepinus	5 <i>β</i> -Pregnane-3 α , 17 α -diol-20-one glucuronide	Attracts and stimulates ovulated females	Resink et al., 1987

small schools, and several days in large ones. The fish move into deeper water, and males and females deposit gametes on submerged vegetation or other substrates, forming trails of sticky eggs or viscous milt. The milt dissipates, rendering the water milky. High milt concentrations may even inhibit spawning and thus regulate density of egg deposition. A male pheromone releases behavior in both males and females. Promiscuous "school spawning" may be the ancestral form of reproduction in teleosts. However, it may be specialized, facilitated by long-lived sperm: maximum fertilization occurs 0.5–2.5 hours after gamete release and sperm is still mobile after 5 days (review by Carolsfeld *et al.*, 1997a,b).

Male brook sticklebacks, *Culaea inconstans*, attract females by chemical cues. Females respond to such cues with courtship and receptivity displays. These consist of raising the head, sinking to the bottom, and remaining stationary. Female brook sticklebacks responded strongly to the male cue during ovulation. Their response waned after that and increased again at the next ovulation, 4–5 days later (McLennan, 2005).

Male bullhead (Cottocomephorus grewingki) urine contains a sex attractant for females. It is still active when it had been diluted by 1 \times 10⁻¹² (Dmitrieva and Ostroumov, 1986). The male African catfish, Glarias gariepinus, produces in his seminal vesicles a sex pheromone that attracts and stimulates ovulated females. One particularly active compound is 5β -pregnane- 3α , 17α , diol-20-one glucuronide (Resink et al., 1987). Among several steroid glucuronides, this compound released the strongest electrical responses in the olfactory epithelium of females. Holding water from males with their seminal vesicles intact was also active, while absence of the organ or lower levels of glucuronides diminished the response (Resink et al., 1989a). Groups of ovulated catfish are attracted by the steroid conjugate fraction from the seminal vesicle fluid. This fraction contained eight different steroid glucuronides. Without these, the activity was lost. A synthetic mixture of seven steroid glucuronides had a strong dose-dependent effect. A multicomponent pheromone from the seminal vesicle fluid probably attracts the females to the males shortly before spawning (Resink et al., 1989b).

Testosterone, 11β -hydroxytestosterone, and a polyene alcohol, probably farnesol, were found in the urine of male yellowfin Baikal sculpin (*C. grewingki*). Synthesized in the testes, these compounds are excreted with milt (Katsel *et al.*, 1992).

In addition to goldfish and African catfish, chemical analysis of male pheromones has advanced in other teleost fish. Examples are the black goby, *Gobius jozo* (Colombo *et al.*, 1980), the roundgoby (Zielinski *et al.*, 2003), and the zebrafish, *Brachydanio rerio* (van den Hurk *et al.*, 1987). Males of the black goby attract ovulated females to their nests by a C₁₉ steroid (etiocholanolone glucuronide). This steroid emanates from a specialized Leydig cell-rich part of the testes, the mesorchial gland (Colombo *et al.*, 1980).

Female pheromones in fish

In various fish species, sexually mature females that are ready to spawn emit chemicals to attract a male and stimulate him to court, release sperm into the water, be more active, build a nest, or assume the courtship coloration. We know more examples than for male pheromones (Liley, 1982). Sources of female pheromones can be ovarian fluid, eggs, mucus, urine, or skin.

Female goldfish (*Carassius auratus*) ovulate in spring, responding to rising temperature, fresh aquatic vegetation, and pheromones. They release eggs within a few hours, requiring close synchronization between the sexes. The female synthesizes the gonadal steroid $17\alpha,20\beta$ -dihydroxypregnenone (Fig. 7.1*a*) which induces final oocyte maturation. This hormone, plus

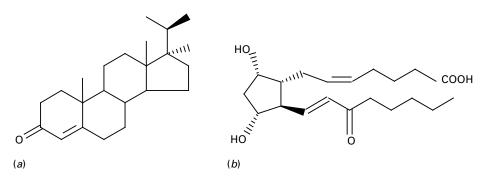


FIGURE 7.1 Female goldfish pheromones. (a) The preovulatory pheromone 17α , 20β -dihydroxypregn-4-en-3 one; (b) the postovulatory pheromone 15-keto-prostaglandin $F_{2\alpha}$.

 17α ,20 β -dihydroxypregnenone sulfate and androstenedione are released into the water and affect the male (Sorensen *et al.*, 1990a). The first two compounds stimulate milt production and sexual activity in the male, while androstenedione inhibits these processes (Sorensen *et al.*, 1992). In a comparative study, only goldfish, carp, and hornyhead chub (*Nocomis biguttatus*), all fish species of the minnow family (Cyprinidae), responded to the three steroids, while two members of the Catastomidae did not. Responses to prostaglandins were also weaker in this family (Sorensen *et al.*, 1992).

After ovulation, the female releases prostaglandin metabolites. The prostaglandin $F_{2\alpha}$ and its metabolite 15-keto prostaglandin $F_{2\alpha}$ (Fig. 7.1*b*) are most active in modulating the electro-olfactogram in mature male goldfish. The detection thresholds are 10^{-10} and 10^{-12} mol/l, respectively. These compounds stimulate different receptors, which, in turn, are different from those responding to other olfactory cues (Sorensen *et al.*, 1988). These prostaglandin F derivatives also act as hormones, modulating follicular rupture and stimulating spawning and they arouse the males sexually. Sorensen and Stacey (1990) proposed that most, if not all, postovulatory releasers are prostaglandins. However, although male Atlantic salmon, *Salmo salar*, are attracted to female urine of their own species and that of brown trout, *Salmo trutta*, prostaglandin $F_{2\alpha}$ and one of its metabolites do not appear to be the critical stimuli. These compounds abound in ovarian fluids of both species but occur at low levels in their urine (Olsén *et al.*, 2002a). Sorensen and Stacey (1999) have discussed the evolution of fish hormones into pheromones.

Female fathead minnows, *Pimephalus promelas*, produce a chemical signal that stimulates males to approach and lead them, both courtship patterns. This signal is produced if a female is injected with prostaglandin. (Cole and Smith, 1987).

Species	Sex of sender	Gland	Function	Compound	Reference
Red-bellied newt Cynops pyrrhogaster	Male	Abdominal gland (cloaca)	Attracts female	Sodefrin (decapeptide)	Kikuyama <i>et al</i> ., 1995
Sword-tailed newt Cynops ensicauda	Male	Abdominal gland	Attracts female	Silefrin (decapeptide)	Yamamoto et al., 2000
Magnificent tree frog <i>Litoria</i> <i>splendida</i>	Male	Parotoid and rostral glands	Attracts female	Splendipherin (peptide: 25 residues)	Wabnitz <i>et al.</i> , 1999

Table 7.3 Pheromones in amphibians

7.1.3 Amphibia

Texas blind salamander *Typhlomolge rathbuni*, a subterranean species, lives in a few caves and wells in the area of San Marcos, Texas. Males and females are attracted by water conditioned by the opposite sex (Bechler, 1986). Males of the salamander *Plethodon cinereus* actually signal their diet quality to the females. Males may feed on termites or ants. Termites provide the better diet because they are rich in lipids, energy, and vitamin B components, and lack the hard cuticle of ants. Female salamanders spend more time near fecal pellets from a male on a termite diet than one that feeds on ants. Also, in a forest, more males with termites in their diet were found associated with females than were males on an ant diet (Walls *et al.*, 1989).

Salamander courtship pheromones

Male red-bellied newts, *Cynops pyrrhogaster* (Salamandridae), attract females with a pheromone that is released into the water from epithelial cells of the abdominal gland of the cloaca. A decapeptide called sodefrin (Ser-Ile-Pro-Ser-Lys-Asp-Ala-Leu-Leu-Lys) is the first amphibian pheromone with female-attracting properties ever chemically identified (Kikuyama *et al.*, 1995). Silefrin in the related sword-tailed newt, *Cynops ensicauda*, is a similar decapeptide and differs from sodefrin in only two amino acid residues (Yamamoto *et al.*, 2000) (Table 7.3).

Another salamander courtship pheromone, identified in 1999, is a protein. During courtship, males of the Appalachian woodland salamander, *Plethodon jordani*, (Plethodontidae), a terrestrial species, actively deliver a pheromone to the female. In the mating season, males develop a "mental gland," located

under the chin. During courtship, the female straddles the male's tail, with her chin touching the male's tail base. Together, they walk forward. During this "tail-straddling walk" the male repeatedly turns back and touches the nares of the female with his mental gland, applying pheromone. This pheromone increases the female's receptivity and has been termed plethodontid receptivity factor. Chemically, it is a 22 kDa protein and shows homology with cytokines of the interleukin-6 family (Rollmann et al., 1999). Non-volatile pheromones such as this protein require close contact between the communicating animals, insuring privacy vis-à-vis predators or competitors. While pheromones in the aquatic-breeding salamanders, such as the red-bellied newt mentioned above, attract the female, in terrestrial salamanders, they increase the receptivity of the female. Furthermore, terrestrial salamanders have a prolonged mating season, and insemination can occur several months before oviposition. Therefore, plethodontid courtship pheromones are unique among vertebrates in that they induce changes in receptivity in females that are inseminated long before they lay their eggs (Rollmann et al., 1999). The authors recommend these salamanders as a model system to study changes in female receptivity that occur separately from ovulatory processes. Both the plethodontid receptivity factor, and a second protein (7 kDa, named P_i-7), occur in several isoforms. Salamander populations differ in the composition of their mental gland secretions. Some isoforms may be present or absent, and their ratios often differ between males in one population and also between populations. This divergence of courtship pheromone composition may be the result of sexual selection by conferring different rates of mating success (Rollmann et al., 2000).

Frogs and toads

The first anuran sex pheromone has been chemically identified. Males of the magnificent tree frog (*Litoria splendida*) of Australia produce a sex attractant in the parotoid and rostral glands on their head. One of several peptides secreted by the glands, splendipherin, attracts females. The peptide comprises 25 amino acids residues (Gly-Leu-Val-Ser-Ser-Ile-Gly-Lys-Ala-Leu-Gly-Gly-Leu-Leu-Ala-Asp-Val-Lys-Ser-Lys-Gly-Gln-Pro-Ala-OH) and in the breeding season, the frog produces 10 times as much splendipherin as at other times. In laboratory tests, females approach a pad scented with splendipherin, stay on it, and refuse to budge (Wabnitz *et al.*, 1999).

7.1.4 Reptiles

Odors are important in sexual behavior of snakes, but less so in lizards, which use primarily visual signals.



FIGURE 7.2 A "mating ball" of garter snakes. The large head (center left) belongs to the female, which is surrounded by males. (Photograph courtesy Robert Mason.)

Snakes

Female snakes leave odor trails as they move through vegetation. Their body odor adheres to the anterolateral surfaces of vertical objects. Males then are able to determine the direction of a female's path. The plains garter snake, *Thamnophis radix*, extracts information in this way (Ford and Low, 1984).

Male adders (*Vipera berus*) fight for access to unmated females. Courtship starts in spring only after sexually active males have shed their skins. Males tongue flick at females and court them but attack other shed males. If a recently shed male meets a non-shed male or female, he will ignore both. In summary, the male adder needs a chemical cue from the skin to court or fight (Andren, 1982).

Working with garter snakes, Robert Mason identified the first reptilian sex pheromones. Canadian red-sided garter snake (*Thamnophis sirtalis parietalis*) emerge from their winter dens and form clusters termed "mating balls" (Fig. 7.2). Many males compete for a female. Females attract males with nonvolatile saturated and monounsaturated long-chain methyl ketones (Fig. 7.3a). Males have squalene (Fig. 7.3b) as part of their sex recognition system. Squalene inhibits courtship of females. Males that behave like females to evade male aggression when pursuing females, so-called "she-males", have no or little squalene (Mason *et al.*, 1989a). Thirteen methyl ketones have been identified for females. The major unsaturated ketones are (*Z*)-24-tritriaconten-2-one and

(a) O
$$CH_3(CH_2)_7CH \longrightarrow CH \longrightarrow (CH_2)_{n+9} CH_3$$

$$n = 12: (Z)-24-Tritriaconten-2-one$$

$$n = 14: (Z)-26-Pentatriaconten-2-one$$

$$CH_3(CH_2)_{n+18} CH_3$$

$$n = 12: 2-Tritriacontanone$$

$$n = 14: 2-Pentatriacontanone$$
(b)
$$Squalene$$

FIGURE 7.3 Sex pheromones in garter snakes. (a) Methyl esters in the attractiveness pheromone of females. (b) Squalene, which forms part of the male sex recognition system.

(*Z*)-26-pentatriaconten-2-one (Fig. 7.3*a*). These and their saturated analogues, 2-tritriacontanone and 2-pentatriacontanone, were synthesized and tested in the field. A blend in natural proportions released courtship behavior and tongue flicking (Mason *et al.*, 1989a). The pheromone originates in the skin, which has seven cell layers. The most basal layer has complete cells with all organelles functioning. As the cells differentiate into the upper (outer) skin layers, they degrade and the organelles decompose. The outermost (surface) layer has flattened cells with no organelles left and the degraded cell constituents oozing out into the interstitial space. As the outer skin is shed, the new skin underneath is coated with the methyl ketones and squalene. Therefore, newly shed female snakes are most attractive to males in terms of their pheromones (R. T. Mason, personal communication, January 2005). The pheromone is a recycled product, stemming from constituents of former cell organelles. Since pheromone is derived from lipids of the epidermis, it qualifies as a protoadapted or exapted (Gould and Vrba, 1982) chemical body constituent.

Male red-sided garter snakes court larger females more than small ones. Even the skin lipid extracts from large females elicit courtship from more males than those from small females. Larger females possess more unsaturated methyl ketones, while small females have more saturated methyl ketones (LeMaster and Mason, 2002).

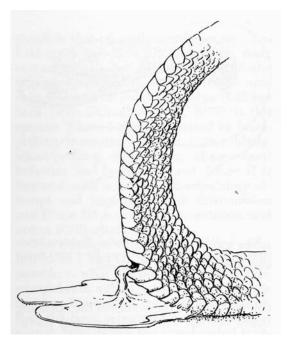


FIGURE 7.4 A female brown tree snake discharges cloacal defense secretion. (From Greene and Mason, 2000.)

Male garter snakes do not pursue already mated females. These females carry a gelatinous mating plug in their cloaca for about 2 days. Copulatory fluids around her cloaca contain a "copulatory pheromone" that inhibits males' advances. Males detect this copulatory pheromone on a female's trail by tongue flicking from a distance (O'Donnell *et al.* 2004).

Female brown tree snakes (*Boiga irregularis*) appear to employ a pheromone to discourage males' courtship. Males start courting a female by tongue flicking her dorsal integument, which is typical for snakes to detect pheromones by the vomeronasal organ. Then the male jerks his head sideways while rapidly tongue flicking, followed by chin rubbing, mounting, and chasing females that move. Aligned side by side, or mounted, the male aligns his cloaca with that of the female, whereupon the male intromits one of his hemipenes. During this process, the female tongue flicks, bobs her head, then mounts the male, and positions her body in front of male's head. This elicits tongue flicking and courtship from the male. The female moves away with head-lifting movements; the male chases and mounts her until their bodies align. If a female rejects a certain male or is not sexually receptive, she may lift her tail and release a clear liquid with a yellow or white precipitate from her cloaca (Fig. 7.4). This secretion, plus the visual signal of the raised tail, stops courtship by males (Greene and Mason, 2000). Female cloacal secretion shortens the duration of a male's

courtship and lowers its intensity. This cloacal pheromone specifically inhibits males' courtship of females, as male cloacal secretion has no effect, and ritualized combat between males is not affected by either male or female secretion (Greene and Mason, 2003).

Lizards

Lizards probably rely less on odors for sexual communication. Instead, sex is recognized visually by its typical pigmentation. Males of the viviparous (or common) lizard *Lacerta vivipara* courted males if these were painted with the color pattern of females. However, females were treated the same when painted like males, uniformly black, or left untreated. Males did not change their behavior vis-á-vis females washed with alcohol, especially at their cloacal and femoral regions – which would remove sexual pheromones in garter snakes, *T. radix* (Ross and Crews, 1978). Thus pheromones may not be important in lizards (Bauwens *et al.*, 1987). However, male lizards (*Gerrhosaurus nigrolineatus*) discriminate male from female cloacal secretions (Cooper and Trauth, 1992).

Geckos use chemical cues in mating. In the western banded gecko, *Coleonyx variegatus*, the male cannot determine the gender of another individual from a distance. Rather, he approaches it and grips its tail. Only then will he court a female, but ignore or attack a male. Surgically exchanged tails can guide the response: a female with a male tail is attacked, while the reverse preparation is courted (Greenberg, 1943). Chemical cues in the skin of female leopard geckos (*Eublepharis macularius*) release courting by males before and after the female's skin shedding. During shedding, a female is attacked by the male, presumably because chemical cues are unavailable. The male has the steroids cholestanol, stigmasterol, and stigmastanol, while the female has long-chain saturated and monounsaturated methyl ketones (Mason and Gutzke, 1990).

Tortoises

Tortoises can distinguish sex by chemical cues. A mixture of fatty acids has been extracted from the chin glands of males of several species of *Gopherus*. Applied to the head of a plaster tortoise, these fatty acids trigger head bobbing in females, a component of courtship, while males rammed the models in an aggressive manner (Rose, 1970).

Turtles

Freshwater turtles appear to use waterborne cues to find members of the oposite sex. During the mating season, males of the Iberian peninsula terrapin,

FIGURE 7.5 Both (Z)-4-decenal and octanal are found seasonally elevated in feathers of crested auklets. Auklets are attracted to these compounds.

Mauremys leprosa, are attracted to water from females, while females prefer water from other females. Outside the mating season, both females and males avoid water with cues from the opposite sex (Muñoz, 2004).

7.1.5 Birds

Birds are not exactly known for pheromone communication. However, there is some indication that ducks might use sex pheromones. As mentioned in Section 6.6, domestic ducks altered sexual and other behavior after olfactory nerve section and scenting with odorants (Balthazart and Schoffeniels, 1979). The chemical composition of the secretion of the preen (uropygial) gland of mallards, *Anas platyrhynchos*, differs between the sexes during the breeding season, but neither in the quiescent stage nor in ducklings (Jacob *et al.*, 1979).

The crested auklet (*Aethia cristatella*), a social, monogamous seabird of northeast Asia and western Alaska, emits a tangerine-like odor from its feathers. The odor appears strongest on the nape. There the birds smell each other during courtship in what is called "ruff sniff" (Hagelin *et al.*, 2003). The odor is partly caused by (*Z*)-4-decenal and octanal (Fig. 7.5). These two compounds or odor from conspecific feathers attract crested auklets in a two-way choice apparatus. The compounds are present primarily during the breeding season (Hagelin *et al.*, 2003). At a breeding colony in the Aleutian Islands, Alaska, experimental odor on auklet models attracted males or females only when presented on male models. Odor on female models had no effect on males or females (Jones *et al.*, 2004).

7.1.6 Non-human mammals

Chemical cues attract the sexes and modulate sexual behavior in many or most mammal species. Chemical signals also often reveal the quality and reproductive potential of individuals. In addition to chemical cues, multiple cues in different sensory modalities guide the complex reproductive behavior of mammals.

Male responses to female odors in mammals

Rodents

Mice vary their urination pattern according to urine cues from other mice they find in their environment. Adult male mice urinate most near cues from prepubertal and adult females, while adult females urinate more near cues from males than those from females (Drickamer, 1986). This proximate mechanism is consistent with a communication function.

Male Mongolian gerbils (*Meriones unguiculatus*) scent mark more when a female urine odor is present. The odor cue is hydrophilic, non-volatile, and resides in the non-hydrolyzable high-molecular-weight polypeptide fraction of female gerbil urine. Within a species where body size, form, or coloration is sexually monomorphic, sexual selection may have resulted in odor dimorphism (Blaustein, 1981). Urine of male gerbils and female laboratory mice are not active (Probst and Lorenz, 1987). Experienced males of the golden hamster prefer the odor of a female on the day before maximal receptivity. This is also the day of maximal scent marking by the female. Males least preferred the female odor on the day before diestrus. This is the day when the female chases and attacks the male. Ecologically, this system provides for quick detection and response to impending estrus in a solitary and promiscuous species. The mating advantage for the first male is reflected in pronounced male–male competition (Huck *et al.*, 1989).

Comparative studies have shed light on differences in chemical communication that reflect differences in *social organization*. Examples are the kangaroo rats *Dipodomys merriami* and *Dipodomys spectabilis* of the western United States. In *D. merriami*, one female shares her home range with several males that have separate, smaller home ranges. The reproductive condition of females can be communicated in close contact by anal–nasal circling, and males are not attracted to the urine of estrous females. In *D. spectabilis*, by contrast, the single males and single females defend territories, and females are not in close contact with males. Here, males are attracted to urine of estrous females. The urine seems to be needed as a signal over a greater distance (Randall, 1986).

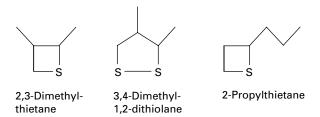


FIGURE 7.6 Structures of sulfur compounds in anal gland secretion of ferrets.

Carnivores

Ferrets, *Mustela furo*, distinguish between anal sac secretions of males and females. Males did not discriminate between the anal gland odors of estrous and anestrous females. Chemical sex differences were found, but no seasonal differences. Males had high concentrations of 2,3-dimethylthietane and/or 3,4-dimethyl-1,2-dithiolane (Fig. 7.6). Most individuals had 2-propylthietane. Clapperton *et al.* (1988) concluded that the odor of the anal gland provides sexual and territorial signals and cues to individual identity.

Ungulates

Sex pheromones in large mammals have been difficult to demonstrate. For instance, in white-tailed deer, *Odocoileus virginianus*, females were assumed to leave olfactory signals at the scrape sites that bucks had created on the ground, although females also sometimes scrape the ground with their front hooves (Sawyer *et al.*, 1982). Females rub their forehead at trees near scrapes, urinate near scrapes, walk through and sniff them. The males' responses are not clear. Females possibly also obtain information on the dominance status of bucks. For about 24 hours around conception, the does are found 1.6–2.3 km away from the core area of their own home range. Do they undertake mating excursions to the males' display areas (Sawyer *et al.*, 1989)?

Flehmen (Fig. 5.12) is a behavior shown by male ungulates to primarily female urine. In the stallion, it serves to monitor the estrus cycle, rather than being a step in the actual courtship sequence (Stahlbaum and Houpt, 1989). Table 7.4 lists some examples of the time in the estrus cycle when flehmen is most frequent.

Lek species

In lek species, the male's mating success can depend upon female odors, which attract the females to males. In both Uganda kob (*Kobus kob thomasi*) and Kafue lechwe (*Kobus leche kafuensis*), males attract estrous females to small breeding territories within a lek. Mating success of a male is site specific: it is predicted by the success of the previous occupant of the same territory and not by

Species	Time (or peak) of flehmen	Reference
Goat	More during diestrus than estrus	Ladewig et al., 1980
Elephant	More during estrus than diestrus	Rasmussen et al., 1982
Ram	24 h before estrus	Signoret, 1975
Black-tailed deer	Equally often during estrus and diestrus	Müller-Schwarze, 1979

Table 7.4 Timing of flehmen during estrus cycle

the male's own previous success. Males do not scent mark, but breeding territories are dotted with yellow urine patches from females. Transfer of soil from a successful to an unsuccessful territory increased mating success in the latter more than tenfold. The more successful the soil-donating territory had been, the greater the increase in success at the receiving territory. The mean number of females on the treated territory also increased. On the territories from which soil had been taken, mating success decreased. Successful territories had about 50% more oestrone 3-sulfate in the soil than unsuccessful ones. This is a metabolite of estrogen that occurs in urine of estrous female ungulates. Chance events such as predation by lions, or environmental factors such as predator-harboring cover, may funnel breeding activity to certain territories where females will concentrate and affect males' mating success without enhancing sexual selection (Deutsch and Nefdt, 1992).

Livestock: cattle and sheep

For obvious practical and commercial reasons, the sex attractant(s) of livestock have received considerable attention. In *cattle*, experiments have shown that bulls' responses to sex odors depend on the breeding regimen. Free-ranging bulls with access to cows will prefer to mount a cow that had been scented with urine from an estrous cow to one carrying the urine odor of an anestrous cow. Bulls kept tied up indoors and encountering cows in stalls mount cows indiscriminately (Sambraus and Waring, 1975).

Vulval skin secretion and blood serum contain an active principle, which attracts the bull and releases smelling, salivating, urinating, licking, flehmen, vocalizations, penis protrusion, and mounting. It is not clear whether the blood transports the active principle to or from the vulval skin tissue (Rivard and Klemm, 1989).

The systemic presence of estrus odor in domestic cows was confirmed experimentally: if estrous and diestrous cows are scrubbed with detergent, a bull cannot distinguish between the two. Furthermore, the response of a bull to a diestrous cow was not affected if the cow was marked with vaginal mucus of an

FIGURE 7.7 Structure of the acetate in urine of female elephants.

estrous cow. These results suggest that the estrus odor is not restricted to the vaginal mucus, urine, or feces but emanates from the body surface, particularly from the hindquarters and genital region (Umemura *et al.*, 1988).

Candidate compounds for sex pheromones have been isolated from cervico-vaginal mucus of domestic cows. Several diols, ketones, and amines were identified in fractions that released sexual responses in bulls, such as sniffing and licking the sample, flehmen, penile contraction, and preputial secretion (Klemm *et al.*, 1987).

Elephants

Preovulatory urine of female Asian elephant (*Elephas maximus*) contains (*Z*)-7-dodecen-1-yl acetate (Fig. 7.7). This acetate releases flehmen, and sometimes erections, in males and signals approaching ovulation (Rasmussen *et al.*, 1997). During musth, an annual event, Asian elephants have elevated levels of testosterone and are more aggressive. They secrete from their temporal gland and rub the secretion on trees. They dribble urine, leaving it on their hindlegs and on trails, for others to smell. Before temporal secretion appears, volatiles emanate from the gland. Temporal gland secretion contains frontalin, known as aggregation pheromone in bark beetles. Especially subadult males investigate musth urine, and particularly that of strange bulls. In the current view, musth odors boost males' chances of mating and inform other males of the breeding condition of a bull in the manner of "honest signals" (Schulte *et al.*, 2005; Scott and Rasmussen 2005).

Primates

Some female primates do not advertise estrus by behaviors or physical changes, such as genital swellings. Much sociobiological significance had been attached to this so-called "concealed ovulation." However, chemical cues may still signal ovulation to males. A case in point is the cotton-top tamarin, *Saguinus oedipus*. Scent marks from females during their periovulatory phase were transferred to males' cages. The males showed more penile erections, and mounting of test females, when periovulatory odor was present than in the presence of scent marks from the follicular or luteal phase of the donor females' cycles (Ziegler *et al.*, 1993).

FIGURE 7.8 Structures of boar pheromone compounds.

Responses of females to male odors

Odors of male mammals attract females and modulate their behavior in a variety of ways.

Ungulates

The well-known *boar pheromone* was not only one of the first mammalian pheromones identified, but also the first one applied commercially. The saliva from the submaxillary gland contains two steroids. These are 5α -androst-16-en-3-one and 5α -androst-16-en-3 α -ol (Fig. 7.8). They are emitted during head-to-head contact in courtship. Both individually stimulate the sow to assume the mating stance, but a mixture of the two is not more active than either compound (Melrose *et al.*, 1971). This may be a case of adaptive redundancy.

The scrapes and rubs produced by bucks of the white-tailed deer, *O. virginianus*, have been interpreted as olfactory attractants for does (Sawyer *et al.*, 1989).

Rodents

In their mate choice, estrous female house mice can detect small genetic differences between males. They prefer to mate with males that are dissimilar in their major histocompatibility complex (MHC). They also spend more time near the odor of dissimilar males than near that of males sharing their own MHC. Females seem to avoid mating with close relatives, and thus maintain genetic diversity (Egid and Brown, 1989). Familial imprinting is an important factor for the development of mating preferences between MHC-dissimilar mice (Yamazaki *et al.*, 1988).

Wild female house mice, *Mus musculus*, prefer the odor of males with the t-complex wild genotype +/+ over males with deleterious mutations (+/t) at the t complex. This preference depends on the estrus of the female and her genotype being +/t (Williams and Lenington, 1993). Lethal factors within the t complex play a role in production of the male cue and the expression of preference

by females. There may be a second gene (*s*) within the t complex that is involved in expression of female preference (Lenington and Egid, 1985). Preferences by males for +/t females are less dependent on environmental factors and may actually have a strong genetic basis (Egid and Lenington, 1985). No specific chemical compounds characterize mice of different t-locus genotypes, but at certain ages particular volatiles in the urine may be more concentrated. This applies to males and females.

Learning of the parental odor may be important for odor preferences (Jemiolo *et al.*, 1991). Estrous white-footed mice prefer males of intermediate relatedness, or their odors. The levels of reproductive success (i.e. litter size at weaning and offspring weight at weaning) indicated inbreeding depression. Non-estrous females showed no preferences (Keane, 1990).

Female mice are attracted to urine and preputial secretion from male mice. Preputialectomy renders males less attractive. The combination of an androgen-dependent factor in the urine and an androgen-independent factor in preputial secretion is essential for attracting females (Ninomiya and Kimura, 1986, 1988).

The function of the flank gland of the golden hamster, *Mesocricetus auratus*, is not clear but it appears to be involved in signals of sexual and social status and familiarity of the male to the female. Sexually receptive females spend more time near flank scent marks of intact males than castrates, or clean controls. They also stay longer near marks from familiar males than novel males. Finally, these females spend more time near marks of dominant males (compared with subordinate males) (Montgomery-St. Laurent *et al.*, 1988).

Seasonal variation of responses in mammals

Rodents vary seasonally in their responses to odors from the same and opposite sex. The meadow vole, *Microtus pennsylvanicus*, provides an example. Odor preference in a Y-maze and social behavior in dyadic encounters both differed between breeding and non-breeding season. During the breeding season, females are exclusive, and there is more aggression between females than between males or in male–female encounters. Breeding males prefer the odor of females to that of males, and breeding females prefer their own odor and male odor to that of other females. However, non-breeding females prefer each other's odor and are less aggressive. Outside the breeding season, the females' territoriality is relaxed, and they nest in groups. Non-breeding males have no odor preferences. They are solitary, and overwintering groups are, therefore, female biased. Thus, seasonal changes in social organization are correlated with changing odor preferences (Ferkin and Seamon, 1987).

Variation of response with individual reproductive cycles

Male meadow voles, *M. pennsylvanicus*, respond strongly to the anogenital area odor of females only immediately after parturition and for 2 days afterwards, the time of postpartum estrus (Ferkin and Johnston, 1995b). Females, by comparison, responded to male odor throughout their pregnancy and lactation (Ferkin and Johnston, 1995b).

Odor-stimulated flank marking by the female hamster shows a rhythm with a period of 4 days. This may reflect 4-day fluctuations of estrogen and progesterone levels (Albers and Rowland, 1989).

Reproductive status and odor preferences

Young female mice are able to exert some control over their own sexual development and reproductive condition by seeking or avoiding exposure to certain social odors. Prepubertal females avoid odors of adult males, which would hasten their maturation, and prefer the odors from grouped female mice, which slow down sexual development. At puberty, however, their odor preferences are reversed: now they are attracted to male odors and avoid odors of grouped females. Finally, as adults, females strongly prefer male odors and avoid odors of grouped females. Ultimately, the females' changing stimulus preferences during different ontogenetic phases may have important consequences for their reproductive (and inclusive) fitness (Drickamer, 1989b). The specific priming effects will be discussed in Chapter 8.

Chemistry of mammalian sex pheromones

The first two mammalian sex pheromones that were chemically characterized are those of the domestic pig (see p. 54) and of the golden hamster.

The mating behavior of the golden hamster, *M. auratus*, includes two pheromonally guided steps. The female attracts the male with dimethyl disulfide in her vaginal secretion (Singer *et al.*, 1976). Vaginal secretion contains 0.25 ng/mg (0.005 µg/20 mg) dimethyl sulphide and only approximately 200 molecules are needed to attract a male hamster. Dimethyl trisulfide suppresses the response to the former compound (O'Connell *et al.*, 1979). A second, closerange pheromone, *aphrodisin*, triggers mounting by the male. This non-volatile pheromone is a protein of the $\alpha_{2\mu}$ -globulin superfamily and has a molecular weight of 17 kDa. Its structure is similar to the retinal binding protein and the pyrazine-binding protein in the nasal mucosa. The pheromone is very specific: three related globulins that share the same overall molecular shape and similar amino acid sequences were not nearly as active: the second most abundant

protein in hamster vaginal secretion, the female mouse major urinary protein, and the bovine β -lactoglobulin from cow's milk. Specific structural characteristics, instead of merely the overall molecular shape, appear to be decisive for the high specificity of aphrodisin. Volatiles were experimentally removed from the protein by adsorption chromatography. Therefore, aphrodisin is probably not acting by transporting active ligands to the receptors in the vomeronasal organ (Singer *et al.*, 1989). Aphrodisin produced in *Escherichia coli* by molecular cloning is slightly active, but considerably less active than aphrodisin isolated from the vaginal discharge of the golden hamster. A ligand that is missing in bacterial aphrodisin could be responsible for most of the biological activity (Singer and Macrides, 1990). The gene for aphrodisin is expressed in the vagina, uterus, and Bartholin's gland, even in parotid glands, and can be detected in vaginal discharge before the females reach fertility (Mapert *et al.*, 1999).

In male goats, 4-ethyl-octanoic acid has the most intense goaty odor. This compound also attracts estrous does, and may be a releaser and primer pheromone at the same time (Sugyiama *et al.*, 1981, 1986; Sugiyama, 1983). Another team (Smith *et al.*, 1984) identified 6-transnonenal as typical for the odor of male goats. 4-Ethyl-oct-2-enoic acid gives goat's milk its "goaty" flavor. In the wolf, *Canis lupus*, the volatiles isopentyl sulfide and 3,5-dimethyl-2-octanone occur in male urine, while acetophenone was found in female urine (Raymer, *et al.*, 1984).

The Asian elephant has farnesol, 4-ethylphenol and 4-methylphenol in its temporal gland secretion while the African elephant has only 4-methylphenol. Farnesol levels are inversely related to testosterone levels (Rasmussen and Perrin, 1999). Preovulatory female urine contains (Z)-7-dodecen-1-yl acetate (Rasmussen *et al.*, 1996).

Interspecific detection in mammals

Animals other than conspecifics have been used to detect female sex odors, especially as a bioassay for isolating sex pheromones. Rats distinguish estrous from diestrous urine odor of domestic cows. This discrimination is specific to cows; it is not transferred to urines of goat, pig, or women. Estrous and diestrous urine was discriminated even when diluted 25-fold but not at 1:50 (Dehnhard and Claus, 1988). Dogs have also been used for detecting estrus in cows (Kiddy and Mitchell, 1981). Trained dogs detected estrus odors not only in vaginal mucus or vulva–vestibule samples but also in urine (both naturally voided and catheterized), milk, and blood plasma of cows (Kiddy *et al.*, 1984). The odor can be detected by dogs as early as 3 days before estrus but disappears abruptly on the first day after estrus (Kiddy and Mitchell, 1981).

7.1.7 Humans

Odors are important in the attraction of the sexes, and men and women differ in this. Women (American college students) ranked body odor as more important than any other factor in attraction to men, except "pleasantness." Men, however, ranked "good looks" highest, except for "pleasantness" (Herz and Inzlicht, 2002).

Verbal ratings of strength, pleasantness, and sex attribution have been used as measures of human responses to conspecific sex odors. Several studies have shown that sex, and within the same sex even one's spouse, can be correctly identified by smelling axillary odor on a T-shirt (Russell, 1976; Hold and Schleidt, 1977; Schleidt *et al.*, 1981). Generally, odor intensity and pleasantness are negatively correlated, and strong odors, whether from men or women, are labeled "male." German couples who maintained their regular personal hygiene were less able to distinguish sex by odors on T-shirts. This indicates that cultural personal hygiene patterns can suppress olfactory distinctions between the sexes (Schleidt, 1980).

The apocrine glands in the axilla can secrete enormous amounts of steroids such as dihydrotestosterone and pregnenolone (Brooksbank, 1970). Three single steroid compounds have also been tested on T-shirts. Surprisingly, both sexes attributed androstenol to females. Two other synthetic compounds were attributed to one or the other sex, depending on concentration, and one was perceived as very negative. Androstenol and the two synthetic compounds have very low olfactory thresholds for humans.

The compound responsible for much of the underarm odor is 3-methyl-2-hexenoic acid in both its (*E*) and (*Z*) isomers (Zeng *et al.*, 1991). *Breath* can also communicate gender information. However as with axillary odor, breath from men tends to be classified as stronger and more unpleasant than that of women (Doty *et al.*, 1982).

7.2 Alarm and alert odors

Alarm and alert responses triggered by conspecifics encompass behaviors ranging from increased vigilance to outright fleeing. The cue-sending individuals, in turn, typically are alarmed, disturbed, pursued, caught, or injured by a predator, but also possibly by a conspecific, or perhaps a competitor of another species. Alarm signals exist in all sensory modalities. Alarm calls and visual signals such as raising or flicking the tail in deer and antelope are well known. Chemical alarm signals are more difficult to detect by humans. Therefore, little is known about them in "higher" vertebrates, while fish provide the best-investigated examples.

We have to distinguish alarm signals emitted by a disturbed or stressed, but not captured, animal from "disturbance signals" and chemical cues from injured or captured individuals, termed "damage-released alarm signals" (Chivers and Smith, 1998; Mirza and Chivers, 2001).

7.2.1 Fish

Karl von Frisch (1941) observed minnows, *Phoxinus laevis*, at the edge of an Austrian lake, flee from an injured group member after an attack by a predator such as a pike, Esox lucius. Von Frisch induced the minnows to flee by adding macerated minnow skin to a feeding station in the lake (Fig. 7.9). In the laboratory, he introduced skin extracts into a fish tank at the minnows' feeding station. The minnows escaped, swam to the bottom of the tank, and froze. A chemical factor, the "alarm substance" or "Schreckstoff," is released from the skin's club cells (named after their shape) when a fish is injured. Many later experiments have uncovered details, mechanisms, and the distribution of this chemical alarm response among various taxa of fish. Solitary fathead minnows, P. promelas, respond to extracts from the skin of injured conspecifics with dashing, freezing and slowing, exploring, or no response (Smith and Lawrence, 1989). Exploring behavior resembles "predator inspection behavior" described for European minnows, Phoxinus phoxinus, and is thought to provide information about the predator (Magurran, 1986). Figure 7.10 shows the behavior sequence in the chemical alarm response.

Alarm substances in fish are extremely potent: extract from 1 cm² minnow skin creates an active space of over 58 000 liters of water (Smith and Lawrence, 1989).

Guppies avoid a model of a predator such as a pike more when it is paired with guppy extract containing alarm odor. The specific avoidance behaviors of the Trinidadian guppy, *Poecilia reticulata*, include shoaling, dashing, and freezing (Brown and Godin, 1999). Fish can learn to associate the alarm odor with sight of a specific predator, or even a non-predatory fish. Fathead minnows, *P. promelas*, can be conditioned to avoid goldfish or pike by pairing minnow alarm odor with the sight of either species. However, after 2 months, the response to pike, a natural minnow predator, was stronger than that to goldfish, a non-piscivorous exotic. The conditioning is species specific: minnows conditioned to pike did not respond to goldfish and vice versa (Chivers and Smith, 1994). Fathead minnows also learnt a predator odor without seeing the predator at the same time. Fish first exposed to the odor of yellow perch, coupled with alarm substance from other fathead minnows, later survived attacks by yellow perch better than

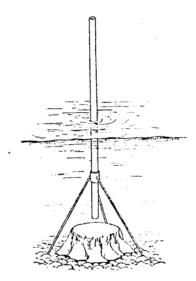


FIGURE 7.9 A field experiment to test alarm pheromone in minnows. The minnows were attracted to a "feeding table" and worms were fed through the tube. When minnows were feeding, macerated minnow skin, containing alarm pheromone, was dropped through the pipe. (From von Frisch, 1941.)

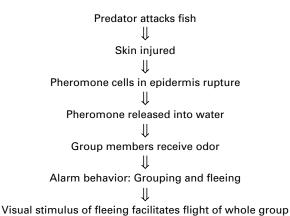


FIGURE 7.10 The sequence of fish responces to an alarm pheromone.

"untrained" fish. This advantage existed only when no cover was available. Simulated aquatic plants in the test tank negated the effect (Gazdewich and Chivers, 2002).

Even a neutral chemical cue can trigger alarm responses in fish if they had experienced it together with a true alarm signal. A coral-reef dwelling goby, *Asterropteryx semipunctatus*, learned to associate a novel chemical cue from a

non-predatory fish (water from the tank of the planktivorous damselfish *Acanthochromis polyacanthus*) with danger after this neutral cue was presented together with skin extract of freshly killed conspecifics that contained alarm pheromone. Later the fish responded with alarm (reduced movement and feeding) to the neutral cue alone. Moreover, this response was formed after only one single exposure to the two cues together (Larson and McCormick, 2005).

In addition to behavior changes, exposure to the alarm odor also has physiological effects. For instance, in pearl dace, *Semotilus margarita*, the levels of plasma cortisol and glucose increase 15 minutes after the alarm and are back to normal after 5 hours. The brain concentrations of dopamine, norepinephrine, 5-hydroxytryptamine, or tryptophan did not change (Rehnberg *et al.*, 1987). The fish recovered physiologically much sooner than the behavioral activation; For example, Von Frisch (1941) observed that minnows avoided the site of their encounter with alarm substance for many hours, even days.

Levels of predation risk influence the strength of the alarm response: in the laboratory, female Trinidadian guppies, *Poecilia reticulata*, from a population with much predation, shoaled, dashed, and froze more in response to skin extract from sympatric females than did females from a population that experienced less predation (Brown and Godin, 1999).

The alarm substance (Schreckstoff) has served to test Hamilton's "selfishherd" theory. Fourteen dace, *Leuciscus leuciscus*, were habituated to minnow schreckstoff, until they no longer responded. They were then joined by a single, naive minnow. Upon adding schreckstoff to the water, the single minnow was alarmed while the school was not. The single minnow moved into the school and became surrounded by other fish on all sides (Krause, 1993). Among alarmed fish, it is "everybody for himself."

The minnow's (*Phoxinus* sp.) alarm substance has been isolated and identified as hypoxanthine 3*N*-oxide (Pfeiffer and Lemke, 1973; Fig. 2.1). The pteridine derivatives isoxanthopterin, 6-acetonylisoxanthopterin, and 2,6-diamino-4-oxodihydropteridine elicit the fright reaction, produce bradycardia, and enhance the dorsal light reaction in giant danios, *Danio malabricus* (Pfeiffer, 1978). The oxide group linked to a nitrogen atom appears to be essential for the alarm response to occur (Brown *et al.*, 2000). Another, structurally different compound with the same group, pyridine *N*-oxide, also stimulates alarm responses such as "fast movement" and remaining "motionless" in juvenile channel catfish (*Ictalurus punctatus*), a species in the superorder Ostariophysae. Non-ostariophysan species such as a cichlid and rainbow trout did not show antipredator behavior vis-à-vis hypoxanthine 3*N*-oxide (Brown *et al.*, 2003).

7.2.2 Amphibia

Tadpoles of the European toad *Bufo bufo* release an alarm pheromone when injured (Eibl-Eibesfeldt, 1949). Alarm pheromones of toad tadpoles may indirectly affect capture efficiency of predators. The giant waterbug, *Lethocerus americanus*, feeds on tadpoles of western toads (*Bufo boreas*) of North America. When a tadpole is damaged, a substance is released that triggers alarm in conspecifics. The tadpoles increase their activity and avoid the feeding site, but only if the injured tadpole is of the same species. The alarm substance released after attack by the giant waterbug also reduces the success rate of the other predator of these tadpoles, the preying naiads of the dragonfly *Aeschna umbrosa*. The success rate is low because the prey is warned, and most likely not as a direct effect on the predator by the released substance since latencies to first contact, first attack, and first capture are not affected by the alarm odor. The tadpoles have giant cells in their skin that may contain bufotoxin (Hews, 1988).

The long-toed salamander, *Ambystoma macrodactylum*, a terrestrial form, avoids areas contaminated with the aqueous extract of ground skin and muscles of conspecifics (Chivers *et al.*, 1996).

7.2.3 Mammals

Alarm odors are very difficult to demonstrate in mammals. Black-tailed deer, *Odocoileus hemionus columbianus*, produce an odor in their metatarsal gland when alarmed, disturbed, or stressed (Fig. 7.11). This odor alerts group members (Müller-Schwarze *et al.*, 1984). Domestic sows stressed by restraining them in a food dispenser without access to food leave an odor that deters other gilts from approaching the feeder. Urine appears to carry this alarm odor (Vieulle-Thomas and Signoret, 1992).

Feral hogs (*Sus scrofa*) in dense scrub vegetation in Florida emitted a typical "pig odor" when fleeing an observer in 36% of 53 episodes, while such an odor was noticeable in only 2 of 18 "control episodes" of feeding or slowly moving ("nonfleeing") hogs. Fleeing pigs often grunted, hissed, or did both, and sometimes did not vocalize: 88.9% of the hissing episodes were accompanied by an odor and 60% of the episodes with both hissing and grunting; 79% of all incidents of hissing, whether by itself or combined with grunting, evidenced a simultaneous odor. When no sound was heard, an odor was noticeable in only 12% (D. Müller-Schwarze, personal observations). Emitting an odor in the face of danger, and then fleeing into dense vegetation, may confuse a potential predator. The predator can be distracted by the lingering odor, while the now invisible prey animal



FIGURE 7.11 Bioassay of the alert odor from the metatarsal gland of black-tailed deer. In foreground compressed air cylinder that creates air current to release MT odor in vessel under tree at feeding station. Left: Deer feeding undisturbed before odor release. Right: Deer become alert when air stream opens valve in odor container literally under their noses. (Photograph: D. Müller-Schwarze.)

can change direction and slip away. Like the severed tail of a lizard, the pig odor might amount to a *pars-pro-toto* diversionary tactic, an antipredator ploy.

7.3 Trail odors

Snakes follow odor trails, presumably to find one another and to migrate to hibernation sites. Male red-sided garter snakes, *Thamnophis sirtalis parietalis*, tested in the field, follow female tracks in spring but not in autumn. Females failed to follow conspecific trails in both spring and autumn (LeMaster *et al.* 2001). In mammals, footprints, especially by animals that also have glands on their feet, can carry information for group members or other conspecifics. Naturalists and hunters have observed deer sniffing tracks of other deer, especially when scattered from a group. Reindeer, *Rangifer tarandus*, have interdigital

glands and their secretion has been analyzed (Brundin *et al.*, 1978). In an experimental pen, reindeer sniffed pegs with interdigital secretion that were arranged in a line, resembling a track (Müller-Schwarze *et al.*, 1978b). Details of following non-pheromone trails of other species are discussed in Section 13.4.5.

7.4 Information about food

7.4.1 Rat's breath: exhaled carbon disulfide as food cue

Rats learn from group members about new food sources, and clans may develop "food traditions." How does one rat transmit to another this information about food? This type of intraspecific communication employs both body and foreign odors. Bennet Galef and coworkers (1985) showed that "observer rats" who encounter "demonstrator rats" with food odor on their heads will prefer that food over another when given a choice later. It is important that the food odor is on the head portion of a live rat; if it is applied to the rear end of a rat or to the head of a dead rat, it has no effect (Galef and Stein, 1985).

The food odor conveys information only if associated with breathing. Is breath simply moving air carrying the food odor, or has it a signal odor of its own? Air from the nostrils was sampled and analyzed. It contains carbonyl sulfide and carbon disulfide (Galef et al., 1988). Carbon disulfide proved to be an effective semiochemical to facilitate diet selection in rats: powdered food on cotton was offered to rats, either moistened with a dilute carbon disulfide solution or water only. They were permitted to eat one of these samples and a second type of food in succession. After that, they were injected with lithium chloride. Finally, they were given a choice between the two types of food. They always preferred the food coupled with carbon disulfide during the initial experience (Galef et al., 1988). Adding carbon disulfide and carbonyl sulfide to bait prevents "bait shyness." This procedure has been patented (Patent 4,861,585 to J. R. Mason and B. G. Galef). In field experiments, wild Norway rats consumed three times more bait if carbon disulfide was added. The effect is stronger in females than males. Carbon disulfide looks promising as a rodenticide enhancer (Mason et al., 1988).

House mice, *M. musculus*, also respond to carbon disulfide. Two drops of solutions of different concentrations were added to food pellets. The mice entered an enclosure with treated bait more often, spent more time there, and consumed more food than in an enclosure with water-treated pellets. As in rats, females responded more than males. Higher concentrations of carbon disulfide attracted the mice more than lower ones. Carbon disulfide was more effective than butanol, suggesting that it is a specific cue (Bean *et al.*, 1988).

7.5 Evolutionary considerations

7.5.1 Olfactory isolation mechanisms

Reproductive pheromones are powerful isolating agents between species, as is well known for insects such as moths and drosophilas (Wyatt, 2003).

Fish

Are bony fish (teleosts) reproductively isolated by pheromones? This may depend on the sympatric presence of similar species. Two Southeast Asian species of *Trichogaster* (Belontidae) differed in their discrimination of chemical cues. *T. pectoralis*, the snakeskin gourami, distinguished chemical cues from the two species. It came from stock originally sympatric with three congeners. *T. trichopterus*, the blue gourami, did not discriminate between the two species' odors. It came from a population isolated from congeneric species (McKinnon and Liley, 1987). In general, a given sensory mode should be critical in reproductive isolation if it is (a) an important and efficient mode of communication in the group in question, and (b) involved in the early stages of courtship behavior (Liley, 1982).

Amphibia

Males and females of the North American slimy salamander *Plethodon glutinosus* prefer substrates previously occupied by a conspecific over their own, but they do not discriminate between the sexes. However, in an olfactometer with live salamanders as odor source, male and female *P. glutinosus*, male *Plethodon jordani*, and a related phenotype ("species A") all preferred female over male odors. Male *P. glutinosus* and species A chose conspecific over heterospecific female odors in the olfactometer. This indicates that airborne odors may constitute an important pre-mating isolation mechanism (Dawley, 1984).

Reptiles

Red-sided garter snakes from different regions of Manitoba, Canada show signs of isolation by chemical cues. In choice tests, males from a hibernaculum (overwintering den) in central Manitoba preferred females from their own population to females from western Manitoba. Males from western Manitoba showed no preference. When confronted with experimental trails, males made the same choices. This demonstrated that a chemical factor is involved. Furthermore, the sexual attractiveness pheromone of females, a series of ω -9-cis-unsaturated methyl ketones, varies between the populations. Specifically, the

populations differed in the relative concentrations of particular methyl ketones that the females expressed (LeMaster and Mason, 2003).

Mammals

Discrimination of species odors, and by inference, reproductive isolation between species by means of odors, has been demonstrated for many species. Early examples are bank voles, *Clethrionomys glareolus* (Godfrey, 1958), *Peromyscus* spp. (Moore, 1965; Doty, 1972), *Mus* spp. (Bowers and Alexander, 1967), and gerbils (Dagg and Windsor, 1971). Male naked mole rats (superspecies *Spalax ehrenbergi*, Spalacidae) preferred odors from estrous females of their own to a different karyotype in a two-choice apparatus. The odors used were those of soiled bedding and urine from females (Nevo *et al.*, 1976).

Female house mice prefer the odors of males that do not carry *t*-haplotypes. These odors qualify as signal and primer pheromones. (The t complex is a region on chromosome 17.) Males heterozygous for a t-haplotype produce over 90% t-bearing functional gametes; males homozygous for the t-haplotype suffer sterility and homozygosity is generally lethal to embryos of both sexes (Drickamer and Lenington, 1987). The priming issues are discussed on p. 210.

7.5.2 Evolution of pheromone communication

Odors – as any other body features or social signals – are both product and agent of evolutionary processes. Are there extreme results of "runaway evolution"? As such examples are not immediately accessible to our senses, we have yet to discover the olfactory equivalents of the bizarre Irish elk's antlers, the wings of the Argus pheasant, or the antics of the bowerbirds.

We distinguish the evolution of sensory sensitivity, behavioral responses, and of the signals themselves. Without any accompanying changes in signal chemistry, individuals, populations, or species may change their sensitivity or responsiveness to certain compounds or mixtures.

When considering evolution of chemical signaling, the spatial and temporal modulation that the environment provides has to be taken into account. Stimulus propagation characteristics in different environments shape communications patterns. In the acoustical arena, this is true for bird song (Morton, 1975). It also applies to odors. Air currents being most important for odor transport, their presence, force, direction, timing (time of day, for instance), regularity, turbulence, and layering all determine the way an animal transmits its signals coded by volatiles. In addition, temperature, humidity, and other environmental factors, as discussed in Chapter 1, affect the chemical communication system. As

a result, animals as disparate as subterranean rodents, small primates of dense tropical forests, or the caribou of the treeless Arctic tundra communicate in vastly different ways purely on environmental considerations. Additional factors such as varying body size or social systems (which, in turn, reflect the environment) leads to enormous potential diversity of signaling systems.

Several comparative studies show that chemical cues may be emphasized in one species, while a closely related species relies more on other modalities. We shall examine why such differences exist.

Fish

In fish, sensory tracking of preexisting cues in evolutionary time permits "spying" by monitoring steroidal pheromones that most likely existed first as hormones and indicate hormonal state. In this way, "high-pass filters" can evolve. These are sensory systems that emphasize certain compounds at low concentrations. Sorensen and Stacey (1999) have discussed the possible evolution of such specific communication channels from early stages of general spying.

The "hormone-to-pheromone hypothesis" proposed for arthropods by Kittredge and Takahashi in 1972 appears to apply to fish. Since fish pheromones are related to steroid hormones, Sorensen and Stacey (1990) favored the "hormone-to-pheromone" hypothesis for three reasons: the preexisting signals are already in synchrony with discrete reproductive events, they are readily excreted into the water, and the evolution of pheromone receptors is simplified by existing internal endocrine receptors. In foraging behavior, fish responses to amino acids as feeding stimuli provide another example of sensory systems becoming attuned to useful stimuli that already exist independently of any other fish "tuning in."

On a larger taxonomic scale, the opposite "pheromone-to-hormone hypothesis" (Haldane, 1955) suggested that the interindividual pheromones of unicellular organisms such as protozoans enabled metazoan development and led to internal chemical communication in and between tissues by hormones in multicellular organisms.

Evolution of a signaling system can also be driven by preexisting sensory biases. The signaler adapts to the receiver's sensory abilities ("sensory drive;" Endler and MacLellan, 1988). In acoustic communication of frogs, such an evolutionary process has been termed "sensory exploitation" (Ryan *et al.*, 1990).

Some taxa may possess pheromone-producing tissues, while others have similar tissues but without pheromones. At the cellular level, extensive comparative studies of fish alarm responses show that cyprinids possess club cells, which release an alarm odor when ruptured by a predator attack. Polypteriformes have

very similar club cells. If these cells are homologous, the alarm function in cyprinids is a secondary adaptation (Hugie and Smith, 1987).

Amphibia

European newts of the genus *Triturus* are attracted to the water flowing from a courting pair of the same species. In *T. carnifex*, the crested newt, the response is weak and occurs only in females. This species is highly sexually dimorphic visually. *T. alpestris*, the Alpine newt, is less dimorphic and both sexes respond strongly to the water from the courting pair. Finally, the Italian newt, *T. italicus*, shows the strongest responses. It is not dimorphic at all and exhibits an extended tail-fanning phase during courtship (Belvedere *et al.*, 1988). In summary, each newt species is the more responsive to courtship odors, the less sexually dimorphic it is. This appears to be an example of shifting sensory modalities.

Mammals

In mammals, many chemical signals derive from metabolites in urine or feces or from maintenance compounds such as lipids in skin. In many cases, natural selection appears to alter *not* the signals themselves but rather the *behaviors* of donors and recipients of olfactory cues. This includes depositing, approaching, or avoiding signals. In addition, the *sensitivity* of recipients can evolve (Drickamer, 1986). In other words, evolutionary changes are thought to take place at the level of the central nervous system. Existing chemical cues, such as "body odors" or excretions, can be utilized by the signaler, the recipient, or both for their respective benefits. The response may be genetically anchored, or learned. Chemical signals can derive from excreted metabolites, as in urine of mice and carnivores, or beaver castoreum.

The first step in the evolution of elaborate patterns of odor emission such as scent marking was probably the regular elimination of solid or liquid waste and the inadvertent brushing of the body against vegetation or ground. From this evolved a more deliberate placing of scents at certain places and times with a certain frequency and intensity, resulting in a ritualized dosage of "typical intensity." Finally, supporting bodily structures for the odor signal evolved, such as osmetrichia (scent hairs or bristles). Behavior can also evolve for building supporting structures in the environment, such as the scent mounts that the beaver (*Castor canadensis*) builds from mud.

Intersensory elaboration can improve a signal. Scent release can be combined with visual signals into conspicuous displays. Genets, *Genetta genetta*, rub their flanks on the substrate, a movement accompanied by piloerection. This latter

visual display is considered to be *derived* from scent marking. Both are particularly frequent in agonistic encounters (Roeder, 1980). Pronghorn flare the hair of their large white "rump patches" when they release alarm odor from the sciatic glands (Müller-Schwarze, personal observation).

In mammals, males usually have stronger odors, larger and/or more scent glands, scent mark more often, and respond more to alien scent marks. Selection on the basis of odor differences can take place at the level of the individual, deme, population, or subspecies. In polygamous species, intrasexual selection via male–male or female–female competition and sexual selection can be most intense. Most mammals are polygamous, so odor dimorphism is probably widespread (Blaustein, 1981).

Variations of chemical cues

The chemical composition of a glandular secretion may vary considerably between species. Very little is known yet on such differences in vertebrates. Where known, it is not clear whether the differences are genetically controlled, the result of different diets or other ecological or phenotypical variation, or, finally, a combination of different factors.

At the evolutionarily critical level of the subspecies, divergence of the chemical cueing systems of different populations may be not only indicative of but also instrumental in accelerating incipient speciation. Deer mice (Peromyscus maniculatus) have lived in North America for many thousands of years, while house mice (M. musculus) were introduced only a few hundred years ago. Accordingly, the two species differ in their diversity: House mice have not (yet) developed subspecies, while there are 56 species in the genus Peromyscus, and one species of deer mouse has about 20 subspecies. In the laboratory, deer mouse subspecies from locations 3000 km apart cannot effectively communicate by olfactory signals, nor prime each other's reproductive processes chemically. In deer mice, males and female nest together. However, male deer mice of the subspecies P. m. borealis from Alberta, brought into the laboratory in Texas, instead killed and cannibalized prepupertal females of the Texas subspecies P. m. pallescens, and also P. m. bairdii females from Michigan. Priming had also grown disparate: male deer mice from Alberta stimulated uterine growth most in young females from Alberta, and less so in the other two subspecies. House mice from Alberta and Texas, however, were compatible in both signaling and priming (Perrigo and Bronson, 1983).

Intraspecific signals: priming pheromones

As the males of most animals search for the females, these odoriferous glands probably serve to excite or charm the female, rather than to guide her to the spot where the male may be found.

CHARLES DARWIN (1896) Descent of Man and Selection in Relation to Sex, p. 352

In the rat, olfactory communication results in the synchronization of estrous cycles within a female social group: the majority of females are likely to be at the same phase of their estrous cycles on the same day.

MARTHA McCLINTOCK, (1983) Chemical Signals in Vertebrates, vol. 3, p. 159

Priming pheromones are intraspecific chemical stimuli that act on the endocrine system and set in motion slower and often long-lasting processes of growth, maturation, or reproductive state. The difference between signaling and priming pheromones is one of degree. Behavioral responses to pheromones can also be slow and/or are based on physiological changes. Finally, the same compound(s) may release a behavior and also set in motion a physiological response, as occurs in fish and in pigs, for example.

8.1 Fish reproduction

8.1.1 Effects of female pheromones on males

In fish reproduction, the best-investigated pheromone system is that of the goldfish (*Carassius auratus*). Here, sex steroids and prostaglandins play important roles. The female produces two pheromones sequentially: a preovulatory primer pheromone and a postovulatory prostaglandin pheromone that act on the male.

Preovulatory primer pheromone

Female goldfish have an increased level of $17\alpha,20\beta$ -dihydroxy-4pregnen-3-one (17,20 β P) in their blood at the time of final maturation of their oocytes (i.e. 10 hours before ovulation and spawning). The pheromone is released across the gill. If 17,20 β P is added to aquarium water of male goldfish, they feed less, their level of endogenous gonadotropin rises after 0.5, 1, or 2 hours, and their milt (sperm and seminal fluid) production increases within 4 hours. In females the blood levels of gonadotropin do not increase. Lesions in the anterior telencephalon that disrupt olfactory input and male sexual behavior also block the increase of gonadotropin in response to 17,20 β P. Given the presence of $17,20\beta P$ in females during oocyte maturation, it is thought that this steroid is a primer pheromone that synchronizes increased male fertility with the time of ovulation (Dulka et al., 1986, 1987, Sorensen and Stacey, 1990). Response to this pheromone is an important factor in reproductive success in male goldfish. Preovulatory 17,20 β P has both physiological and behavioral effects. Within 15 minutes, males respond to this stimulus: their blood gonadotropin increases and milt production is stimulated. Thirty minutes after exposure, males are sexually aroused and remain aroused for 10 hours. They have more spawning success, more sperm are produced, and the sperm is more motile (Defraipont and Sorensen, 1993). Males exposed to $17,20\beta P$ fertilize a greater percentage of eggs than control males. In competitive spawning, males exposed to this steroid outcompete other males, as shown by microsatellite DNA fingerprinting (Zheng et al., 1997). Sperm quality, rather than spawning activity, appears responsible for this effect: in vitro fertilization experiments also showed that sperm from pheromone-exposed males fertilized more eggs than that from non-exposed males (Zheng et al., 1997).

The olfactory system of the male is extremely sensitive to $17,20\beta P$. The fish respond to a concentration of 5×10^{-10} mol/l. This amount (3×10^{14} molecules) is released by a 90 mm female fish into 1 liter water. The females are also very sensitive to $17,20\beta P$. It may stimulate ovulation. Of 47 vitellogenic females 13 ovulated when $17,20\beta P$ was added to the water, while only 1 of 43 did so in untreated water (Dulka *et al.*, 1987). Both sexes probably release $17,20\beta P$. Ecologically, this "bisexual" pheromone is thought to synchronize milt production with ovulation and thus coordinate spawning in local populations (Dulka *et al.*, 1987; Sorensen and Stacey, 1990).

Postovulatory prostaglandin pheromone

At the time of ovulation – which is about 12 hours after onset of the gonadotropin II surge in the female – females release less pheromonal steroids than before. They now become sexually active, receptive, and attractive to males.

These phenomena result from the hormonal function of prostaglandin $F_{2\alpha}$, which is produced by the female, and released via urine and across the gill. It stimulates follicular rupture and female spawning. The ovulated eggs are in the oviduct for several hours. During that time they stimulate a 100-fold rise in blood levels of prostaglandin $F_{2\alpha}$.

Released into the water, the prostaglandin also acts as a pheromone. It synchronizes male courtship with female reproductive behavior. Males are very sensitive to this prostaglandin and become sexually aroused (Sorensen and Stacey, 1990). Males detect receptive females by this and a related compound, 15-keto-prostaglandin $F_{2\alpha}$. Females release the latter in urinary pulses that grow more frequent as they enter the vegetation to spawn. The two prostaglandins also act synergistically: both are needed to raise the level of gonadotropin II in males. Sorensen and Stacey (1999) have summarized these studies.

Female and male goldfish also discharge steroids that inhibit milt production in other males. Most potent was androstenedione, having a threshold of 10^{-11} mol/l. These androgens could be functioning as antagonists for $17,20\beta$ P receptors (Sorensen *et al.*, 1990b).

Female rainbow trout, *Oncorhynchus mykiss*, also release in their urine 17,20 β P. As in goldfish, this pheromone increases the plasma levels of gonadotropin II and testosterone in spermiating males (Scott *et al.*, 1994). Levels of 17,20 β P rises within 1 hour of exposure and peak at 3–4 hours. Milt production also increases (Vermeirssen *et al.*, 1997).

Milt production in male carp (*Cyprinus carpio*) can be similarly stimulated by females and possibly a pheromone (Billard *et al.*, 1989). Table 8.1. summarizes some priming pheromones in fish.

The priming effect is not necessarily species specific: ovarian fluid and urine from mature ovulated Atlantic salmon, *Salmo salar*, and brown trout, *Salmo trutta*, affected males of both species. After exposure, maturing males (parr) of both the salmon and the trout developed higher levels of steroid hormones than did controls. Salmon parr had significantly more $17,20\beta$ P, 11-ketotestosterone, and testosterone in response to stimuli from both species, while brown trout males only increased their $17,20\beta$ P. The amount of strippable milt was elevated only in salmon. Finally, neither species had a changed spermatocrit value (Olsén *et al.*, 2000). Such interspecific effects of priming pheromones bear on the observed hybridization in these and other sympatric species.

8.1.2 Pheromone effects on females

In angelfish, *Pterophyllum scalare*, chemical cues from males stimulate and accelerate spawning in the female. Simultaneous chemical and visual cues stimulate oviposition even more, suggesting an additive effect of the two modalities (Chien, 1973).

Species	Source and target sex	Pheromone	Effect	Reference
Goldfish Carassius auratus	$F \Rightarrow M$	Oocyte-maturation-inducing steroid $(17,20\beta P)$	 ↓ Feeding ↑ Interaction with females ↑ Blood gonadotropin II ↑ milt (within 4 h) 	Sorensen and Stacey, 1999
	$F \Rightarrow M$	Androstenedione	Inhibits above effects; prevents premature milt release	Sorensen and Stacey, 1999
Rainbow trout Oncorhynchus mykiss	$F \Rightarrow M$	Urinary pheromone	↑ gonadotropin II ↑ 17,20βP ↑ Testosterone ↑ Milt production	Vermeirssen et al., 1997
European minnow phoxinus	Both sexes	Alarm pheromone	Bradycardia	Pfeiffer and Lamour, 1976
Pearl dace Semotilus margarita	Both sexes	Alarm pheromone	↑ Plasma cortisol, glucose	Rehnberg et al., 1987

 $17,20\beta$ P, $17\alpha,20\beta$ -dihydroxy-4-pregnen-3-one; M, male; F, female.

Males of the zebrafish, *Danio rerio*, release a primer pheromone that stimulates ovarian growth and ovulation in females (Chen and Martinich, 1975; van den Hurk *et al.*, 1987).

Under certain conditions, ovulation and plasma gonadotropin levels can be stimulated by ovarian fluid from other ovulated females. The African catfish, *Clarias gariepinus*, is an example. It lives in muddy water and spawns at night, conditions expected to favor pheromone communication. Moreover, this species is a seasonal breeder, spawning after rainfall. The sudden mass spawning requires synchronization. Water from males or ovulated females, and also ovarian fluid of ovulated females, experimentally induced ovulation in 67% of females. Plasma gonadotropin was also increased by these treatments (Resink *et al.*, 1989c).

For fish, it is thought that pheromones derived from hormones. "Hormones are pre-existing signals produced in temporal synchrony with discrete reproductive events" (Sorensen and Stacey, 1990). These hormones are readily excreted into the water. The evolution of olfactory reception mechanisms for pheromones may have been facilitated by already existing internal endocrine receptors. Sorensen and Stacey (1990) argued that "for pheromonal functions to arise mutations are initially required only in the recipient." Specificity of the signal

results either from admixing other compounds or from a slight change in the steroid molecule. Such specificity is the result of the selective pressure for reproductive isolation mechanisms (Sorensen and Stacey, 1990).

8.2 Amphibia

Male courtship pheromones can increase the receptivity of the female. In the plethodontid salamander *Desmognathus ochrophaeus* the male courts the female by scraping her dorsum with his specialized premaxillary teeth and swabs the same area with secretion from the "mental gland" on his chin, amounting to an "injection." In an experiment, filter paper with an extract from the excised mental glands of males was placed on the dorsum of females. These treated females mated 28% (59 minutes) sooner than controls (Houck and Reagan, 1990).

The terrestrial salamander *Plethodon jordani* applies courtship pheromone to the female by rubbing or slapping his mental gland directly on the female's nares. This way the pheromone can stimulate the vomeronasal organ and accessory olfactory system. Experimental application of mental gland extracts to the nares of females accelerated the time until active courtship started (Houck *et al.*, 1998).

8.3 Reptiles

In iguanas, *Iguana iguana*, chemical signals from adult males elevate the levels of corticoid steroids in juveniles, indicating stress. Together with visual cues, these cues suppress growth and assertion displays in juveniles (Alberts *et al.*, 1994).

8.4 Mammals

Priming pheromones would be expected to operate foremost in nocturnal and/or subterranean mammals that are precluded from communicating visually, or in species that inhabit enclosed spaces that permit high concentrations of odor to build up. Such a lifestyle would be analogous to those of many social insects, notably termites. The most bizarre social organization found in mammals fits almost exactly that description. Naked mole-rats, *Heterocephalus glaber*, of Kenya, Somalia, and Ethiopia live in large underground colonies of as many as 60 or 80 individuals. Their eyes are very small. Only one female, the "queen," breeds. She is also the colony's largest individual, with prominent teats

and a perforate vagina. The remainder of the colony form two or three castes, each with both sexes. The castes are of different body size and perform different tasks. "Frequent workers" dig, forage, and build nests, and are the smallest animals. "Infrequent workers" work less than half as much as frequent workers. Both worker castes carry young about when alarmed. "Non-workers" are the largest animals. The males in this class may mate with the reproductive female and may also assist in the care of the young. The young beg food from all colony members, except the breeding female. Individuals enjoy a long lifespan, and generations overlap. They cooperate in brood care and possibly pass through age-specific roles (polyethism).

All these features parallel those found in eusocial insects, and the naked molerat has been recognized as the only eusocial vertebrate (Jarvis, 1981). Naked mole-rats resemble termites more than hymenopterans: they are diploid, male and females form the worker castes, the young contribute to the colony labor, some workers reproduce if the breeding female is removed, and the young obtain food by coprophagy (Jarvis, 1981). A similar social system exists in the Damaraland mole-rat, *Cryptomys damarensis* (Bennett and Jarvis, 1988).

The breeding female needs physical contact to suppress reproduction in colony members. It is possible that her urine carries a regulating pheromone. After urinating at the communal toilet areas, mole-rats scratch their body with their hind feet. This way they may distribute through the colony a pheromone from the breeding female. However, evidence for chemical cues in pheromonal colony regulation of naked mole-rats remains elusive. While soiled bedding and litter do not suppress reproduction in males and females, removal of non-reproducting females from the colony restores ovarian cyclicity. In males, removal from the colony raises urinary testosterone and plasma luteinizing hormone (LH) levels (Faulkes and Abbot, 1993). Even when together with high-ranking non-breeding colony members, these isolated females increased their progesterone levels and their sexual and aggressive behavior. Reproductive suppression probably requires direct contact with the breeding female (Smith *et al.*, 1997). This suggests that primer pheromones may not suppress reproduction, but we do not know the active cues at this time.

8.4.1 Growth and development

Neonatal female mice are *retarded* in their growth if urine from virgin adult female mice is applied to their nostrils, while urine from pseudopregnant females *accelerates* growth (Cowley and Wise, 1972). Both sexes of prairie voles (*Microtus ochrogaster*) arrest their growth when held together as littermates or exposed to air from littermates (Fig. 8.1; Batzli *et al.*, 1977).

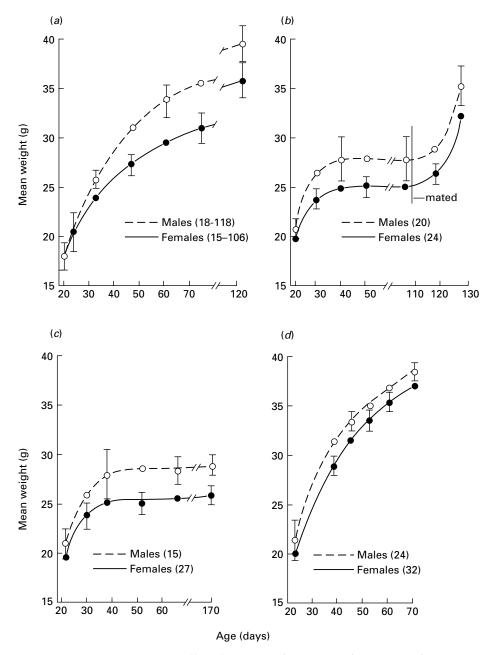


FIGURE 8.1 Priming effect of group members on growth in prairie voles, *Microtus ochrogaster*. (a) Normal growth curves for males and females when animals lived in individual cages from age 21 days. (b) Growth curves of littermates when kept together, then mated at age 12 weeks. (c) Littermates held separately but with air circulation among the cages. (d) Growth curves of voles living separately, with no air circulation among cages. (From Batzli *et al.*, 1977.)

5,5-Dimethyl-2-ethyltetrahydrofuran-2-ol 6-Hydroxy-6-methyl-3-heptanone

FIGURE 8.2 The active compounds in the urine of dominant male mice.

8.4.2 Puberty Acceleration

Males

Adult females or their odor slightly accelerate the puberty of young male mice while the presence of a male inhibits puberty (Vandenbergh, 1971).

Females

A male odor, contained in urine and linked to the presence of testosterone, accelerates puberty in female house mice. Vaginal opening and first estrus take place about 15 days earlier (Vandenbergh, 1969; Lombardi et al., 1976). This pheromone is effective at very low doses (Drickamer, 1982, 1984a) and is detected primarily by the vomeronasal organ (Lomas and Keverne, 1982). Urine from dominant males is effective while that of subordinates is not (Vandenbergh, 1969). Very early on, the pheromone was found to have low volatility (Vandenbergh et al., 1975, 1976). Researchers invoked a proteinpheromone complex that has to be contacted. A male urinary protein had been shown to accelerate puberty in females (Mucignat-Caretta et al., 1995). We now know that the active volatile compound, a lactol, and its hydroxyketo tautomer are complexed to a protein in male mouse urine (Novotny et al., 1999). The lactol (a cyclic vinyl ether) is 5,5-dimethyl-2-ethyltetrahydrofuran-2-ol. It exists in equilibrium with its open form, 6-hydroxy-6-methyl-3-heptanone, a hydroxyketone (Fig. 8.2). Both are implicated in the pheromonal effect of enlarging the uterus in young mice. The lactol has a strong affinity for urinary proteins. Complexing with proteins protects these labile volatiles and provides a slow-release mechanism (Novotny et al., 1999).

The presence of pregnant or lactating females can reduce the effect of the male pheromone (Lombardi and Vandenbergh, 1977). Wild house mice show the same effect (Drickamer, 1979).

Single-locus genetic differences between males can change the urine signal. Maturing female mice attained their first estrus sooner (at 29.9 days) when exposed to urine from males that were +/+ at the t-locus than with that from

+/t males (32.6 days). Control mice (treated with water) had their first estrus at 34.7 days (Drickamer and Lenington, 1987).

Puberty acceleration occurs in a number of rodent species. In the Djungarian hamster, *Rhodopus sungorus*, an adult male kept with young females accelerates their uterine and ovarian development. The effect is androgen dependent, as castration eliminates the effect and exogenous testosterone reinstates it. Urine is effective, while ventral sebum is not. There are neural effects of exposure to adult males: an area in the stria terminalis (lateral division of bed nucleus) is smaller in male-exposed females than in females with spontaneous onset of puberty (Reasner and Johnston, 1988). Soon after sexual maturation, at the age of 50 days, females of the Levant vole, *Microtus guentheri*, respond to male urine by increased uterus weight (Benjamini, 1987). Puberty acceleration by a male urine factor in female meadow voles, *Microtus pennsylvaticus*, manifests itself as earlier vaginal perforation, heavier uterus and adrenals, and ovaries with more and larger Graafian follicles. The pheromone responsible for these effects appears to originate at the level of the kidneys rather than the sexual accessory organs (Clulow and Baddaloo, 1987).

Male odor accelerates puberty in the domestic pig. The two steroids 5α -androstenone (5α -androst-16-en-3-one) and 3α -androstenol (5α -androst-16-en-3-ol) play a role in this effect (Booth, 1984).

Puberty acceleration has also been found in a marsupial. The odor left behind in a cage by a male can accelerate the time of first estrus in a female gray, short-tailed opossum, *Monodelphis domestica*, from 161 to 126 days of age (Stonerook and Harder, 1989).

Female odors can also accelerate puberty in young female mice. Urine from singly caged estrous females (Drickamer, 1982) or from pregnant or lactating mice (Drickamer, 1979; Drickamer and Hoover, 1979; Drickamer, 1984b) accelerates puberty in young females and also facilitates estrus in adult mice.

Other factors

Some effects depend on more than the urine stimulus. Reproductively naive females of the prairie vole, *M. ochrogaster*, do not cycle spontaneously. They need a male for stimulation. If male urine is applied to the nares, the uterus weight increases threefold within 24 hours. However, urine alone is not sufficient to induce behavioral estrus as well. This requires pairing with an unfamiliar male. Even though male urine and bedding soiled by males can induce behavioral estrus in some females, most require actual contact with a male (Carter *et al.*, 1980, 1986).

Diet may counteract or substitute for chemical cues that influence sexual maturation. Male California voles, *Microtus californicus*, for instance, are inhibited in their sexual development by odors from their mothers or unrelated females. This effect is eliminated if the regular diet is enriched with fresh lettuce. In young females, uterus growth is stimulated by the odor of males but, in the absence of males, fresh lettuce added to the diet can accomplish the same effect (Rissman and Johnston, 1986). This can be seen as another case of *adaptive redundancy* of cues for an important function.

In primates, effects can be complex. Female cotton-top tamarins, *Saguinus o. oedipus*, require three conditions to start sexual behavior and ovarian cyclicity. These are release from the suppression by their mother, release from suppression by other family members, and direct contact with, or stimulation by, an unfamiliar male (Widowski *et al.*, 1990).

8.4.3 Puberty Delay

Males

Exposing young male prairie deer mice, *Peromyscus maniculatus*, to soiled bedding from adult male conspecifics retards the growth of their testes and seminal vesicles. Male, but not female, urine applied to the nose has the same effect. Removal of the olfactory lobes at the age of 3 weeks blocks this effect (Lawton, 1979). The reproductive development of male California voles, *M. californicus*, is suppressed by chemical cues from the mother (Rissman *et al.*, 1984).

In rabbits, *Oryctolagus cuniculus*, urine of an adult female delays growth and sexual development in unrelated young males, while mother's urine also delays body growth, but it accelerates sexual development in her own sons. This can be seen as females manipulating the reproductive success of nearby conspecifics (Bell, 1986).

Females

Puberty in female mice can be delayed by an odor from adult females that live in groups (Drickamer, 1977). This effect is mediated by the vomeronasal organ (Reynolds and Keverne, 1979). Endocrinologically, the adrenals, and not the ovaries, are required for the pheromone to occur (Drickamer and McIntosh, 1980).

Puberty delay has been documented for feral mice by a clever experiment. Females in populations delimited by a freeway cloverleaf produced urine that delayed puberty in laboratory mice only if their population was very dense (Massey and Vandenbergh, 1980). In another experiment, foreign females were

added to a population in a highway cloverleaf. The urine from females in this population delayed puberty in juvenile laboratory females only after the introduction of the additional females. Ecologically, the urine signal in denser populations reduces population growth by extending the generation time. Even though the added females emigrated or died off soon, they stayed long enough to increase social interactions. This, in turn, presumably led to the production of puberty-delaying pheromone in the females' urine. The pheromone was induced rapidly after the population increased, and declined in potency after the population stabilized (Coppola and Vandenbergh, 1987). In addition to timing of puberty, other life-history effects, such as altered litter size or the size of the young, are expected (Coppola, 1986). Crowded female mice do not produce the puberty-delaying pheromone if their vomeronasal organ is experimentally removed (Lepri *et al.*, 1985).

Puberty delay also occurs in other species. Female white-footed mice, *Peromyscus leucopus*, mature later and weigh less if exposed to urine and feces from conspecifics of both sexes (Rogers and Beauchamp, 1976). Bank voles, *Clethrionomys glareolus*, have lower numbers of ovarian follicles and smaller uteri and ovaries if they mature in the presence of several other females. The European pine vole, *Pitymys subterraneus*, lives in sparse populations and urine from even a single female delays puberty in females (Marchlewska-Koj and Kruczek, 1986). Two factors keep subordinate female prairie voles, *M. ochrogaster*, prepubescent: (a) the odor of familiar males, usually father and male siblings; and (b) the inhibitory effect of females such as the mother and female siblings on the young females' response to the odor of unfamiliar males. (The odor of unfamiliar males would help the young female to mature) (Carter and Roberts, 1997).

Female mice have volatile components in their urine that are depressed after adrenalectomy. Six components have been detected, of which three (2-heptanone, *trans*-5-hepten-2-one, and *trans*-4-hepten-2-one) have no apparent effects either as a group or when added to the other three components. The active three components are *n*-pentyl acetate, *cis*-2-penten-1-yl acetate, and 2,5-dimethylpyrazine (Fig. 8.3). If 2,5-dimethylpyrazine is painted daily on the external nares of young female mice from day 21 on, the time of the first vaginal estrus is delayed. These three compounds act in redundant fashion: the two acetate esters delay vaginal estrus by 1.5 days, on average; 2,5-dimethylpyrazine alone delays it by 2.4 days, and a mixture of all three delays it by 1.7 days. (Novotny *et al.*, 1986a).

Stimuli other than chemical cues can play an important role in puberty delay in females. In the California mouse, *Peromyscus californicus*, which is monogamous, physical contact with the mother, and not a urinary chemosignal *per se*, is necessary for delay of puberty of females (Gubernick and Nordby, 1992).

$$^{\mathring{\downarrow}}_{\circ}$$

n-Pentyl acetate

(Z)-2-Penten-1-yl acetate

2,5-Dimethylpyrazine

FIGURE 8.3 Puberty-delaying compounds in female mouse urine.

8.4.4 Priming effects in adults: fine-tuning of breeding cycles

Reproductive inhibition in females

In grouped female mice ovulation and estrus are suppressed (Champlin, 1971) and pseudopregnancies occur (van der Lee and Boot, 1955). The odor of crowded female mice (eight per cage) inhibited ovulation in other females. The hormonal control of this effect has been difficult to explain. Early studies indicated that ovariectomized mice were not able to inhibit other females but in other studies, ovariectomized females still inhibited estrus in other females (Marchlewska-Koj, 1990).

Voided or bladder urine from *single* wild house mice also inhibited estrus cycles in females. These two types of urine reduced the normal rate of 2.4 cycles per female in 18 days to only 1.66 and 2.00, respectively. Urine from spayed females was not effective (Pandey and Pandey, 1986).

Synchronization of cycles in females

Urine odor of adult males induces and synchronizes estrus cycles in adult female mice (Whitten, 1958). The cycles grow shorter and more regular when a male is nearby. A male shortens cycles of 7–8 days to 4–5 days. Exposure to male urine for 48 hours has the same effect. This phenomenon is known as the Whitten effect. The estrus-inducing activity of male urine is retained even after storage at $-4\,^{\circ}\text{C}$ or freeze-drying (Gangrade and Dominic, 1986). This has

ecological implications. Marsden and Bronson (1964) synchronized estrus cycles by applying male urine to the nasal region of females. The estrus-stimulating pheromone has multiple sources. It has been found in salivary and preputial glands (Marchlewska-Koj *et al.*, 1990).

Nerve transection experiments have shown that normal estrus cyclicity and behavioral estrus in mice relies on sensory input through the main olfactory bulbs and does not require the accessory olfactory system (Rajendren and Dominic, 1986).

The estrus-synchronizing effect of adult male urine described for female rats has puzzled investigators. Recent experiments and more sophisticated statistical analysis have indicated that apparent synchronization may be actually chance (Schank, 2001).

The wallowing, pawing, and urine-soaking of the body by some deer species is thought to synchronize estrus in the females. If this is so, these behaviors should be more prevalent in alpine and high-latitude species than in tropical species with a long or permanent breeding season.

Induced ovulation

The prairie vole, *M. ochrogaster*, is an induced ovulator. Adult male urine, placed on the nose of females, stimulates weight increase of the uterus. The vomeronasal organ plays a role in this effect (Carter *et al.*, 1980).

Rats ovulate when exposed to soiled bedding from males. The stimulus is non-volatile, as a wire screen can eliminate this response. Vomeronasal organ occlusion reduces the response, implicating this pathway for priming pheromone reception in this species (Johns *et al.*, 1978). Table 8.2 summarizes the role of the vomeronasal in priming effects in rodents.

In domestic sheep, ewes will ovulate in response to reencountering a ram after 2 or more weeks of separation from adult males. This is true for prepubertal, lactating, and seasonally anovulatory ewes. The primary signal from the ram is chemical in nature, androgen dependent, and resides in "suint," the mixture of secretions from the sebaceous and apocrine glands of the skin and extractable from wool fat (Knight and Lynch, 1980). Urine or preorbital secretions have little effect. The main olfactory system, and not the vomeronasal system, primarily mediates this effect. However, other sensory modalities also modulate the response, as the sexual behavior of the ram intensifies the effect. Neural connections between the main olfactory tract and the anterior hypothalamus play a crucial role. More frequent pulses of luteinizing hormone (LH) are the first measurable event, occurring within minutes of exposure to the ram. This high pulse frequency stimulates follicle growth and estradiol secretion by the ovaries.

Table 0.2 The release the warmanese		::	rin Mammala
Table 8.2 The role of the vomeronasal	organ m	քпппп	g iii Maiiiiiiais

Species	Target sex	Context	Stimulus	Effect	Reference
Rat Rattus norvegicus	F	Estrus cycles	M urine	Shortening of cycles	Sanchez-Criado, 1982
	F	Ovulation	M urine	Induced ovulation	Johns <i>et al.</i> , 1978
Prairie vole Microtus ochrogaster	F	Reproductive activation	M odors	Maintain uterus and ovary weight	Lepri and Wysocki, 1987
	F	Ovulation	M urine	Uterus weight increase	Carter et al., 1980
	F	LHRH release	M urine	Presence of LHRH containing looping fibers in AOB	Reger et al., 1987
Mouse Mus domesticus	M	Exposure to F	F urine	Surge of LH and testosterone	Wysocki et al., 1983
	F	Pregnancy block	Strange M urine	Prevention of implantation	Bellringer et al., 1980
	F	Sexual maturation	Dominant M urine	Earlier vaginal opening and first estrus	Lomas and Keverne, 1982

M, male; F, female; LH, luteinizing hormone; LHRH, luteinizing hormone-releasing hormone; AOB, accessory olfactory bulb.

Estradiol subsequently builds up in the blood and first (within 2 to 12 hours) reduces the levels of follicle-stimulating hormone (FSH) and the amplitude of LH pulses, then (within 12 to 48 hours) causes preovulatory surges of LH and FSH. The former promotes ovulation and development of a corpus luteum (reviewed in Martin *et al.*, 1986). Two compounds have been indicated in the effect of the odor of ram's fleece on LH secretion in anestrous ewes. These are 1,2-hexadecanediol and 1,2-octadecanediol. In Merino sheep at least, maximum stimulation of ovulation requires full exposure to a ram, such as "fenceline contact" in pastures. Olfactory cues from the ram's wool, presented in a facemask for the ewe, are ineffective by themselves; visual and tactile stimuli are also important. The Merino breed does not rely as much on olfactory cues as other breeds of sheep (Pearce and Oldham, 1988). The effect is not necessarily species specific: hair extract from male goats stimulates LH release in ewes. For this effect, the accessory olfactory system is not necessary (Signoret *et al.*, 1989).

In domestic cattle, non-estrous cows may become attractive to bulls when in the company of estrous cows. It has been suggested that estrous cows influence their penmates via pheromone(s) (Hradecky, 1989) but other mechanisms may be involved.

An extract of secretion from the sebaceous glands on head and neck of sexually active feral billygoats increases the number of does ovulating. The extract was placed on cotton wool and worn in facemasks. 4-Ethyloctanoic acid and 4-methyloctanoic acid, responsible for the "goaty" odor, were not active but both the free fatty acid and lipid-free non-acid fractions were. The 4-ethyl branched fatty acids are present in the active fraction (Birch *et al.*, 1989).

8.4.5 Stimulation of reproductive activity in males

Male prairie deer mice (*Peromyscus maniculatus bairdii*) in very dense (i.e. asymptotic) populations cease to reproduce. Their testes remain abdominal and are small (20–25% of normal). The testes can assume normal size if such males have contact with reproductively proven females for 30 days. They also recover reproductively if exposed to female urine for 30 days, but contact with females is more effective (Creigh and Terman, 1988).

8.4.6 Pregnancy Block

Inseminated female laboratory mice may fail to implant if they are exposed within 24 hours of coitus to a male (or his odor) that is different from the stud male. The pheromone is contained in urine of intact adult males. It lowers prolactin production by the pituitary of the female, and corpora lutea fail to develop properly, so that pregnancy is not supported endocrinologically. Females with pregnancy thus blocked lose the embryo at the blastula stage and return to estrus within 4–5 days after the original mating (Bruce, 1959). This preimplantation pregnancy block (termed the *Bruce effect*) occurs in at least 12 rodent species such as wild house mice, Mus musculus (Chipman and Fox, 1966), deer mice, P. maniculatus (Eleftheriou et al., 1962), and seven species of voles, including field voles, Microtus agrestis (Clulow and Clarke, 1968), meadow voles, M. pennsylvaticus (Clulow and Langford, 1971), and prairie voles, M. ochrogaster (Stehn and Richmond, 1975). Pregnancy can also be blocked if the female is exposed to a strange male after implantation. This occurs in P. maniculatus, M. pennsylvanicus and particularly (up to 15 days of gestation) in M. ochrogaster (Stehn and Richmond, 1975; Kenney et al., 1977).

In addition to males, females can also induce pregnancy block. An example is the chemical cue from older, probably dominant, female white-footed mice, *P. leucopus* (Haigh, 1987).

Male mouse urine applied directly to the external nares of the female produces pregnancy block (Dominic, 1964). In mice, the vomeronasal mediates this block (Bellringer *et al.*, 1980). The pregnancy-block pheromone of the male may be a product of androgen metabolism, and not one of androgen-dependent

tissue. The steroidal anti-androgen cyproterone suppresses the ability of males to induce pregnancy block if given for a long time (42 days). If injected for only 14 days, there is no effect (Rajendren and Dominic, 1988).

The impregnated female mouse retains an *olfactory memory* of the stud male. This memory depends on cervico-vaginal stimulation at mating (Keverne and de la Riva, 1982). For the effect to occur, the female has to be exposed to the stud male's odor for 4–6 hours (Rosser and Keverne, 1985). The memory for the stud male lasts for about 30 days (Kaba *et al.*, 1992). Memory formation is accompanied by synaptic changes in the accessory olfactory bulb (Kaba *et al.*, 1992). The major urinary proteins, currently subjected to intense study, may provide individual information about the male, in addition to the protein's pheromone-binding role.

If the stud male remains near the pregnant female during encounters with a strange male, implantation failure will be prevented. Also, pregnancy block does not occur if the original stud male is reintroduced after 24 hours of separation (Parkes and Bruce, 1961). The active chemical stimulus from the stud male is probably not volatile and may act synergistically with bodily contact between the mates (Thomas and Dominic, 1987a,b). Furthermore, exposure to the original stud male *after* an alien male has caused pregnancy block results in pseudopregnancy. In prairie voles, *M. ochrogaster*, presence of the original mate also prevents strange male odor from terminating the female's pregnancy. In the wild, the pair would share a home range and a nest, but intrusion by strange males could be common, especially when a population is dense. Avoiding pregnancy block is adaptive, considering the short lifespan (45–61 days) of a prairie vole (Hofmann *et al.*, 1987).

The females play an active role in exposing themselves to certain male odors. In wild stock house mice, recently inseminated females avoid the odor of a strange male and prefer that of the stud male, but only up to 8 of the 18 days of gestation. Implantation occurs 4–5 days after insemination. By their behavior, the females protect themselves from pregnancy blockage (Drickamer, 1989b). Non-pregnant female mice do not avoid the odor of a strange male (C. J. Wysocki, unpublished data). The pregnancy block has not been found in rabbits, *O. cuniculus* (Bell, 1986).

8.4.7 Proximal mechanisms: hormonal responses to odors

All the stimulating primer effects in mammals analyzed so far share a common hormonal pathway: the first measurable event is stimulation of LH-releasing hormone (LHRH) release. This, in turn, stimulates LH levels and leads to increases of estrogens in females and testosterone in males. In females, changes in size and function of uterus and ovaries follow, while males respond

with the usual testosterone-dependent processes, including production of male pheromone(s).

The LHRH release appears to depend on the vomeronasal organ. Its extirpation in female prairie voles (*M. ochrogaster*) results in the absence of LHRH-containing looping fibers in the accessory olfactory bulb several months later (Reger *et al.*, 1987).

A single drop of male urine, applied on the upper lip of female prairie voles (*M. ochrogaster*) leads to changes of LHRH and norepinephrine concentrations in the olfactory bulb within 1 hour. Rapid increases in serum LH were also observed in these females (Dluzen *et al.*, 1981).

In mice, strange females or their urine increase the levels of plasma testosterone in males (Macrides *et al.*, 1975). Experienced male golden hamsters, on the other hand, do not depend on specific pheromonal odors for this testosterone surge induced by estrous females. Other cues, possibly learned odors from a female, perceived via the main olfactory system, appear to activate the neuroendocrine reflex that results in increased testosterone release (Johnston, 2001).

Anal gland secretion of male short-tailed voles, *M. agrestis*, stimulates plasma testosterone, body weight, and anal gland size in other males. The biological significance of this effect is unclear (Khan and Stoddart, 1986).

Goat pheromones

Most goats in the northern hemisphere breed in fall and winter. Decreasing day length stimulates ovarian cycles. In addition to photoperiod, male pheromones stimulate pulse frequency of LH in the blood plasma of females (Martin *et al.*, 1986). This not only enhances the seasonal onset of ovarian function, but also synchronizes breeding and subsequent lambing.

Male pheromones originate in glands of the parietal region. Scent-saturated hair from billy-goats stimulates LH secretion pulses in females within minutes. For a long time, practitioners have used this method to stimulate female goats that live without access to bucks. Follicular maturation and behavioral estrus require sustained high LH pulse frequency. Follicles in anestrous goats will mature if the animals are exposed to buck hair for 3 days (Claus *et al.*, 2001).

Active compounds have been found in the neutral fraction of buck hair extract (Over *et al.*, 1990). The carbonyl group and a hydroxyl group appear to be important for the biological activity (Claus *et al.*, 2001).

8.4.8 Inhibition of hormones

In a primate, *Microcebus murinus* (Prosimia) male odor depresses the level of plasma testosterone in other males. The lipid fraction (ether extract) of male

urine has this effect, while the aqueous fraction does not (Perret and Schilling, 1987). Female tamarins, *Saguinus oedipus*, increase their anogenital scent marking when removed from their natal group or from the company of other cycling females. Their urinary estrone and estradiol also increase. The cyclical pattern of estrone excretion also becomes more pronounced. Males, removed from their natal group or other males, increase sexual behavior, but not scent marking (French *et al.*, 1984).

8.4.9 Adrenocortical activator

Male mice crowded, or "stressed" otherwise, develop enlarged adrenals. For this response to occur no direct contact is needed; odors from crowded male mice suffice (Ropartz, 1968; Wuensch, 1982).

8.4.10 Ecological significance of priming pheromones

The priming pheromones in mice – and presumably other mammals – are crucial signals in feedback mechanisms that synchronize the sexual readiness of the sexes and adjust breeding behavior to ecological conditions and optimize reproduction within the constraints of the environment (Bronson, 1979). Sexual readiness can be adjusted by delaying or accelerating puberty, by modulating estrus cycles in adult females, and by stimulating testosterone production in adult males. Urine odors of adult males accelerate estrus in adult females and accelerate puberty in young females, while urine odors of adult females prime the testosterone in males and delay the puberty and estrus cycles of females, as described above. The Bronson model of priming in mice is shown in Fig. 8.4.

Pregnant and lactating females also produce a pheromone that accelerates puberty in females. Under favorable conditions, this factor is released throughout the year (Drickamer and Hoover, 1979; Drickamer, 1986).

Population dynamics are greatly affected by changes in generation time, especially in fast-breeding small animals. In the female house mouse, the onset of puberty can vary from 4 to 8 weeks of age, depending on whether a young animal is exposed to accelerating male pheromone or inhibitory female pheromone. Field studies have shown that the age of puberty can vary with population density: crowded females inhibit puberty in younger females. The signal seems to be mediated by the vomeronasal organ (Vandenbergh, 1987).

In outdoor enclosures treated with urine and soiled bedding from male mice, mouse populations grew to larger numbers than in water-treated control enclosures. Populations in enclosures treated with urine and soiled bedding from group-caged females, however, grew less than the water controls. Females

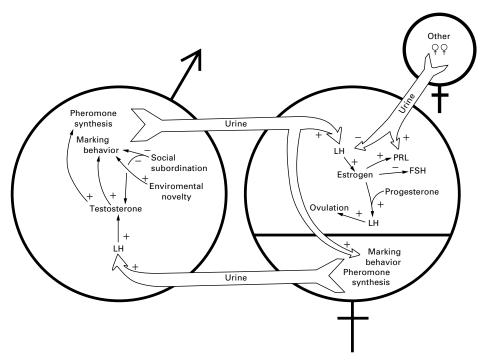


FIGURE 8.4 The Bronson model of priming pheromone actions in rodents. Chemical signaling between males and females constitutes a feedback mechanism that results in accelerated maturation and reproduction. This, in turn, permits the mice to adjust their reproduction and population size quickly to respond to environmental conditions such as sudden food abundance at harvest time. FSH, follicle-stimulating hormone; LH, luteinizing hormone; PRL, prolactin. Stimulation and inhibition are marked by + and -, respectively. (From Bronson and Coquelin, 1980.)

exposed to male cues were in "higher states of reproductive condition," experienced more pregnancies, and matured earlier sexually (Drickamer and Mikesic, 1990).

Small mammals undergo population cycles that each last several years. Mathematical modeling of the influence of several life-history traits showed that, among several factors, changes in age at maturity affected the population cycles most (Oli and Dobson, 1999). These authors suggested that adult females might release puberty-delaying pheromone when the environment is perceived as unfavorable or risky. Signaling poor environmental quality, the pheromone inhibits the maturation of young females. A lower population density ensues, which, in turn, decreases competition for resources (Oli and Dobson, 1999).

Priming pheromones affect the life history of a species. Coppola (1986) pointed out that the recipients benefit from the chemical signals while the signals' producers are little affected. The puberty delay pheromone, in particular, does not seem to be part of a mutually evolved signal system, but rather "unwitting release of metabolic correlates of physiological state." Coppola (1986) proposed to look for pheromonal priming effects on litter size and size of young. Inclusive fitness will depend on such life-history effects of primer pheromones as well as behaviors that regulate exposure to these pheromones, as described by Drickamer (1989c).

Drickamer (1986) suggested that natural selection has had little effect on the chemical *signals* that modulate puberty. Instead, natural selection has shaped the odor sensitivity of the young females and the donors' and recipients' behaviors of depositing, approaching, or avoiding the chemical cues.

8.4.11 Evolutionary significance of pregnancy block

Traditional explanations of the pregnancy block have included insurance of parental care, male competition, protection against infanticide, and promotion of outbreeding. Keverne and Rosser (1986) called such explanations a "blinkered view of evolution."

Instead, the pregnancy block can be seen as one of several possible consequences of the fact that primer pheromones lower the level of prolactin. When the mated female is exposed to a strange male, the male odor is perceived via the vomeronasal organ. Mediated by the accessory olfactory bulb, tuberoinfundibular dopaminergic neurons of the hypothalamus are activated. Dopamine increases and this starts the final common pathway for several pheromonal effects. Release of β -endorphin is inhibited and prolactin levels fall. In the mouse, prolactin is luteotrophic. Lowered prolactin leads to lowered progesterone release from the corpus luteum. This, in turn, releases the hypothalamic–pituitary axis from the inhibitory action of progesterone. LH production is disinhibited and the increase in its levels causes estrogen levels to rise. In an anovulatory female, cyclicity is restored. In short, pregnancy block within 3 days after mating results from lowering of prolactin. Implantation is prevented, the blastocyst is lost, and estrus can occur again (Keverne and Rosser, 1986).

The dopamine agonist bromocriptine produces the male-odor induced effects, while they are prevented by pimozide, which blocks dopaminergic transmission (Kaba *et al.*, 1992).

According to this scheme, the major difference between male and female odor effects is that the hypothalamic dopamine system is inhibited by female odors instead of activated, as described for male odors. Therefore, the converse

reproductive effects occur even though they are controlled by the same hormonal system (Keverne and Rosser 1986). All that is needed to produce the various effects is a "maleness" and a "femaleness" pheromone, or simply "odor."

The inverse relationship between dopamine and the β -endorphin neurons is of fundamental evolutionary significance. It is the final common pathway for inhibition of estrus during pregnancy and lactation, "nature's own means of contraception."

How does the female recognize the stud male as familiar? Keverne and Rosser (1986) suggest that the vomeronasal organ is involved, but not the main olfactory system. There are noradrenergic projections from the brain to both the olfactory bulb and the accessory olfactory bulb. If these fibers are lesioned, the accessory bulb is depleted of norepinephrine. The females still suffer the block but fail to recognize the stud male. In this experimental case, the stud male's own pheromones block the pregnancy induced by him. The noradrenergic pathways must be activated by mating. Coitus activates the brain's noradrenergic neurons and those fibers that terminate in the olfactory bulb. The noradrenergic neural system "imprints" recognition. Natural selection has favored recognition of familiar males. Olfactory block to pregnancy circumvents this recognition. This effect is unlikely to occur in the wild, and some consider it a laboratory artifact.

The *memory* that is necessary for the olfactory block to occur involves the synapses between granule cells and mitral cells in the accessory olfactory bulb. During the critical period immediately following copulation, the female is exposed to male pheromone. At the same time, noradrenergic fibers are active and *reduce* the inhibition that granule cells exert on mitral cells. This is considered to improve the signal-to-noise ratio. Such prolonged excitation of the mitral cells, in turn, activates a population of granule cells for the 4 hours when male pheromone and internal norepinephrine impinge on the female's accessory olfactory bulb. This forms an olfactory memory.

In summary, the synapses between the dendrites of the mitral and granule cells of the accessory olfactory bulb appear to be critical for forming the olfactory memory necessary for the pregnancy block. The association of norepinephrine and pheromonally induced activity during the critical period following mating produces a lasting change in the synapse. Once mating and the associated high level of norepinephrine are over, the affected subset of mitral cells experiences more inhibition by their associated granule cells. From then on, when the female encounters the familiar odor of the stud male, the pattern of activated mitral cells will match that of the population of mitral cells whose synapses had been modified and are now more inhibited by their granule cells. Hence the odor will fail to have the effect of blocking pregnancy. Different male odors, however, will not match this subset of mitral cells. They will act on (unmodified) mitral cells

that lack the increased feedback inhibition. Because of this, the neuroendocrine mechanisms that lead to pregnancy block will be set in motion (Brennan *et al.*, 1990). There are many different forms of memory. In the pregnancy block process, the changes that constitute memory occur at the most economical place in the nervous system, namely the *first neural relay* in the sensory system (Brennan *et al.*, 1990).

8.5 Priming pheromones in humans?

8.5.1 Puberty

Possible pheromone effects in humans never cease to fascinate us. Are there puberty-accelerating pheromones in humans? Altered pheromonal ambience for girls over the past 150 years has been suggested as an explanation for the decline of the age at menarche in Western industrialized countries (Bierich, 1981). Applying the animal model to humans, close contact with adult females should delay puberty while the presence of an adult male should accelerate it. The socio-economic changes over the last 150 years – so the argument goes – resulted for a girl in less contact with the mother because she worked increasingly more outside the home, and more contact with the father because his work hours declined and he spent increasingly more time at home. Contact with older sisters also declined because families became smaller, and older sisters became less available because they tended to be at school or work. Presence or absence of mother, father, or older siblings of either sex, as well as size of living space (close exposure to more concentrated putative pheromones), should then predict age at menarche in contemporary families (Burger and Gochfeld, 1985). It should be kept in mind that, even if contact with adults indeed shifts puberty in the assumed directions, stimuli in other sensory channels (or combinations) may be responsible. Also, environmental changes, such as nutrition, altered circadian activity rhythms or increased illumination, as in large cities, may affect onset of puberty.

8.5.2 Estrus synchronization

Ever since McClintock (1971) described synchronized estrus cycles in 135 women who roomed together in a women's college dormitory, the search for possible olfactory cues, as occur in other mammals, has continued. The longer the women in McClintock's study spent time together during the academic year (October through March), the closer the times of onset of their menstrual cycles

became. In addition, women who had more contact with men had shorter cycles (McClintock, 1971), reminiscent of the Whitten effect.

In a second study at a coeducational college, women who lived without roommates in bedrooms of residence halls or apartments developed menstrual synchrony with "close friends" (Graham and McGrew, 1980). These authors did not find a correlation between cycle length and amount and type of interaction with men.

A third study of 85 20-year-old women found an effect on menstrual synchrony of living together with one, two, or three other women, and of physical activity (Quadagno *et al.*, 1981).

These correlation studies stimulated experiments to identify the active chemical cues. Russell *et al.* (1980) rubbed underarm perspiration from a single woman onto the upper lip of five women aged 19–39 years. After 5 months, the odor-exposed women differed from one another in their onset of the menstrual cycle by 3.4 days, on average, compared with 9.2 days in the control group. Before the experiment, the mean differences had been 9.3 and 8.0 days, respectively. The volunteers were aware of the purpose of the experiment.

The first systematic, double-blind experiments on the influence of male and female odors on women's menstrual cycles were reported by Cutler *et al.*, (1986). and Preti *et al.* (1986), respectively. Neither their subjects nor the administering technician were aware of the true purpose of the study.

Male axillary odor, thought to arise from volatile acids and volatile steroids, was collected on pads worn in the armpit for 6–9 hours. Alcohol extract of these samples was applied to the upper lip of each subject and left there for at least 6 hours. The cycles of women receiving the axillary extract three times per week for an average of 13.5 weeks became less variable and the number of aberrantly long cycles was reduced.

The axillary odor of women, collected over 10 different 3-day sections of the menstrual cycle and applied to the upper lip of recipients, had a different effect. The cycles of the receiving women became more synchronized with the cycle of the donor female (Preti *et al.*, 1986), measured as difference in menses onset in terms of days. Wilson (1987) and Weller and Weller (1993) have criticized these experiments but still they remain pioneering studies in an extremely difficult field.

In a later experiment, two pheromones from axillary odor of young women influenced the estrus cycle of other women (Stern and McClintock, 1998). Armpit secretion that appears odorless to humans was applied daily between the upper lip and nose of healthy young women and had two opposite effects depending on the menstrual phase of the odor donors. Secretion from nine donor women, collected on pads in their armpits during the follicular phase of their cycles

(immediately before ovulation), shortened the menstrual cycles in 20 recipients aged 20–35 years. Specifically, the follicular phase of the treated women became shorter. In other words, ovulation was hastened. By contrast, odor from the ovulatory phase of the donors lengthened the follicular phase of the recipients and, therefore, the entire menstrual cycle. Ovulation, and by implication the preovulatory surge of LH, was delayed. In short, interpersonal chemicals can manipulate the timing of ovulation (Stern and McClintock, 1998).

Chemically, androstenol and dehydroepiandrosterone sulfate, found in male axillary secretion (Brooksbank *et al.*, 1974), occur in active samples. Aliphatic acids with 2 to 18 carbon members may also contribute to the axillary odor (Preti *et al.*, 1987).

Development of intra- and interspecific chemical communication

The infant, the adult, the aged person, each has is own kind of smell, and ... it might be possible, within certain limits, to discover the age of a person by his odor. In both sexes puberty, adolescence, early manhood and womanhood are marked by a gradual development of the adult odor of skin and excreta, in general harmony with the secondary sexual developments of hair and pigment.

H. H. ELLIS Sexual Selection in Man

The nature–nurture problem revisited: in most vertebrates, early experience of certain odors, interwoven with genetically anchored developmental processes, produces lasting, often irreversible odor recognition, preferences, or avoidance. Such behavioral development often occurs during more or less defined critical windows in time. The development of responses to odors often precedes that of odor production. Neonates already orient towards odors, while many pheromones are not produced until adulthood. Even before hatching or birth, the journey of chemical communication starts in the egg or the uterus. Knowing how chemical communication and chemosensory responses to food or danger develop is essential in areas such as animal husbandry or human behavior.

9.1 Fish

Young fish are attracted to conspecific odors without prior experience. Examples are European eels, *Anguilla anguilla* (Pesaro *et al.*, 1981), Arctic charr, *Salvelinus alpinus* (Olsén, 1987, 1990), and Atlantic salmon (*Salmo salar*). Kin recognition depends on early experience, as in Arctic charr. Fish reared with siblings later preferred water scented by unfamiliar siblings to water scented by non-siblings. By contrast, fish reared singly did not discriminate between water from siblings and water from non-siblings, even though they preferred sibling water to unscented tap water. At 15 months of age, the isolated fish were reared with siblings for 50–62 days. After this, they still made no choice between sibling

water and non-sibling water. Thus, sibling preferences appear to be established during the first 15 months of life (Winberg and Olsén, 1992).

9.2 Amphibia

Toad (*Bufo americanus*) tadpoles will recognize their siblings, but this depends on earlier experience. Here, the prehatching environment influences the post-hatching preferences (Waldman, 1981). Tadpoles were reared under three conditions: with siblings only, completely isolated from conspecifics, and sequentially exposed to both siblings and non-siblings. In subsequent choice tests, the first two groups preferred to associate with siblings. The third group did not associate with siblings unless they had been reared with siblings during early development. Tadpoles reared in isolation discriminated paternal, but not maternal, half-siblings from full siblings. In sum, sibling preference can develop without exposure to conspecifics, but early experience is important for normal development of sibling recognition. Table 6.1 (p. 129) contains other examples.

9.3 Reptiles

Numerous studies have shown that neonate or naive reptiles can already show complete species-specific chemical responses to conspecifics or rely on some form of social imprinted responses to food organism or predators, developed by interactions of genetic factors and experience, with each contributing.

9.3.1 Social odors

Neonate garter snakes, *Thamnophis sirtalis*, and brown snakes distinguish conspecific from heterospecific odors (Burghardt 1977, 1983). Newborn timber rattlesnakes, *Crotalus horridus*, are able to follow conspecific odor trails (Brown and MacLean, 1983). Neonate water snakes are attracted to conspecific odor (Scudder *et al.*, 1980) and neonate prairie rattlesnakes, *Crotalus viridis*, to lipoids from the epidermis of adult conspecifics (Graves *et al.*, 1987).

Neonate prairie rattlesnakes, *C. viridis*, show tongue flicking, mouth gaping, and face wiping frequently after birth. These patterns are virtually absent at 2–3 months of age. Face wiping and tongue flicking are directed towards the body of the mother. It has been suggested that some type of olfactory imprinting is served by this behavior.

Odor trails lead from the birthing rookeries of pregnant snakes to their ancestral winter dens. These trails probably help the neonates to find shelter (Graves *et al.*, 1987). Socially naive neonate prairie rattlesnakes were tested for odor

preferences in a two-way choice apparatus. The snakes had a choice between clean substrate and substrate marked with exudates from conspecific neonates, conspecific adults, and the predatory bullsnake *Pituophis melanoleucus sayi*. The hatchlings spent more time exploring or resting on areas with conspecific odor and avoided heterospecific odors. They possibly use the conspecific odor to locate the communal hibernacula for overwintering (Scudder *et al.*, 1992).

9.3.2 Antipredator responses

The response to predator odor is little modifiable by experience as two examples illuminate. First, neonate pygmy, *Sistrurus miliarius*, and timber, *C. horridus*, rattlesnakes respond to a predator odor: They "bodybridge" (raise the midsection of their body) when exposed to the odor of ophiophagous colubrid snakes (Marchisin, 1980). Second, king snakes, *Lampropeltis getulus*, prey on pine snakes, *Pituophis melanoleucus*. Pine snake hatchlings kept on soiled bedding from king snakes still prefer pine snake odor over king snake odor despite being familiar with the latter. The predatory king snakes also avoid the king snake odor (Burger *et al.*, 1991).

9.3.3 Feeding behavior

As the feeding behavior of snakes develops, genetic as well as experiential factors come into play. Genetic predispositions have been found at several levels. At their first exposure to food and food odors, neonate snakes respond to odors of foods that are typical for their species. Aquatic forms such as *Thamnophis* elegans aquaticus respond more to fish extracts, and the midland brown snake Storeria dekayi wrightorum to extracts from worms and slugs. Thamnophis r. radix, which lives on a varied diet, responds to extracts from worms, leech, fish, salamanders, and frogs, while the western smooth green snake, Opheodrys vernalis blanchardi, responds to cricket extract, reflecting its diet of insects, spiders, and other arthropods (Burghardt, 1967). Naive snakes from different populations of even the same species can differ behaviorally: in California, neonate coastal Thamnophis elegans responds to slugs, while the inland neonates do not (Arnold, 1981a). Arnold also showed that these two populations represent different genotypes. Hybrids between the two populations varied more in their behavior than snakes of either (Arnold 1981b). Within each population, Arnold also found genotypic polymorphism. There is no prenatal food imprinting: the maternal diet from the gastrulation stage of the young onwards had no influence on food preferences in neonates of Th. sirtalis.

Table 9.1 The shaping of feeding behavior in garter snakes

Genetic factors

- 1. Neonate food-naive snakes prefer odors of their "natural" food: eastern plains GS respond to worm, leech, fish, and tadpole odors while western smooth green snakes respond to cricket odor (Burghardt, 1967); coastal neonate *Thamnophis elegans* respond to slugs but the inland form does not (Arnold, 1981a)
- 2. Population differences: there is geographic variation in genotypes between coast and inland forms of *Th. elegans* (Arnold 1981a)
- 3. Hybrids between coastal and inland snakes vary more in behavior (Arnold 1981b)
- 4. There is genotypic polymorphism within populations (Arnold 1981a)

Role of experience

- 1. No prenatal food imprinting: maternal diet from gastrulation stage on had no effect on preferences by neonate *Th. sirtalis* (Burghardt, 1971)
- 2. Early postnatal experience modifies feeding in Th. sirtalis (Fuchs and Burghardt, 1971; Arnold, 1978)
- 3. Effect of experience depends on prey type: occurring for fish but not for tadpoles (Arnold, 1978)
- 4. Injection of LiCl after an earthworm meal induces food aversion in Th. sirtalis (Burghardt et al., 1973)

Experience modifies feeding behavior in garter snakes. Early postnatal experience can change feeding behavior in *T. sirtalis*. When two groups of neonates were fed fish (guppy) or redworm, they learnt to prefer the diet they were given (Fuchs and Burghardt 1971). The effect of experience depends on the diet: the snakes learn fish odors, but not those of tadpoles (Arnold 1978). In the same species of snake, injection of lithium chloride after an earthworm meal can produce a food aversion (Burghardt *et al.*, 1973). Table 9.1 summarizes the factors that determine formation of feeding behavior in garter snakes.

Odors can have delayed and unexpected effects. Garter snakes, *T. sirtalis*, were presented with either live earthworm or mosquito fish, *Gambusia affinis*, in a screen-covered bowl for several days. One day after transferring the snakes individually to a box free of prey odors, they were tested with aqueous extracts of fish and worms on cotton swabs. Snakes exposed to fish odor attacked fish extract less, and those exposed to worm attacked worm odor less. This is interpreted as habituation with a possible switch to other prey. This also demonstrates that in any experiment with chemical cues an odor not experienced for 22 hours may still have an effect (Burghardt, 1992).

Experience can further hone behaviors that are already present in naive animals. From 28 to 138 days of age, neonate rattlesnakes (*Crotalus viridis* and *C. horridus*) recognize and attack prey. They also show chemosensory searching and trailing. The initial trailing is "jerky and erratic," but after several feeding experiences becomes more "methodical" (Scudder *et al.*, 1992).

9.3.4 Olfactory "imprinting"

Some reptiles have been successfully imprinted to odors early in life. The hallmarks of classical imprinting are: primacy of exposure to a stimulus is more important than recency, a short (or one-time) exposure suffices, there is a critical (or sensitive) period during development (a "window of opportunity") when imprinting is possible and the effect is irreversible during the life of the animal. Green sea turtles, *Chelonia midas*, were exposed to an odor (morpholine or 2-phenylethanol) either during incubation plus 5 days in the nest, or 3 months after hatching in the holding water, or both. They later preferred the familiar odor in a choice experiment only if they had experienced the odor in the nest and the post-hatching holding water. Early exposure has no effect if the odor was added only to the nest or only to the holding water. The turtles needed a long period of exposure to the odor (Grassman and Owens, 1987). Therefore, the acquisition of the preference for the odor may be different from classical imprinting, which occurs during a brief period. Similarly, snapping turtles, Chelydra serpentina, require 2 weeks of exposure to food odors to be imprinted (Burghardt and Hess, 1966). It is debatable whether we should maintain the term "imprinting" for these "large window" acquisitions of later preferences.

9.4 Birds

Goslings of the greylag goose, *Anser anser* at 10 hours old and food naive will shake their heads when presented with the odors of sage, peppermint, dill, or lavender. All goslings responded at the age of 18 hours. They discriminated different plant odors at 4 to 9 days of age (Würdinger, 1979).

Even bird embryos are sensitive to odors. If domestic fowl embryos were exposed to odors 1 day before hatching (including dichloroethane, formic acid, cineole, and amyl acetate) they responded with increased heart rate, beak clapping, and head shaking to the first three compounds. Amyl acetate had inconsistent results. Blocking the nostrils with wax eliminated the responses (Tolhurst and Vince, 1976).

9.5 Mammals

First, I will discuss the development of *responses* to odors and follow this by the development of *odor production*. Mammals spontaneously respond to odors *in utero* and can also be conditioned to chemical stimuli before birth. Much research has focused on the chemical ecology of the fetus. *Experience* plays an

important role in many aspects of sensory development. It will be discussed throughout this section.

9.5.1 Prenatal development

"Spontaneous" preferences

Olfaction

Both *social* and *dietary* odor preferences may be acquired *in utero*. Rat pups at 8 hours of age orient more toward the amniotic fluid of their mother than to that of an unrelated female rat. This preference must be acquired prenatally, as Caesarean-born pups also prefer the mother's amniotic fluid. At birth, rats already know the odor of their kin (Hepper, 1987). The vomeronasal organ probably monitors the chemical quality of the intrauterine environment (Pedersen *et al.*, 1983).

Prenatal olfactory and taste experiences may create standards against which postnatal odors are matched (Schaal, 1988a). Many chemicals, including volatiles, cross the placental barrier and enter the fetal blood. Volatile compounds may diffuse out of the blood and stimulate the olfactory receptors. For instance, the essence of garlic, allyl sulfide, is carried from the mother's blood to that of the fetus. Pregnant rats will readily eat garlic if provided. One clove of garlic consumed daily from gestation day 15 to 21 affects the preferences of a female's offspring. Her pups, tested at 12 days of age, preferred garlic over onion, while pups of mothers without garlic in their diet did not discriminate between garlic and onion. The response is acquired prenatally, since pups of garlic-eating mothers still show that preference even if they have been crossfostered to a "garlic-free" mother. As to the pathway of the chemical cue, the odor is assumed to diffuse from the capillaries in the nose to the olfactory receptors (Hepper, 1988). Sheep fetuses on gestational day 144 were "externalized" from the uterus and implanted with intranasal catheters and heart rate electrodes. Later, odorants were delivered to the nose through the catheters by means of a syringe while the lambs were in the womb. The odorants were citral (lemon-like odor) and 2-methyl-2-thiazoline, which has a foul odor. The latter significantly reduced the heart rate in both lambs, while citral only slightly accelerated it, and saline (control) had no effect (Schaal et al., 1991). Rabbits also "learn" prenatally odors of aromatic foods eaten by their mother. Postnatally, they prefer such odors (Hudson and Altbäcker, 1994).

Late in gestation (days 19 and 20), rat fetuses respond to intraorally applied lemon solution or citral, a tasteless odor, with behavioral activation. No such response is observed after surgical transection of the olfactory bulb of the fetus

(Smotherman, 1987). To conclude, prenatal olfaction is possibly adaptive by preparing the animal for diet preferences, successful suckling, and socialization (Hepper, 1990).

Taste

The sense of *taste* is also functional in the fetus at the end of the gestation period. In one experiment, rat fetuses received intraoral infusions of milk (a biologically relevant stimulus) or lemon (a novel chemical stimulus) on days 19, 20, or 21 of gestation. They were then observed *in utero*, that is in an "externalized" uterus in saline, removed from the mother's body but still connected to it. Their motor activity showed that they discriminated these stimuli. They responded to milk with delayed fetal movements of low magnitude, while lemon provoked high-magnitude, spiked patterns. Late in gestation, milk elicited stretching, as at the nipple, while lemon caused face wiping, a motor pattern used by rat pups or adults to aversive gustatory stimuli (Smotherman and Robinson, 1987).

A similar study examined the role of *experience*, using the movements of rat fetuses in an "externalized" uterus in response to intraoral infusion of lemon or mint extracts on day 19 of gestation. Prior experience does indeed affect fetal responses. There is *habituation* to repeated stimulation, *central processing* of sensory information, fetal *orienting reflexes* to novel stimuli, and prenatal states exist that are associated with different response patterns (Smotherman and Robinson, 1988).

Aversive conditioning

Rat fetuses exposed to apple juice injected into the amniotic fluid and lithium chloride intraperitoneally on gestational day 20 will avoid the odor of apple juice after birth. They will not attach to nipples that have been treated with apple juice (Stickrod *et al.*, 1982). They will also reduce their wheel-running speed when the odor of apple juice is presented (Smotherman, 1982). This fetal conditioning may help the pups to avoid foods that made their mother sick. Prenatal learning occurs even earlier: a combination of mint odor and lithium chloride given to rat fetuses on day 17 of gestation leads to conditioned aversion to mint 2 days later (Smotherman and Robinson, 1985). The heart rate can be conditioned in response to intrauterine application of apple juice. This olfactory conditioning ontogenetically precedes visual and auditory conditioning of the heart rate, just as somatomotor conditioning does (Sananes *et al.*, 1988; Fig. 9.1).

Particularly alarming are fetal effects of *alcohol* and *drugs* on food-related odor responses in humans. Apart from the severe fetal alcohol syndrome, alcohol can affect the chemosensory behavior of a fetus. Alcohol administered to pregnant female rats impaired odor aversions and preferences in their offspring. A

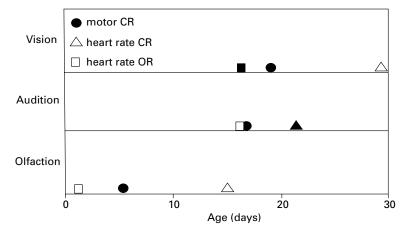


FIGURE 9.1 In rat pups, sense of smell matures first among different sensory modalities. Within the sense of smell, conditioning of somatomotor responses to odors occurs earlier (\bullet) than conditioning of heart rate (\triangle). The earliest occurrence of "orienting response," (i.e. a heart rate deceleration [\square]) in response to the first appearance of a conditioned odor. (Redrawn from Sananes *et al.*, 1988.)

liquid diet provided the mothers with either 35 or 0% of the calories in the form of ethanol. The offspring of the group receiving 35% ethanol did *not* develop an odor *aversion* to lemon odor at 10 days of age when this odor was followed by a lithium chloride injection, making the rat sick. Furthermore, if the pups were infused with milk through a cannula, paired with banana odor, the ethanol group failed to develop a preference for banana odor. However, these effects appeared to fade with age. As adults, animals in the ethanol group managed to associate banana odor with drinking water laced with lithium chloride if the odor was present when they became ill (Barron *et al.*, 1988).

Maturation of chemosensory systems

Although the vomeronasal system is specialized to detect stimuli in a liquid environment, it probably is not functional *in utero*, at least in mice. Fluorescent microspheres were not taken up by the vomeronasal organ as the access canal is not open yet *in utero*. In rats, by contrast, the canal is open before birth and the microspheres can be taken up. The olfactory epithelium of the main olfactory system plays a greater role prenatally, as evidenced by the uptake of radiolabeled 2-deoxyglucose (Coppola and Coltrane 1994). Fetal mice respond to amyl acetate and isovaleric acid delivered into the nasal cavity through a tiny cannula (Coppola, 2001). In both rats and mice, the main olfactory system, and not the vomeronasal system, appears to mediate prenatal olfaction (Coppola, 2001).

In utero priming: freemartins

Androgen from the mother or embryonic siblings affects the development of embryos, a form of priming. Males produce testosterone from about day 12 of gestation and male neighbors *in utero* androgenize female siblings. Animal breeders have called such females with male features "freemartins." The effect depends on intrauterine positions. Males on both sides of the female fetus in the uterus (2M condition) have a stronger effect than a single male sibling on only one side (M). Female fetuses with no male neighbor (O condition) provide convenient controls. The androgenizing effects include larger anogenital distance, lower uterine weight, less body fat, delayed puberty, longer and irregular cycles, and an altered sex ratio. In one experiment, control females had 40% male pups, M females 50%, and 2M females 60% while the overall population had 50% males (Vandenbergh and Hotchkiss, 2001). The androgenized females differed in their postnatal behavior from normal females, showing altered odor preferences, urine marking, and olfactory priming of other mice, especially puberty delay (Drickamer, 2001b).

Male mice prefer O or M females or their odor when confronted with soiled traps in the field, while females prefer 2M females and their odor in such contaminated traps. Also, the larger the anogenital distance (i.e. the more masculinized a female), the more urine marking it performed in an arena. The 2M females were less responsive to chemical puberty-delay cues from crowded female mice and also less sensitive to puberty-accelerating male chemosignals (Drickamer, 2001b). They had high levels of serum testosterone and were more infanticidal, while 2M males were less so, instead showing more parental care. Finally, both males and females of the 2M type were more aggressive in encounters with mice of the same sex of the O type (Drickamer, 2001b).

The production of odors in utero can have social and reproductive consequences for the adults: Male mice can smell the fetal MHC odortype as part of the mother's odortype (Beauchamp *et al.*, 2000).

Prenatal humans

The rich chemical ecology of the uterus plays an important role for the human fetus. The chemical senses and metabolism of the fetus are bombarded with its own waste products; the aromas, alcohol, or drugs in the mother's diet; and her inhaled substances such as perfumes or tobacco. Preterm neonates respond reliably to the odor of mint at the gestational age of 7–8 months, and some responses are observed at 6 months of gestational age (reviewed in Schaal, 1988b). Premature newborns, aged 28 to 33 gestational weeks, distinguish the

odors of vanilla and butyric acid (Marlier *et al.*, 2001). Prenatal exposure to maternal food flavors via the amniotic fluid is thought to prepare the infant for neonatal preferences (Schaal and Orgeur, 1992). Infants born to mothers who had consumed anise flavor during pregnancy preferred anise odor during their first 4 days after birth. By contrast, offspring of mothers on a diet without anise flavor displayed aversion or neutral responses to anise odor at the same age (Schaal *et al.*, 2000).

Human infants do not discriminate between their mother's amniotic fluid and her milk until 3 days after birth. However, they distinguish well between their mother's amniotic fluid and milk formula. Thus, chemical information acquired *in utero* is used after birth (Schaal *et al.*,2001). Three-day old infants discriminate familiar from unfamiliar amniotic fluid, whether they are being breast-fed or bottle-fed (Schaal *et al.*,1998).

In humans, just as in mice, hormones can act as priming pheromones. Androgens from the mother in the condition known as congenital adrenal hyperplasia masculinize daughters. With increasing use of assisted fertility, heterozygous (fraternal) twins are becoming more frequent. A male sibling will androgenize a female fetus. Both maternal and sibling masculinizing influences have behavioral effects. These include altered spatial ability, sensation seeking, selection of more masculine toys, more masculine-type sexual orientation, and fantasies more related to lesbian or bisexual lifestyles. Moreover, we are just starting to understand how endocrine disruptors in the general environment affect human behavior. So far most of these compounds are estrogen mimics (Vandenbergh and Hotchkiss, 2001).

9.5.2 Postnatal development

Sensory development points to the importance of chemical cues very early in life after birth. From the fourth day of life, mouse pups respond to the odor of their nest (Schmidt *et al.*, 1986). In rats, the main olfactory system processes odor responses during the first days of life.

Newborn mammals use odor cues to find the nipples of their mother and to attach there. Piglets deposit saliva on the teats of their mother that contains 16-androstenes. These androstenes are already present in the saliva of pig fetuses (Gower and Booth, 1986). Piglets learn the odor of their mother within 12 hours after birth. They discriminate between mother and non-mother female odors and are most attracted to odors of feces and skin secretions. Novel odors such as orange or banana are not preferred. The putative maternal pheromone deoxycholic acid was not preferred by piglets. In a Y-maze, piglets were attracted to feces of the mother, not to her colostrum or ventral washings. Ventral washings

may be important at close range for nipple finding, however. The critical period for acquiring an attraction to maternal fecal odor lies between birth and 12 hours of age. The preference persists in 14- and 21-day old piglets. Two important reasons for being attracted to maternal feces may be to obtain iron and other nutrients, and to be protected by bile acids against *Escherichia coli* toxins (Morrow-Tesch and McGlone, 1990).

Newborn rabbits (*Oryctolagus cuniculus*) are guided to the nipples of their mother by an odor, the "nipple pheromone" (Hudson and Distel, 1983). The vomeronasal organ is not necessary for this response (Hudson and Distel, 1986). Once mobile, the pups discriminate and are attracted to the anal gland secretion of their mother. They discriminate mother's odor from those of other females or males after the age of 10 days. Behavioral responses are not always obvious; the frequency of nostril movements, indicative of changed respiration rate, is higher in response to mother's anal gland odor and there were fewer "rejection" behaviors in the presence of this secretion. It was concluded that rabbit nestlings acquire the preference for the odor of the mother during the first days of their life (Mykytowycz and Ward, 1971).

The *amniotic fluid*, deposited by a rat on her own teats, can guide her pups to the teats to initiate suckling (Teicher and Blass, 1980; Pedersen and Blass, 1982). In domestic sheep, only inexperienced (primiparous) ewes need amniotic fluid on the lamb's coat for normal development of maternal behavior, such as licking the young and accepting it at the udder. Experienced (multiparous) ewes lick their lambs less if amniotic fluid is removed but otherwise accept them (Levy and Poindron, 1987). The amniotic fluid links mother and offspring chemically, as it stimulates in both directions. In the rat, it serves kin recognition, since pups prefer their mother's amniotic fluid over that of other females, even after caesarean section (Hepper, 1987), as mentioned earlier. The responses by sheep to amniotic fluids of sheep, cows, and goats show some specificity: cow amniotic fluid was always repulsive to sheep, while caprine and ovine amniotic fluid were repulsive only before parturition. Ovine amniotic fluid becomes attractive after parturition while caprine fluid merely loses its repulsiveness (Arnould *et al.*, 1991).

Spiny mice, *Acomys cahirinus*, are the only murid species with precocial young. This permits study of the postnatal development of odor preferences as expressed by active (locomotor) behavior. When given choices of own (familiar), unfamiliar conspecific odor, and "no odor" (clean shavings), infant spiny mice prefer (i.e., stay longer on) odor from another litter (unfamiliar odor) but avoid clean shavings at the age of 3 days. By day 5, they slightly prefer to remain near their parents; by day 7 and 9 they clearly prefer their parents (Birke and Sadler, 1987b).

Odors can affect the survival of newborn rat pups: 100% of pups normally born on gestation day 21 survive, and so do pups delivered by Caesarean section on day 20. If the pups are exposed to odors for 1 hour after birth, the survival rate varies greatly: 9% survive in the presence of the odor of amniotic membranes and placenta; 80% with dimethyl disulfide (which occurs in rat saliva); 75% with no odor; and only 50% with mint odor. In addition to survival, the activity levels of the pups differ. Pups are more active in the presence of odor of amniotic fluid and placenta than they are in the presence of mint odor (Smotherman *et al.*, 1987).

One-day old rat pups show strong *negative* responses to strong taste stimuli such as strong acids or quinine solutions. They gape and flail with their forelimbs. At 12 days of age, they scrape their chin and tread with their paws (Johanson and Shapiro, 1986).

Humans

General odor responses

Newborns respond to odors by rapid breathing and activity changes. They can detect odors, rapidly develop their sensitivity over the first 4 days of life (Lipsitt *et al.*, 1963), discriminate odor qualities and intensities, memorize odors for 1 day (and possibly for life), prefer odors they had experienced earlier, and localize odor sources (e.g. newborns aged 1 to 5.5 day olds turn away from a cotton swab with ammonia; Rieser *et al.*, 1976). Neonates younger than 12 hours show olfactory preferences and aversions even before they experience their first food.

Infants are able to acquire odor preferences on the first day of life. In one experiment, 12 male and 12 female white, healthy, full-term neonates were exposed to the odors of cherry or ginger on a pad taped to the inside of their crib for 24 hours. After this exposure, they were tested for preferences during active sleep (stage II). The behavior was videotaped and the duration of time oriented to each odor measured. Only the female neonates showed a preference for the familiar odor (Balogh and Porter, 1986). Therefore, even on the first day of life, females outperform males, as often described for children and adults (e.g. Yousem *et al.*, 1999).

Development of responses to conspecific odors

In human infants *social odors* are important from a few days after birth, as indicated by head turning in the direction of odors. In 1877, Darwin observed that an infant with its eyes closed would turn its head towards its mother when her breast was brought near. Darwin suggested that the infant might be attracted

by the mother's odor or body heat. Hungry sleeping infants turn their heads and show the rooting reflex. At 2 days of age, head turning is random; while some infants orient toward odors with 6-8 days, they all do by 6 weeks. Babies discriminate breast odors: they choose the odor of the mother's breast areola over no odor or odor from another lactating woman (MacFarlane, 1975). Head and arm movements decrease when the infant is exposed to its mother's breast odor (Schaal et al., 1980). Sleeping infants respond with mouth and head movements to bottles with mother's milk but not to empty bottles (Schaal et al., 2001). Infants also discriminate neck and axillary odors. It was concluded that infants are not able to identify the odor of their mothers at 2 days of age but can discriminate her from an unfamiliar mother by the second week after birth. This may be biologically adaptive. Historically, in humans, maternal death rate was high. Therefore, it was adaptive for human babies to form irreversible attachments later than in other mammals. This would permit adoption of alternative parents (MacFarlane, 1975). Breast-fed infants learn to discriminate their mothers' axillary odor, while bottle-fed babies are not able to do so (Cernoch and Porter, 1985). The question arises whether bottle-fed children will subsequently suffer any other, lasting deficiencies in their social behavior.

In summary, postnatal learning, *in utero* experiences, and genetic predispositions play a role in individual olfactory recognition in humans. Learning is demonstrated by the fact that breast-fed, but not bottle-fed, neonates discriminate their mother's odor from that of other lactating women. Artificial odors worn by their mothers are also preferred by infants. Inborn preferences account for the attraction of breast-fed infants to odors of unfamiliar lactating females, although they prefer their own mother's odor. Even bottle-fed infants orient to breast odors from unfamiliar lactating women when presented with the choices of axillary odor, the odor of their own formula, or breast odors from non-lactating females (Porter *et al.*, 1992).

Children 28–36 months of age chose T-shirts worn by their *mothers* over those from other women in over 72% of cases. A group of children aged 45–58 months was over 60% correct (Montagner, 1974). Children of that age group also discriminate between T-shirts worn by siblings and non-siblings (Porter and Moore, 1981).

Preschool children are reputed to differ from adults in their odor preferences. For instance children aged 3–4 years were as likely to "like" the odor of amyl acetate (banana) as synthetic sweat or feces odors. By 6 years of age, their preferences resembled adults, liking banana, and disliking sweaty and fecal odors (Stein *et al.*, 1958). At 4–5 years of age, a shift occurs from positive or neutral to negative characterization of odors of sweat, feces, asa foetida, or butyric acid. However, it is increasingly becoming clear that responses by very young children

depend much on how or what they are being asked. For instance, 4–6 year olds respond more positively if asked "tell me if this smells pretty" than when asked "tell me if this smells ugly" (Engen, 1974a). The preferences by 3-year olds in a forced choice played as a game did not differ from those of adults. They were asked to place pleasant odors near a popular puppet ("Big Bird") and unpleasant ones near a doll of a "bad" character ("Oscar the Grouch"). This first study not only found hedonic differences in children under 5 years of age but also established that children are more sensitive than adults to the steroid androstenone. Adults liked androstenone more often than children, and 29% of the children liked the odor of pyridine which adults never did (Schmidt and Beauchamp, 1988). The hedonic hierarchy of some odors is similar for children aged 1, 2, and 3–5 years old as for adults. For instance, the following sequence from most to least liked was the same for all these groups: lavender, amyl acetate, butyric acid, dimethyl disulfide (Bloom, 1975; in Schaal, 1988b).

Odors are not easily forgotten. The recognition of an odor depends on the original coding activity. When trying to recall an odor, life episodes related to that odor and the ability to label and define the odor enhance odor memory. Visual imagery, by comparison, did not enhance odor memory (Lyman and McDaniel, 1986).

Learning responses

Adults continue to associate new odors with pleasant and unpleasant situations in social and sex life, work and recreation, and concerning food and drink. The human patterns of odor recognition and preferences do not merely involve the olfactory nerve and its central projections. Learned associations are formed and stored in memory. To retrieve odor information, we need affective and cognitive components, as well as verbal descriptors. Without the latter, an odor appears familiar but cannot be labeled, the "tip-of-the-nose-phenomenon" (Lawless and Engen, 1977).

In old age, humans experience a precipitous drop in olfactory ability (Doty *et al.*, 1984; Doty, 1986; see Section 5.7). Two excellent recent reviews of the ontogeny of human olfaction and olfactory communication are by Doty (1986) and Schaal (1988b).

I conclude this section with a science fiction idea. If nursing an infant (instead of bottle-feeding) fosters olfactory discrimination (Cernoch and Porter, 1985), would it be desirable and possible to train noses early on? Should babies be breast-fed to give them a chance to become gourmets, sophisticates, wine or tea testers or would we be doing them a favour by bottle-feeding them to render them less sensitive to unpleasant odors such as pollution in our air?

9.6 Learning

The odor preferences of rat pups aged 3–6 days can be reversed by training. If the initially avoided odor of orange extract is presented together with maternal saliva, the pups will later orient toward orange odor. Thus, conditioning can enhance the value of an unfamiliar odor (Sullivan *et al.*, 1986).

The newborn rat is a "natural split-brain preparation" for olfactory learning protocols: it can be trained to associate an odor with a milk reward via just one nostril. If the other nostril is tested, the animal shows no preference. However, at 12 days of age or later, the two sides of the brain are connected and a learned preference occurs with either nostril open. The information is stored *uni*laterally: the animal shows unilateral preference if the commissure is cut *after* training. The maturation of the commissure pathways occurs between 6 and 12 days of age. In summary, unilaterally represented memories remain unilateral, even after bilateral retrieval processes have developed. The mnemonic storage capacity of the brain is increased by confining memory to one side (Kucharski and Hall, 1987).

Home odor can modulate certain behaviors in infant mammals. For instance, rat pups aged 7, 9, and 11 days learned better to escape from a shock if the odor of soiled bedding from their own cage was present than they did in the presence of clean shavings. The familiar odor may reduce a heightened state of arousal in an unfamiliar environment (Smith and Spear, 1981). At the age of 17 days, rats spend less time in the dark half of an apparatus if home nest odor is present, but at 30 days no effect of home nest odor was noted (Richardson and Campbell, 1988). Similarly, 16-day-old rats take 12 trials to learn not to enter the black compartment of a black-and-white shuttle box if trained with clean shavings placed under the apparatus, but only seven trials with soiled litter from their home cage available (Smith and Spear, 1978).

9.6.1 Critical periods

The critical period for developing a preference for the *maternal pheromone* in rat feces occurs at 14 to 16 days of age (Leon and Behse, 1977). Non-rat odors such as pure mint or peppermint can be made as attractive as maternal feces to rat pups under the age of 21 days. At that age, simple exposure suffices. A preference is acquired whether the odor is simply presented or pasted on the mother. But the older the pup, the more important the maternal context becomes, and pups up to 33 days old can acquire peppermint preference only if it is presented on the mother. The waning of the response is also much slower for the odor experienced on the mother. After 20 days of age, the rats acquire preferences less easily (Galef, 1982; Galef and Kaner, 1980). Postnatal conditioning to odors is facilitated not

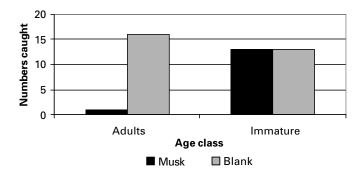


FIGURE 9.2 Immature muskrats do not avoid live traps scented with musk from adult male muskrats, as adults do. (After data from van den Berk and Müller-Schwarze, 1984.)

only by the mother as background but also by *maternal stimulation*. If neonatal rats are exposed to an odor and at the same time receive tactile stimulation that mimics maternal contact, they will prefer that odor later. That odor will also increase uptake of 2-[14C]-deoxyglucose by the olfactory bulb. Various types of moderate stimulation, such as an odor coupled with high humidity, can bring about this preference (Do *et al.*, 1987). This early olfactory learning depends on noradrenergic factors.

Woodmice, *Apodemus sylvaticus*, are able to walk when 11 days old and increase their locomotion between postnatal days 11 and 19. When moving about, they spend more time on home (nest) and male odor than on female odor and clean bedding. This difference is not observed if the woodmice are tested for the first time on day 15, suggesting a critical period for acquiring these odor preferences (Pontet and Schenk, 1988).

Sexually immature mammals often ignore territorial odors that are important signals for adults. For instance, juvenile muskrats, *Ondatra zibethica*, were caught equally often in clean live traps and traps scented with musk (anal gland) secretion from adult male muskrats, while adults avoided the musk (van den Berk and Müller-Schwarze, 1984) (Fig. 9.2).

9.6.2 Long-term effects of early experience: olfactory social imprinting

In mammals, early experience with social odors will have lasting effects on odor preferences. We have to distinguish *filial imprinting* (attachment of young to mother) from *maternal imprinting* (attachment of mother to young) in precocial mammals.

Filial imprinting

In a classical experiment Mainardi (1965) exposed male mouse pups in their nest to an artificial odor, Chanel No. 5. As adults, these mice preferred females treated with Chanel No. 5 over those with regular mouse odor. Mammals differ from birds in that learning plays a critical role in acquiring social preferences; the sensitive period is more drawn out than a narrow "critical period," and effects are not necessarily permanent, and they can be overridden more easily by subsequent relearning.

Even the body odors of mice that genetically differ only at the major histocompatibility complex (MHC) are learnt by their offspring. MHC, called locus H2 is located on chromosome 17 (out of 20 chromosomes in the mouse). Normally, mice of both sexes prefer to mate with partners of a different MHC type. In one experiment, entire litters of mouse pups were fostered onto parents of the same or a different MHC type. In males, the mating bias appears to be acquired during early development through MHC-controlled signals from the parents: males fostered onto parents of a *different* MHC type mated more often with females of the *same* type. The latter now represented a "strange" odor. The foster rearing produced a reversal of preference; males mated with a female whose odor was different from that of their foster parents. In females, the mating bias was unaffected by the fostering history (Yamazaki *et al.*, 1988).

The MHC odor types are present as early as day 1 after birth. The odor type of a pregnant female is a combination of the fetal and maternal odor types, and a male mouse can smell the fetal odor (Beauchamp *et al.*, 2000).

Long-lasting and specific effects of early odor experience have not yet been reported for many mammal species. In one experiment, rat mothers that raised male pups had their nipples and vagina scented with citral. When tested at 100 days of age, these males ejaculated sooner with sexually receptive, citral-scented females than with controls. Conversely, the latency to ejaculation of males reared with saline-treated mothers was shorter in matings with "normal" females than with citral-scented ones (Fillion and Blass, 1986).

Rabbit pups learnt an odor of their mother in one trial. This odor emanates from the belly of the dam and releases suckling. This rapid learning is considered a form of olfactory imprinting tied to an early sensitive period (Hudson and Altbäcker, 1994).

Maternal imprinting

Maternal imprinting occurs more rapidly than filial imprinting. An ungulate mother such as a goat (Klopfer and Gambale, 1966) or black-tailed deer (Müller-Schwarze and Müller-Schwarze, 1971) will irreversibly and exclusively

attach to the odor of her newborn within a few minutes, while the young can still be bonded with their mother up to several days later.

Birth odor of lambs is used by ewes to discriminate their own from alien lambs (Alexander *et al.*, 1987), while in goats the mother appears to "label" her kid with her own odor (Gubernick and Klopfer, 1980).

Imprinting of prey odors

Ferrets, *Mustela furo*, acquire knowledge of prey odors as young animals. This search image is not modifiable at later ages (Apfelbach, 1973).

9.6.3 Odor learning in adults

Even adults can still develop olfactory preferences that contravene those acquired before sexual maturity. Female laboratory mice "imprinted" by the odor of one mouse strain will prefer this odor even more if they are exposed to males of this strain as adults. However, if they are exposed to males of a different strain when sexually mature, their original odor preference will be reversed (Albonetti and D'Udine, 1986). Naturally occurring sex or body odors may assume their sexual significance after association with sexual activity: male mice were aroused by a perfume that they had experienced earlier on scented females they had interacted with (Nyby et al., 1978). Practitioners have known that adult mammals can acquire responses after exposure to certain animals. For instance, bulls of the Asian elephant that had been housed near African elephant bulls respond to temporal gland secretion and its three components phenol, 4-methylphenol, and (E)-farnesol from the latter species. Asian bulls that had not been associated with African bulls did not respond (Rasmussen, 1988).

Short-term memory for odors

Adult rats investigate a juvenile rat they have encountered 30 minutes before much less than a completely new individual. This "recognition memory" lasts about 80 minutes and is termed a *transient* memory. The anticholinergic drug scopolamine (hyosine) abolishes the decrease of investigation during the second encounter. It was postulated that scopolamine "disturbs chemosensory receptivity in rats" (Soffie and Lamberty, 1988).

Neural mechanisms

The brain sites that are concerned with early olfactory learning of social cues include the olfactory bulb. In *rat pups* blocking the glutamate receptors

of the N-methyl-D-aspartate type in the olfactory glomeruli by 2-amino-5-phosphonovaleric acid prevented both the takeup of 2-[14C]-deoxyglucose in the glomeruli and the development of a preference for a *learned maternal odor* (Lincoln *et al.*, 1988).

The organization of the olfactory bulb during the first 2 postnatal weeks may be affected by specific olfactory experiences. Early experience may be incorporated into the firing pattern of olfactory neurons with the result that the mature brain responds in a particular, conditioned way to the now-familiar odor. Such response changes of the mammalian brain to naturally occurring odors as a consequence of specific early experience may be an important component of individual variation (Coopersmith and Leon, 1986).

The processing of biologically relevant odors often changes drastically around the time of giving birth. In sheep, parturition has profound effects on the ewe's response to lamb odors. Within 3 hours of parturition, a lasting bond between mother and lamb is formed. Before the lamb is born, ewes reject the smell of amniotic fluid, but they are attracted to it immediately after giving birth. During parturition, the vagina and cervix are stimulated, leading to effects on the brain that render the ewe maternally responsive and capable of bonding with her lamb. Single mitral cells in the olfactory bulbs of ewes respond only to food before parturition but respond also to lamb odors after parturition. Between 3 and 4 days after giving birth, the number of cells responding to lamb odors increases dramatically until 70% of the cells respond to any lamb odor, and the remaining 30% preferentially to the odor of the own lamb. The odor of wool was almost as active as that of the whole lamb. *In vivo* sampling of the external plexiform layer revealed neurotransmitter interaction between granule and mitral cells. Both the excitatory amino acid glutamate, and the inhibitory γ -aminobutyric acid increased during the first 5 minutes of exposure to lamb odor (Kendrick et al., 1992).

Ontogeny of odor production

Odor types distinctive of *H2b* and *Hdh* genotypes already appear in the urine of 1-day-old mouse pups (Yamazaki *et al.*, 1992a). At that age, the normal intestinal bacterial flora is not yet present and hence is not necessary for the odor (Yamazaki *et al.*, 1992b). In addition to MHC, gene(s) on the X and Y chromosomes and other autosomal genes contribute to individual odors (Yamazaki *et al.*, 1986, 1990).

Allomones I: chemical defense by animals

The 'bonnacon' [bison] "... emits a fart with the contents of his large intestine which covers three acres, any tree that it reaches catches fire and its pursuers are driven off with the noxious excrement..."

[Clearly, an exaggeration] ANON (c. 1150) Bestiary (transl. and ed. T. H. White) New York: Putnam.

Allomones are interspecific semiochemicals that primarily benefit the sender. Animals as well as plants defend themselves chemically against predators. Myriads of ways to deter predation have evolved. Many chemicals have more than one function, being aimed not only at predators but also at parasites, prey, or conspecific competitors. As common denominator of allomones we assume that the inclusive fitness of the sender – rather than the receiver – is enhanced. Chapter 10 deals with defenses by animals, and Chapter 11 with plant defenses against herbivores.

Numerous ingenious chemical defense mechanisms of vertebrates protect against predators, but also against competitors and fierce prey. These range from innocuous odors of prey animals or plants that signal that they are unpalatable to some of the most toxic secretions known in nature. Slower animals tend to rely more on chemical and mechanical defenses. Humans have known animal toxins since time immemorial and medical research has elucidated the mechanisms involved in great detail. However, ecologists have investigated why animals and plants have poisons and venoms in the first place only since the 1950s.

10.1 Fish

Among fish we find some of the most potent toxins in animals. We distinguish passively toxic fish from actively toxic fish. The former simply have toxins in their tissues, typically taken from some other source such as their diet. The latter produce the poison and have evolved apparatus to discharge, deliver, or inject the toxins.

FIGURE 10.1 Ciguatoxin. It passes through food chain from dinoflagellates to herbivorous and carnivorous fish, and humans.

10.1.1 Passively toxic fish

Over 500 species of fish can poison humans (and presumably other consumers) when eaten. The *ciguatoxin* ciguatera moves through the food chain, starting from dinoflagellates (specifically *Gambierdiscus toxicus*). The dinoflagellates reside on macroalgae that herbivorous fish will eat. Carnivorous fish then acquire the toxin from their prey, herbivorous fish, and store it in liver, testes, and intestines, but less in muscles. Humans are poisoned when consuming fish such as mackerel, coral trout, and cod. The toxin is odorless and tasteless, and it survives cooking and freezing. Several hours after the meal, breathing becomes difficult. Poisoning does not lead to immunity. On the contrary, a second poisoning usually proves more serious than the first one. The first report of *ciguatera* poisoning from eating yellowtail tuna appeared in China around AD 700. Ciguatoxins are polyether (polycyclic) compounds that contain a series of rings (Fig. 10.1).

Among the most poisonous of all marine animals are the puffer fish (Tetraodontidae). The ancient Egyptians knew puffer fish to be poisonous and depicted it on the tomb of Pharao Ti in 2500 BC. Puffer fish take up alkaloid toxins such as tetrodotoxin (Fig. 10.2) from algae (*Shewanella* sp.) and bacteria (Yasumoto and Murata, 1993; Yasumoto *et al.*, 1986). The nerve poison resides in liver, gonads, intestines, and skin, but not in the musculature. Therefore, the flesh is generally edible, provided other organs do not contaminate it during processing. The nerve damage starts as a tingling feeling on tongue and lips and spreads down the body. Over 60% of those poisoned die, and death is rapid and violent. Captain James Cook nearly died of (presumably) tetrodotoxin after eating fish in New Caledonia. Eating puffer fish amounts to a game of Russian roulette. In Japan, puffer fish is known as fugu and is prepared by skilled fugu chefs, demanding highest prices. Some toxic species are *Fugu rupripes*, *E. vermicularis*, *F. pardalis*, and the white-spotted ("death") puffer, *Arothron hispidus*.

$$H_2N^+$$
 H_2N^+
 H

FIGURE 10.2 Tetrodotoxin, a toxin from pufferfish and newts.

Besides the Tetraodontidae, the order Tetraodontiformes has very toxic fish in the families Diodontidae, Canthigasteridae, and possibly Molidae and Triodontidae.

Soles of the genus Pardachirus possess one of the better-understood chemical defense systems in fish. The Red Sea Moses sole, Pardachirus marmoratus, dubbed "the Red Sea's sharkproof fish," and the related peacock sole, Pardachirus pavoninus, of the western Pacific secrete from the mucus glands on their dorsal and anal fins a milky substance that is retained in the mucus coat of the skin. This secretion repels sharks, kills starfish and sea urchins on contact, and is hemolytic. From these secretions, two groups of ichthyotoxic compounds have been isolated. The pardaxins are peptides consisting of 33 amino acid residues and physically and pharmacologically resemble melittin, the toxic peptide in the venom of the honeybee, Apis mellifera. They are strongly surfactant, owing to hydrophobic and hydrophilic parts of the molecule. Several pardaxins kill killifish, Oryzias latipes, within 30 minutes at a concentration of 25 μg/ml. Pardaxins act on the gustatory sense: white-tip reef sharks, Triaenodon obesus, escape from pardaxins (100 mg dissolved in 5 ml seawater) that are placed into their mouths. The toxin affects the gills, where osmoregulation is disturbed (Primor et al., 1978). Steroid monoglycosides from P. pavoninus have been named pavoninins (Fig. 10.3) and those from P. marmoratus are mosesins (Fig. 10.4). Both together are also known as pavoninins. These pavoninins are both hemolytic and strong repellents for dog sharks, Mustelus griseus, and lemon sharks, Negaprion brevirostris. They act on the shark's olfactory sense (Tachibana et al., 1984). Needless to say, pardaxins and pavoninins offer fascinating prospects as shark repellents.

Moray eels (Muraenidae) of tropical reefs possess a water-soluble poison. Poisoning occurs from eating the flesh.

10.1.2 Actively toxic fish

About 250 fish species possess various kinds of venom apparatus. The stingray and scorpion fish belong in this category. Stingrays have a

OR

OB-glcNAc

Pavoninin 1:
$$R = COCH_3$$

Pavoninin 2: $R = H$

OH

OB-glcNAC

Pavoninin 4

Pavoninin 4

OB-glcNAC

Pavoninin 5

FIGURE 10.3 Pavoninins from the peacock sole.

Mosesin 1: R^1 = OH R^2 = H R^3 = 6-Acetyl- β -D-galactose R^4 = OH R^5 = COCH $_3$ Mosesin 5: R^1 = OH R^2 = H R^3 = β -D-galactose R^4 = OH R^5 = COCH $_3$

FIGURE 10.4 Mosesins from the Red Sea Moses sole.

sting apparatus at the end of their tail, with poison glands at the teeth of the spine. The active venom contains proteins with a molecular weight of 100 kDa.

The perfectly camouflaged stonefish (Scorpaenidae) blend into rocks and dead corals and are extremely poisonous. On their dorsal fins, they have 13 needle-like spines with poison glands at their bases, used for defense. As the fish are sluggish, poisoning occurs by stepping on the half-buried fish. It is painful and can be fatal within a few minutes or hours. Divers need to exercise care, because the stings even penetrate rubber. There are about 20 species of stonefish, all occurring in shallow waters and coral reefs of the Indo-Pacific. Two better-known species are *Synanceja trachynis* and *Synaneichthyes verrucosus*. Table 10.1 summarizes some chemical defenses in fish.

Predatory fish may also be affected by alarm pheromones (Section 7.2) of the prey, both directly and indirectly. The alarm odor may act as defense compound that inhibits predator attack or reduces capture rate by inducing predator avoidance in school members of the prey species.

Table 10.1 Defense compounds in fish

Emitting species	Compound	Class of compound	Target species	Effect
Stingrays Dasyatidae	Not a phospholipase	Protein	Defense in general	Pain, death (1%)
Box fishes Ostraction spp.	Pahutoxin etc.	Esters of palmitic acid	Predators?	
Red Sea Moses sole Pardachirus marmoratus	Pardaxins	Peptides (33 amino acid residues)	Sharks	Affect taste: shark withdraws
	Mosesins	Steroid monoglycosides	Sharks	Affect olfaction, repellent
Peacock sole Pavonininus pavonininus	Pavonins	Steroid monoglycosides	Sharks	Shark repellent, affect olfaction
Puffer fish Fugu spp.	Tetrodotoxin (found in Vibrio, Pseudomonas spp.)	Alkaloid	Predators?	Local anesthetic (5–30 minutes); death in 6–24 hours (60% mortality)

10.2 Amphibia

Amphibian skin proliferates with mucus and granular glands. The granular glands produce defense compounds. The chemistry of these secretions is diverse, including biogenic amines, peptides, steroids, and alkaloids. Some of the most toxic compounds known are found in amphibians. Among amphibians, chemical defense is found in sluggish species such as salamanders, larval amphibians, and toads.

10.2.1 Urodeles

Many urodeles produce irritating or even toxic secretions in their skin glands, and some are extremely toxic. The rough-skinned newt (*Taricha granulosa*) is the most toxic of newts from several genera. Its dorsal skin and ovarian eggs contain the very potent tetrodotoxin. Only 0.00005 ml of newt skin is needed for a lethal peritoneal injection in mice. The skin of one newt can theoretically kill 25 000 mice. When bothered, the newt assumes a characteristic defense posture: it arches, lifts its tail and releases an odor (Brodie *et al.*, 1974).

The level of tetrodotoxin in the skin of a particular female rough-skin newt correlates with that in her eggs. The amounts in the eggs of one clutch vary little, but there is considerable variation between clutches. These findings suggest that the mother passes on the toxin to her eggs. As the amount of toxin is not correlated with egg size, it may not be transferred automatically (passively) with maternal resources to the egg. Rather, the mother may exert some control over the amount allocated to the eggs (Hanfin *et al.*, 2003).

In certain geographical areas, termed "hotspots of snake-newt coevolution," rough-skinned newts are very toxic, and garter snakes (*Thamnophis sirtalis*) have adapted by tolerating high tetrodotoxin levels. The snakes accumulate tetrodotoxin in their livers where it remains for 1 month or longer. After eating one newt, snakes averaged 42 µg tetrodotoxin in their livers. This amount would severely affect avian predators of garter snakes such as harriers, hawks, bittern, or crows, and possibly kill them (Brodie *et al.*, 2002). Mammalian predators would be less sensitive to the toxin (Williams *et al.*, 2004).

Are these toxic compounds of urodeles effective against predators? Short-tailed shrews (*Blarina brevicauda*) prey on these animals; their burrows often contain the remains of salamanders. Laboratory tests confirmed that their chemical defenses are effective: noxious species of salamanders took the shrews longer to kill and had a higher survival rate than non-toxic species. The shrews needed more time to kill the toxic salamanders, and wiped their mouths more often (Brodie *et al.*, 1979).

A non-toxic species of salamander may derive protection from predators by visually resembling a toxic form so closely that predators cannot distinguish between them (*Batesian mimicry*). Free-ranging birds avoid both the toxic red eft (*Notophthalmus viridescens*) and the similar-looking non-toxic red morph of the red-backed salamander (*Plethodon cinereus*). The red-striped morph of *P. cinereus*, which does not resemble the red eft, is eaten (Brodie and Brodie, 1980).

Palatability may change during individual development: the premetamorphic larvae (tadpoles) of seven species of North American frogs, toads, and salamanders are palatable to the predaceous diving beetle *Dytiscus verticalis*, while the metamorphic stage (with limbs) of five of these species is unpalatable. The palatability is inversely correlated with the number, but not the size, of skin glands. Before metamorphosis, the tadpoles can flee quickly, and this primary defense suffices for survival. With legs, the larvae are slower and probably for this reason have to depend more on secondary, chemical defense (Formanowicz and Brodie, 1982).

Predators *learn* to avoid distasteful salamanders. When molested, spotted salamanders (*Ambystoma maculatum*) discharge a white slime, mostly in the tail region. They also raise and wag their tail when a predator is near. In one experiment, four out of five chickens learned to avoid these salamanders by *sight* after

FIGURE 10.5 Batrachotoxin from dart-poison frogs.

14 trials. It is important to know that these salamanders are merely distasteful: the chickens ate 6 of 70 salamanders that were presented without lethal effects (Howard, 1971). The predators probably suffer other, more subtle effects. Mammalian predators also learn to avoid distasteful salamanders: in rats, both primary taste aversion and postingestional cues are important for acquiring a conditioned aversion to tiger salamanders, *Ambystoma tigrinum* (Mason *et al.*, 1982). The gastrointestinal/postingestion cue is the unconditioned stimulus and the food cue the conditional stimulus.

10.2.2 Anurans

The powerful toxins of toads and frogs are well known. The famous dart-poison frogs (Dendrobatids) from South America produce in their rather inconspicuous granular glands some of the most powerful poisons known. Indians have used the toxins on their arrows for many years; the poison of one tiny frog can be used for 50 arrows. The most toxic genus is *Phyllobates*, and the most toxic species *Phyllobates terribilis* of Western Colombia. The hunters merely draw the tip of the dart over the back of *P. terribilis* frogs but impale other species such as *P. bicolor* and *P. aurotaenia* to stimulate a copious flow of skin secretion (Daly and Spande, 1986). Dart-poison frogs are also brightly colored, as warning to potential predators, commonly termed *aposematic coloration*. However, the levels of poison and coloration in some Panamanian poison frogs of the genus *Dendrobates* are not correlated (Daly and Meyers, 1967).

The active principle of dart frog poisons is alkaloids. The study of the dendrobatid poisons led to the discovery of over 200 new alkaloids, including *batrachotoxins* (Fig. 10.5), pumiliotoxins, histrionicotoxins, gephyrotoxins, and decahydroquinolines (Daly *et al.*, 1994). The most common compounds have the basic structure of *piperidine* and include histrionotoxin. In *Phyllobates*, the synthesis of other alkaloids is suppressed in favor of batrachotoxins. These are

related to steroids and are some of the most poisonous compounds known. Batrachotoxins affect the Na⁺ permeability of mammalian membranes of nerves and motor end plates of muscles irreversibly. This causes arrhythmias, fibrillations, and ultimately failure of the heart. The specific sites of action are the Na⁺ channels. Binding of batrachotoxin prevents the normal closing of the channels, resulting in massive influx of Na⁺ and permanent depolarization of cells. The nerve cells can no longer transmit signals, and the muscles remain contracted. Histrionotoxin affects the K⁺ channel, and pumiliotoxin B the movement of Ca²⁺ (Meyers and Daly, 1983). The poisons do not affect the frogs themselves (Daly *et al.*, 1980) but probably originated from enzymes that regulate water and salt balances in the skin of frogs (Flier *et al.*, 1980).

The level of toxic compounds parallel the diet of the frogs: ants constituted 50 to 73% of the diet of aposematic, toxic species of *Dendrobates*, but only 12–16% of that of non-toxic, cryptic frogs of the genus *Colosthetus*. The lipophilic alkaloids of frogs actually occur in ants (Caldwell, 1996). Some dendrobatid alkaloids appear to be derived from the diet: precoccinelline "most certainly" from small beetles, and pyrrolizidine oximes "most likely from small millipedes" (Daly *et al.*, 1994). In captivity, the toxicity of the frogs declines over time. Panamanian poison dart frogs (*Dendrobates auratus*) produce at least 16 alkaloids but only when feeding on ant-containing leaf litter, while frogs in an aquarium and eating only fruit flies developed no alkaloids. The main alkaloids taken up from ants are pyrrolizidines and indolizidines (Daly *et al.*, 2000). The search is on for prey that might contain other dendrobatid compounds. *D. auratus* from central Panama were introduced into Hawaii and after only about 30 generations, their array of alkaloids had changed, presumably because of the change in diet (Daly *et al.*, 1992).

Frogs of the genus *Dendrobates* are ant specialists. Of the more than 20 structural classes of lipophilic alkaloids found in the frogs, six occur in myrmicine ants. However, many dendrobatid alkaloids such as the batrachotoxins, histrionicotoxins, and pumiliotoxins, have not yet been found in insects and other leaf-litter prey such as beetles and millipedes (Daly *et al.*, 2000). The snake *Liophis epinephelus* feeds on *Dendrobates* and may further bioaccumulate alkaloids.

The African clawed frog, *Xenopus laevis*, well known as a laboratory animal, produces mucus in its granular (poison) glands that affects predatory snakes. The most common frog-eating snakes in the clawed frog's habitat are the African water snakes, *Lycodonomorphus rufulus*, and *Lycodonomorphus laevissimus*. Experiments with snakes from the Cape Town area in South Africa demonstrated the potent effect of the frog's mucus. Live frogs caused the snakes to "yawn" and "gape" more often and to climb away from the prey. Toxin-laden frogs released these behaviors in snakes more often than frogs that had been depleted of toxin



FIGURE 10.6 Skin glands of American toad, *Bufo americanus*. There are large parotoid glands (PG) behind eye. On the back are macroglands ("dorsal warts" [DW]), consisting of clusters of mucus, granules ("poison"), and lipid glands. (Photograph: D. Müller-Schwarze.)

by epinephrine injections. The white, viscous mucus from the granular glands was also active by itself. When given directly to the snakes' mouths, it triggered the same responses as the whole frog, and more so than porcine stomach mucine, which was used as control. The northern water snake of eastern North America, Nerodia (Natrix) sipedon, responded even more to the mucus than Lycodonomorphus (Zielinski and Barthalmus, 1989).

Aquatic frogs and toads probably need less toxin for defense than terrestrial forms. The compounds in the frogs' mucus are thought to be neuroleptic, blocking dopamine receptors. They also possibly are antibacterial and aid wound repair, and they are known to elevate the level of prolactin, the amphibian juvenile hormone (Barthalmus and Zielinski, 1988).

Toads have skin glands that produce irritating secretions (Fig. 10.6). Mucus glands keep the skin moist. As mucus is an excellent medium for microorganisms, toads and frogs produce toxic compounds with antibiotic properties. Indeed, removal of skin toxins leads to skin infections and death (Habermehl, 1994). Granular glands produce a different kind of secretion. Clusters of both types of gland form macroglands, such as the large *parotoid* glands, located near the ear (*para*, beside or near, and *otid*, ear). (There is no similarity with the mammalian parotid salivary glands.) Numerous "dorsal warts" also contain glands of both types.

FIGURE 10.7 Bufotalin, found in toad secretion and samandarin, a neurotoxin from salamanders.

The secretions of toad tadpoles affect the behavior of predators. Largemouth bass, *Micropterus salmonides*, even when starved for 1 day, almost totally reject tadpoles of *Bufo americanus* (Fig. 10.6) and *Bufo woodhousei*. However, the hungrier they become, the more tadpoles they will eat. With increasing experience with these tadpoles, the bass take fewer into their mouths and spit more out. In choice experiments, they prefer tadpoles of the spring peeper, *Hyla crucifer*, to those of *Bufo* spp. (Kruse and Stone, 1984).

Predators eat the narrow-mouthed toad, *Gastrophryne carolinensis*, less than other anurans. They contact the toad, bite it, and release it again. Several reptilian and avian predators have been tested, among them the yellow-bellied water snake, *Nerodia erythrogaster*, garter snakes, *T. sirtalis*, snapping turtle, *Chelydra serpentina*, and the black-crowned night heron, *N. nycticorax*. The toad's skin contains a potent toxin: mice injected with extract from 3 mg toad skin per gram body weight of the mouse, died within 15 minutes. Skin from the American bullfrog, *Rana catesbiana* had no effect. Skin secretions also protect the toad from counterattack of ants, its main prey. The granular and mucus glands and their pores appear in tadpoles when the forelimbs emerge. At the same time, the tadpoles become toxic (Garton and Mushinsky, 1979).

The many compounds in skin gland secretions of toads fall into two groups: biogenic amines and steroidal bufogenins. There are two types of biogenic amine: the catecholamines (epinephrine and norepinephrine) and indole alkyl amines. The latter include bufotenin, serotonin, bufotenidin, bufoviridin, bufothionin, and others. One example of steroidal bufogenins is *bufotalin* (Fig. 10.7). If esterized with suberyl arginine, these steroids are called bufotoxins. They act on the heart, similar to digitalis toxins and are potentiated by epinephrine and norepinephrine. Bufotalin occurs in the European toad, *Bufo vulgaris*. *Samandarin* from salamanders (genus *Salamandra*) is structurally similar to bufogenins but acts as a neurotoxin (Fig. 10.7).

Bufo marinus, the giant, or cane toad, of South and Central America has been introduced to control pests, particularly in sugar cane fields, in areas from Florida and Cuba to Hawaii, Fiji, and the Philippines. Mortality of pets poisoned by eating giant toads varies by region. In Hawaii, only 5% of dogs and cats exposed to *B. marinus* die if untreated, while nearly 100% die in Florida. The animal may die within 15 minutes of mouthing or eating the toad (Fowler, 1992).

10.3 Reptiles

10.3.1 Snakes

Naturalists have known peculiar, strong, and persistent body odors of reptiles for some time. For instance, the mamba has a powerful curry-like, not unpleasant, smell that clings to the handlers' skin (Wingate, 1956). Some reptilian secretions possibly serve in defense against predators. The odor of the rat snake *Elaphe climacophora* affects mammalian predators. Brown bears (*Ursus arctos*) retreat from a dead rat snake and drool. They react similarly to a rope treated with the snake's odor (Kano, 1976). There is no indication that canids are repelled by snake odors. Coyotes (*Canis latrans*) are attracted to stations with secretion from the paired glands in the tail base of the western diamondback rattlesnake (*Crotalus atrox*). They rub and roll at these scent stations. Dogs lick, bite, and eat paper treated with the secretion (Weldon and Fagre, 1989). Domestic cats salivate and rub on secretion from a gland on the tail base of the gray rat snake, *Elaphe obsoleta*. The response was stronger than to skin samples of this species and cats ate less of food treated with the secretion (Wright and Weldon, 1990).

The blind snake *Leptotyphlops dulcis* is a specialized burrower that feeds on termites and ant brood. It follows the ant pheromone trails to find its prey (Section 12.1). When attacked by ants, this snake tilts its scales individually so that the skin appears silvery. While it writhes, it covers itself with feces and a clear viscous fluid, discharged from the anus. It may also assume a stationary coiled position. When it resumes searching and feeding, it is no longer attacked by ants. The effect lasts from 3 to 30 minutes (Gehlbach *et al.*, 1968). Here a predatory species protects itself from the defensive actions of its prey.

The cloacal secretion of *L. dulcis* also repels its predators, ophiophagous snakes. All five species of the tested colubrid snakes are repelled, including the ophiophagous species *Diadophis punctatus* and *Lampropeltis triangulum*. However, the cloacal secretion attracts conspecifics. This attraction possibly evolved first as the most important function of the secretion for the blind snakes (Watkins *et al.*, 1969).

Aposematically colored, the yellow-bellied sea snake, *Pelamis platurus* (Hydrophiidae), of the eastern Pacific has venom and is distasteful. It has no known aquatic predators, although remains were found in murray eels and sharks. Predatory fish such as snappers refuse the snake. They reject its meat even when hidden in palatable squid. Predatory fish of the Atlantic ocean, however, ate the sea snake in experiments, and died after 1 of 12 meals (Rubinoff and Kropach, 1970).

Snake venoms

Since predators of snakes (and humans) have to deal with snake venoms as defenses, they are included here, even though they serve in predation. Snake venoms are primarily enzymes (proteins), especially of the phospholipase A_2 type, which breaks down cell membrane phospholipids hydrolytically. Other snake venoms such as *cobrotoxin* contain peptides with 60–70 amino acid residues. Pharmacologically, they have neurotoxic, cytotoxic, anticoagulant, and other effects. The neurotoxins, in turn, can have pre- or postsynaptic effects. Snake venoms with both neurotoxic and hemolytic effects on the heart are known as cardiotoxins. Cytotoxins attach to the cells of blood vessels and cause hemorrhage. Snake venom factors may stimulate or inhibit blood clotting. Finally, platelet-active factors can contribute to hemorrhage.

Snakes themselves are immune to their own venom. Rattlesnakes have antibodies in their serum. Indeed, serum from snakes is more effective in protecting mice from rattlesnake venom than is commercial antivenin. Non-venomous snakes such as the king snake, Lampropeltis getulus, which also prey on crotalid rattlesnakes, are resistant to crotalid venom. Mongooses are very skilled in avoiding bites by cobras, but they also are resistant to cobra venom. A large dose, however, will kill even a mongoose. The Virginia opossum is also resistant to crotalid venom and the European hedgehog, Erinaceus europaeus, neutralizes some of the hemorrhagic factors in viper poison. Domestic and feral pigs are also impervious to venom by virtue of their thick skin and the subcutaneous fat layer, which retards absorption of venom into the blood. If this protection is circumvented by intramuscular injection, the snake venom can kill a pig. Cats resist snake venom very well. They show local symptoms, vomit, and defecate, but recover after 2 days. Dogs are more sensitive, and horses are more sensitive than cattle. Birds appear not to be protected against snake venom, though raptors take snakes. The feathers are thought to protect against penetration of the skin by the fangs of the snake. Some people such as snake worshippers and individuals who have been repeatedly bitten claim immunity against snake venom. However, there is no evidence for long-term immunity. A herpetologist in Australia had himself hyperimmunized by repeated injections of snake venom over 1 year. At the end of that period, he received eight times the lethal dose of tiger snake, *Notechis scutulatus*, venom and survived. However, just 1 year later, the titer had dropped, and he was probably unprotected. Attempts to vaccinate people in high-risk areas have not been successful. On the contrary, there exists the danger of anaphylaxis (hypersensitivity) from acquired sensitivity to snake venom.

10.3.2 Lizards

Gila monsters (Heloderma horridum and H. suspectum) bite when bothered. They hold on for 10–15 minutes. Their powerful mouth can be pried open only with a tool such as a screwdriver. Fortunately, we know of no human fatalities. (Cardiac arrest has occurred through excessive fear, alcohol overdose, or infection.) The gilatoxin from the oral glands contains phospholipase A₂ and hyaluronidase, which breaks down hyaluronic acid, a bonding agent between cells. This permits more rapid spread of venom between cells. The Mexican bearded lizard, Heloderma h. horridus, produces a gilatoxin component, a serine protease with 245 amino acid residues. This compound lowers b lood pressure (Utaisincharoen et al., 1993). Monitor lizards (Varanidae) share nine toxin types with snakes and helodermatids. Oral venoms of the Australian lace monitor, Varanus varius, lower blood pressure and slow blood clotting. Prey lose consciousness and bleed to death (Fry et al., 2005).

10.3.3 Turtles

Most aquatic and semiaquatic turtles have two or more pairs of Rathke's glands, which release a secretion through duct openings in the axillary, inguinal, and inframarginal areas. If a probe is applied to the axillary or inguinal regions of loggerhead (*Caretta caretta*) or Kemp's ridley (*Lepidochelys kempi*) turtles, the glands near the stimulation discharge secretion, and those on the opposite side of the body do not (Weldon and Williams, 1988). Disturbed stinkpot turtles (*Sternotherus odoratus*, Kinosternidae) of the eastern USA exude a musky secretion from a pair of glandular openings. The discharge may signal the undesirable taste of the turtle's flesh (olfactory aposematism). Four phenylalcanoic acids in this secretion have been identified. These are phenylacetic, 3-phenylpropionic, 5-phenylpentanoic, and 7-phenylheptanoic acid (Eisner *et al.*, 1977). Table 10.2 lists some chemical defenses in amphibians and reptilians.

Table 10.2 Defense compounds in amphibians and reptiles

Species	Compounds	Class of compound	Effect
California newt Taricha torosa	Tetrodotoxin	Alkaloid	Local anesthetic, death
Toad	Epinephrine, norepinephrine,	Biogenic amines	Sympathomimetic
	Bufotalin	Bufogenins (steroids)	Digitalis like (stimulates heart contraction)
	Bufotoxins	Bufogenins esterified with suberylarginine (steroids)	Digitalis like
Salamanders Salamandra, Triturus spp.	Samandarin	Steroid	Spasms, inhibits breathing, death
Dart poison	Batrachotoxins	Alkaloids	Na+ channels
frogs,	Pumiliotoxins	Alkaloids	Ca ²⁺ channels
Phyllobates spp.	Histrionicotoxins	Alkaloids	K ⁺ channels
Snakes	Phospholipases A ₂	Proteins, peptides (cobrotoxin), amino acids	Neurotoxins
Gila monster	Phospholipase A ₂	Protein	Neurotoxin
Heloderma horridum,	Hyalurinodase	Protein	Breaks down
H. suspectum	-		hyaluronic acid

10.4 Birds

Judging by their variable and often disagreeable taste, birds and their eggs may be chemically defended. In extensive experiments, H. Cott (1954–55) tested birds' eggs and flesh with humans, rats, cats, hedgehogs, ferrets, and hornets. To rats and hedgehogs, the eggs of Galliformes and Pelicaniformes were most palatable, least palatable those of Falconiformes and Passeriformes. Humans resembled hedgehogs and rats: all three most preferred eggs of chicken, coot, and kittiwake, while the eggs of the songbirds Carduelis cannabina, A. schoenobaenus, and Sylvia communis ranked at the bottom of all three species of egg tasters. The feeding habits of the birds played a role: eggs of piscivorous, piscivorous/invertebrate eating, and omnivorous bird species were most palatable. Least palatable were eggs of insectivorous and insectivorous/herbivorous birds. For humans and hedgehogs, no close relationship between the taste of the flesh of a bird species and the palatability of its eggs was found.

Four findings are particularly relevant to the question of chemical defense. First, the smaller the eggs the less palatable they are. Smaller eggs have more predators. Second, the eggs of solitary nesters are more unpalatable than those of colonial nesters. Third, palatability to the rat varied with nest site location: the most-palatable eggs came from birds that nest on cliffs or stacks, or over water. The least-palatable eggs belonged to birds nesting in vegetation less than 90 cm above ground or in holes of trees or masonry. Birds nesting in ground burrows or trees were intermediate. Fourth, shell coloration correlated with palatability: cryptic eggs, such as those of ducks and game birds, were considerably more palatable than white/"immaculate" eggs. Combined, these four variables provide a vulnerability score. Cott's composite vulnerability index correlated negatively with palatability. Four of the five most palatable species – herring gulls, lesser black-backed gull, great black-backed gull, and gannet – are also the four least-vulnerable species. At the other end of the scale, the six species with the most repugnant eggs, all passerines, fall all in the most vulnerable class. These are the lesser white throat, linnet, great tit, eastern house wren, blue tit, and white throat.

A combination of egg flavor, color, and induced gastrointestinal illness can condition egg predators to avoid eggs on sight. Free-ranging ravens (Corvus corax) were presented with surrogate eggs in their breeding territories. The eggs had been injected with a tasteless, illness-producing cholinesterase inhibitor (2,3,5- and 3,4,5-trimethylphenylmethyl carbamate [trade name U.C. 27867]). The treated eggs had one color, the controls another. Of the treated eggs, 70% survived intact, but only 38% of the controls. Such conditioned taste aversion most likely plays a major role in the evolution of aposematism and mimicry. Non-toxic mimics may benefit and survive better if they appear after a conditioned taste aversion has been established in local predators (Nicolaus, 1987). For application in conservation efforts, it is important that the visual cue (here egg color) suffices for avoidance since "sampling" the eggs would obviously destroy them. Many natural defense compounds have a taste, usually bitter, that plays a role in establishing an aversion. In Nicolaus' study, illness rather than a noxious taste alone was necessary to produce an aversion. Furthermore, predation was lower within raven territories, presumably because they defended their food resources against non-territorial ravens and other predators. Lower predation pressure within breeding territories than outside has been demonstrated for other birds, such as the south polar skua, Catharacta maccormicki (Müller-Schwarze and Müller-Schwarze, 1973, 1977).

The palatability of the *flesh* of birds to humans is broadly correlated with their visibility, including plumage color and behavior. Cott (1947) noted in Egypt that hornets (*Vespa orientalis*) fed on a carcass of a dove but ignored that of a kingfisher



FIGURE 10.8 Hornets, *Vespa orientalis*, feed on carcass of a palm dove, *Streptopelia senegalensis aegyptiaca*, left, while ignoring that of a pied kingfisher, *Ceryle rudis rudis*, on the right. (From Cott, 1947.)

(Fig. 10.8). In Zambia, he asked people to rate cooked meat of the pectorals of 200 species of birds from 57 families for their palatability. Humans ranked these birds from "ideal" to "inedible." Hedgehogs, rats, ferrets, and cats followed the same rank order. Birds (and their eggs) that are very vulnerable to predation, such as auks, turacos, kingfishers, starlings, and woodpeckers, tasted the worst. These birds were also more conspicuous than better-tasting species, with blue, red, rufous, white, and black predominating. A general pattern emerged: visibility of the bird was negatively correlated with palatability (Cott, 1947, 1952, 1953, 1954; Cott and Benson, 1969).

Birds may actually defend themselves by toxins. One of the most toxic animal compounds known, a homobatrachotoxin, was found in a New Guinea bird, the hooded pitohui, *Pitohui dichrous* (Dumbacher *et al.*, 1992). These birds are known in New Guinea as "rubbish birds" because the skin tastes bad. Two species of pitohui also emit a "strong sour odor." Three species of the genus *Pitohui* proved to contain homobatrachotoxin, a steroidal alkaloid. The same compound is found in a South American poison-dart frog. The frog and the pitohui are brightly colored (aposematism). People who handle pitohuis suffer watery eyes and runny noses. Five species of *Pitohui* carry batrachotoxins, in some species particularly in the contour feathers of the belly, breast, or legs. The most toxic species are *P. dichrous* and *P. kirhocephalus*. Batrachotoxins also occur in feathers and skin of a bird in a second genus, the blue-capped ifrita, *Ifrita kowaldi*. Populations differed in batrachotoxin levels, suggesting a dietary origin (Dumbacher

et al., 2000). A beetle that was found in pitohui stomachs also contains batrachotoxins. The possible targets of the toxins are feather lice, Phthiraptera. They choose a non-toxic over a highly toxic feather. On toxic feathers, the lice die sooner (Dumbacher 2003). The pitohuis' predators are snakes, hawks, such as the brown goshawk, and human hunters, who do not eat them because of the poison (Dumbacher 2003).

The preen gland (uropygial gland) of young hoopoes (*Upupa epops*) is largest at 12 days of age. It possibly has a repellent effect on intruders. A ring of specialized feathers carries and dissipates its odor. Upon approach, a predator is sprayed with a liquid from the large intestine, accompanied by a hissing sound (Sutter, 1946). Petrels (Procellariidae) defend themselves by squirting stomach oil with undigested food at predators and intruders. The aldehydes octanal and decanal, compounds with citrus-like odor in the feathers of crested auklets, appear to repel and kill ectoparasites such as ticks (Douglas *et al.*, 2004). The hoatzin (*Opisthocomus hoazin*) of South America is a sluggish, vulnerable large bird that predators appear to avoid. The bird has a strong, unpleasant smell and Peruvians call it *chancho* (pig). The odor possibly derives from the unique leaf diet of the hoatzin and perhaps protects it from predators.

10.5 Mammals

The duck-billed platypus, *Ornithorhynchus anatinus*, uses a slightly curved poison spur on its ankles (tarsus) in male—male fights over females. It possibly uses it also against predators. In the resting position, the spur is retracted against the leg. Activated, it is raised at a 90 degree angle to the leg. The platypus kicks to insert the spur into the victim. The spur connects to a duct that, in turn, comes from a reservoir downstream from a kidney-shaped crural venom gland located on the medial aspect of the thigh. The venom contains proteins and toxicity varies seasonally, possibly coinciding with the breeding season. Humans struck on their hand will be crippled for months (Vaughan, 1978). Echidnas have a spur but a non-functional gland.

The secretion of the anal glands of skunks epitomizes mammalian defense secretions. Sulfur compounds in the scent of striped skunk (*Mephitis mephitis*) include *trans-*2-butene-1-thiol, 3-methyl-1-butanethiol (Andersen and Bernstein, 1975), *trans-*2-butenyl thioacetate, 3-methylbutanyl thioacetate, and 2-quinolinemethane thiol (Wood, 1990; Fig. 10.9). The thiols, once called mercaptans (because they captured mercury atoms), are like alcohols with a sulfur atom in place of the oxygen. In the striped skunk, the thioacetates break down in water. They yield thiols, which have the more potent, characteristic skunk odor and linger for up to 2 weeks. In the spotted skunk, by contrast, the

(E)-2-Butene-1-thiol (38-44%)

3-Methyl-1-butanethiol (18-26%)

(E)-2-Butenyl methyl disulfide (?)

(S)-(E)-2-Butenyl thioacetate (12-18%)

(S)-3-Methylbutanyl thioacetate (2-3%)

FIGURE 10.9 Major sulfur compounds in the anal sac of the striped skunk *Mephitis mephitis*. (After Andersen and Bernstein [1975], Wood [1990], and Wood *et al.* [1991].)

thiols are not tied up in slow-release compounds, and the odor dissipates in a few hours (Wood *et al.*, 1991).

The black-and-white contrasts of the skunks are thought to function as a warning color. Intriguingly, a marsupial in New Guinea displays convergent features: the long-fingered triok, *Dactylopsila palpator*, is black and white and also smells like a skunk (Flannery, 1998).

A toxin, blarinatoxin in saliva from the submaxillary and sublingual glands of the North American short-tailed shrew, *Blarina brevicauda*, serves in defense and for paralyzing the prey. Ducts lead to the incisor teeth, which may be modified with channels to direct venom in the wound. Shrew venom is proteolytic and neurotoxic. The poison is not dangerous to humans, who experience a burning sensation and minor swelling that can be painful for several days. To small mammals it can be lethal. It dilates blood vessels and leads to irregular respiration, paralysis, convulsion, and eventually death. In one experiment, one third of bitten mice died. Small mice (body weight under 10 g) died in larger

numbers (80%), and often within minutes (average 157 minutes). They were most affected when bitten near the head, and least at tail or feet. The severity of the wound determines mortality. Venom from one shrew can kill 200 mice. The shrews also bit earthworms and cached them comatose. The salivary toxin seems to function in stunning or paralyzing the prey and facilitating storage. It is not important in killing rodents (Tomasi, 1978). Chemically, blarinatoxin is a high-molecular-weight, water-soluble protein with 253 amino acid residues (Kita *et al.*, 2004).

10.6 Pars pro toto: decoy odors

Some of the seemingly contradictory observations that a predator is actually attracted to a presumed "defense secretion" may be explained by the pars pro toto principle. A prey animal in danger, when it is being detected, pursued, or handled by a predator, or simply sneaking away before detection, discharges an odor that attracts the curiosity of the predator. While the odor cloud lingers or is moved by air currents, the prey animal moves off in another direction. This would work especially well when vision is impaired as in the dark, or in heavy vegetation. The emitted odor would serve as decoy, the olfactory equivalent of the severed tail of the lizard, and an energetically much cheaper one. Possible examples are the metatarsal odor of the black-tailed deer, *Odocoileus* hemionus columbianus, which is discharged during alarm (Müller-Schwarze et al., 1984), and the secretions from anal sacs in snakes that are sniffed by predators. Similarly, the discharge of stomach oil in procellariid seabirds may also distract predators. Further candidates for chemical pars pro toto defense are the odors noticeable to humans when feral pigs flee in dense cover (Section 7.2), and a puff of odor from a fleeing deermouse, Peromyscus sp. (Müller-Schwarze, personal observations).

10.7 Invertebrate allomones that deter vertebrate predators

10.7.1 Taste aversion to invertebrate prey

At overwintering sites of the monarch butterfly (*Danaus plexippus*) in Mexico, only one of the three local mouse species, *Peromyscus melanotis*, actually feeds on the butterflies. The monarchs contain cardiac glycosides (CG) and pyrrolizidine alkaloids (PA). All three species of mice have similarly low avoidance thresholds to PA (specifically, monocrotaline). But *P. melanotis* is less sensitive to CG (specifically, digitoxin) than the other two, *Reithrodontomys sumichrasti* and *Peromyscus aztecus*. Laboratory tests indicate that PA is toxic to young mice,

but CG is not; even so, PA does not reduce food intake. The mice do not know how to avoid specifically compounds that will damage them so, instead they generally avoid bitter compounds. Since they cannot vomit, mice benefit from avoiding eating many bitter, potentially harmful foods but, by "being on the safe side," they may forgo feeding on potentially useful foods. The three mouse species differ in their ways to overcome the chemical defenses of their prey. Therefore, the effectiveness of the monarch's defenses depends on the species of predator: *P. melanotis* selects monarchs with low CG concentrations, learns to reject the CG-laden cuticle of the butterfly, and is insensitive to the bitter taste and toxic effects of CG. *R. sumichrasti* uses only the first two mechanisms, and *P. aztecus* only the first (Glendinning *et al.*, 1990).

Fish, such as the reef predator *Thalassoma bifasciata*, avoid sponges. Organic extracts of sponges of the sponge genus *Ircinia*, mixed with food, deterred *T. bifasciata* from feeding. Non-volatile furanoseterpene tetronic acids in the extracts are responsible for that effect. By contrast, volatile compounds from the sponges, such as dimethyl sulfide, methyl isocyanide, and methyl isothioyanate, do not help to defend the sponge against fish (Pawlik *et al.*, 2002).

Table 10.3 lists some defense compounds, mostly alkaloid, that invertebrates use to deter vertebrate predators.

10.8 Recycled animal and plant materials

10.8.1 Birds

Ornithologists have known for a long time that certain birds use materials from other animals or plants in or around their nests. The burrowing owl (*Speotyta cunicularia*) of the Western hemisphere lines its burrow with cow dung. It is believed that the dung deters badgers from preying on the owls. Of nests lined with dung, only 8% were lost to badgers, but 54% of the unlined nests (Green, 1988). It is unclear whether the dung masks bird odor or acts as a repellent. Redbreasted nuthatches, *Sitta canadensis*, smear pine resin around their nest cavities. This is assumed to serve a hygienic function, as pine resin is antiseptic, but it is also toxic to some insects (Bent, 1964).

Birds that breed in cavities and reuse their nest sites often incorporate green parts of aromatic plants into their nests. Starlings, *Sturnus vulgaris*, prefer certain species of plants to others. House sparrows, *Passer domesticus*, incorporate neem (margosa) tree, *Azadirachta indica*, leaves into their nests. Extracts from neem leaves repel arthropods and inhibit oviposition (Sengupta, 1981). The aromatic plants are thought to fulfill an important function by keeping down populations of microbes in the birds' nests (Mason and Clark, 1986).

Table 10.3 Allomones: invertebrate compounds that deter vertebrate predators

Vertebrate predator	Invertebrate prey	Compound(s)	Reference
Reef fishes	Orange sponge <i>Axinella</i> sp.	Indoles: herbindole A	Herb <i>et al.</i> , 1990
Reef fishes Thalassoma bifasciatum, Eupomacentrus partitus	Caribbean ascidian Tridemnium sodium	Macrolides: didemnin B and nordidemnin B	Lindquist et al., 1992
Reef fishes	Sea hares Stylocheirus longicauda	Alkaloids: malyngamide A and B, and B acetate, from cyanobacteria	Paul and Pennings, 1991
Blind snake Leptotyphlops dulcis	Formicid ant Neivamyrmex sp.	Skatole (attracts, but repels other preying snakes and ants)	Watkins <i>et al.</i> , 1969
Lizards	Butterflies, months, beetles	Senecionine and lycopsamine type PAs, sequestered from Asteraceae and Boraginaceae	Various authors
Anole lizard Anolis carolinensis	Bug Neacoryphus bicruris (Heteroptera)	Senecionine type PAs sequestered from Senecio smallii (Asteraceae)	MacLain and Shure, 1985
Birds	Taiwan stick insect Megacrania tsudeii	Monoterpenoid alkaloid actinidine (3500 \times conc. in defense gland)	Chow and Lin, 1986; Ho and Chow, 1993
Gray jay Perisorius canadensis	Eumaeus atala floridana (Lepidoptera, Lycaenidae)	Cycasin, from <i>Zamia floridana</i> (Cycadaceae), stored in larvae	Bowers and Larin, 1989; Bowers and Farley, 1990
Glaucous-winged gull Larus glaucescens and sea otter Enhydra lutris	Clams	Paralytic shellfish toxins (Guanidins [saxitoxins]), from dinoflagellates	Pawlik, 1993; Kvitek, 1991
Tree sparrow Passer montanus	Swallowtail butterfly Atrophaneura alcinous	Aristolochic acids (from <i>Aristolochia debilis</i>), in test applied to rice grains	Nishida and Fukami, 1989
Mice Peromyscus spp., Reithrodontomys sumichrasti	Monarch butterfly	Cardiac glycosides and PA	Glendinning, et al., 1990

PA, pyrrolizidine alkaloid.

In a North American study, starlings preferred agrimony (*Agrimonia* sp.), wild carrot (*Daucus carota*), flea bane (*Erigeron* sp.), yarrow (*Achillea millefolium*), red-dead nettle (*Lamium purpureum*), and rough goldenrod (*Solidago rugosa*). In laboratory tests, plants preferred by starlings retarded the hatching of eggs of the louse *Menacanthus* sp. that lives on the feathers but none of the tested plants

was effective against the fowl mite *Ornithonyssus sylviarum*. Five of the selected plant species inhibited the growth of at least one bacterial pathogen (*Streptococcus* sp., *Staphylococcus aurealis*, and *Pseudomonas* sp.). They were never effective against *Escherichia coli*.

The total concentration of volatiles was higher in preferred than in other plants. The volatiles in the preferred plants are mostly mono- and sesquiterpenes such as myrcene, α - and β -pinene, and others. One preferred plant, the rough-stemmed goldenrod *S. rugosa* of old fields, contains the sesquiterpenes 2-bornyl acetate and farnesol. These compounds are implicated as potent analogues of insect juvenile hormone, which suppresses molting in arthropods. They possibly impair reproduction of ectoparasites by delaying their development to terminal instars. Reduced populations of parasites would benefit the nestling birds. Volatiles from the plants in question elicit strong responses from the olfactory nerve of the birds. Tests with aromatic plant parts underneath food containers showed that starlings can discriminate different plants by odor (Clark and Mason, 1985; Mason and Clark, 1986). If plant odors were paired with gastro-intestinal malaise caused by the bird repellent methiocarb, a conditioned avoidance was produced. If both olfactory nerves were sectioned before the conditioning, no avoidance occurred (Clark and Mason, 1987).

Experiments have shown that aromatic herbs in the nest do not reduce the ectoparasite load in starlings but may stimulate parts of the immune system that mitigate the effects of ectoparasites (Gwinner *et al.*, 2002). Corsica blue tits (*Parus caeruleus ogliastrae*) select 6–10 species of aromatic plants out of the approximately 200 species that occur in their habitat and place them in the nest at the nestling stage. The birds will replenish the aromatic plants when the volatile components have been lost. The chosen plants had proven antibacterial antiviral, antifungal, and insecticidal properties. Interestingly, 40% of the plants involved possess known activity against human immunodeficiency virus 1. (Petit *et al.*, 2002; see p. 379).

10.8.2 Mammals

Both the African hedgehog, *Atelerix pruneri*, and the long-eared hedgehog of Asia, *Hemiechinus auritus*, use defense secretions of toads. The animal takes the secretion up into its mouth and then licks its spines. The spines then become covered with frothy saliva. This behavior does not depend on learning and is released by several species of toads and salamanders, and even by substances such as tobacco juice (Brodie, 1977).

Wedge-capped capuchin monkeys (*Cebus olivaceus*) in the llanos of central Venezuela appropriate the defenses of millipedes to repel mosquitoes. They

rub the 10 cm long millipede *Orthoporus dorsovittatus* over their fur, transferring benzoquinones from the millipede. The behavior is most prevalent during the rainy season when mosquitoes are particularly numerous. The mosquitoes also transfer eggs of parasitic bot flies into the skin of the monkeys. We do not yet know whether rubbing milliped benzoquinones into the fur prevents bot fly infestation. Rubbing is a social event: up to four monkeys may share a millipede (Valderrama *et al.*, 2000; see also p. 377 and p. 382).

10.9 The question of coevolution between predator and prey

Coevolution between predator and prey has often been invoked. In one well-investigated case, support for coevolution is weak. In Mexico, the blackheaded grosbeak, Pheucticus melanocephalus, and the black-backed oriole, Icterus galbula abeilli, feed on monarch butterflies, D. plexippus, which defend themselves with high levels of cardenolides. The cardenolides are particularly concentrated in the cuticle of the butterfly. The grosbeaks are insensitive to the cardenolides. The oriole bypasses the cuticle and thus avoids emetic dosages of the toxin. It pries open the cuticle and feeds on the contents of the abdomen, which contain less cardenolides than the cuticle. Has the oriole's feeding technique coevolved to breach the defenses of the butterfly? Is the grosbeak insensitive to cardenolides because it responded to these compounds in a coevolutionary process? The answer is no in both cases. These birds are exapted (formerly "preadapted") (Gould and Vrba 1982; Brower et al., 1988): Other orioles that do not feed on butterflies also use the prying movement in feeding. The northern oriole, Icterus galbula, pierces fruit and then gapes and eats the pulp without the skin. The blackthroated oriole uses the same movements and then laps up the juice with its bifurcated and brushy tongue. The grosbeak is also exapted: early in evolution it raised the amount of toxins it can tolerate (Brower et al., 1988).

10.9.1 Competition

One animal species can suppress a competing species by means of chemical signals. An example of such *competitive exclusion* (*sensu* Gause, 1934), or "interactive segregation," are two species of spiny mice of the genus *Acomys* in southern Israel. They are sympatric and occupy the same rocky habitat. The common spiny mouse, *A. cahirinus*, is nocturnal and forces the golden spiny mouse, *A. russatus*, to be active in the daytime heat of the Negev desert. If *A. cahirinus* is removed from the field, *A. russatus* becomes nocturnal (Shkolnik, 1971). Kept by itself in the laboratory, *A. russatus* is also nocturnal. Odor of feces and urine of the

competing species *A. cahirinus*, added to the air supply shifts the activity rhythm of *A. russatus* if the mice are kept in 12 hours darkness and 12 hours light. Under "free-running conditions" (i.e. in constant dim red light), *A. russatus* lower their activity level, as measured by oxygen consumption, in response to these chemical cues from *A. cahirinus* (Haim and Fluxman, 1996).

Allomones II: plant chemical defenses against herbivores

The plant world is not colored green; it is colored morphine, caffeine, tannin, phenol, terpene, canavanine, latex, phytohaem-agglutinin, oxalic acid, saponin, L-dopa, etc.

JANZEN (1978)

Herbivores select certain plant species or parts and reject others. Plant defenses determine food choices as much as nutritional value does. Plants can defend themselves mechanically as with thorns, hairs, waxes, or structural fibers, and chemically with secondary plant compounds. Mammals have had to cope with plant defenses since they adapted to an herbivorous lifestyle approximately 85–100 millions years ago (Archibald, 1996).

In 1888, Stahl suggested that plants use toxic chemicals as defense against herbivores based on his feeding experiment with snails. Fraenkel (1959) postulated that "secondary compounds in plants exist solely for the purpose of repelling and attracting insects." We now know that these compounds are aimed at vertebrates, other invertebrates, and microbes as well, and in many cases their roles are still being debated. Here I apply the term "allomone" in its widest sense: compounds that benefit the "sender," even though many are not "signals" in the strict sense. Therefore, "donor" or "originator" organism is a better term. In the metabolism of the herbivore, the receiving organism, such foreign compounds are termed xenobiotics, whether natural or synthetic.

The interactions of plants and vertebrate herbivores can be broken down into several questions.

- What defense compounds do plants use?
- What are their effects on mammalian and avian herbivores?
- What defense strategies do plants use? This includes species differences based on ecological needs; variations of plant defenses between plant parts (leaf, bud, internode, terminal shoot, bark, root) or growth stages that correlate with vulnerability; and seasonal variation of plant defenses.

Steroidal saponins, triterpene saponins

Dhurrin, taxiphyllin

Trypsin inhibitors

Cardenolides, butadienolides

Selenocystathionin, L-mimosine

Class	Examples
Phenolics	
Simple phenolics	Phloroglucinol, gentisic acid, vanillin, caffeic acid, ferulic acid
Complex phenolics	Coumarins, phenolic quinones, lignins, flavonoids, stilbenes, hydrolyzable tannins, condensed (or catechin) tannins, phenolic lipids
Terpenoids	Mono-, sesqui-, di-, and triterpenes
Alkaloids	Nicotine, guinine, morphine, colchicine, strychnine

Table 11.1 Classes of plant compound with antiherbivore defense functions

- What are the mechanisms of effective defense? Is the plant chemistry systemic and genetically "hardwired," or is it programmed to respond to need, termed "induced defense"?
- Is intensity of herbivory on specific plants correlated with levels of secondary plant metabolites?
- How do herbivores avoid or deal with ill effects of plant defense compounds?
- Is there evidence of coevolution between herbivorous vertebrates and their food plants?

This first section presents the "cast of characters," the compounds involved in defense.

11.1 Classes of plant defense compound

Saponins Mustard oils Cyanogens

Proteins

Cardiac glycosides

Non-protein amino acids

Phytohemagglutinins (lectins)

Many thousands of secondary plant compounds have been identified; and 400000 are suspected to exist. These numbers provide a great incentive for "chemical prospecting" (Eisner, 1989). The most prevalent, broad classes of plant secondary compound are phenolics, alkaloids, and terpenoids (Table 11.1).

FIGURE 11.1 Structures of simple phenolics.

11.1.1 Phenolics

In abundance second only to carbohydrates (Levin, 1971), phenolics occur in plants as complex mixtures. Their name derives from "phen-ol" (benzene), from Greek *phen*- (phainein): "to show." They range from simple molecules such as phloroglucinol to very complex ones such as lignins. Many phenolics are harmful to herbivores, e.g. by binding to proteins, while others are beneficial as antioxidants. Compounds with more than one phenol unit are designated polyphenols. Gallic acid is most readily oxidized and may be the best antioxidant in the context of human nutrition. Salicylic acid is also an antioxidant, but less so than gallic acid. Both may contribute to the "French paradox" – good health despite unhealthy components in diet (de Lorgeril *et al.*, 2002).

Simple phenolics

Simple phenolics are based on a six-carbon ring. Examples are salicylic acid and salicylaldehyde, vanillin, and thymol (Fig. 11.1).

Complex phenolics

An example of more complex phenolics is coniferyl benzoate (Fig. 11.2), which inhibits feeding in birds (Jakubas *et al.*, 1989), as discussed below. It binds to skin proteins and dietary proteins.

Pinosylvin methyl ether

FIGURE 11.2 Structures of complex phenolics.

Flavonoids

We know of thousands of flavonoids. They have a C6–C3–C6 skeleton and are responsible for the bright colors in flowers. The bitter taste in citrus fruits is caused by flavonone glycosides.

Isoflavonoids have a substituent at C-3. Examples are genistein and formononetin (see Fig. 11.11, below). Isoflavones exert estrogenic activity in mammals, interfere with reproduction in birds, and may render farm animals infertile. For example, sheep feeding on isoflavonoid-containing *Trifolium subterraneum* suffer reduced lambing (Harborne, 1993).

Stilbenes (from the Greek stilbos, glistening, after shiny crystals), are two-ring structures of the C6–C2–C6 type; examples are pinosylvin and pinosylvin methyl ether (Fig. 11.2). They play a role as antifeedants for mammals.

Tannins

Tannins have molecular weights of 0.5–20 kDa, with no upper limit. They are responsible for bitter or astringent taste of plants. The dry, astringent sensation brought about by complexation with mucoproteins of the mouth is thought to be repellent to herbivores (Harborne, 1993) while a moderate level of astringency can be pleasing to the human palate. Unripe fruits such as bananas, apples, or persimmon contain high levels of tannins (Mehansho *et al.*, 1987). This is thought to delay consumption by frugivores until the seeds have matured and can benefit from dispersal by animals and possibly from improved germination after passing through their digestive tracts (Section 12.2).

Tannins derive their name from their ability to tan (i.e. they combine with protein). They render plants less palatable and impair digestion by binding with the buccal mucosa, dietary proteins, and digestive enzymes of the animal. Tannins are thought to bind to proteins upon destruction of plant tissue by herbivores. This reduces the nutritive value of the plant to the herbivore. Some tannins such as oak gallotannins, are even toxic to livestock and rabbits (Meyer and Karasov, 1991). However, Martin and Martin (1983) have questioned the role of tannins as plant defense against herbivores.

Two different types of tannin, condensed and hydrolysable tannins, play important roles.

- 1. **Condensed or catechin tannins** (catechol tannins, Fig. 11.3) are the most common tannins in vascular plants, occurring in three quarters of gymnosperms and over half of the angiosperms. They are unbranched, linear polymers of flavonoid compounds (flavan-3-ols), linked through acidlabile carbon–carbon bonds. Condensed tannins may protect plant cell walls against microbial attack and so may affect microbial fermentation of plant cell walls in herbivores.
- 2. Hydrolyzable tannins, also known as tannic acids, are restricted to the dicotyledons. They have an astringent taste and the ability to tan leather. Their basic building block is gallic acid or ellagic acid (Fig. 11.4). These acids are esterified to the hydroxyl groups of glucose. Since one glucose unit can accommodate esterification with five molecules of gallic acid, the simplest hydrolysable tannin is pentagalloylglucose (Fig. 11.4). Tannins usually occur as mixtures. For instance, "tannic acid" of commerce is a mixture of free gallic acid and various galloyl esters of glucose.

Differences between hydrolyzable and condensed tannins are listed in Table 11.2.

Phenolics are responsible for the coloration of "black-water rivers," particularly in the tropics. An example is the Rio Negro, which remains distinct in its black color for many miles as it flows into the milky brown Amazon. In and near such "black" rivers, fauna and flora are considered impoverished (Swain, 1979).

Lignins are high polymers of phenolic alcohols such as coniferyl alcohol (Fig. 11.2). They increase the toughness of the plant and form wood, obviously useful for defense against herbivores.

11.1.2 Terpenoids

Terpenoids are the most diverse class of organic compounds in plants and the largest group of allelochemicals. The word is derived from "terpentin"

	Hydrolyzable tannins	Condensed tannins
Building blocks	Gallic acid, galloyl ester, ester-linked with glucose	Flavanoid units (Procyanidins), catechin
Occurrence in plant	Cell interior	Structural; tightly bound to cellulose wall of plant cell
Possible functions	Anti-herbivore: inhibition of digestion	Protect cell wall against microbes Retard decomposition of leaf litter: slow down release of nutrients (assumed to benefit mother plant) Mostly aimed at microbes, not herbivores
Mechanism	Bind to digestive enzymes of herbivores; variable molecules, specific for proteins	Inhibit pathogen's exo-enzyme system
When abundant	In earliest leaves but low levels in late summer and autumn	Late summer and autumn leaves
Metabolic cost	High: ties up phenylalanine for phenolic portion, plus glucose	Lower

Table 11.2 Differences between hydrolyzable and condensed tannins

(German for turpentine) and there are approximately 15 000 terpenes. Terpenes are lipophilic, and the building blocks are five-carbon units with the branched carbon skeleton of isopentane. The basic units are sometimes called isoprene (Fig. 11.5*a*), because heat decomposes terpenoids to isoprene. Depending on the number of C_5 units fused, we distinguish mono- (C_{10}) , sesqui- (C_{15}) , di- (C_{20}) , tri- (C_{30}) , tetra- (C_{40}) and polyterpenoids $[(C_5)_n$, with n > 8]. Alpha-Pinene and borneol (Fig. 11.5*b*) are examples of monoterpenes.

Nepetalactone (Fig. 11.5*c*), a monoterpene lactone (or iridoid) from the volatile oil of catmint, *Nepeta cataria*, excites domestic cats and other felids (Todd, 1962; Palen and Goddard, 1966; Hill *et al.*, 1976).

Three terpenoids (the monoterpenes carvacrol and β -thujaplicine, and the sesquiterpene thujopsene) from the japanese tree *Thujopsis dolabrata* inhibit gnawing by mice. A 1200-year old pagoda in Nara, Japan, made from wood of *Thujopsis*, shows no damage by rodents, insects, or microorganisms. Carvacrol is recommended as a long-term repellent against rodents (Ahn *et al.*, 1995).

The great diversity of terpenes helps to counteract tolerance by herbivores. In all, terpenes are not very toxic to vertebrates. Many mammals ingest a significant amount of terpenoids with their diet. Monoterpenes from pine oil added to the diet reduces food intake in red deer, *Cervus elaphus*, calves (Elliot and Loudon, 1987). The brush-tailed possum, *Trichosurus vulpecula*, detoxifies (+)- α -pinene to alcohol and carboxylic acid derivatives.

Basic structure of a condensed tannin

HO
$$\frac{8}{7}$$
 $\frac{0}{6}$ $\frac{1}{3}$ OH OH

Catechin

FIGURE 11.3 Condensed tannin. Catechin, a flavan-3-ol, is one of the most common building blocks.

Sesquiterpene lactones (e.g. glaucolide A) are "bitter principles" in plants such as the genus *Vernonia* (p. 382). Cottontail rabbits, *Sylvilagus floridanus*, and white-tailed deer, *Odocoileus virginianus*, eat the one species of this genus that does not contain glaucolide A (Mabry and Gill, 1979). Examples of sesquiterpenes are germacrone (Fig. 11.5*d*) in Labrador tea, *Ledum groenlandicum*, and isovelleral in mushrooms.

Pentagalloylglucose

Gallic acid

FIGURE 11.4 Hydrolyzable tannin. In the structure shown, five gallic acid units surround a sugar (hexose).

Diterpenes from Euphorbiaceae are toxic to livestock, and so are daphnane and daphnetoxin, diterpenes from *Daphne mezereum* (Thymelaeaceae). Toxins from *Rhododendron* are diterpenes.

Triterpenes (C30) are common in birches, especially in bark. Papyriferic acid from paper birch, *Betula resinifera*, is a feeding deferent for snowshoe hares, *Lepus americanus* (Fig. 11.5) (Reichardt *et al.*, 1985).

Glycosides of compounds with a triterpenoid origin include saponins, cardiac glycosides, and glycoalkaloids such as solanine (in potatoes).

Saponins

Saponins occur in Liliaceae such as asparagus, in legumes, spinach, and yams. They are triterpenoid glycosides with soap-like properties. Many are glycosides of steroid alcohols, and all have a bitter taste. Two types are

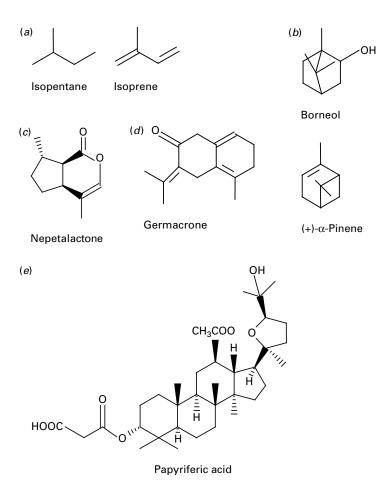


FIGURE 11.5 Terpenoids. (a) The basic five-carbon units isopentane and isoprene. (b) The monoterpenes borneol and α -pinene. (c) A monoterpene lactone, nepetalactone. (d) A sesquiterpene, germacrone. (e) A triterpene, papyriferic acid, from paper birch.

distinguished: steroidal saponins (C_{27} skeleton) and triterpene saponins (C_{30}). An example of the former is medicagenic acid (Fig. 11.6) in alfalfa (luzerne), Medicago sativa.

Saponins are foaming agents that hemolyze red blood cells. Injected into the bloodstream, they cause rapid lysis of erythrocytes. They also cause bloat in cattle by altering the surface tension of rumen contents and by trapping gas. Poultry are the most sensitive monogastric animals; on a saponin-containing alfalfa diet, they reduce feeding and grow more slowly (Applebaum and Birk, 1979). Mammals avoid saponin-containing plants, although livestock poisoning has been reported. Elephants avoid trees were the foliage contains steroidal saponins

FIGURE 11.6 Medicagenic acid, a steroidal saponin.

FIGURE 11.7 Cardiac glycosides consist of a sugar, a steroid skeleton, and a lactone ring. A five-membered lactone ring as here occurs in the cardenolide type.

(Jachmann, 1989). Saponins are used as fish poisons and also used as starting materials for steroid hormones.

Cardiac glycosides

Cardiac glycosides are bitter-tasting compounds that consist of three parts: steroid, lactone, and sugar. There are two groups, the cardenolides (Fig. 11.7) and the butadienolides. They are characterized by a 5- or 6-membered lactone ring, and a C_{23} or C_{24} steroid, respectively. Both are common in the Asclepiadaceae (milkweeds). Over 70 species of milkweeds in this family have cardenolide-laden latex. Examples are labriformin and labriformidin in the milkweed species Asclepias eriocarpa and Asclepias labriformis. These two compounds are toxic to sheep and other livestock (Benson et al., 1979; Nelson et al., 1981). Animals benefit from the warning signal afforded by the bitter taste. If ingested, vomiting removes the toxic agent. Livestock in Africa and North America leave milkweeds alone. Nerium and oleandrin are cardiac glycosides from Nerium oleander (Apocynaceae), digitoxin from Digitalis sp. (Scrophulariaceae).

Butterflies such as the monarch (*Danaus plexippus*) recycle cardenolides from milkweed (*Asclepias curassavica*), taken up by the caterpillar, as defense against bird predators. Blue jays, *Cyanocitta cristata*, learn to avoid monarchs after becoming sick from eating one. Within 12 minutes the birds become "violently ill." They vomit as often as nine times in 30 minutes but recover fully within half an hour (Brower and Brower, 1964). The birds will also avoid the similarly colored viceroy butterfly, although it is not toxic. Similarly, a grasshopper, *Poekilocerus bufonius*, ingests cardenolides from milkweed and then incorporates them into its noxious foam, which it sprays from a dorsal poison gland against vertebrate predators, such as mice. African hunters used to coat their arrows with extracts of cardenoloid-rich plants.

Cardiac glycosides are medicinally important steroids. Cardenolides are used in heart therapy, as emetics, diuretics, and purgatives. They affect the Na⁺/K⁺-ATPase and are very toxic at high doses.

Alkaloids

Alkaloids (compounds with basic properties, like ashes of saltwort, *al*qili) are amino acid-derived compounds that contain nitrogen as part of a heterocyclic ring. They are water soluble, bitter tasting, and occur in about 20% of species of flowering plants. "An alkaloid is a cyclic compound containing nitrogen in a negative oxidation state which is of limited distribution among living organisms" (Pelletier, 1983). Over 10000 compounds belong to this very diverse group. Well-known examples of alkaloids are nicotine, quinine, morphine, colchicine, and strychnine. Coniine in poison hemlock, Conium maculatum, is believed to have put to death the philosopher Socrates. Alkaloids are concentrated in vulnerable plant parts such as growing tissues, flowers, seeds, and peripheric tissues such as root bark. In contrast to most other secondary plant compounds, alkaloids may be primarily directed against mammals (Swain, 1977). Alkaloid-rich plants important for livestock are lupines and reed canary grass. Alkaloid-caused livestock losses are high in autumn when alkaloid-rich seeds such as those of lupines mature. Lupinine (Fig. 11.8) is one of the active compounds. In the North American west, tall larkspur (Delphinium barbei) is toxic to cattle and sheep. The main toxic compounds are the diterpene alkaloids methyllycaconitine and 14-deacetylnudicauline. They block acetylcholine receptors in the central and peripheral nervous systems. This results in muscular paralysis and eventually death (Pfister et al., 1996).

Solanine (Fig. 11.8) occurs in potatoes, particularly in peel (to 1.5 mm depth), eyes, and repaired tissue, and in small and immature, and light-exposed

FIGURE 11.8 The alkaloids lupinine and solanine.

green tubers. Tomatine is found in tomatoes, particularly in green and storeripened specimens. *Coprine*, from the mushroom *Coprinus atramentarius*, the "alcohol inky," is an alkaloid that inhibits aldehyde dehydrogenase, one of two enzymes that metabolize alcohol. By itself not toxic, the mushroom causes poisoning through accumulation of aldehyde when taken together with alcohol.

Cyanogenic glycosides

Cyanogenic glycosides are water soluble and occur in the vacuoles of plants cells. They consist of glucose attached to an aglycone, are intermediately polar, and are derived from amino acids. Upon damage to the plant tissue by an animal, enzymes release hydrogen cyanide from cyanogenic glycosides. For example, in manioc (see below), linamarin and lotaustralin come in contact with the enzyme linamarase after cells of the outer layer of the tuber are injured.

Hydrogen cyanide is toxic because the cyanide ion has high affinity for metal ions and so binds to the metal-containing cellular respiratory enzymes. Heme proteins such as cytochrome oxidase are complexed, resulting in asphyxiation at the cell level.

Dozens of cyanogenic glycosides occur in higher plants (in 2700 species from 130 families). Examples are dhurrin and taxiphyllin in grasses, and prunasin and amygdalin in the family Rosaceae. The latter two hydrolyze to form benzaldehyde, the "bitter almond" odor. The odor is probably a signal that vertebrate herbivores pay attention to. Proportionally more cultivated plants (perhaps two thirds) are cyanogenic than flowering plants at large (11%; Jones

1998). Seeds of cherries, plums, apples, pears, and other members of the rose family contain cyanogenic glycosides. Deermice consume cherry pits regularly, presumably with no adverse consequences. Intact glucosides are not toxic unless they are ingested along with β -glucosidase. Animals detoxify cyanide by converting it to thiocyanate. While this process protects the body from lethal levels of cyanide, the resulting thiocyanate competes with iodine in the thyroid and can lead to thyroid insufficiency. The body trades an acute, severe danger for chronic poisoning.

Common bracken, *Pteridium aquilinum*, and other ferns are rarely eaten by wildlife. Bracken is cyanogenic, but polymorphic for that trait. Sheep and fallow deer feed only on the acyanogenic form (Cooper-Driver and Swain, 1976). The African savanna grass, *Cynodon plectostachus* (Poaceae), after defoliation by army worms (*Spodophora exempta*), becomes toxic, even deadly, to cattle, because of its cyanide content. The golden bamboo lemur, *Hapalemur aureus*, of Madagascar eats growing tips of cyanogenic bamboo, *Cephalostachyum* ef. *uiguieri*. So far, it is not known how this animal avoids cyanide poisoning. The entire pathway for synthesis of the cyanogenic glucoside *dhurrin* has been transferred from *Sorghum bicolor* to *Arabidopsis thaliana*. The genetically engineered *Arabidopsis* sp. proved resistant to feeding by the flea beetle *Phyllotreta nemorum* of the Chrysomelidae family (Tattersall *et al.*, 2001).

Cyanide content of manioc (Manihot esculenta) clones varies with herbivore pressure and soil quality. The more pests and diseases, the more bitter the manioc grows, and "stressed" plants on infertile soil and/or lacking water, also are more bitter. (They do not have the option of outgrowing damage to their tissues.) However, only a few of the about 100 species of Manihot have been analyzed. Along the major rivers in Amazonia, cyanide-containing, bitter varieties of manioc dominate, while sweet forms are grown at the Amazon rim and in Central America. Riverine areas have more herbivorous mammals, and larger, semipermanent fields of more sedentary communities attract these herbivores. Frequently shifted small fields are not as vulnerable to herbivory (McKey and Beckerman 1993). Bitterness also protects against human theft in hit-and-run warfare: intruders on the run cannot easily provision themselves from local fields (in McKey and Beckerman, 1993). Current efforts to remove the cyanogenic glycosides, mainly linamarin and lotaustralin, by genetic engineering rely on the fact that the leaves retain cyanogenic compounds (6-40% linamarin) although these compounds are removed from the roots. As little as 1% linamarin remains (Pickrell 2003). It remains to be seen how resistant against pests and pathogens the genetically engineered manioc will be in the long run.

$$CO_2H$$

Azetidine 2-carboxylic acid

Proline

FIGURE 11.9 A Non-protein amino acid that interferes with proline metabolism.

$$NH_2$$
 NH_2 NH_2

FIGURE 11.10 Non-protein amino acids.

Non-protein amino acids

About 300 different non-protein amino acids occur in plants. They may be incorporated into proteins in place of the "correct" amino acids. If they are incorporated into enzymes, they can prevent them from functioning. This often leads to death of the animal. For example, azetidine 2-carboxylic acid in lily-of-the-valley, *Convallaria majalis*, and several legumes interferes with synthesis or utilization of structurally similar proline (Fig. 11.9).

Selenocystathionin occurs in seeds of the Central and South American *Lecythis ollaria*. It causes abdominal pain, nausea, vomiting, and diarrhea. *Lathyrus odoratus* contains aminopropionitrile (Rosenthal and Bell, 1979). 2,4-Diaminobutyric acid (Fig. 11.10) in *Lathyrus latifolius* and *Lathyrus sylvestris* causes muscular weakness, paralysis, and death in humans and other mammals. In India, 25000 cases of this neurolathyrism were reported in a population of 634000 people (Rosenthal, 1991). 3-Cyanoalanine (Fig. 11.10) from *Vivia sativa* and 15 other *Vivia* species causes convulsion, rigidity, and death.

Heterocyclic compounds such as L-mimosine (Fig. 11.10) cause hair loss in mules, horses, and sheep. Mimosine interferes with the enzyme that forms cysteine from methionine. Cysteine, in turn, is needed in growing hair follicles. Mimosine has been tried as a deflecting agent in sheep. After 2 days of intravenous infusion of mimosine, it takes 15 days for a sheep to drop its entire fleece (Reis *et al.*, 1975). However, it has not yet proven practical in sheep ranching.

L-Canavanine, another non-protein amino acid, occurs in high concentrations in the tropical vine *Dioclea megacarpa* (sea purse, a legume) and is very toxic to mice. Its structure is $NH_2C(NH_2)=NO(CH_2)_2CH(NH_2)CO_2H$. This is very similar to arginine, with which it interferes.

Oxalic acid

Oxalic acid occurs in sour-tasting plants such as soursob, *Oxalis pescaprae*, or the better-known *Oxalis acetosella*. Sheep that have ingested too much oxalate suffer labored breathing, "depression," weakness, and death. Rumen microbes degrade oxalate, but excessive doses result in insoluble calcium oxalate, which damages organs such as the rumen or kidneys. Oxalate may reduce calcium levels, and also inhibit respiratory enzymes (Burrit and Provenza, 2000).

11.2 Physiological effects of secondary plant metabolites

At the cellular level, plant secondary metabolites have five major effects on herbivores: (a) alteration of DNA replication, RNA transcription, and protein synthesis; (b) alteration of membrane transport processes; (c) enzyme inhibition and activation; (d) blocking of receptor sites for endogenous chemical transmitters; and (e) affecting the conformation of other macromolecules (Robinson, 1979).

The following will discuss effects at the organ level because this is what the biologist, farmer, zoo keeper, pet owner, or veterinarian deals with. From the viewpoint of the animal, specific life functions, such as digestion, muscle contraction, are affected. While harmful plant compounds mostly act in the digestive tract and the circulation beyond, they can enter cells and interfere with feeding as soon as they enter the mouth. For example, flavonoids in the Mediterranean rockrose *Cistus ladanifer* can prevent skeletal muscles of the mouth in rabbits from relaxing by disturbing the Ca²⁺ transport into the sarcoplasmatic reticulum of the muscle cells, preventing chewing. An avoidance response will result. The active flavonoids are apigenin, quercetin and 3,7-di-*O*-methyl kaempferol (Sosa *et al.*, 2004).

11.2.1 Inhibition of digestion

Secondary plant compounds can interfere with any stage of digestion. Here we focus on effects in the digestive tract rather than postabsorptive processes such as those taking place in the liver. Phenolics produced the following measurable responses in the voles *M. Microtus ochrogaster* and *Microtus pennsylvanicus*: reduced food digestibility, increased daily food requirements (15–20% with a 6% phenol diet), decreased growth rates, altered organ masses, caused organ lesions, increased excretion of uronic acid, reduced fat deposition, and increased basal metabolism (Lindroth and Batzli, 1983, 1984).

The same diet affects species differently. For instance, mountain hares, *Lepus timidus*, consume much birch in their winter diet, while European hares, *Lepus europaeus*, do not. A high concentration of birch phenolics in the diet causes massive sodium loss via the urine in European hares, but not in mountain hares (Iason and Palo, 1991). Further, among laboratory rodents, hamsters are extremely sensitive to tannins while rats and mice easily adjust within 3 days to doses of condensed tannins that are lethal to hamsters (Mehansho *et al.*, 1987).

Tannins reduce digestion in at least three ways. They reduce the available protein of the plant, as shown for moose (Robbins *et al.*, 1987); inhibit the activity of digestive enzymes (but Blytt *et al.*, suggested in 1988 that condensed tannins may not bind appreciably to digestive enzymes *in vivo*); and inhibit gut microorganisms. A very drastic example is the effect of the condensed tannin quebracho on weanling prairie voles (*M. ochrogaster*). Quebracho inhibits feeding and lowers the digestibility of proteins. Most weanling voles died when fed quebracho (Lindroth and Batzli, 1984).

Ruminants are particularly vulnerable to digestion inhibitors because the short pathway to the rumen provides much less opportunity for detoxification than the longer passage in hindgut fermenters. A phenol from a European birch ($Betula\ pendula$), platophylloside (5-hydroxy-1,7-bis-(4-hydroxy-phenyl)-3-heptan-one-3-O- β -D-glucopyranoside), inhibits rumen digestion. A significant effect was observed even with amounts below those found in birch (0.8% of dry matter) (Sunnerheim *et al.*, 1988). In black-tailed deer (*Odocoileus hemionus columbianus*), 1.2% of essential oils, containing monoterpenoids and sequiterpenoids, inhibit rumen microbes (Oh *et al.*, 1967).

North American prairie grasses contain polypeptides that act as inhibitors of the proteinase trypsin in mammalian herbivores. The grasses with these trypsin inhibitors are western wheatgrass (*Agropyron smithii*), big bluestem (*Andropogon gerardii*), little bluestem (*Andropogon scoparius*), and blue grama (*Bouteloua gracilis*) (Ross and Detling, 1983).

11.2.2 Growth suppression

The flavonoid quercetin suppresses growth somewhat in prairie voles, *M. ochrogaster*, while "tannic acid," a hydrolyzable tannin, has an even stronger effect (Lindroth and Batzli, 1984). Quercetin is toxic, but does not bind with protein and, therefore, does not affect protein digestibility. Tannic acid has some effect on protein digestibility but is also toxic.

Hamsters, unlike mice and rats, are not able to adjust to high levels of tannins in their diet by producing more proline-rich proteins in their saliva. A diet high in tannin has no effect on the salivary glands, and proline-rich proteins are not

induced. Hamsters on a diet containing 2% tannin do not grow. On this diet, at 60 days of age they still weigh the same as at 3 days. After switching diets, the former control animals, now on a 2% tannin diet, lost 20% of their body weight, while the former tannin-fed group now gained weight at almost the normal rate for young hamsters. A 4% quebracho tannin diet killed 75% of hamsters within 3 days, while rats and mice were not adversely affected (Mehansho *et al.*, 1987).

11.2.3 Reproductive inhibition

Dry salt grass, *Distichlis stricta*, has high levels of phenolics after fruiting. Cinnamic acids and their vinyl phenols lower uterus weight, inhibit follicular development, and lead to cessation of reproductive activity in the montane vole *Microtus montanus*. Specific active compounds are 4-vinylguaiacol and 4-vinylphenol, and also *p*-coumaric acid (4-hydroxycinnamic acid) (Berger *et al.*, 1977).

Phytoestrogens

Many plants contain estrogenic compounds. Estrone (Fig. 11.11) is found in seeds of date palms, pomegranates, and apples, and estriol in willow. These may be merely by-products of sterol metabolism, or serve a particular function. Harborne (1993) proposed that plants synthesize steroid hormones to deter feeding by mammals. Estrogenic compounds in plants are thought to upset the delicate hormone balance of mammals.

In Australia, lambing in sheep has dropped to as little as 30% after grazing on subterranean clover, *Trifolium subterraneum*. The clover contains the isoflavonoids formononetin and genistein (Fig. 11.11). These compounds mimic the steroidal nucleus of the natural female hormone estrone. Alfalfa, *M. sativa*, and ladino clover, *Trifolium repens*, contain the even more potent isoflavonoid coumestrol. It is 30 times more active than genistein or formononetin, but occurs in lower concentrations in the plant (Shutt, 1976).

In general, isoflavonoids in clovers and other legume pasture plants pose a danger to reproduction in farm animals because they lead to difficult labor, infertility, and lactation in unbred ewes. In Australia, an estimated one million ewes fail to lamb each year because of "clover disease" (Harborne, 1993).

During the Second World War, women in the Netherlands had to resort to eat tulip bulbs. They blamed their frequent menstrual upsets and ovulation failures on this diet. Harborne (1993) listed garlic, oats, barley, rye grass, coffee, sunflower, parsley, and potato tubers as having effects on estrus in women, but also cows. The active principle may not be a hormone but rather compounds that

FIGURE 11.11 Phytoestrogens.

mimic hormones. One such compound is *miroestrol* (Fig. 11.11). It was isolated from the roots of the legume *Pueraria mirifica* (Bounds and Pope, 1960). Pregnant women in Burma and Thailand use a root extract of this tree for abortions. Miroestrol is as potent as 17β -estradiol, and more than three times as active as the synthetic drug diethylstilboestrol.

Humans have used a range of plant species to manipulate their own fertility. A test of 10 plant species that bedouins of the Negev desert use for birth control showed that herbivorous rodents are also affected; 6 of the 10 species reduce fertility in laboratory rats. The fruits and shoots of one of these, the tree Ziziphus spina-christi (Christ's thorn; Rhamnaceae), delayed puberty and reduced the number of surviving offspring in its natural herbivore, the gerbil Meriones tristrami. This effect is thought to be part of a regulatory mechanism: the seeds eaten by the herbivore are dispersed, thus benefiting the plant. The ingested plant material reduces the population of the herbivore, keeping food supply and herbivore population in balance (Shappira et al., 1990). Specifically, the effects of the isoflavonoids formononetin and genistein (Fig. 11.11) may aid in adjusting animal populations to the available food supply. California quail (Lophortyx californicus) feed on legumes that vary in growth with rainfall. In good years, the plants are large, with low relative isoflavone concentrations, and the quail breed vigorously. The seeds carry the young through the winter. In dry years, the plants are stunted but rich in isoflavone on a fresh weight basis. The female quail suffer an estrogenic effect and lay fewer eggs (Leopold et al., 1976).

It has been suggested that metabolites resulting from detoxification of plant compounds such as ferulic acid, a detoxification by-product of coniferyl benzoate and analogous compounds, may interfere with reproduction. However, experiments have shown that coniferyl benzoate in the diet of Japanese quail (*Coturnix coturnix*) had no hormonal effects. Rather, costs of detoxication and reduced nutrient utilization deter wild birds such as ruffed grouse, *Bonasa umbellus*, from feeding (Jakubas *et al.*, 1993).

FIGURE 11.12 Androgens found in plants.

Androgens

Androgenic compounds occur in various plants. Celery stalks and parsnip roots contain trace amounts of androstenedione (Fig. 11.12), the same compound found in boar odor (Claus and Hoppen, 1979). Testosterone is also found in pollen of Scots pine (*Pinus sylvestris*).

11.2.4 Embryonic malformations by Teratogens

Steroidal alkaloids can cause malformations of fetuses during development. Domestic sheep feeding on *Veratrum californicum* often give birth to lambs with grotesquely shaped one-eyed heads. This *cyclopia* is caused by the steroidal alkaloids jervine, cyclopamine, and cycloposine (reviewed by Keeler, 1986).

11.2.5 Carcinogens

For obvious reasons, plant carcinogens have attracted extensive research. Here we review some examples that shed light on the effect of certain plant secondary metabolites on vertebrates, including humans. Tea is one of the best-investigated plant commodities. It contains up to 30% (of dry weight) *tannins* (polyphenols). These may cause esophageal cancer. Esophageal cancer used to be more prevalent in Japan than in Britain. By contrast to the Japanese, the British use milk in tea, which binds tannins and thus protects the esophagus. The Dutch also experienced much esophageal cancer during the nineteenth century when they used to drink tea without milk. The cancer rate dropped when they switched to coffee after they had established coffee plantations in Java and coffee became widely affordable (Morton, 1989). Esophageal cancer is more prevalent in northern Iran, southern Russia, and Turkey, where much tea is consumed (Morton, 1970). Brown and green tea are produced in different ways: brown tea, after withering for 1 day, is rolled which creates flavor and

astringency. Enzymes, cell contents and oxygen are mixed during the following fermentation at 27°C for 1–3 hours. Polyphenoloxidase transforms colorless phenolics into astringent tannins. Now the flavor and astringency is fully developed. Finally, the tea is fired at 93°C, resulting in a 5% moisture content. By contrast, green tea is not fermented. The enzymes are destroyed by steam, and the tea is then rolled and fired.

Maté (*Ilex paraguensis*), widely used as tea in Brazil, Paraguay, Uruguay, and Argentina, contains 9–12% tannin and up to 1.5% caffeine. Esophageal cancer rates are high in the Brazilian state of Rio Grande do Sul (Prudente, 1963) and Uruguay (Kaiser and Bartone, 1966), areas where consumption of maté is particularly high. Khat (*Catha edulis*) leaves, commonly chewed in Aden, Yemen and eastern Africa from Ethiopia to Madagascar, contain from 7 to 15% condensed tannins. Khat is chewed for several hours a day; an Ethiopian can chew as much as 2.5 kg per day. In some areas of heavy khat use, the rate of esophageal cancer is high (Morton, 1989).

Esophageal cancer rates were found to be correlated with consumption levels of sorghum in the Transkei, red wine in Western Europe generally, and high tannin apple cider in northern France (Morton, 1970). In Japan, esophageal cancer occurs particularly often in people who are eating tea-rice gruel, accompanied by tea drinking (Segi, 1975).

Some secondary plant compounds have proven carcinogenic in laboratory rodents. These include D-limonene in orange juice; 5-8-methoxypsoralen in parsley and parsnips; and caffeic acid in coffee, but also in smaller amounts in apples, apricots, broccoli, Brussels sprouts, cabbage, cherries, kale, peaches, pears, and plums (Ames and Gold, 1990; Ames *et al.*, 1990). How dangerous these compounds can be to human health, is still being debated.

11.2.6 Neurotoxicity

Some plants regularly eaten by humans contain neurotoxins that pose serious health problems. On Guam, for example, the seeds of *Cycas circinalis* used to be an important source of carbohydrates. Seeds of *Cycas rumphii* were ground into flour for tortillas. However, the seeds contain β -N-methylamino-L-alanine, a suspected excitotoxin that overstimulates and destroys nerve cells. This compound causes a parkinsonism-like disease in macaques (Spencer *et al.*, 1987). Other toxins have been proposed to be responsible for the disease, among them cycasin, another cycad toxin (Stone, 1993).

Among the Guam population, the disease known locally as *lytico-bodig* has been common for hundreds of years (Kurland, 1972). The disease is one of paralysis, tremors, and dementia and is an amyotrophic lateral sclerosis—parkinsonism

dementia complex (Lou Gehrig's disease); few cases are now seen (Stone, 1993). The Chamorros of Guam soak the cycad seeds for days and leach out up to 99% of the toxin by repeated rinsing (Duncan, 1991). The disease occurred most often during the time after the Second World War when the Chamorros ate flying foxes, typically prepared in coconut cream. Flying foxes subsist on cycad seeds and accumulate cycasin in their bodies. With depletion of the flying foxes, fewer humans suffered the disease. It has been suggested that the flying foxes were an important link (Cox and Sacks, 2002), but cycasin is water soluble and unlikely to accumulate in the bat body, if indeed it is the cause of the disease. Another, fat-soluble neurotoxin also remains under discussion (Stone, 2002).

11.2.7 Hallucinogens

Many plants contain compounds that cause hallucinations in people. In the book *One River*, Wade Davis describes the work he and his mentor Richard Evans Schulte carried out on the many Amazon plants with hallucinogenic effects in humans. They affect the central nervous system and may be designed to derail the behavior of herbivores, although the precise ecological role of these compounds remains uncertain. The effects of the active compound in cannabis (marijuana), Δ^9 -tetrahydrocannabinol (THC), on the mammalian brain promise new insights into its ecological function. Many parts of the brain (but not the brainstem) and the immune and reproductive systems carry cannabinoid receptors. These receptors mediate the action of endogenous cannabinoids, leading to pain relief, loss of short-term memory, sedation, and mild cognitive impairment. External cannabinoids act through this pathway. Although THC may protect a plant from detrimental ultraviolet radiation (cannabis plants contain more THC at higher altitudes), it may induce short-term memory loss in herbivores and thus reduce repeated attacks.

Rodriguez *et al.* (1982) noted that psychoactive compounds also often kill gastrointestinal parasites. They suggested that humans first used the plants to control parasites and discovered the effects on the central nervous system in the process.

The better-known and widely used hallucinogenic plants are San Isidro mushroom (*Psilocybe cubensis*), ergot (*Claviceps*), soma (*Amanita muscaria*), peyote (*Cactus lophophora*), yagé (or ayahuasca), the "vision vine" of the Amazon (*Banisteriopsis caapi*), cannabis (*Cannabis sativa* and *indica*) and perhaps coca (*Erythroxylum novogratense*),

An example of a hallucinogen affecting mammalian herbivores is the isox-azole muscimol (Fig. 11.13), a mushroom toxin. It is a γ -aminobutyric acid agonist in the central nervous system of vertebrates. Muscimol's role in nature

$$CH_2$$
 NH_3^+ HO CH_3 CH_3 CH_3 CH_3 CH_3 CH_3 CH_3 CH_3 CH_3

$$\begin{array}{c|c}
O > P & OH \\
O & \\
O & \\
O & \\
CH_2 & CH_2 & \\
CH_3 & \\
CH_3
\end{array}$$

Psilocybin

FIGURE 11.13 Hallocinogens.

is defense against fungivores. Virginia opposums, *Didelphis virginiana*, are fungivorous but avoid muscimol-containing mushrooms such as *A. muscaria*. They develop an aversion after having eaten the mushroom and falling ill (Camazine, 1983).

Another well-known hallucinogen is psilocybin from the mushroom genus *Psilocybe* (Fig. 11.13). Muscarine is an isoxazole derivative (Fig. 11.13). Table 11.3 lists some of the hallucinogens in plants.

11.2.8 Electron transport inhibitors

Inhibition of cytochromes of electron transport system can be caused by cyanogenic glycosides, such as amygdalin (Fig. 11.14) in bitter almonds, *Prunus amygdalus*, linamarin and lotaustralin in clover and birdsfoot trefoil, or dhurrin (Fig. 11.14) in *Sorghum vulgare*. The potent effect of cyanide on cell respiration has given rise to a recent serious conservation problem. In Southeast Asia, divers stun fish on coral reefs with a blast of cyanide to collect them for the aquarium trade. In the process, many fish are killed and the corals bleached, because their symbionts die (e.g. Payne, 2001).

Under special conditions, such as hot weather followed by a freeze and drought, cyanogenic compounds can accumulate in cherry leaves. Caterpillars such as the Eastern tent caterpillar, *Malacosoma americanum*, concentrate the toxic substances further in their bodies. Finally livestock inadvertently ingest their feces and carcasses on pastures or from water tanks. In 2001, this resulted in over

Table 11.3 Hallucinogens from Plants

Plant	Part	Compound	Use	Effect	Users	Place
Yopo tree Anadenanthera peregrina	Beans	Cohoba	Snuff	Hallucinogenic	Taino Indians	Hispaniola
Snake plant Turbina corymbosa	Seeds	Ololiuqui: LSD-like compounds	Drink	Hallucinogenic	Aztecs	Mexico
Coco plant Erythroxylon	Leaves	Cocaine	Chewed	Stimulant, analgesic	Incas, South Americans	Peru etc.
Cactus Lophophora williamsii	Top: mescal button	Mescaline	Swallowed	Hallucinogenic	Native American Church	Mexico, Texas
Teonancatl Psilocybe mexicana	Mushroom	Psilocybin	Eaten	Hallucinogenic	Aztecs, modern Native Americans	Mexico, Central America
Hemp Cannabis sativa	Flowers, leaves	Marihuana: tetrahydrocannabinol	Smoked	Sedative	Worldwide	Origin: China
Khat Catha edulis	Leaves	Alkaloids: cathinone, cathine	Chewed, tea	Stimulant	Yemenis, Ethiopians, Somalis' etc.	East Arabia, East Africa, Madagascar
Ayahuasca (yagé) Banisteriopsis caapi	Bark of vine	Harmine, harmaline	Drink	Hallucinogenic	Shamans, various indigenous groups	Amazon basin
Ebéna Virola theiodora	Inner bark	Several tryptamines, including the most potent hallucinogen known from nature: 5-methoxy-N,N-dimethyltryptamine	Snuff	Hallucinogenic	Yamomami	Amazon basin

$$H_2COH$$
 H_2COH
 H

Amydalin

FIGURE 11.14 Cyanogenic glycosides inhibit the electron transport chain.

500 stillbirths among thoroughbred mares in Kentucky, caused by the blocking of oxygen delivery to the cells by cyanide (Holden, 2001).

11.2.9 Renal effects of chronic poisoning

Voles (*M. pennsylvanicus*) suffer renal lesions (interstitial nephritis) when fed extracts of white clover, *T. repens*. Milder lesions were observed after feeding on reed phalaris (*Phalaris arundinacea*) and timothy (*Phleum pratense*). Many varieties of reed phalaris contain the toxic compounds gramine and tryptamine (Fig.11.15). In summer and autumn, protein levels in the leaves decrease, fiber content goes up, and secondary compounds increase in concentration. Therefore, second growth plants have more toxic effects on voles than the spring plants that grow fast and have lower levels of secondary compounds (Bergeron *et al.*, 1987).

11.2.10 Lethal light: toxicity linked to ultraviolet radiation, or why mammals are nocturnal

Alarmingly, the ozone hole has turned mammalian food into food for thought. The ozone hole permits an increase of ultraviolet radiation in the range 290 to 320 nm, the same wavelengths that induce furocoumarin (also known as

FIGURE 11.15 Compounds causing renal lesions.

Angelicin

Psoralen

FIGURE 11.16 The furocoumarins.

furanocoumarin) production in certain plants. Many plants contain compounds that are toxic to herbivores after ingestion *only if* the animals are exposed to sunlight. These include the quinone hypericin from St. Johnswort, *Hypericum perforatum*.

A well-investigated group of potent phototoxic compounds are the *fura-nocoumarins* (psoralens) (Fig. 11.16), compounds that are also used as fish poisons. Furocoumarins absorb ultraviolet at around 330 nm and are converted to triplet stages; these, in turn, produce singlet oxygen that damages amino acids (Fig. 11.17).

Photosensitization by plants containing furocoumarins has been described for geese, chickens, turkeys, ducklings, sheep, dairy cattle, horses, hyrax, and humans, among others. Herbivorous insects and mammals can avoid the toxic effects if they stay in vegetation cover, underground, or lead a crepuscular or nocturnal life. Leaf-rolling behavior in insects has been seen as protection against phototoxic effects of constituents of their diet. How do reptiles that need daylight or direct solar radiation to be active avoid damage by ultraviolet radiation? Does their skin armor screen out the radiation sufficiently?

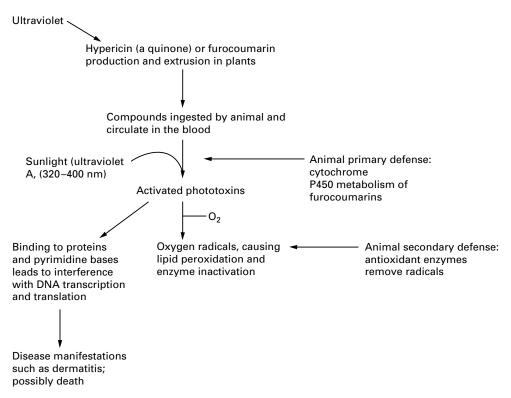


FIGURE 11.17 Phototoxic effects of plant compounds on mammalian herbivores.

Furocoumarins occur particularly frequently in the Leguminosae, Moraceae, Rutaceae, and Umbelliferae (carrot family) (Larson and Berenbaum, 1988). One better-investigated species of the latter family is *Pithuranthos triradiatus*, a desert shrub in the Middle East. Hyrax, *Procavia capensis syriaca*, feed on this plant. After eating branches and staying in the sun, hyraxes develop symptoms of photosensitization: they become apathic, photophobic, and develop injuries around the eyes, and on the back. The animals ate less of older branches, but ingested more furocoumarins, because, although older shoots contain more of the phototoxic compounds imperatorin and isoimperatorin than young shoots, the young shoots are "loaded with higher concentrations of secondary metabolites." Plants seem to avoid autotoxic hazards; there are less furocoumarins in shoots that are still growing and so DNA strands are not damaged in mitotically active tissue (Ashkenazy *et al.*, 1985).

Plants that can cause photodermatitis include carrots, celery, parsnip, dill, fennel, caraway, parsley, lovage, anise, and chervil among the Apiaceae (Umbelliferae, or carrot family), figs (Moraceae), and citrus fruits (grapefruit, lemon, lime,

and orange) among the Rutaceae (Berenbaum, 1991). In arid and semiarid areas of North America such photosensitizers are widespread in plants and occur particularly in the Asteraceae (Downum *et al.*, 1989).

Any increase in ultraviolet radiation will exacerbate the danger to herbivores. Ozone depletion by 25% leads to an increase of 50% in ultraviolet B over ambient levels (El-Sayed, 1988). Will livestock be drastically affected if levels rise more in the future? Will animals be able to shift to nocturnal activity? If not, will large portions of the vegetation be rendered useless for domestic stock and wild species or will animals have to be fed indoors? Will humans be affected?

Furocoumarins in the diet can cause photophytodermatitis in humans. Carrots, for instance, contain furanocoumarins. Humans touching leaf surfaces of a variety of plants can pick up the coumarins and produce handlers have suffered from photophytodermatitis. In addition to diet, perfumes can be a source of these compounds. Oil of bergamot contains a furocoumarin and used to be an ingredient of some perfumes. Such perfumes can cause skin rashes upon exposure to sunlight. Eight species of Umbelliferae (carrot family) and Rutaceae (citrus family) were shown to contain several photoactive furanocoumarins. These include xanthotoxin, psoralen, and bergapten. They are more concentrated on younger, spring leaves and least on old, autumn leaves. Zobel and Brown (1990) have suggested that a leaf has an "atmosphere" of sublimating compounds that is toxic to approaching bacteria or fungal spores; furocoumarins are actually antibacterial.

11.2.11 Metabolic costs of detoxification

Secondary plant compounds can be a substantial burden for a herbivore. Nutrients are diverted for conjugation (p. 330) and this can have severe consequences for the organism at low nutrient levels. Malnourished people in developing countries often treat their ailments with herbal cures that, unfortunately, aggravate their problems because precious carbohydrates or amino acids are diverted for detoxification and excretion of plant compounds. Oxidation and conjugation, the two phases of biotransformation (detoxication), require the input of energy. The basal metabolic rate in the meadow vole, *M. pennsylvanicus*, increased by 13.6 to 22.6% when 6% (by dry mass) gallic acid, a phenolic, was added to its diet. This is thought to be the metabolic cost of detoxification (Thomas *et al.*, 1988).

Sheep that are given orcinol intravenously increase their energy expenditure by 5%. (Orcinol is a non-tannin phenolic that occurs in Ericaceae, including heather, *Calluna vulgaris*.) If orcinol and quinol are infused into the rumen,

they have no effect on energy expenditure but the intake of digestible energy is reduced. This shows that the effectiveness of the detoxication system is crucial for utilization of toxin-containing plants (Iason and Murray, 1996). It is assumed that microbes in the rumen modify toxins, reducing their systemic effects.

Koalas (*Phascolarctos cinereus*) increase their glucose intake by 20% if they eat *Eucalyptus* spp. foliage, which requires conjugation reactions with glucuronic acid for excretion (Eberhard *et al.*, 1975). Goats and sheep eat more of a toxic diet if given surplus food, which helps detoxication (Provenza, 2004).

11.2.12 Interactions of secondary compounds

Complex interactions may occur if a herbivore ingests a variety of defense compounds from one or several plant species as the various compounds may counteract each other. For instance, tannins can help to reduce the toxic effects of cyanogenic plant compounds. In the papaya, *Carica papaya*, the cyanogenic glycosides tetraphyllin B and prunasin (also present in the passion fruit, *Passiflora edulis*) produce hydrogen cyanide upon enzymatic hydrolysis. The plant also contains protein-precipitating tannins that inhibit enzymes and thereby the enzymatic production of cyanide from tetraphyllin B, prunasin, and also amygdalin. If low levels of hydrogen cyanide are measured in a plant, this may be because tannins are present to interact with the enzymes (Goldstein and Spencer, 1985).

Secondary plant compounds can have negative and positive effects on a herbivore, depending on the form in which they are ingested. In an *in vitro* experiment, rumen microbes from sheep were inhibited by the simple phenols orcinol and quinol: production of gas and volatile fatty acids were reduced. Both compounds reduced digestion of the substrate. By contrast, arbutin, a phenol glycoside, stimulated production of gas and volatile fatty acids. This was probably because of the availability of the fermentable sugar moiety of the glycoside (Murray *et al.*, 1996). Many simple phenolics exist in plants in the form of glycosides. The sugar moiety is liberated in the gastrointestinal tract on hydrolysis by glucosidase enzymes. Therefore, beneficial effects of the sugar moiety may counteract the inhibitory effects of phenolics (Murray *et al.*, 1996).

11.2.13 Effects of toxic plants on herbivore populations

Vole cycles are thought to be influenced, at least in part, by the ratio of toxic plants in the food (Freeland, 1974). According to this hypothesis,

increasing numbers of voles deplete the more palatable plant species. At high population levels, their diet would then contain a high percentage of "toxic" plants. This would affect health and growth, especially of young animals, and the population would decline. Data presented by Batzli and Pitelka (1971) in response to this hypothesis, however, did not substantiate the notion that California voles (*M. californicus*) resort to more toxic plants when population levels are high.

Endangered herbivorous mammals that compete with large numbers of live-stock may be forced to feed on less-palatable or outright toxic plants, especially if fenced in and poisoning could result in population declines. This has been suggested for the endangered pampas deer, *Ozotoceros bezoarticus*, which shares its – often severely grazed – range with large numbers of cattle and sheep (D. Müller-Schwarze and D. E. Moore, unpublished field observations). High population levels, consumption of tannin-rich browse, and high mortality were correlated in kudu, *Tragelaphus strepsiceros*, in fenced areas (van Hoven, 1991).

Researchers currently debate whether terrestrial ecosystems are regulated "bottom up" by secondary plant metabolites that limit food available to herbivores or "top down" by predators that keep herbivores in check, resulting in lush plant growth. One recent study ruled in favor of the second. Islands of varying sizes, created by the hydroelectric impoundment of Lake Guri in Venezuela, permitted these hypotheses to be tested. On predator-free islands 10 to 100 times more herbivores were living, such as rodents, howler monkeys, iguanas, and leaf-cutter ants, than on the nearby mainland. The densities of seedlings and saplings of canopy trees on these islands were severely reduced, owing to these abundant herbivores (Terborgh *et al.*, 2001).

11.2.14 Potential beneficial effects of secondary plant compounds

Plant secondary metabolites in the diet may actually benefit consumers of plant materials. Many phenolics are antioxidants that mop up free radicals. Gallic acid is particularly active, and resveratrol in grapes and red wine has been in the news for that reason. Long-term ingestion of undesirable plant compounds may boost the immune system. In Liberia, a diet of cassava exposed adults chronically to medium to high levels of cyanogenic glycosides. They had elevated levels of immunoglobulins, especially IgM. Dietary cyanide bound to proteins, and bound forms of cyanide in the diet, may act as antigens. IgM is strongly antibacterial and may play a beneficial role in resistance to tropical bacterial diseases such as Hanson's disease (leprosy), louse-born relapsing fever, yaws, and cholera (Jackson 1994).

Plant species	Plant part	Herbivore species	Compound	Reference
Green alder Alnus crispa	Buds, catkins	Snowshoe hare Lepus americanus	Pinosylvin, pinosylvin methyl ether, papyrific acid	Bryant <i>et al.</i> , 1983; Clausen <i>et al.</i> , 1986
Green alder	J versus M internodes	Snowshoe hare	J has more pinosylvin and pinosylvin methyl ether	Clausen et al., 1986
Paper birch Betula resinifera	J growth versus winter-dormant internodes	Snowshoe hare	J has 25 \times concentration of papyriferic acid	Reichardt <i>et al.</i> , 1985
Quaking aspen Populus tremuloides	Scales of flower buds	Ruffed grouse Bonasa sp.	Coniferyl benzoate	Jakubas <i>et al.</i> , 1989
Balsam poplar Populus balsamifera	Buds	Snowshoe hare	Mono- and sesquiterpenes	Reichardt et al., 1990b
White spruce Picea glauca	J stages	Snowshoe hare	Camphor	Sinclair et al., 1988
Balsam poplar	J twigs	Snowshoe hare	2,4,6-Trihydroxy- dihydrochalcone	Jogia <i>et al.</i> , 1989
Willow Salix spp.	J stages	Mountain hare Lepus timidus	Phenolic glycosides, catechin	Tahvanainen <i>et al.</i> , 1985

Table 11.4 Plant parts differ in chemical defense against vertebrate herbivores

J, juvenile; M, mature.

11.3 Chemical defense strategies by plants

11.3.1 To the bitter end: distribution of defense compounds within plants

Plant *parts* often differ in their levels of secondary metabolites (Table 11.4), some containing extremely high levels. For example, the creosote bush, *Larrea tridentata*, of the western United States has phenolic resins concentrated in leaves, amounting to as much as 18% of dry weight. In experiments, desert woodrats, *Neotoma lepida*, selected plant parts of creosote bush with low levels of resins (Meyer and Karazov 1989).

Mountain hares, *L. timidus*, selected birch twigs of intermediate diameter (1.5 to 3.0 mm) and attained the best body weight on such a diet. Smaller twigs are high in phenolic glycosides, and larger twigs are least digestible (Palo *et al.*, 1983). In the genus *Populus*, twig bark usually has higher concentrations of phenolics than trunk bark. Also, the levels of these compounds in twigs vary more, and the spectrum of compounds is narrower than in trunk bark (Pearl and Darling, 1968; Palo, 1984).

Particularly valuable and/or vulnerable parts are protected best. Because of their size and seasonal changes, trees provide instructive examples. In green alder, *Alnus crispa*, defense compounds, such as pinosylvin, pinosylvin methyl ether and papyriferic acid, are concentrated in buds (2.6% of dry weight) and catkins (1.7%), and least abundant in the internodes (0.05%). Snowshoe hares, *L. americanus*, reject buds and catkins, while they eat second-year internodes (Bryant *et al.*, 1983; Clausen *et al.*, 1986).

In quaking aspen, *Populus tremuloides*, the scales of the flower buds have the highest concentrations of coniferyl benzoate (Fig. 11.2). Ruffed grouse avoid trees particularly rich in this compound (Jakubas *et al.*, 1989). Experiments have shown that coniferyl benzoate inhibits food consumption (Jakubas and Gullion, 1990). Balsam poplar, *Populus balsamifera*, has redundant compounds, practicing "defensive overkill" against snowshoe hares, who do not eat its buds. The buds contain resins, mainly made up of mono and sesquiterpenes based on cineol, benzyl alcohol, and (+)- α -bisabolol. Juvenile internodes of shoots from stumps are protected by salicaldehyde, while the current annual growth of mature plants is protected by 6-hydroxycyclohexenone (Reichardt *et al.*, 1990b). The buds are defended by 2,4,6-trihydroxydihydrochalcone, but the internodes of juvenile trees by three volatiles: salicaldehyde, 6-hydroxycyclohexenone, and 1,2-cyclohexadione. These volatiles may signal that other noxious compounds are present, constituting an example of chemical aposematism (Eisner and Grant 1981).

The *age* of plant tissues influences palatability in other species as well: Younger plant parts generally have higher concentrations of defense compounds. An example is green alder, *A. crispa*. Juvenile internodes have three times as much pinosylvin and pinosylvin methyl ether as mature internodes. Snowshoe hares, *L. americana*, prefer to feed on mature internodes (Clausen *et al.*, 1986). Similarly, snowshoe hares prefer internodes of the mature growth phase of winter-dormant paper birch, *B. resinifera*, to juvenile growth phase internodes. In perhaps the most dramatic example, the internodes of the juvenile phase contain as much as 30% of the dry weight (25 times as much) as the triterpene papyriferic acid (Fig. 11.5) than mature internodes and this acts as a feeding deterrent (Reichardt *et al.*, 1985). Snowshoe hares also rarely browse juvenile stages of white spruce, *Picea glauca*, a slow-growing late succession tree, but utilize mature stages heavily. Juvenile spruce contained four times as much camphor as mature stages. In experiments, camphor deterred feeding (Sinclair *et al.*, 1988).

Strong chemical defenses of reproductive units such as seeds, bulbs, or tubers, as opposed to the vegetative parts, are commonplace. For instance, potatoes, which have to survive in the ground from one growing season to the next,

contain glycoalkaloids such as solanine and chaconine, saponins, a phytohemagglutinin, proteinase inhibitors, sesquiterpenes phytoalexins, and phenols (Johns, 1990). These effectively deter fungi, invertebrates, and rodents.

Female flowers of dioecious trees are better defended than males. Ruffed grouse, *Bonasa umbellus*, prefer male flowers of quaking aspen, *Populus tremuloides*, in winter, and leaves of male specimens in summer (Svoboda and Gullion 1972). Ruffed grouse also feed more on male trees of balsam poplar (Bryant *et al.*, 1991).

11.3.2 Seasonal variation of levels of chemical defenses

Are plants best defended during the season when they are most vulnerable to herbivory? Specifically, do bark and twigs contain their highest level of plant secondary metabolites during the northern winter when they are browsed by mammals small and large, and cannot respond by regrowth? Do they contain leaves with most plant secondary metabolites during the months when insect attacks are most intense? The answers to these questions are complex. It has been suggested that woody boreal plants allocate the largest amount of "defense" compounds to tissues that are essential to photosynthesis and growth and, therefore, contribute significantly to plant fitness (Palo, 1984).

In the European birch, *B. pendula*, the relative concentration of protein, and with it the *in vitro* organic matter digestibility, increases in spring and early summer, while the proportions of cell walls (neutral detergent fiber) and water-soluble phenols, especially catechin, decrease during that time. Such seasonally changing allocation within the plant results in increased palatability to ruminants. The plant can afford to lose biomass to herbivores during the time of rapid growth (Palo *et al.*, 1985). Bark of the birch, *B. pendula* has low tannin levels (4.1–4.8%) in winter, but a maximum (5.8–6.2%) in September (Chernayeva *et al.*, 1982). In the same species, total phenolics reach their maximum (3%) in winter twigs, and are lowest (0.9%) in June (Palo *et al.*, 1983). Leaves of *Betula tortuosa* contain only 7% total phenolics when they emerge, but up to 15% in autumn (Palo, 1984).

Levels of phenolic glycosides in bark of *Salix* and *Populus* spp. generally increase towards the winter. Concentrations are highest during winter dormancy and lowest in September/October (Thieme, 1965). Buds, by comparison, contain the highest levels in May immediately before leaf burst. The leaves contain the least phenolic glycosides in September (Thieme, 1965).

The levels of phenolic glycosides can also vary diurnally. In *Salix fragilis* and *Salix pupurea*, their concentrations decrease by 20% and 40%, respectively, from early morning to late in the day (Thieme, 1965).

FIGURE 11.18 Caffeine.

11.3.3 Plant secondary metabolites and nutrients in fruits and seeds

Fruits typically are more chemically defended when still unripe, thought to deter frugivores before the seeds have matured. However, in some cases high levels of secondary plant metabolites persist during the ripe stage. For example, among North American nightshade species, ripe fruits of Solanum carolinense have high levels of glycoalkaloids, while in Solanum americanum they are negligible (Cipollini and Levey, 1997). The specific compounds are α -solasonine and α -solamargine. Among birds and mammals eating the seeds, American robin and opossum, D. virginiana, pass almost all seeds intact and are possibly major seed dispersers, while bobwhite quail and deer mice, Peromyscus maniculatus, destroyed most of the seeds. All four species, however, preferred the low-glycoalkaloid species and were deterred by the high-glycoalkaloid species. This supports the general toxicity hypothesis as opposed to the directed toxicity hypothesis, which posits that toxins are directed against specific consumers. Highly preferred, nutritious fruits of S. americanum have lower levels of glycoalkaloids. This supports the removal rate hypothesis: highly nutritious fruits that consumers remove quickly and predictably will show lower levels of secondary plant metabolites. (The opposite is the nutrient-toxin-titrationhypothesis: the higher the nutritional quality of a fruit, the more secondary metabolites it will have.) Cipollini and Levey (1997) concluded that the primary function of the glycoalkaloids in these Solanum fruits is defense against pests and pathogens. This, in turn, will influence the interactions between fruits and seed dispersers. In general, the plants appear to strike a compromise: the fruits contain enough alkaloid to avoid rotting but not so much that animals avoid eating them.

As these examples show, different defenses among plant parts and seasonal differences have to be considered together. Coffee (*Coffea arabica*) is a well-investigated example of increased chemical defenses when the plant is most vulnerable to herbivory. The concentration of the alkaloid caffeine (Fig. 11.18) varies between plant parts and with the growth cycle. In the germinating seed, the

developing shoot has little alkaloid as long as it is protected by the endocarp. The caffeine concentration in the seedling is more than twice as high as in the seed (Baumann and Gabriel, 1984). In the young still-soft leaf, the concentration of caffeine increases to the high level of 4% of dry weight. The daily formation of this alkaloid decreases rapidly from 17 to 0.016 mg/g tissue as the leaf matures. The fully expanded leaf remains at that low level (Frischknecht *et al.*, 1986). Leaves are essentially alkaloid free before they die off. The caffeine is probably recycled within the plant, and its nitrogen used for protein synthesis. In the developing fruit, the alkaloid content is high (2% dry weight) early on when the pericarp is still soft but it decreases as the endocarp matures and hardens and is only 0.24% when the fruit is fully ripe. Harborne (1993) emphasized the advantage to the plant of reclaiming nitrogen from the alkaloid molecule as soon as the latter is no longer needed for defense.

The effect of caffeine has been tested with insect herbivores. Tobacco hornworms, *Manduca sexta*, die from a diet with 0.3% caffeine, and the beetle *Caliosobruchus chinensis* becomes sterile at a concentration of 1.5% (Nathanson, 1984). However, experiments showed that caffeine does not protect the coffee plant against the leaf miner *Perileucoptera coffeella*, one of the major pests of coffee (Filho and Mazzafera, 2000).

Humans, the world's greatest caffeine-consuming vertebrates, have tested their tolerance limits of toxic effects in often extreme ways. Along the lower Putumayo in southwest Colombia, the Indians cut meter-long lengths from the vine *yoco* (*Paullinia yoco*) and keep it at their homes for up to 1 month. As needed for drinks in the early morning, they rasp the bark and squeeze the milky sap into cold water. They drink the extract from 100g bark in one serving. The bark contains three times as much caffeine per weight as coffee beans. Even with the incomplete extraction in cold water, one dose delivers as much caffeine as more than 20 cups of coffee! The strong potion suppresses hunger for up to 5 hours, and energizes both men and women for marches of 20 miles or more (Davis, 1996). Too much of a good thing can prove fatal: The French novelist Honoré de Balzac died in 1850 of coffee poisoning. He was 51 years old. To keep writing day and night, he downed four dozens cups of coffee every day (Nathan, 1998).

11.4 Feeding or avoiding? herbivores vis-à-vis plant defenses

11.4.1 Secondary plant compounds, food preferences, and consumption

Herbivores and their food plants live in a delicate balance. We distinguish generalist herbivores from specialists. It is thought that generalism

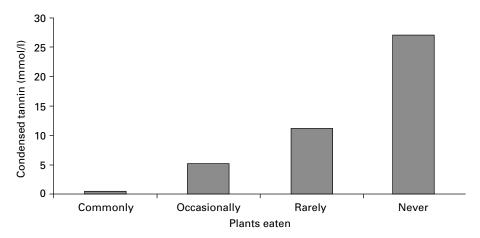


FIGURE 11.19 Condensed tannin concentrations in plants eaten to various degrees by giant tortoises on Adabra Island. (After Swain, 1979.)

evolved to avoid overloading of the herbivore's system with toxic compounds from only one or two plant species (Lindroth, 1988). In temperate and arctic regions, herbivores enjoy a larger number of food plants during summer than in winter. This also reduces the herbivore pressure on any one plant species (Palo, 1984). During the growing season, a plant may be able to outgrow herbivore damage more or less rapidly. This is particularly true for tropical plants. Depending on season and latitude, plants are exposed to different predation pressures: boreal plant species, such as deciduous and coniferous trees, suffer vertebrate herbivory for most or all of the year, while insect and pathogen damage is limited to the short growing season. The following covers feeding patterns of reptiles, birds, and mammals in relation to plant defenses.

Reptiles

Giant tortoises (*Geochelone gigantea*) on Aldabra Island in the Seychelles archipelago are more likely to feed on certain plant species, the less tannins they contain (Swain 1979; Fig. 11.19).

Birds

Ruffed grouse, *B. umbellus*, feed on staminate flower buds and extended catkins of trembling aspen, *P. tremuloides*. In winter these birds select specific trees or clones. Analysis for alkaloids, tannins, and other phenolics showed that feeding preferences were not related to the levels of tannin or total phenolics in

flower buds or catkins. However, trees with no history of feeding had buds with higher concentrations of coniferyl benzoate and lower levels of proteins. Catkins have less coniferyl benzoate than buds, but they do not differ in protein content. Extended catkins are probably eaten in spring because they have little bud scale material. Bud scales contain almost all of the non-tannin phenolics found in catkins and dormant buds. Avoiding bud scales appears to be the feeding strategy of the grouse and possibly other birds (Jakubas *et al.*, 1989).

Canada geese, *Branta canadensis*, feed on coastal marsh plants that are low in soluble phenolics. The less phenolics (in percentage dry weight) the leaves of a species contained, the more often geese were observed to feed on it. Nutrient content of the plants was not related to consumption. Tannic acid or quebracho from the tropical American tree *Schinopsis lorentzii* (from *quebrar*, to break, and *hacha*, ax, because of its hard wood) added to *P. pratensis* rations inhibited feeding, and phenolic acids (1% ferulic acid and 1.5% *p*-coumaric acid) were slightly inhibitory. Isolated extracts or pure secondary compounds were never as active as the plant itself. The fractionation may have abolished synergistic effects in addition to changing other factors such as texture or water content (Buchsbaum *et al.*, 1984). Subarctic ptarmigans and grouse also avoid plants with a surface cover of phenolic resins. The resins are antimicrobial and interfere with microbial digestion in the caecum of these birds (Bryant and Kuropat, 1980).

Adult greylag geese, *Anser anser*, respond to non-food plant odors such as sage, violet, lavender, onion, or peppermint with head shaking before they contact the material with their bill. This indicates the role of olfaction in food selection (Würdinger, 1979). The response is present on the first day of life (Section 9.4).

The *ratio* of nutrients to plant secondary metabolites may be more important for food choice by birds and other herbivores than the level of the metabolites per se. Leaves of Arctic willow, *Salix arctica*, develop each spring from closed buds to open buds, rolled leaves, and finally open leaves. During that progression, phenol levels rise, fiber content decreases, and protein content peaks during the open-bud phase. The plant is most vulnerable during the open-bud and rolled-leaves stages when the combination of these three factors favors the herbivore. Greater snow geese, *Chen caerulescens atlantica*, feed on the open-bud and rolled-leaf stages, but not the closed-bud and open-leaf stages. The geese feed during these stages because the high protein content, declining fiber levels, and not-yet-high phenols level provide an optimal compromise. A high phenol:protein ratio lowered feeding more than large concentrations of phenolics alone (Gauthier and Hughes, 1995).

Bullfinches, *Pyrrhula pyrrhula*, feed on ash tree seeds with a high fat content and low tannin levels (Greig-Smith, 1988). Similarly, red-winged blackbirds, *Agelaius phoeniceus*, prefer sunflower seeds with higher oil concentrations, but avoid all

levels of anthocyanin. Cultivars with high levels of anthocyanin and low levels of oil may be resistant to these birds (Mason *et al.*, 1989a). Zebra finches, *Taeniopygia guttata*, feed less on lettuce leaves or seed stands of *Setaria italica* that have been scented with oils of peppermint, lavender, sage, eucalyptus, anis, caraway, or crystalline thymol than control leaves (Würdinger, 1990). Finally, house sparrows, *Passer domesticus*, prey less on seeds of those hybrids of the cereal crop sorghum that contain high levels of tannins (Tipton *et al.*, 1970). Table 11.5 summarizes some of these studies.

Thrushes show distinct preferences and aversions for flavors of fruits. European blackbirds, *Turdus merula*, and songthrushes, *Turdus philomelos*, learned to associate the flavor and color of an artificial "fruit" made from flour and lard after only one experience (Sorensen, 1983). Among 11 fruit extracts tested, the birds preferred ivy (*Hedera helix*) most and almost always rejected buckthorn (*Rhamnus cathartica*). Preference for dogrose (*Rosa canina*), elder (*Sambucus nigra*) and hawthorn (*Crataegus monogyna*) was intermediate. The birds were wild caught and, therefore, had experience with wild berries.

Mammals

How much a mammal eats of a given plant often depends on the levels of different classes of chemical constituent, notably nutrients and plant secondary metabolites. As in birds, it is not the plant defense compounds alone, but rather complex balances between nitrogen and carbohydrate contents, levels of defense compounds, and fiber that determine palatability.

Marsupials

The western grey kangaroo (*Macropus fuliginosus*), the major indigenous herbivore in southwestern Australia, avoids plants of the family Myrtaceae (which include *Eucalyptus* spp. cloves, and guava), which includes 3500 species on that continent. Essential oils characterize the Myrtaceae; many species contain two to nine of these. In one experiment, seven Myrtaceae species were not browsed, while comparable species of other families were. All seven species contained the monoterpene 2,5-dimethyl-3-methylene 1,5-heptadine. The kangaroos also browsed one non-Myrtaceae species (*Sollya heterophylla*, Pittosporaceae) very little. This species contained the same monoterpene (Jones *et al.*, 2003).

The brushtail possum, *T. vulpecula*, feeds on eucalyptus leaves. Among the terpenes these contain, 1,8-cineole is the most abundant. Feeding experiments with increasing levels of cineole showed that this compound limits food intake. In the brushtail possum, multiple pathways oxidize cineole, and this total system, rather than any one enzyme, limits the amount of cineole that the animal

Bird species	Plant species	Compound(s)	Reference
Red-winged blackbird Agelaius phoeniceus	Sunflower seeds	Anthocyanin	Mason et al., 1989b
Bullfinch P. pyrrhula	Seeds of ash, Fraxinus sp.	Tannins	Greig-Smith, 1988
House sparrow Passer domesticus	Sorghum: unripe seeds	Tannins, "astringents"	Tipton <i>et al.</i> , 1970
Ruffed grouse Bonasa umbellus	Aspen Populus tremuloides	Coniferyl benzoate	Jakubas et al., 1989
Canada goose Branta canadensis	Coastal marsh plants	Phenolics	Buchsbaum <i>et al.</i> , 1984
Greater snow geese Chen caerulescens atlantica	Arctic willow Salix arctica	Phenols	Gauthier and Hughes, 1995

Table 11.5 Plant secondary metabolites inhibiting feeding by birds

can process (Boyle and McLean, 2004). Levels of the terpenes 1,8-cineole and limonene correlate significantly with the those of the formylated phloroglucinols, the active antifeedants. This suggests that, as a result of aversive conditioning, the volatile terpenes signal to the mammalian herbivore how unpalatable a particular plant is (Moore *et al.*, 2004).

Rodents

The voles *M. pennsylvanicus* and *M. ochrogaster* did not feed at all on some of 15 tested plant species in the laboratory. Others were eaten to different degrees, and 73% of the variance was explained by the nitrogen contents. The selectivity in the field corresponded to the palatability ratings in the laboratory: *Taraxacum* (dandelion), *Medicago* (alfalfa), and *Trifolium* (clover) spp. were the most palatable. *Andropogon* (salsify) and *Pastinaca* (parsnip) spp. were unpalatable in field and laboratory (Marquis and Batzli, 1989). The unpalatable species contain toxins, specifically alkaloids in *Penstemon digitalis* (Lindroth *et al.*, 1986), glucosinolates in *Barbarea vulgaris* (van Etten and Tookey, 1979), and furocoumarins in *Pastinaca sativa* (Berenbaum, 1981).

Turning to conifers, the meadow vole *M. pennsylvanicus* attacks seedlings of the introduced Norway spruce, *Picea abies*, and the North American red (Norway) pine, *Pinus resinosa*, but not white spruce, *Picea glauca*, and white pine, *Pinus strobus*. The last two have in common β -myrcene and bornyl acetate (Fig. 11.20) which are absent in the former two tree species. These two compounds are also toxic to insects and may be the agents protecting the trees (Roy and Bergeron, 1990a). Total levels of phenolics also tended to be higher in the two species

$$CH_2$$
 CH_3
 $CH_3COO_{1/1/1}$
 $CH_3COO_{1/1/1$

FIGURE 11.20 Secondary metabolites in some conifers.

of conifers that are avoided, but monoterpenes are seen as more important in defense against voles than phenolics (Bucyanayandi *et al.*, 1990).

In winter, Abert's (tassel-eared) squirrels (*Sciurus aberti*) of the southwestern United States feed on the inner bark of ponderosa pine (*Pinus ponderosa*) twigs, besides seeds of pines and piñons. The squirrel selects specific, so-called "feed" or "target" trees that are low in α -pinene while ignoring other nearby trees (Farentinos *et al.*, 1981). Snyder (1992) found low levels of β -pinene and β -phellandrene in the xylem of such "feed trees." Trees defoliated by squirrels suffered up to 90% loss in fitness indicators such as growth, cone production, and seed quality (Snyder 1993). Monoterpene levels in xylem are under strong genetic control. Therefore, Abert's squirrels may play an important role in shaping monoterpene variation in pines (Snyder 1998).

Gray squirrels, S. *carolinensis*, possibly use the high levels of tannins (6–10%) in acorns of red oaks (subgenus *Erythrobalanus*) as cue for burying them as winter cache. Acorns of white oak (subgenus *Lepidobalanus*) are low in tannins (0.5–2.5%) and germinate in autumn, making them unsuitable for caching. Concentrations of tannins and lipids covary in these acorns. Experimental doughballs, prepared from acorn flower, were avoided when tannin was added, but additional fat did not increase feeding unless tannin had been added. Addition of fat attenuated the negative effect of tannin. By avoiding perishable acorns and burying more durable ones, the squirrels optimize their food supply for the entire (winter) season, rather than for shorter periods. The tree may signal to small-mammal scatter hoarders by means of tannin, with the result of improved dispersal (Smallwood and Peters, 1986). In addition, the moist soil may enhance palatability by reducing the tannin level. When acorns of the red oak are abundant, gray squirrels eat only the basal part and discard the tip of the acorn, which contains the better defended radicle (Fig. 11.21)

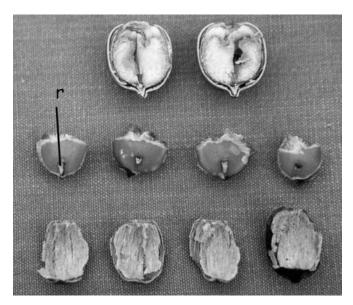


FIGURE 11.21 Gray squirrels have partly eaten acrons of red oak, *Quercus rubrum*. Top shows cross-section through untouched acorn. Two bottom show the rows remains of acorns after feeding. In the middle, there are discarded bottom parts of acorn, which is thought to have a high tannin concentration. The radicle (r) remains intact and such pieces often still germinate. (Photograph: D. Müller-Schwarze.)

Meadow voles, M. pennsylvanicus, and sheep eat reed canarygrass according to the alkaloid content of the particular sample (Kendall and Sherwood, 1975; Robinson, 1979). Meadow voles also consume the more alfalfa the less saponins it contains (Kendall and Leath, 1976). Beaver, Castor canadensis, feed on the usually avoided red and Scots pines only in late winter (D. Müller-Schwarze, personal observations). It is assumed that the ratio between carbohydrates and terpenes is optimal at that time.

Lagomorphs

Phenols deter Eurasian mountain hares, *Lepus timidus*, from debarking Scots pine, *Pinus sylvestris* (Rousi and Häggman, 1984). In Finland, these hares prefer some willow species (*Salix* sp.) over others in winter, and mature shoots to juvenile shoots. Feeding differences are correlated with phenolic glycoside (Fig. 11.22) content. In feeding experiments in the field, hares ate less oat grains treated with phenylglycoside or catechin (Tahvanainen *et al.*, 1985). The phenolic compound pinosylvin and its monomethyl ether in green alder are highly repellent to snowshoe hare, *L. americanus* (Bryant *et al.*, 1983).

$$CH_2O$$
 O-glucose O -glucose O -glucose

FIGURE 11.22 Phenyl glycosides.

Ruminants

Mule deer, *Odocoileus hemionus hemionus*, prefer juniper (*Juniperus* spp.) that contains lower amounts of volatile oils (i.e. oxygenated monoterpenes). Of three species tested, alligator juniper (*J. deppeana*) had the lowest level of volatile oils and was preferred, while Utah juniper (*J. osteosperma*) and Rocky Mountain juniper (*J. scopulorum*) ranked higher in volatile oil content and lower in preference by deer (Schwartz *et al.*, 1980).

Sagebrush, Artemisia sp., covers 422000 square miles in 11 western states of the USA and Canada. It is important as winter diet of mule deer, O. h. hemionus, pronghorn, Antilocapra americana, and sage grouse, Centrocercus urophasianus. Mule deer prefer sagebrush taxa with low levels of the oxygenated compounds methacrolein and arthole. However, only one fifth or less of utilization of eight taxa of sagebrush leaves is attributable to the levels of the volatiles methacrolein, α -pinene, camphene, β -pinene, arthole, 1,8-cineole, p-cymene, and camphor. In October, the value is 15.9%, in winter (March) 20.7%. Therefore, the volatile components may have their greatest effect during intense winter browsing but the sagebrush volatiles actually decrease during winter (Scholl et al., 1977). When chopped alfalfa hay was treated with various sagebrush terpenoids, methacrolein deterred mule deer most. It may guide preferences among different subspecies of big sagebrush, Artemisia tridentata, and p-cymene between big sagebrush and black sagebrush, Artemisia nova. The crude terpenoid fraction with its sesquiterpenes closely correlated with feeding in all four tested taxa of Artemisia (Bray et al., 1991).

Moose (*Alces alces*) in northern Europe discriminate between two birch species. They prefer the faster-growing *B. pendula* over *B. pubescens*, most likely because the former has less defense compounds (Danell *et al.*, 1985). This is consistent with the hypothesis of Bryant and Kuropat (1980) that faster growing tree species are less defended chemically. Phenols may defend birch in winter against vertebrate browsers by reducing digestibility and being toxic. Moose also feed less on Scots pine, *P. sylvestris*, if it contains higher levels of pinifolic acid, a diterpene (Danell *et al.*, 1990). Moose choose to utilize particular Scots pine trees

FIGURE 11.23 (1*S*,2*R*,4*S*,5*S*)-Angelicoidenol-20-glucopyranoside (also known as (-)-5-exohydroxyborneol.

according to phenol content, with the concentrations of most phenols internally correlated (Sunnerheim-Sjöberg and Hämäläinen, 1992). A glycoside of the terpene (1*S*,2*R*,4*S*,5*S*)-angelicoidenol, also known as (–)-5-exohydroxyborneol (Fig. 11.23), is a moose feeding deterrent found in Scots pine (Sunnerheim-Sjöberg, 1992). However, no correlation between moose browsing and the composition of ether extracts of Scots pine was found in several studies (Löyttiniemi and Hiltonen, 1978; Löyttiniemi, 1981; and Njemelä and Danell, 1988).

On the South African savanna, large generalist browsers among the ruminants feed on many species. The greater kudu, *T. strepsiceros*, was observed to feed on 60 species of plants in 12 hours. The kudu and two grazer–browsers, the impala (*Aepyceros melampus*) and the Boer goat rejected all plants with more than 5% of condensed tannins in their leaves during the wet season. Otherwise there was no correlation with plant contents (Cooper and Owen-Smith, 1985). The condensed tannins are attached to the plant's cell walls and are thought to protect against microbial and fungal attack (Zucker, 1983). Cooper and Owen-Smith (1985) postulated that the plant defense is really aimed at the microbes, originally in the environment and now in the rumen. Obviously, ungulates need the rumen flora for digestion but become "third parties to the chemical warfare between higher plants and pathogenic microorganisms." Domestic cattle also prefer food with less tannin (Marks *et al.*, 1988), and sheep avoid grazing on lupines with high alkaloid content (Arnold and Hill, 1972).

Proboscids

African elephants avoid plants with high contents of total phenols, steroidal saponins, and lignin (Jachmann, 1989).

Primates

Free-living primates respond to plant chemistry: black-and-white colobus monkeys, *Colobus polykomos*, avoid plant leaves with alkaloids, biflavonoids, and milky latex and prefer to eat leaves with a better ratio of nutrients to digestion inhibitors (McKey *et al.*, 1981). The howler monkey, *Alouatta palliata*, a generalist herbivore, depends on more than one factor in its choice

Table 11.6 Some plants and their secondary compounds that inhibit feeding in mammals

Mammal species	Plant species	Compound(s)
Meadow vole, Microtus	Penstemon digitalis	Alkaloids
pennsylvanicus, and Microtus	Barbarea vulgaris	Glucosinolates
ochrogaster	Pastinaca sativa	Furocoumarins
Meadow vole	White spruce Picea glauca, White pine Pinus strobus	β -Myrcene and bornyl acetate
Mountain hare Lepus timidus	Scotch pine Pinus sylvestris	Phenols
Snowshoe hare <i>Lepus</i> americanus	Green alder Alnus crispa	Pinosylvin, pinosylvin ether
Mule deer Odocoileus hemionus	Juniper <i>Juniperus</i> sp.	Oxygenated monoterpenes
Moose Alces alces	Scotch pine	Pinifolic acid (diterpene), exohydroxyborneol (terpene)
Greater kudu, Tragelaphus strepsiceros, and impala Aepyceros melampus	60 species	Condensed tannins
Sheep, meadow vole	Reed canary grass	Alkaloids
Meadow vole	Alfalfa	Saponins
Snowshoe hare	Labrador tea Ledum groenlandicum	Germacrone (sesquiterpene)
Opossum Didelphis sp.	Salix sp.	Salicin derivatives (phenolic glycosides)

of leaves. Young leaves of the monkeys' food plants had, on average, 11% more total phenolics than mature ones. The ratio of protein to fiber is most important, with secondary plant compounds indeed playing a "secondary" role in this case (Milton, 1979). Vervet monkeys, *Cercopithecus aethiops*, select plants (*Acacia tortilis* and *A. xanthophloea*) with low level of tannins for feeding (Wrangham and Waterman, 1981). Rhesus monkeys (*Macaca mulatta*) in Pakistan select food with low tannin content (Marks *et al.*, 1988). Table 11.6 summarizes some of these studies.

Response differences between mammal species

Some defenses are so effective that herbivores do not consume any amount of a plant. Snowshoe hares are deterred from feeding on Labrador tea, *Ledum groenlandicum*, by germacrone, a sesquiterpene, the major compound of its essential oil. Leaves and fresh growth internodes contain large amounts of germacrone (Reichardt *et al.*, 1990a). Yet beaver, *C. canadensis*, in Saskatchewan,

Canada, have been observed to feed on Labrador tea and store it in their food caches (Gunson, 1970). Storing in water may make Labrador tea more palatable.

Neighbor effect

Mammals often avoid palatable plants growing or placed next to effectively defended species. Western grey kangaroos, *M. fuliginosus*, usually browse *Hardenbergia comptoniana* (Fabaceae) heavily. But when grown next to a lesspalatable plant from the Myrtaceae family – characterized by essential oils, notably volatile monoterpenes – *Hardenbergia* sp. suffered less browsing by these animals. By comparison, a palatable non-Myrtaceae plant as "nurse plant" next to *H. comptoniana* resulted in heavy browsing of the latter (Jones *et al.*, 2003).

Similarly, Arctic ground squirrels, *Spermophilus parryi plesius*, browse plants with palatable neighbors more than those without (Frid and Turkington, 2001). Field experiments with plants or plant parts treated in different ways have to take into account these neighbor effects. While statistical designs call for random distribution of treatments in experimental plots, the behavior of animals may dictate otherwise. Since the taste or odor of one specimen can overshadow its neighbors, field bioassays work better when same-treatment specimens are clustered. Such "pseudobushes" were used in food choice experiments with beavers (Müller-Schwarze *et al.*, 2001) and field bioassays of plant extracts with Eastern cottontails (Müller-Schwarze and Giner, 2005). We can also recruit these neighbor effects for reducing herbivory of crops since treatment of only certain plants with repellent may also protect untreated neighbours.

Mushrooms

Mushroom poisoning in wild animals is little understood. Many mammals, including squirrels, other rodents, and deer, readily eat mushrooms, even toxic species. Gray squirrels, *S. carolinensis*, feed on *Amanita phalloides*, the death cap. A large portion of the diet of flying squirrels, *Glaucomys sabrinus*, and the California red-backed voles, *Clethrionomys occidentalis*, are hypogeous fungi (Fogel and Trappe, 1978). Red squirrels, *Tamiasciurus hudsonicus*, are famous for consuming toxic mushrooms, among them *Amanita* spp. In laboratory experiments, opossum, *D. virginiana*, readily ate the toxic fly agaric *A. muscaria*, which contains the hallucinogenic muscimol. Opossums vomited after ingesting muscimoltreated, otherwise palatable mushroom species such as *Calvatia gigantea* and *Panellus serotinus*. After one day of eating poisoned mushrooms, the opossum avoided the same and a second, non-poisoned, mushroom species for several days (Camazine, 1983).

Tracking changes in toxicity

A generalist herbivore can base the amount of food it eats on flavor concentrations, if these are correlated with toxicity. Sheep are able to adjust their food intake when the toxicity of food changes. Experimental animals identified plant toxins by associating food flavor added to oats, with post-ingestion consequences. First, if the concentration of a tastable toxin such as lithium chloride (LiCl) is increased, lambs decrease consumption; conversely, they increase consumption if the LiCl concentration is decreased. Second, lambs that had experienced two concentrations each of a bitter and a sweet compound in their barley chow, choose the *lower* concentration after receiving a low dose (125 mg/kg body weight) of LiCl. Finally, lambs were fed oregano-flavored ground barley. Following that, they received LiCl in gelatine capsules orally, in medium, high, or variable doses. The lambs with the medium dosage consumed the most barley, while those receiving high or variable dosages consumed less and by the same degree (Launchbaugh *et al.*, 1993). When toxicity cannot be detected by flavor, their intake depends on the maximum dose of toxin they had experienced.

Herbivores possibly regulate the amount eaten from toxic plants and thus avoid ill effects. Cattle cycle between consuming large and small amounts of tall larkspur, *D. barbeyi*. The principal toxic alkaloid in tall larkspur is methyllycaconitine (Pfister *et al.*, 1997).

Effects of nutritional status on consumption of toxic plants

Toxic plants may kill or harm herbivores if few other plants are available, because the animals are forced to be less discriminating, and because they are more susceptible to the toxins. In an experiment by Wang and Provenza (1996), three groups of lambs were maintained on three different levels of protein. In addition, they received every morning rations that varied in energy (barley) and a toxin (LiCl). The proportions of barley changed every 5 days, for five such periods. In the absence of the toxic LiCl, the lambs, consumption correlated with barley content. However, as LiCl concentrations increased, the lambs ate less of foods high in energy. Remarkably, with increasing food deprivation the lambs ate less and less of the LiCl-containing food, even if it had a high level of energy. This study shows how important interactions between nutritional status and toxicosis can be in herbivores.

11.4.2 When predators cannot cope with toxins of their animal prey

The bearded dragon, *Pogona vitticeps*, from Australia enjoys increasing popularity as a pet in the United States. The species comes from a geographical area lacking very potent chemical defenses in insects. Consequently, the bearded

dragon is vulnerable to toxins from prey in its new homeland, fireflies (*Photinus* sp.), particularly, can be deadly as they contain lucibufagin, a steroid resembling digitalis steroids and with the same effect on the heart. The reptile will eat fireflies, fails to neutralize the toxin, and dies as a result. Other pets such as African chameleons and lizards from Eastern Europe suffer similar poisoning by fireflies (Knight *et al.*, 1999).

Absence of plant secondary metabolite effects

In some studies, feeding intensity did not vary with the concentration of the monitored plant defense compounds. Mule deer, *O. hemionus*, use from 0 to 83% of the current year's growth of sagebrush, *Artemisia* sp. The monoterpenoid content varies from 0.75 to 3.62% dry matter. However, no clear correlation between these two has been established in a study of 21 accessions from five taxa (Welch *et al.*, 1983). Penned black-tailed deer, *O. h. columbianus*, consumed Douglas fir of different genetic origins regardless of essential oil contents, even though their preference was correlated with the chlorogenic acid content (Radwan and Crouch, 1978). Similarly, pygmy rabbits, *Brachylagus idahoensis*, did not feed differentially on 15 populations from two subspecies of sagebrush, *A. tridentata* (White *et al.*, 1982). Nevertheless, Welch and McArthur (1981) suggested that sagebrush could be selectively bred for reduced monoterpenes to allow better utilization by mule deer.

In Sweden, phenols did not protect introduced lodgepole pines, *Pinus contorta*, particularly well against debarking by the vole *M. agrestis*. Being an alien tree species may be an important factor (Hansson *et al.*, 1986). The authors pointed out that, in general, any natural defense will work only at low browsing pressure by voles.

11.4.3 Coping: how herbivores overcome plant defenses

They put arsenic in his meat
And stared aghast to watch him eat;
They poured strychnine in his cup
And shook to see him drink it up:
They shook, they stared as white's their shirt:
Them it was their poison hurt.
—I tell the tale that I heard told.
Mithridates, he died old. A. E. HOUSMAN

Herbivores have evolved many offensive adaptations that counteract the defensive adaptations of plants, including antifeeding compounds (Rhoades, 1985).

Herbivores deal with potentially toxic plant secondary metabolites in three different ways: they can avoid particular plants altogether; process the food to improve its palatability; or ingest it and deal with the plant secondary metabolites by biotransforming them. This is where King Mithridates of Pontus (today's Turkey) comes in. Fearing poisoning by his enemies, he ate toxins in increasing amounts and built up his resistance. He survived an assassination attempt and has given his name to a procedure to build up immunity to toxins: "to mithridate."

Food avoidance

Preferences and avoidance such as seeking sweet foods and rejecting bitter-tasting plants can be acquired in three ways. Avoidance acquired in evolutionary time may not require individual learning and is termed *primary aversion*. It occurs to unpleasant taste or odor. A conditioned aversion can be acquired by an individual after a negative experience such as illness by food poisoning. Finally, an individual can learn from relatives or group members to avoid a food, without ever trying out the food itself.

Primary aversion linked to taste

Alkaloids and cyanogenic glycosides taste bitter to humans and have been thought to be universally repellent to higher animals (Bate-Smith, 1972). However, guinea pigs, *Cavia porcellus*, tested with nine bitter substances, reduced their feeding to only two (quinine and sucrose octaacetate), and that only slightly (Nolte *et al.*, 1994a).

Primary avoidance of toxic mushrooms by the fungivorous oppossum, *D. virginiana*, is a good example. Many mushrooms have toxins, including emetics, cathartics, hallucinogens, and liver poisons. The pungent-tasting mushrooms of the family Russulaceae contain *isovelleral*, a sesquiterpenoid dialdehyde. Opossums feed little or not at all on isovelleral-containing species. If isovelleral is added to morsels of the palatable species *Agaricus*, consumption declines by 67 to 22%, depending on concentration. The opossums rapidly eject the morsel, salivate profusely, froth at the mouth, and clean their muzzle. Fatty acids may enhance the peppery sensation of isovelleral and may help to solubilize isovelleral in the saliva (Camazine *et al.*, 1983). However, red squirrels actually prefer peppery mushrooms such as *Russula emetica* or *Lactarius* sp. Similarly, the opossums ate the peppery *Boletus piperatus*. Preference for pepper taste by humans and other mammals may moderately stimulate the induction of detoxication mechanisms to keep them ready for dietary contingencies.

Primary aversion linked to odor

The avoidance of the odor signal may be primary. An odor can be a negative signal acquired during evolution, with no conditioning necessary. However, even anosmic animals can show food aversions. Therefore, taste alone can be sufficient for the aversion. So far, there is no evidence for an intrinsic repellent effect of odor alone. In all studied cases, odor had to be associated with ill effects, including nociception (damage to receptors) or toxicosis (gastrointestinal malaise) (e.g. Provenza *et al.*, 2000).

In *neophobia*, defined as "an individual's avoidance of a novel object" (Barrows, 1995), an animal may totally reject a new food, or take minute amounts to await the consequence of this ingestion. Neophobia, therefore, forms the transition between primary and conditioned aversions.

Conditioned Taste Aversion

Conditioned taste aversions occur after food produced an illness. Both the chemical senses and gastrointestinal feedback are involved.

Postingestive consequences of eating toxic plants may lead to an aversion. For instance, tall larkspur (*Delphinium* sp.) is toxic to cattle, even though it is palatable. Cattle that had been fed larkspur consumed less in subsequent feeding trials. However, animals with esophageal fistulae that had tasted larkspur but not experienced it in their rumen did not reduce their consumption. This shows that chemosensory input alone is not sufficient for producing an aversion (Pfister *et al.*, 1990a). Goats, however, were shown to discriminate between two novel foods with different postingestive consequences. This discrimination depended on both the relative amounts of the two foods eaten and the salience of the flavors of these foods (Provenza, and Lynch, 1994).

An intriguing question is how many plant odors and tastes a herbivorous mammal is able to memorize at any one time, and whether it increases feeding efficiency by grouping several plant species under one odor or taste. In one experiment, lambs were given a meal of five foods, one of which was novel. After 1 to 6 hours, they experienced toxicosis. Subsequently, they decreased their intake of the novel but not the familiar food (Provenza, 1995).

Conditioned aversion to taste plus odor

In a two-step response, an additional, volatile (olfactory) stimulus may signal over a distance that the plant is unpalatable and/or toxic.

Olfactory aposematism (Eisner and Grant, 1981) means associating an odor by conditioning (experience) with an odorless toxin such as nicotine, morphine, or strychnine. This is probably widespread among mammals. First the animal tastes the plant and finds it either unpalatable or suffers ill effects. After that,

a different, more volatile compound becomes the signal for immediate avoidance. Opossums learn to associate the typical mushroom odor of the innocuous volatile 1-octen-3-ol with delayed illness from toxic mushrooms (or edible mushrooms experimentally poisoned with the odor- and tasteless toxin muscimol) (Camazine, 1985). Plant odors deter lambs from feeding only when associated with, or followed by, gastrointestinal malaise induced by LiCl. A novel plant odor, such as that of *Astralagus bisulcatus*, an unpalatable, toxic, and sulfursmelling herb of the North American west, reduces feeding by lambs briefly, but for more than 5 days only if the toxicosis continues (Provenza *et al.*, 2000).

Two speculative possibilities relate to olfactory aposematism. The first is whether there are non-toxic plants that smell or taste like toxic ones. In other words, do plants practice Batesian mimicry? Such mimicry is unlikely, as mammalian herbivores constantly sample plants and thereby test for favorable and adverse effects of eating a particular species. Furthermore, given the keen sense of smell of mammals, two plant species would have to exactly smell alike for mimicry to work. Second, do two distasteful or toxic plant species smell or taste alike so that herbivores can more easily classify dangerous plants and avoid them (Müllerian mimicry) (Eisner and Grant, 1981; Lindroth, 1988; Augner and Bernays, 1998).

Potentiation describes the additive effects of taste and odor that allow a conditioned responce to both that would not occur to either alone. Odor is not necessarily an effective conditioned stimulus, and it has been said that "the nose learns from the mouth". After illness from a poisoned meal, taste is a more potent conditioned stimulus than odor, but taste can potentiate odor. An animal will first smell, then lick a food, and only then reject it. During foraging, odor is perceived before taste. The opposite sequence is true in the intrinsic equilibrium responses of the body: a taste experience leads to an odor becoming a signal (Garcia and Rusiniak, 1980). Learning is extremely quick and lasting: one exposure to a toxic food (single-trial learning) often suffices, and the effect may be lifelong (long-delay learning).

Several sensory modalities can be involved in complex interactions involving at least three cues: rats learnt better to associate the bitter taste of quinine in water with a "context" such as a black or white box if a pyrazine was also present (the specific compound used was 2-methoxy-3-isobutyl pyrazine) (Kaye et al., 1989). It is said the odor "potentiates" learning the connection between taste cue and "context."

A case of taste potentiating a visual cue is that of hawks feeding on mice. If the hawks were routinely fed white mice, and only occasionally a black mouse followed by LiCl injection, the hawks would not eat either black or white mice. However, when a distinctive taste was added to the black mouse, hawks learnt to avoid black mice on sight after experiencing only a single meal of a black mouse that resulted in toxicosis (Brett *et al.*, 1976).

Observational learning

Young animals can learn food choices by observing a parent or other group member without ever eating the plant itself. Many young mammals, such as deer fawns (D. Müller-Schwarze, personal observations), forage close to experienced conspecifics – usually the mother – for long periods of time so they have ample opportunity to learn food preferences. Lambs with their mothers consume more palatable food and become poisoned less often than lambs without mothers (Provenza, 1995).

Stephen's woodrat, *Neotoma stephensi*, specializes on juniper (*Juniperus* sp.) foliage that contains high levels of tannins and terpenoids. Compared with other woodrat species, the young are weaned late, grow slowly, and associate long and closely with their mother. Mothers and young are "sitting nose to nose and eating the same food item" (Vaughan and Czaplewski, 1985). The authors have suggested that during the long preweaning period the young learn to select leaves from particular trees that are low in defensive compounds.

Processing food before ingestion

Some animals "process" plants to reduce defense compounds even before eating. The meadow vole *M. pennsylvanicus* cuts winter branches of white spruce, Norway spruce, white pine, and Norway pine and leaves them on the snow for 2–3 days before eating them. This reduces the levels of condensed tannins and other phenolics by one half, to their summer levels. A high level of protein (12%) and reduced phenolics (1.5% of dry matter) now render the food acceptable. It is not clear how the phenolics are being lost, possibly by polymerization or oxidation (Roy and Bergeron, 1990b).

Pikas (Ochotona princeps) of the North American west also practice food conditioning. They store winter food in "hay piles." These food caches consist primarily of Alpine avens, Acomastylis (Geum) rossii, which the pikas do not eat during the summer. The Alpine avens contain three to six times more phenolics than the summer diet. These compounds preserve the food. After weeks or months of storage from October to January, the toxins in the cached plants decrease to summer diet levels. As the stored plants lose phenolics, the pikas consume increasingly larger amounts. In short, the pikas practice food conditioning in addition to using natural food preservatives (Dearing, 1997).

Animals that bury acorns and other seeds as winter cache may also practice food processing, tannins being lost during the weeks in the moist soil. This



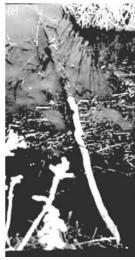




FIGURE 11.24 Leaching: (a): Standard pieces of different trees leach out different amounts of phenolics and other compounds. During 24 hours, red maple and witch hazel (left) stain water a darker brown than willow and quaking aspen (right). (b) and (c): Beavers leave pieces of branches in the water for 1–3 days with some or all bark intact before eating the bark. This presumably leaches out water-soluble plant secondary compounds.

possibly applies to gray squirrels (Smallwood and Peters, 1986) and blue jays (Dixon et al., 1997).

Beavers, *C. canadensis* store branches for weeks and months in the water as winter food caches. They also soak logs and sticks of less-palatable trees in their pond, often for 2–3 days, before they eat them (Fig. 11.24). Field and laboratory experiments suggest that in this way undesirable compounds leach out from the bark (Müller-Schwarze *et al.*, 2001).





FIGURE 11.25 Leaching by humans. (a) A cassava plant. (b) Ground manioc (cassava) is mixed with water and pressed through tube woven from palm fibers to remove toxic cyanogenic compounds. Segments are collected in a tub. The liquid with toxins is in a pot in the foreground. (Photograph: D. Müller-Schwarze; Paraitepuy Village, Gran Sabana, Venezuela, 2000.)

Humans obviously have devised effective methods of leaching toxins from staples such as bitter manioc, also known as cassava or yucca, Manihot esculenta (Euphorbiaceae) (Fig. 11.25). Manoic is native to tropical South America and was introduced to Africa by the Portuguese. The cyanogenic compounds discourage herbivores and protect the crop against microorganisms. Cooking destroys enzymes, but not linamarin, and manioc remains toxic. Therefore, the bitter manioc must be detoxified by first shredding or grating it to bring enzyme and substrate into contact, and then removing the generated hydrogen cyanide as gas or dissolved in water. Many Amazonian societies press a mixture of water and grated manioc through a hose of woven plant fiber (Fig. 11.25b) and discard the expressed whitish, toxic liquid. A woman needs 4 to 6 hours daily or every other day for this time-consuming labor (McKey and Beckerman, 1993). Even so, improper processing results in sometimes tragic poisonings. On March 9, 2005, schoolchildren in a village on Bohol Island in the Phillipines were poisoned by deep-fried caramelized cassava and 27 of the children died immediately (USA Today March 10, 2005, p. 13A). As sweeter manioc varieties are being

introduced, loss to pests will most likely increase. Anecdotal evidence suggests that pigs, baboons, and rodents attack sweet manioc more than bitter varieties. Dried chips of bitter manioc roots store better than those made from sweet varieties (in McKey and Beckerman 1993). Bitter varieties are also said to be more productive. People living along rivers rich in fish use bitter varieties of manioc. One reason may be that fish provides sulfur-containing amino acids, needed for detoxifying hydrogen cyanide with thiosulfate. More nomadic peoples, by comparison, grow sweet manioc and hunt agoutis and other bothersome rodents in their gardens. In general, sweet manioc is grown as a vegetable, while groups who produce manioc flour select for more bitter varieties (McKey and Beckerman 1993).

Biotransformation (detoxication)

Herbivores biotransforms xenobiotics from natural or artificial sources to render them harmless. Disarming potentially toxic compounds is termed detoxication, while the term detoxification refers to correcting a state of toxicosis (Johns, 1990). Herbivores degrade secondary plant compounds in a variety of ways, starting in the mouth, and leading to excretion.

Birds

The diet of some birds contains high levels of monoterpenoids. Sage grouse feed on sagebrush, *A. tridentata*, which contains the monoterpenoids α -pinene, camphene, 1,8-cineole, β -thujone, camphor, and α -terpineol. These compounds constitute 3.23% of the dry matter in *Artemisia* leaves. During passage from the beak to the gizzard, the levels of these compounds are reduced to 64% of that in the leaves, but this is still high (Welch *et al.*, 1989). Further reduction of monoterpenes takes place in the gizzard where the leaves are ground into fragments. Normal body heat volatilizes monoterpenoids. Experiments showed α -pinene to be very volatile under these conditions while camphor is not. Drastic reduction of monoterpenoids occurs between gizzard and duodenum. No monoterpenoids were found in the ceca; therefore the ceca microbes are not endangered. In summary, the bird's body is exposed to far less monoterpenes than the plant content would suggest (Welch *et al.*, 1989).

Detoxification of plant secondary compounds has been suggested as one of the nutritional benefits responsible for the evolution of the unique foregut microbial fermentation system in the hoatzin, *Opisthocomus hoazin*, a South American obligate leaf-eating bird (Fig. 11.26). In this ruminant-like digestive system, crop and lower esophagus produce volatile fatty acids whose concentrations and ratios resemble those of the sheep rumen (Grajal *et al.*, 1989; Grajal, 1995).



FIGURE 11.26 The hoatzin is a leaf-eater that is known in Peru as "chancho" (pig) because of its smell. This bird has few predators, presumably because it sequesters plant compounds that render it unpalatable. (Photograph: D. Müller-Schwarze, Rio Napo, Ecuador, 2003.)

Mammals

Mammals neutralize toxic plant compounds at all stages of ingestion and digestion, aided by both microbes and specialized enzymes. Smaller species can detoxify xenobiotics faster because of their high metabolic rate. Therefore, they are more likely to evolve specialized food habits (Freeland, 1991). As in birds, the fight against ingested plant compounds starts in the mouth. Monoterpenes such as α -pinene are very volatile at body temperature and so their concentration can be reduced by mastication, rumination, and eructation, in addition to the traditionally considered absorption and excretion (Welch *et al.*, 1989).

Browsers such as mule deer, *O. hemionus*, have *proline-rich salivary proteins* that bind tannins; this enables them to feed on tanniniferous plants (Robbins *et al.*, 1991). By contrast, grazers such as sheep and cattle do not possess tannin-binding proteins (Austin *et al.*, 1989; Robbins *et al.*, 1991), and neither do mixed feeders such as goats (Distel and Provenza, 1991). Rat and mouse saliva also contain proline-rich proteins that bind tannins. In human saliva, about 70% of total protein is rich in proline (Fig. 11.9) (Mehansho *et al.*, 1987).

Proline-rich proteins have an affinity for tannins at least three orders of magnitude greater than other proteins, protecting dietary proteins and digestive enzymes (McArthur *et al.*, 1995). The salivary proteins form hydrogen bonds with tannins. They also increase the accessability of the peptide backbone for hydrogen bonding. The interaction is very specific for the protein but not for the tannin. Condensed tannins are slightly more effective as protein precipitants than hydrolyzable tannins. The parotid glands of rats can be induced to produce more proline-rich proteins in response to a tannin-rich sorghum diet. This requires about 3 days (Mehansho *et al.*, 1983). However, meadow voles, *M. pennsylvanicus*, do not appear to produce tannin-binding proteins in response to dietary quebracho tannin (Dietz *et al.*, 1994).

Various mammal species differ in their processing of tannins. Laboratory rats are more resistant than hamsters or prairie voles, and goats more than sheep or cattle. In one experiment, tannin from the quebracho tree (*Schinopsis* sp.) was fed to mule deer, domestic sheep, and black bear (*Ursus americanus*). Mule deer, black bear and laboratory rats have salivary tannin-binding proteins that are very effective. These species excrete up to 98.3% of the tannin in their feces, while in sheep 25% are not found in the feces and so are presumed metabolized with all the physiological consequences. Salivary proteins minimize fecal nitrogen loss by maximizing amounts of tannin bound per unit of protein, minimize tannin absorption and toxic effects by forming stable tannin-protein complexes, and prevent or minimize reduction in fiber digestion (Robbins *et al.*, 1991). High levels of protein in the diet also protect against ill effects of tannins as binding of tannins to protein protects the esophagus (Warner and Azen, 1988).

The good earth: geophagy

Ingestion of mineral substances, termed geophagy, occurs in a wide range of herbivorous birds and mammals. Parrots, cracids, pigeons, doves, deer, elk, elephants, giraffes, zebra, sheep, bears, raccoons, monkeys, tapirs, and peccaris regularly eat soil. Geophagy is still little understood. In addition to uptake of minerals or microorganisms, clay can be helpful in detoxification of secondary plant compounds.

Neotropic parrots make daily morning trips to riverside cliffs to eat clay. The clay-licks of Manu National Park in Peru are especially well known (Fig. 11.27). Macaws and parrots visit the clay-lick more often in the dry season when fresh fruit are rare and they have to rely more on seeds with toxic compounds (Brightsmith, 2002). The value of soil was demonstrated using extracts from a variety of ripe and unripe toxic Peruvian seeds that made up the macaw diet. These extracts were less toxic to brine shrimp after mixing them with soil from



FIGURE 11.27 Use of a day-lick by birds. Top shows mealy parrots. Bottom shows macaws in Manu National Park, Peru. (Photograph: D. Müller-Schwarze, 2004.)

Manu. In another experiment, two groups of captive orange-winged macaws, *Amazona amazonica*, were fed quinidine sulfate with or without clay, respectively. The birds treated with clay had less quinidine circulating in their blood (Gilardi *et al.*, 1999). The layers of clay preferred by the macaws contain high levels of sodium; consequently, the birds may enjoy both detoxication and mineral supplements. Macaws even pass on clay to their chicks when feeding them (Brightsmith, 2002).

Chimpanzees daily eat termite mound clay. The habit may have started by ingesting dirt along with termites. Rats, after rotation that causes motion sickness (stomach upset and diarrhea), eat more clay than normal (Mitchell *et al.*, 1977). Geophagy may be instrumental in the success of rats in habitats with unpredictable diets and the resistance of the species to poisoning by humans. Clay can be eaten prophylactically but also in response to toxic food already in the gastrointestinal tract.

Red-handed howler monkeys, *Alouatta belzebul*, studied in eastern Brazilian Amazonia, ate mostly fruits at the peak of the wet season and switched to leaves during the transition from wet to dry season. The percentages of fruits and leaves, respectively, were 53.5% and 40.8% in the wet season and 18.9 and 77.9% in the dry period. When the animals changed their diet, the leaves were mature and relatively rich in secondary compounds. The monkeys then ate soil from termitaria on 26 occasions, all during the drier season. Such soil was richer in calcium, sodium, and organic carbon than soil from the forest floor. The soil probably adsorbed plant secondary metabolites from the leaves in the digestive tract and helped to excrete these rapidly (De Souza *et al.*, 2002).

Humans are also geophageous. Clay or sand is eaten in many countries (Fig. 11.28) and is prevalent in Asia, the Middle East, Africa, Latin America, and the rural south of the United States. Aristotle described this habit, often practiced by pregnant women. The indigenous people of Mexico and Peru eat clay together with frost-resistant potatoes. The Indians in the southwestern United States used to eat clay with wild tubers after crops had failed. Markets in Guatemala sell clay tablets, known as *esquipolas*, particularly to pregnant women (Fig. 11.28). Hunter–gatherers eat clay and acorns together. Eating dirt during famines in China and medieval Europe may have been connected with the eating of toxic wild tubers. Historically, emperors, kings, and popes have eaten clay as precaution against conspiratorial poisoning.

In the Andes, nearly all of the about 160 varieties of wild potatoes, and two of the eight cultivated species are toxic. Some frost-resistant species that grow above 3600 m have high levels of alkaloids, which are bitter and potentially toxic. These are hybrids between *Solanum stenotonum* and wild potatoes such as *Solanum megistacrolobum* and *Solanum acaule*. Other secondary plant compounds in potatoes are saponins, phytohemagglutinin, proteinase inhibitors, sesquiterpene phytoalexins, and phenols.

The Indians of the Peruvian Altiplano eat potatoes with a dip of clay and a mustard-like herb. They say the clay removes the bitter taste and prevents stomach pains or vomiting after eating large amounts of potatoes. The people who eat clay intend detoxication. This may explain how Indians started to utilize and domesticate wild potatoes. Indeed, experiments have shown that four different types of edible clay adsorb the glycoalkaloid tomatine under simulated



FIGURE 11.28 Commercial products for geophagy in humans. Right, Heilerde (literally healing earth) sold in German health food stores. Left, clay tablets (esquipolas or pan de señor) eaten by pregnant women against "morning sickness," purchased by the author in markets in Antigua and Chichicastenango, Guatemala.

gastrointestinal conditions (Johns, 1986). Clay also effectively binds the potato's bitter and toxic glycoalkaloid solanine. The amounts of alkaloids in potatoes can be staggering: 100 g fresh wild potatoes typically contain 100 mg tomatine, five times the safe level of 20 mg/100 g. A glycolalkaloid content above 14 mg/100 g fresh weight is distasteful to humans. Cultivated potatoes contain less than 10 mg/100 g. To effectively reduce the level from 100 to 20 mg/100 g potato, only 50–60 mg clay are needed, a miniscule amount compared with what Peruvians actually eat per meal. For detoxication, adsorption to clay is most important in the stomach. In the higher pH of the intestine, less glycoalkaloids are adsorbed as they are less soluble there and, therefore, not taken up (Johns, 1990).

Clay eating probably was extremely important in human evolution: it enabled hominids who did not use fire to eat plants with toxic antifeedants. However, the glycoalkaloids of the potato are heat stable and insoluble in water. Domestication of tomatoes and potatoes probably went hand in hand with clay eating. Johns (1986) suggested that "geophagy is the most basic human detoxification technique with behavior antecedents that are prehominid."

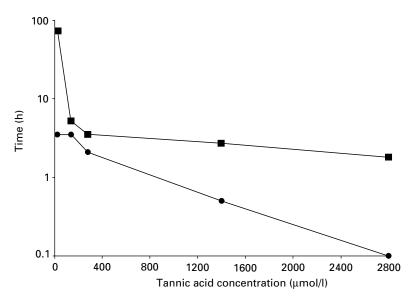


FIGURE 11.29 The effect of tannic acid on time to distress (•) and time to death (■) in fish. (Data taken from Swain, 1979).

Clay may even benefit fish. Swain (1979) found that raising the levels of tannic acid in the water caused increasing discomfort and death to guppies (Fig. 11.29). The addition of clay to the water ($52 \, \text{mg/ml}$ water for a tannic acid concentration of 35 $\, \mu \text{mol/l}$) extended the period the guppies survived over six times. Suspended clay particles in "white rivers" may adsorb phenolics and protect fish that cannot easily live in phenolic-rich "black rivers" (Swain, 1979).

Biotransformation

The continuous shedding of surface cells of the digestive tract protects the digestive and absorptive tissues from toxic effects. Once absorbed by the digestive tract, secondary plant compounds of low molecular weight can be excreted via urine, while larger molecules such as monoterpenoids and sesquiterpenoids are excreted via bile and feces. Perspiration, vomitus, milk, hair, exhaled air, and saliva provide other avenues for excretion. For easy excretion, the compounds have to be *water soluble*, contain *polar groups* and be *ionic* at physiological pH. Highly *lipophilic* compounds move across membranes and accumulate in tissues, exerting their toxic effects. Although they have also polar groups, the non-polar effects prevail, and they are not ionic at physiological pH levels. Therefore, the main task of biotransformation is to render the originally lipophilic, hence membrane-threatening, xenobiotics water soluble for excretion.

FIGURE 11.30 The two phases of biotransformation of xenobiotics. In this example, an amino acid is used for conjugation. A sugar (glucuronyl) or sulfate can also be used.

Metabolism of ingested plant compounds occurs by two kinds of reactions. *Phase I* reactions are catabolic (enzymatic oxidation, reduction, or hydrolysis) and the products are often more chemically reactive and more polar, making them paradoxically sometimes more toxic. *Phase II* reactions are anabolic (synthetic) and involve conjugation, which usually results in inactive products that can be excreted (Fig. 11.30). Functional groups introduced or uncovered in phase I provide reactive sites for conjugation reactions in phase II. Particularly, phase II produces hydrophilic molecules, making the xenobiotics water soluble. The principal enzymes for metabolizing plant secondary metabolites are located in the liver and gut walls, but also in kidneys, intestines and lungs.

Phase I reactions

Phase I renders xenobiotics more polar. *Oxidation* is the most important process of phase I. It is carried out in the endoplasmic reticulum in many tissues by monooxygenases that contain cytochrome P450 (P450-dependent mixed function oxidases) as electron carrier. These enzymes have evolved in the past billion years in response to plant secondary metabolites. There are a number of P450 gene families.

Alcohols are oxidized to aldehydes by the liver enzyme alcohol dehydrogenase, and aldehydes to carboxylic acids by aldehyde dehydrogenase. In mammals, monooxygenases can be induced by plant secondary metabolites such as α -pinene, caffeine, or isobornyl acetate. *Reduction* is less common and plays a role with ketones that cannot be further oxidized. *Hydrolysis*, the degradation of a compound with addition of water, is also less common than oxidation.

Phase II reactions

Conjugation during Phase II is a synthetic reaction such as combining with carbohydrates (Fig. 11.30) or amino acids, resulting in metabolites of reduced biological activity. Conjugation also produces a highly hydrophilic (water-soluble), less toxic, and more readily excretable ionized product (Freeland and Janzen, 1974). The most common conjugate is glucuronic acid. An important enzyme for conjugation is the UDP-dependent glucuronosyltransferase, responsible for glucuronide formation. Other conjugates are glycosides, sulfates, methyl groups (methylation), amides, glutathione, acetyl groups, and thiocyanates. Species can differ in this regard. For instance, common brushtail possums, T. vulpecula, conjugate phenol with glucuronic acid, but carnivorous marsupials use sulfate (Baudinette et al., 1980).

Some mammals depend more on oxygenation (phase I), and others more on conjugation. The common ring-tailed possum, *Pseudocheirus peregrinus*, polyoxygenates terpenes from its eucalyptus diet into highly polar, acidic metabolites that are easily excreted. These metabolites include dicarboxylic acids, hydroxyacids, and lactones. Very little conjugation takes place, possibly to conserve carbohydrate and amino acids (McLean *et al.*, 1993). Generalists appear to conjugate, while specialists oxygenate. Among the generalists, the brushtail possum, *T. vulpecula*, and the laboratory rat excreted more conjugates and less oxygenated metabolites of *p*-cymene, a constituent of eucalyptus leaves. Eucalyptus specialists such as the greater glider, *Petauroides volans*, and the ringtail possum *P. peregrinus*, by comparison, only excreted metabolites with three or four oxygen atoms (Boyle, 1999). These polyoxygenated compounds are more polar and soluble and, therefore, excreted more easily in urine. In this way, the specialists preserve valuable carbohydrates and/or amino acids, which is less critical in the generalist species with their varied diet.

The nutritional value of the diet influences the ability of herbivores to detoxify plant secondary metabolites (Schwass and Finley, 1985). For instance, voles in winter have to consume the bark of birch (*Betula* sp.). This causes stress and leads to poor growth and high mortality among young animals. Birch bark contains phenolics and terpenoids. Both are metabolized by cytochrome P450 monooxidases in phase I and conjugated with glucuronic acid in phase II.

Induced enzymes

In root voles (*Microtus oeconomus*) hepatic phase I detoxication by monooxygenases, specifically ethoxyresorufin-O-dealkylase, can be induced by large doses of birch bark powder in the diet. Conjugation (phase II) is not affected (Harju, 1996).

Degradation speed

The degradation speed differs greatly between species. For instance, the half-life of lysergic acid diethylamide (LSD) is 7 minutes in the mouse, 100 minutes in *Macaca*, 130 minutes in the cat, and 175 minutes in humans. To use another example, ethanol is removed from the blood at a rate of 7 g/h on average, depending on body weight and gender. A quart of beer or 4 oz of whiskey is biotransformed in 5–6 hours. This means that more than 1 gallon of beer or 16 oz of whiskey per day exceeds the enzymatic capacity of the liver. Alcohol would stay in the blood all the time, with destructive consequences.

Acidemia

Many plant secondary metabolites require conjugation, often with glucuronic acid. The conjugation products are strongly acidic and so biotransformation increases the acid load. Accordingly, acidemia is a major consequence of eating diets rich in plant secondary metabolites (Foley et al., 1995). This acid load must be buffered and excreted. An adequate acid balance can be maintained by providing bicarbonate to neutralize excess protons since, at physiological pH, acid metabolites will be almost completely ionized. The hydrogen ion and corresponding anion may be disposed of in different ways. The hydrogen ion can be neutralized in the body by bicarbonate, buffered and retained, or it can be excreted in urine as titratable acid. The anion typically is excreted in urine as ammonium or sodium salts. The bicarbonate needed for buffering the acid load is generated from α -ketoglutarate, which in turn, comes from amino acids (Foley et al., 1995). This requires catabolism of amino acids and results in loss of body protein and depletion of glucose (Illius and Jessop, 1995). To cope with a continuing acid load, the mammal eventually has to break down skeletal muscle to provide glutamine for ammoniogenesis as well. The ammonium ions are needed for excretion of anions as ammonium salts. Any excess ammonium ions are excreted in the urine or combined with bicarbonate to produce urea in the liver (Foley et al., 1995).

Sodium loss

Rabbits form bicarbonate in the gut and absorb it. They do not have to form new bicarbonate in the kidneys and need not excrete ammonium ions in the urine, but they still need to excrete organic anions. These organic anions are accompanied in the urine by sodium or potassium ions, which can generate a severe negative sodium balance for the period that the rabbits are on a browse diet (Iason and Palo, 1991). Therefore, lagomorphs excrete biotransformational

organic anions as sodium salts, while most other mammals excrete them as ammonium salts.

Differences in digestive ability can reflect different "ecological lifestyles" of related species. The mountain hare feeds on birch in winter and grass in summer. When fed birch that is high in phenolics, the hare suffers no sodium loss but detoxifies instead. The European hare does not eat birch in winter but eats grass year round. When forced to eat a high-phenolic diet, it loses much sodium through its urine (Iason and Palo, 1991).

Biotransformation by gut flora

The gastrointestinal flora may modify foreign compounds and either render them toxic to their host or assist in detoxication. An example of the former is *cycasin*, a compound in food made from cycad nuts that by itself is not toxic to rats. Hydrolysis by the rat's microflora generates metabolites, one of which is toxic. Germ-free rats suffer no toxic effects from ingested cycasin.

In ruminants, the microbes of the rumen anaerobically detoxify phytochemicals by hydrolysis, reduction, degradation (dealkylation, deamination, decarboxylation, dehalogenation, and ring fission) (Allison, 1978). It has even been suggested that detoxification may be a more important function of the rumen than unlocking nutrients (Morris and García-Rivera, 1955). Ruminants are particularly vulnerable to toxic compounds in their diet because the digestive action by microbes occurs anteriorly in the digestive tract, before many of the compounds that may harm the microbes can be removed. In hindgut fermenters, the essential oils (mono- and sesquiterpenes) may be absorbed and detoxified before they reach the site of microbial activity in the digestive tract, protecting the herbivores microorganisms from ill effects.

11.5 Plant responses to herbivory

11.5.1 Induced defense

Plants fight back against herbivores in a number of ways. Snowshoe hares, *L. americanus*, browse northern deciduous trees or shrubs often severely during peaks of their 10-year population cycle. After browsing, paper birch, *B. papyrifera*, quaking aspen, *P. tremuloides*, balsam poplar, *P. balsamifera*, and green alder, *A. crispa*, grow adventitious shoots. These new shoots contain high levels of terpene and phenolic resins and are very unpalatable to the hares. They are not eaten, even though they are rich in nitrogen and phosphorus. This new

generation of unpalatable adventitious shoots may play an important role in the 10-year cycle of the hare populations (Bryant, 1981).

There is some evidence for induction in pasture plants. A furanocoumarin, isopimpinellin, increases in floral stands of wild parsnip, *Pastinaca sativa*, after clipping that simulates herbivory (Nitao, 1988).

In some cases, browsing does not appear to lead to induced defenses. Two species of northern European birch (*B. pendula* and *B. pubescens*) do not seem to respond to moose browsing with increased chemical defenses (Danell *et al.*, 1985). After experimental defoliation, the Emory oak of southwestern North America, *Quercus emoryi*, produces in its regrown leaves 2.5 times as much hydrolyzable tannins than in mature leaves, but lower amounts of condensed tannins. Refoliated branches suffered more, not less herbivory. New leaves have more protein, more water, and are tender. These properties may override any effects of induced chemical defenses, at least for the current growing season (Faeth, 1992), but the induced compounds may still protect a plant later on, especially in winter.

The evidence for induced defenses has grown more complex. For example, *Acacia drepanolobium* in Kenya responded to herbivory by large mammals in unexpected ways. Trees in an area with antelope only were compared with trees exposed to antelope plus elephants and giraffe, and an exclosure without any of these herbivores. The tannin concentration in the leaves increased with increasing pressure by herbivores. However, the trees were richer in tannin in the upper levels (2 m high) than at about 1 m, where more browsing takes place. Also, unexpectedly, the more tannin a tree had, the longer its spines. There was no evidence of a trade-off between chemical and thorn defense. Nor was there a trade-off between chemical defense and amount of growth. Furthermore, different species of ants, some more aggressive than others, may have an effect on the herbivores, although no clear effect of the ants was detected (Ward and Young, 2002).

11.5.2 Chemical defense versus growth rate of plants

A plant, especially a tree or shrub, can escape extensive browsing by either defending itself, chemically or otherwise, or by growing so fast that it more than replaces any biomass losses to herbivores. It may also grow rapidly beyond the reach of its herbivores. Slow-growing boreal and arctic trees are more likely to defend themselves chemically, while this would be less essential for rapidly growing tropical trees. Slowly growing plants on poor soil are thought to invest more into plant defense compounds (Coley *et al.*, 1985). One hypothesis is that fast growing trees are more palatable (Bryant and Kuropat, 1980). In the same boreal forest, two related species may differ along this scale: in Sweden,

moose, *A. alces*, prefer the faster-growing birch *B. pendula* over *B. pubescens*. After moderate browsing, *B. pendula* regrows more and larger leaves and larger shoots with more nitrogen than when slightly browsed. This is less true for *B. pubescens*. Experimental clipping also resulted in more browsing during the winter following regrowth. In all, moose find three times more biomass available after browsing (Danell *et al.*, 1985). In *B. pubescens*, phenol levels increase after browsing, while they decrease in *B. pendula* (Palo, 1984).

11.6 The question of coevolution of plants and herbivorous mammals

Coevolution is defined as *reciprocal* stepwise adaptations between at least two species (Ehrlich and Raven, 1964). "Coevolution without the criterion of reciprocity is indistinguishable from evolution and hence a useless concept" (Lindroth, 1988). Consider the following scenario. A plant develops effective antiherbivore defenses. In response, a herbivore counteradapts to circumvent these defenses and is at a competitive advantage over other herbivores. The plant, in turn, responds to this breach of its defenses. In insects, such pairwise reciprocal evolution can take the form of a "chemical arms race" (Dawkins and Krebs, 1979). Coevolution differs from evolution by being narrower, with fewer participants, perhaps even only two species or two populations. In reality, in most ecosystems, many species prey on many other species. Therefore, we can at best speak of *diffuse coevolution*, with a number of participants that exert diluted selection pressures.

Is there coevolution between mammals and their food plants? Herbivorous mammals usually consume many different plant species, and a plant typically is food to more than one herbivore species. Any patterns of coevolution would be diluted, resulting in diffuse rather than narrow coevolution. Even the diets of the koala (eucalyptus) and giant panda (bamboo) are more varied than commonly assumed. The koala feeds on several *Eucalyptus* species and did not thrive in zoos when fed only one species. The giant panda even includes animals in its diet. Nevertheless, Lindroth (1988) saw coevolution as an "attractive hypothesis" for some mammals.

Lindroth (1988) listed five specific criteria for coevolution in mammals:

- long history of close association
- strong mutual selection pressures to produce morphological, physiological, biochemical, and/or behavioral adaptations toward each other
- if the ranges of plant and mammal are not congruent, adaptations should be most pronounced in areas of sympatry

- the *structure of the ecological community* may provide evidence for coevolution
- for mammals, the standard of reciprocal genetic change has to be upheld, but "diffuse coevolution" (groups of populations evolve in response to each other) can be allowed.

Based on these criteria, coevolution between plants and mammals has not been demonstrated satisfactorily. Coevolution is more likely in simpler plant–animal communities. Lindroth (1988) rated coevolution between herbivorous mammals and their food plants an "unsatisfactory 'maybe'."

11.6.1 Possible examples of diffuse coevolution

Three systems possibly result from at least diffuse coevolution: rat kangaroos and legumes in Australia, snowshoe hares and Alaska paper birch in Alaska, and brown and collared lemmings in Arctic North America.

Kangaroos and legumes

The relationship between marsupials and legumes in Australia suggests coevolution: while most mammals die from fluoroacetate concentrations as low as 100 mg/kg body weight, some kangaroos tolerate much higher amounts. Different species of herbivorous marsupials tolerate varying levels of fluoroacetate depending on whether or not they are, or have been, sympatric with the two most toxic plant genera. About 34 species of Gastrolobium and Oxylobium in Western Australia produce fluoroacetate at levels as high as 0.25% of fresh weight. These two genera occur within the range of the rat kangaroo Bettongia penicillata, which tolerates levels of fluoroacetate that would be lethal to other herbivores. By contrast, Bettongia lesueur lives in areas that have been devoid of such plants for at least 7000 years. It is moderately resistant to fluoroacetate. Finally, Bettongia gaimardi, the most primitive living rat kangaroo and limited to southeast Australia in the past, is not resistant. It is suggested that the genus Bettongia radiated westward from its origins in eastern Australia and developed fluoroacetate resistance in its western species (Mead et al., 1985). All three species of gray kangaroos (Macropus spp.), by comparison, tolerate fluoroacetate well. This is consistent with the assumption that they first colonized western areas and spread eastward. The eastern gray kangaroo is still able to detoxify fluoroacetate even though it is not now exposed to plants with this compound. The western gray kangaroo regularly feeds on Gastrolobium spp. but minimizes FA intake by picking species with the lowest concentrations (Mead et al., 1985). While the tolerance of kangaroos, compared with other mammals, is an impressive adaptation,

it is not clear whether the fluoroacetate in the plants is produced in response to herbivorous mammals.

Snowshoe hare and boreal trees

The relationship between the snowshoe hare, *L. americanus*, and birches and willows in the boreal forests of Alaska and Siberia may be an example of moderately diffuse coevolution. In areas of high feeding pressure by hares the levels of chemical defenses in plants are higher than in areas with less pressure. In the high-pressure areas, the hares tolerate higher levels of plant defense compounds. Twigs of birches are covered with phenolic resins such as papyriferic acid, most likely aimed at winter browsers. Birch and willows from Alaska and eastern Asia tend to be particularly resistant to browsing by mammals. These areas were not glaciated during the Pleistocene, allowing ample time for coexisting trees and herbivores to evolve defenses and feeding strategies, respectively (Bryant *et al.*, 1989). On an applied level, trees from these "resistance centers" are a good choice for transplanting to areas with high herbivore pressure.

The 10-year population cycle of hares may be driven, at least in part, by chemical plant defenses (Bryant *et al.*, 1991). During a winter with peak numbers of hares, browsing is severe. Food is reduced so much that the hares must feed on fewer species, which are also of lower quality and heavily defended, such as spruce. The hares start to starve, the population crashes. The growing season after the population crash produces more heavily defended (induced) regrowth of deciduous trees and shrubs. The high levels of defense compounds will persist for two or three winters after the hare population crash. Models show that this can drive the 10-year cycle (May, 1972). The crash of hare populations in response to a diet high in plant secondary metabolites suggests that the animals do not possess effective counteradaptations, weakening the case for coevolution (Lindroth, 1988).

Lemmings

The case of lemmings, according to Lindroth (1988), is an "unsatisfactory 'maybe'" in terms of coevolution. Brown lemming, Lemmus sibiricus, and collared lemming, Dicrostonyx torquatus, dominate the herbivore community in certain areas of Alaska. Brown lemmings live in wet areas and feed on monocotyledons such as sedges and grasses. Collared lemmings have inhabited the tundra for longer than brown lemmings, live in drier areas, and feed on dicotyledons such as forbs and willow. Plant chemistry appears to explain the diet differences. In the laboratory, brown lemmings thrived poorly on willow and

willow-extract-treated lab chow, while collared lemmings ate sedges but grew poorly on rations treated with sedge extract (Jung and Batzli, 1981). The two species of lemming possibly evolved detoxication systems that permit them to specialize on certain food plants but prevent them from dealing with compounds from other species (Lindroth, 1988). In this system, it is not clear what chemicals are involved, nor whether the chemical defenses evolved in response to rodent herbivores (Lindroth, 1988).

11.6.2 Plant defense and wildfires

Wildfires create forest gaps and start new successions of forest plants. Mammals such as snowshoe hares, deer, or beavers take advantage of early successional species. The more hectares burned in an area, the more abundant snowshoe hares become. (In northern mythology, fire "creates hares"). The hares browse birch in winter and the plants respond by induced chemical and other defenses. Thus "selection for anti-browsing defense is greatest where fire has burned the greatest area over evolutionary time" (Clausen *et al.*, 2004). A survey of Alaskan and Canadian sites of lightning-caused wildfires between 1956 and 2002 showed that Alaska paper birch, *Betula neoalaskana*, is more abundant there than is canoe (paper) birch, *B. papyrifera*. The former is better defended chemically than canoe birch and also had more resin glands and higher levels of papyriferic acid in fire areas than the same species from other areas, even when grown together in a garden. So, "selection by an herbivore (snowshoe hare) has resulted in a continental-scale biogeographic pattern in antiherbivore defense" (Clausen *et al.*, 2004).

Kairomones and synomones

Kairomones (from the Greek *kairos*, opportune moment, by stealth) are chemical cues from one species that another uses ("spying"). Primarily the receiver of the signal benefits, as in finding prey by odor, or detecting and avoiding predators by chemical cues. Since the cues are available to another species, they are considered "public" signals, in contrast to "private" signals of restricted pheromone systems, intraspecific by definition. Synomones are chemicals that regulate interspecific relationships where both partners benefit.

Section 12.1–12.7 discuss kairomones and Section 12.8 synomones.

12.1 Predator-prey interactions

12.1.1 Prey odors used by predators

Fish

Not surprisingly, much research in sharks, skates and rays has focused on the responses of sharks to human body odors. Human blood attracts sharks, while sweat does not, and urine was even slightly repellent (Tester, 1963). Practitioners use whale meat and mixtures of fish meal and fish oils as shark attractants. In both carnivorous and herbivorous bony fish (Osteichthyes) smell deals with prey odors, social odors, and chemical stimuli in homing, and it is mediated by the first cranial nerve, the olfactory nerve. By contrast, taste serves in detection and selection of food and avoidance of toxic food, and it employs the facial, glossopharyngeal, vagal, and hypoglossal nerves.

Numerous experiments with prey extracts have elucidated the stimuli that guide fish in their feeding behavior. These studies showed:

- rinses of prey organisms attract predatory fish and release food searching behavior
- predators can distinguish rinses of different prey species
- fractions of prey rinses also release feeding responses

- mixtures of amino acids are also active
- single amino acids may trigger feeding responses
- biological activity decreases from complete rinses to amino acid mixtures to single amino acids
- each predator species responds to a different mixture of compounds
- typically, amino acids with three to six carbon atoms and their derivatives serve as chemical cues for predatory fish.

Marine fish

Among the better known commercial fish, the yellowfin tuna, Thunnus albacares, responds to the odors of several species of anchovies, such as northern anchovies, Engraulis mordax, nehu (inshore anchovy), Stolephorus purpureus, and the surf smelt, *Hypomesus pretiosus*. Tuna respond to surf smelt odor only when hungry, and tuna rendered anosmic by nose plugs lose their response. Whole natural rinses are more effective than fractions of these rinses or synthetic mixtures of amino acids. Amino acid constituents are detected at a concentration of 10^{-11} mol/l. The most active amino acid was tryptophan (Williams et al., 1992b). Tuna possibly form "chemical search images" for the most abundant prey at any given moment (Atema et al., 1980). Yellowfin tuna detect prey by amino acid odors complexed with lipid vesicles, which have an onion-skin structure of liposomes. This was the first demonstration of an extracorporeal biological function for liposomes. Tuna detect prey that is beyond visual range. Complexing with lipids may considerably delay the dilution of amino acids from prey to subthreshold levels and hence extend the range of chemical prey detection (Williams et al., 1992b).

Cod, *Gadus morhua*, also respond to amino acids. Among the cod's prey is the lugworm, *Arenicola marina*. The attractive fraction of lugworm rinse elicits strong searching behavior. It is a blend of threonine, serine, glycine, alanine, valine, leucine, and glutamic acid. Individually, only glycine and alanine released significant responses (Pawson, 1977). Extracts of the shrimp *Pandalus borealis* elicit bottom food search in cod. The most abundant amino acids, glycine, alanine, proline, and arginine, are also the most active ones. Singly, glycine was most potent, followed by alanine, as in Pawson's (1977) study. The four amino acids glycine, alanine, proline, and arginine act together synergistically. These four together are more active than the total amino acid pool in the shrimp extract, suggesting that the pool may contain amino acids with antagonistic effects (Ellingsen and Døving, 1986).

Smell receptors in the marine carnivorous Hawaiian goatfish, *Parupeneus porphyreus* (Mullidae), located on the chin barbels, mediate both arousal and food searching in response to prey homogenate and rinse of intact live prey. The

barbel nerve (seventh cranial nerve) responds, as in the cod, to those amino acids that are most abundant in its prey: proline, glycine, arginine, glutamic acid, and alanine, but not to those that are present only in trace amounts (Holland, 1978).

The pinfish, Lagodon rhomboides, responds three times more strongly to extracts from pink shrimp (Penaeus duoarum) than to extract of blue crab (Callinectus sapidus). Responses to clam (Mercenaria campechiensis), whelk (Busycon contrarium), oyster (Crassostrea virginica), sea urchin (Arbacia punctulata), and striped mullet (Mugil cephalus) were 10 times weaker. Mixtures of various amino acids plus betaine were most active, and betaine was so among the single compounds (Carr et al., 1976). When added to a mixture of five amino acids (aspartic acid, glutamine, glycine, isoleucine, and phenylalanine), however, betaine accounted only for 10% of the activity (Carr and Chaney, 1976). Similarly, a mixture of 19 amino acids, plus betaine stimulates feeding behavior in the pigfish, Orthopristis chrysopterus. Betaine is the most active single compound, accounting for 39% of the potency of the prey extract (Carr, 1976).

Prey fish may mask their own odors. Some marine fish avoid predation by covering their body odors. Some parrot fish (Scaridae) sleep in a mucus cocoon. It is believed that this covers up its scent and protects it from predation. Table 12.1 summarizes some chemical predator—prey relationships in marine fish.

Freshwater fish

One of the earliest experiments on fish responses to prey odors was that by Parker (1911). He showed that catfish (*Ictalurus* sp.) and killi fish (*Fundulus* sp.) rely on their olfactory nerves for responding to earthworm extract. Bullheads (*Ictalurus* spp.) are attracted to cysteine and respond with feeding (Bardach *et al.*, 1967). The single amino acids L-alanine, L-arginine, and L-proline release the entire feeding sequence of turning, biting, snapping, and masticating in channel catfish, *Ictalurus punctatus* (Valentincic and Caprio, 1992).

Carp, *Cyprinus carpio*, are attracted to cysteine, asparagine, glutamic acid, threonine, and alanine. Extracts from *Tubifex* worms contain at least 17 amino acids. Of these, binary mixtures of one non-polar amino acid and one polar uncharged amino acid attracted carp most and led them to explore the area. Alanine, valine, and glycine proved to be the simplest combination to release significant attraction and exploration (Saglio *et al.*, 1990).

Rainbow trout, *Salmo gairdneri*, are primarily visual feeders, but also use their chemical senses for foraging. They prefer a diet flavored with squid extract to non-treated food. A synthetic mixture of 18 amino acids, two amines, and lactic acid was very active. Only L-forms triggered responses; D-forms were even repellent. Only two combinations of amino acids were active: tyrosine, phenylalanine, and lysine; and tyrosine phenylalanine, and histidine (Adron and Mackie, 1978).

Predator species	Prey species	Chemical cue ^a	Reference
Yellowfin tuna Thunnus albacares	Northern anchovy (Engraulis mordax), nehu (Stolephorus purpureus), surf smelt (Hypomesus pretiosus)	Amino acid mixture: tryptophan, on liposomes	Atema <i>et al.</i> , 1980 Williams <i>et al.</i> , 1992 ^b
Cod Gadus morhua	Lugworm, Arenicola marina Shrimp, Pandalus borealis	Mixture: threonine, serine, glycine, alanine, valine, leucine, glutamic acid, proline, arginine	Pawson, 1977 Ellingsen and D ϕ ving, 1986
Pinfish Lagodon rhomboides	Pink shrimp (Penaeus duoarum)	Mixture: betaine, aspartic acid, glutamine, glycine, isoleucine, phenylalanine	Carr <i>et al.</i> , 1976
Pigfish Orthopristis chrysopterus		Betaine and 18 amino acids	Carr, 1976

Table 12.1 Prey odors used by marine fish

Arctic charr, *Salvelinus alpinus*, swim into streams that contain mixtures of amino acids. A whole extract of commercial fish food was active at a concentration of 5×10^{-9} mol/l while mixtures of 7, 11, or 18 amino acids were active only at higher concentrations (2×10^{-6} to 5×10^{-6} mol/l) (Olsén *et al.*, 1986).

Fish discriminate between palatable and toxic prey. For instance, largemouth bass, *Micropterus salmonides*, reject the toxic tadpoles of the toad *Bufo americanus* but eat those of the spring peeper, *Hyla crucifer* (Kruse and Stone, 1984).

Herbivorous fish depend on similar chemical stimuli. The redbelly tilapia, *Tilapia zillii*, of Africa responds to amino acids in romaine lettuce (Adams and Johnsen, 1986). Ten amino acids stimulate the electro-olfactogram of the herbivorous grass carp, *Ctenopharyngodon idella*. The electrical response increases exponentially with logarithmic increases of the stimulus concentration from the threshold to 1×10^{-3} mol/l. The detection thresholds were calculated to be $1 \times 10^{-7.15}$ to $1 \times 10^{-8.7}$ mol/l, similar to known thresholds of carnivorous fish. The relative stimulatory efficacy of the ten amino acids at 1×10^{-4} mol/l was used to distinguish five groups of stimuli: cysteine was the most stimulatory, proline the least. Except for arginine, the relative stimulation efficacy of the amino acids

^a Italics indicate a particularly active compound.

Predator species	Prey species	Chemical cue ^a	Reference
Bullhead Ictalurus sp. Channel catfish Ictalurus punctatus	Earthworm Invertebrates, fish aquatic plants	Cysteine L-Alanine, L-arginine, or	Bardach <i>et al.</i> , 1967 Valentinicic and Caprio, 1992
		L-proline	
Rainbow trout Salmo gairdneri	Insects, plankton, crustaceans, fish eggs, smaller fish	Tyrosine, phenylalanine and lysine; or tyrosine, phenylalanine, and histidine	Adron and Mackie, 1978
Atlantic salmon, Salmo salar, fry	Injured crustaceans	Free amino acids	Holm and Walther, 1988
Carp Cyprinus carpio	Plants, small animals	Cysteine, asparagine, glutamic acid, threonine, alanine	Saglio et al., 1990
Grass carp Ctenopharyngodon idella	Herbivorous	Cysteine, arginine	Johnsen <i>et al.</i> , 1988

Table 12.2 Prey odors used by freshwater fish

did not differ between carnivorous and herbivorous fish. Arginine, however, is very stimulatory for grass carp but had little effect on carnivorous fish. It was concluded that "feeding niche segregation probably is not facilitated by differential olfactory sensitivities to feeding stimuli" (Johnsen *et al.*, 1988).

Fry of the Atlantic salmon, *Salmo salar*, probably rely on olfactory and gustatory stimuli for their first meal. Injured prey such as small crustaceans will leak free amino acids, which can serve as a feeding signal to the fish fry. Such "handicapped" prey will be easier to catch for the fry. If the prey is dead, and/or its free amino acids are depleted, the fry show no interest in them. In this way, the salmon can optimize its capturing efforts as well as its prey digestion. In laboratory experiments, frozen daphnids leaked as much as 35% of its methionine upon thawing. On their first 3 days of feeding, salmon fry typically chose undepleted daphnids first and virtually all spit-out prey were depleted daphnids (Holm and Walther, 1988). Table 12.2 lists some chemical predator–prey relationships in freshwater fish.

Amphibia

Amphibians usually prey visually and detection is better detected if the prey is moving. However, aquatic salamanders and toads (Bufo calamita) use

^a Italics indicate a particularly active compound.

olfaction to locate and catch prey. Vis-à-vis motionless prey, tiger salamanders, Ambystoma tigrinum, rely more on olfaction for foraging in darkness or when experimentally deprived of sight (Lindquist and Bachman, 1980). Other species that find their prey by chemical cues alone include the toads Bufo boreas and Bufo marinus the frog Rana pipiens, the salamander Plethodon cinereus, and two species of Triturus. The newt Notophthalmus viridescens locates pill clams, Musculium rosaceum, by smell alone. Fossorial anurans (Rhinophrynus dorsalis and Myobatrachus gouldii) almost certainly smell out their prey (reviewed by Duellman and Trueb, 1986).

Reptiles

Snakes

Two phenomena of reptilian prey searching are well investigated: responses of various snakes to the odors of invertebrates, and rattlesnakes' trailing of envenomated small mammals.

Responses of snakes to food odors illuminate the interaction of genetically anchored preferences with the modifying role of individual feeding experience (see also p. 229). Newborn, food-naive snakes tongue flick and prey attack cotton swabs soaked with water extracts of skin of small invertebrate and vertebrate prey. Species differ according to their natural feeding habits. For example, ecologically different populations of water snakes, Natrix s. sipedon, respond to chemicals from different prey species but this response can be modified by experience. Previous exposure to prey odors influences chemosensory responses to the odor. Snakes raised in the laboratory on a diet of goldfish later preferred goldfish extracts. Likewise, snakes caught at a fish hatchery and fed goldfish preferred goldfish odor to all others (Gove and Burghardt, 1975). Wild-caught snakes from a mountain stream preferred extracts from stream fishes, including darter (Etheostoma ruflineatum), sculpin (Cottus carolinae), minnow (Notropis coccogenis), and stone roller (Campostoma anomalum). Laboratory-born offspring of such mountain-stream snakes preferred extracts of amphibians and fish from the mountain streams, even though they had not experienced them before. Neonates of garter snakes and a number of other species show such preferences (Burghardt, 1966; Burghardt, 1975). Newly hatched food-naive fox snakes, Elaphe v. vulpine, preferred extract of baby mouse skin to extracts of many other potential prey species. The corn snake, E. guttata, also responded to baby mouse extract, even though it was tested only in comparison with water (Burghardt and Abeshaheen, 1971).

Timber rattlesnakes, *Crotalus horridus*, are ambush hunters. They assume the ambush posture after smelling prey odors. In the laboratory, these snakes recoil the front part of their body into the ambush posture after flicking their tongues

in response to water extracts of the surface of white-footed mice and chipmunks, their natural prey. They tongue flicked and showed the ambush posture in response to control odors, such as water rinses of dog, frog, or skink, no more often than to unscented tap water (Clark, 2004).

Hatchling pine snakes, *Pituophis melanoleucus*, of the Pine Barrens in New Jersey prefer mouse odor in a Y-maze more if they have been incubated at a higher temperature (33 °C versus 28 °C). Furthermore, experience plays a role in this species, too. Snakes that had eaten a mouse detect and follow a mouse odor trail, while naive snakes show no such response (Burger, 1991).

Garter snakes

Garter snakes respond to non-volatiles from prey, as tongue contact with prey samples is necessary. The Eastern plains garter snake, *Thamnophis r. radix*, prefers the odor of worms, leeches, fish, tadpoles, and frogs, while the western smooth green snake, *Opheodrys vernalis blanchardi*, prefers only cricket odor (Burghardt, 1967). This reflects the different habitats of the two species: the first forages in wet areas, and the latter in drier upland habitat. Newborn garter snakes, *Thamnophis sirtalis*, but from mothers of different regions such as Wisconsin, Iowa, and Illinois, differed in their food odor preferences (Burghardt, 1970). However, the diet of the mother does not necessarily influence the chemical-cue preferences of newborn snakes: these newborn always preferred worm extract, whether the mother's diet had been worm or fish (Burghardt, 1971).

Sectioning of the olfactory nerve does not eliminate accurate trailing: garter snakes use their vomeronasal organ for earthworm trailing. When the vomeronasal organ is transected or sutured closed, garter snakes are unable to discriminate prey odor from water (Kubie and Halpern, 1978). For trailing of earthworms, the active components are non-volatile, water-soluble, stable macromolecules (Sheffield et al., 1968). The feeding cue is contained in lyophilized collagen of the cuticle of earthworm, Lumbricus, terrestris, but not in sandworm (Nereis) and is possibly a glycoprotein. Periodate treatment destroys the chemoattractant function, possibly because the carbohydrate residue of the glycoprotein is destroyed (Halpern et al., 1986). The chemical cue is a protein with molecular mass 20 kDa (Wang et al., 1987). It contains glycine, serine, and threonine and has a high hydroxyproline: proline ratio (Kirschbaun et al., 1986). Garter snakes respond to high- and low-molecular-weight fractions of chloroform-methanol extracts from earthworm and fathead minnow, Pimephales, promelas, with a similar pattern of tongue flicking and attacking (Burghardt et al., 1988).

Different populations of the same species may differ genetically in their response to diet and diet odors, reflecting an area's predominant diet. The western garter snake, *Thamnophis elegans*, feeds on slugs in the coastal part of its range, and on fish and frogs in the drier inland areas at higher elevations. Food-naive young coastal snakes readily eat slugs, while food-naive inland snakes refuse them. Inland populations face the problem that a snake might attack dangerous leeches along with desirable slugs, while coastal snakes do not experience leeches at all, hence no such confusion. Moreover, isolated newborn snakes from both coastal and inland populations respond strongly to the *odor* of toad tadpoles, but only coastal snakes responded to slug odor (Arnold 1981a).

Rattlesnakes: prey search after striking and envenomation

Preying rattlesnakes first strike and *envenomate* their prey and then release it and let it wander freely for up to several meters. The snake then searches for the envenomated prey, using its chemical senses. This behavior is termed "strike-induced chemosensory searching" (Chiszar and Scudder, 1980). The sequence of preying is striking, high rate of tongue flicking, locating of prey trails, trail following, finding carcass, and finally ingesting (Chiszar *et al.*, 1988a). In the prairie rattlesnake, *Crotalus viridis viridis*, striking intensifies chemosensory trailing (Golan *et al.*, 1982). The banded rock rattlesnake, *Crotalus lepidus*, discriminates between envenomated and non-envenomated mice that are presented to them in mesh bags (Chiszar *et al.*, 1983). Northern Pacific rattlesnakes, *C. viridis oreganus*, that followed a trail of an envenomated mouse were unable to determine the direction the mouse had moved (Smith and Kardong, 2000). The snake distinguished the odor of the nasal–oral area from that of the anogenital area of the mouse, but the mouse has to be envenomated. This discrimination aids in orienting the prey for swallowing it head first (Duvall *et al.*, 1980).

Comparative studies show that rattlesnakes differ in preying and strike-induced chemosensory searching according to their natural feeding habits. *C. viridis* flicks its tongue for a long time after a strike, even if no prey trail odor is present. The twin spotted rattlesnake, *Crotalus pricei*, specializes as a lizard predator and needs the presence of an odor trail for sustained tongue flicking. The requirement of chemical feedback for a high rate of tongue flicking is considered a primitive trait in rattlesnakes (Cruz *et al.*, 1987).

The rattlesnake *C. viridis* searches for the particular odor it had experienced when striking the prey. In one experiment, snakes were induced to strike perfume-treated mice. Then they were exposed to perfumed, but non-envenomated, carcasses. The snakes preferred a carcass with the same odor as the originally struck mouse. In a second experiment, snakes preferred the carcasses of mice on the same diet as the ones they had struck. Thus rattlesnakes form a

specific chemical search image (Melcer and Chiszar, 1989). They learn a mouse's chemosensory signature, which is independent of envenomation, and follow its scent trail (Chiszar *et al.*, 1990).

The northern Pacific rattlesnake, *Crotalus viridis oregonus*, seems to need a more complex odor to pursue a mouse after striking and envenomating it. They follow neither venom trail, nor mouse odor, if the two odor trails diverge. This was examined using two stimuli: a trail of an unstruck mouse dragged along the surface (but the snake was permitted to strike this mouse later) and an artificially envenomated, different unstruck mouse. The snake uses chemical cues from two possible sources: cues picked up when the snake is striking and cues in, or produced by, the venom. In terms of relative effectiveness, the cues ranked venom > mouse odor > fang puncture (Lavin-Murcio *et al.*, 1993).

In the wild, rattlesnakes migrate from their hibernacula to the feeding grounds, locate rodent colonies, approach their prey, envenomate by striking, release the wounded rodent, tongue flick at a high rate, locate the trail of the prey, follow the trail, locate the carcass, and finally ingest the rodent. Whether or not (and how soon) the carcass is found, depends half and half on striking and tongue-flicking rate. Whatever influences the tongue-flick rate also affects the finding of the carcass (Chiszar *et al.*, 1988a). Migrating rattlesnakes found deer mice, *Peromyscus maniculatus*, placed in their path by odor, and the odor trail emanated from integument rather than urine (Duvall and Chiszar, 1990). Rattlesnakes, *C. v. viridis*, may assess the population density of their main prey, the deermouse, by taking up mouse odor by tongue flicking and mouth gaping (Duvall, 1986). This permits them to find and hunt in more lucrative areas of high-prey density.

Other snakes also use non-volatiles on prey trails as cues for pursuing their quarry. Prey odors from a large variety of taxa elicit tongue flicking in the predatory king snake, *Lampropeltis getulus* (Brock and Meyers, 1979). King snakes can even distinguish the dorsal skin odor of crotaline snakes from that of colubrid snakes when presented on identically looking cotton swabs (Weldon and Schell, 1984).

The brown tree snake

In the 1940s, the brown tree snake, *Boiga irregularis*, of northern Australia and New Guinea was accidentally introduced to Guam and other islands. It poses a serious danger to the survival of other vertebrate species, especially since it now occurs in densities as high as 16 to 50 snakes per hectare. The snake grows to up to 2.4 m in length and has already done much damage to native birds and bats. Of the 18 more common native species of birds, nine are extinct now, six almost gone, and three very low in numbers. The snakes have reduced the populations of

the Mariana fruit bat by preying on their young, and they now feed on chickens and caged birds, their eggs, and other pets.

Efforts to control brown tree snake populations by trapping require knowledge of the cues this species uses for preying. In laboratory experiments, the brown tree snake was unable to locate prey by airborne odors but did follow a trail made by rubbing a rat pup along a tree branch (Chiszar et al., 1988b). Brown tree snakes responded to human blood with an increased rate of tongue flicking (Chiszar et al., 1993a) and 1 in 10 experimental snakes even swallowed a blood-soaked tampon used as a stimulus (Chiszar et al., 1993b). Response to human blood may have serious implications in populated areas on Guam where the brown tree snake is superabundant. Bites by brown tree snakes account for 1 in 1200 emergency room visits here; 80% of these people are bitten while sleeping, and 52% were under 5 years of age. The snakes bite mostly on fingers and hands, chew and try to ingest these body parts while coiling around necks and bodies of their victims. Therefore, these attacks represent attempts at preying, and not defense. Rubbings of human skin are as attractive to brown tree snakes as rubbings from mice. Presumably skin lipids provide a feeding stimulus (Greene et al., 2002).

Dead mice are as attractive as live mice, and dead mouse odor is as active as a dead mouse (Shivik and Clark, 1997). Elucidation of the feeding stimuli used by this species will aid in effective trapping for snake control. Fractions of dead mouse odor are being bioassayed. The best known constituents of rotting carcass (e.g. fish) odor, putrescine (1,4-diaminobutane: $NH_2CH_2CH_2NH_2$) and cadaverine ($NH_2CH_2CH_2CH_2NH_2$) were not active.

Multisensory control of preying in snakes

Visual and chemical cues interact in foraging by natricine snakes. Even visual cues alone can elicit prey attack, especially in aquatic foraging (Drummond, 1985). Aposematic color patterns of prey enhance the learning of prey that induces illness. Garter snakes, *Thamnophis radix haydeni*, were exposed to fish and earthworms presented on black-and-yellow forceps, and then injected with lithium chloride (LiCl). Control prey was offered on green forceps. Later, the snakes avoided food from either forceps, but the aversion to prey paired with black-and yellow was stronger (Terrick *et al.*, 1995).

Learning in snakes

Snakes can learn to discriminate profitable from less-manageable prey. Naive garter snakes, *Thamnophis melanogaster*, attacked both the carrion-eating leech *Erpobdella punctata* and the blood-sucking leech *Haementeria officinalis* even though naive snakes respond less to the odor of *H. officinalis*. The latter thwarted

attack or killed the snake. Over 4 weeks, the snakes progressively learned to avoid live H. officinalis (Drummond and Garcia, 1995).

Lizards

Lizards also increase tongue flicking after striking prey. The savanna monitor Varanus exanthematicus increases its tongue-flicking rate from 25 every 2 minutes to 90 every 2 minutes upon presentation of a mouse if the mouse is removed from its mouth (Cooper, 1989a,b). Among lizards, actively foraging species rely on prey odors, while ambush foragers do not (Cooper, 1994). The vomeronasal organ is important for tongue flicking during foraging in the ocellated skink, *Chalcides ocellatus*. It is assumed that vision is important for attacking fast-moving prey, while chemical identification is required during the subsequent predatory sequence, including consumption of prey (Graves and Halpern, 1990). Indeed, adult males of the broad-headed skink, Eumeces laticeps, increased their tongue flicking after they had bitten a neonatal mouse. Cotton swabs with mouse odor also stimulated tongue flicking (Cooper, 1992). Some scincid lizards such as Eumeces inexpectatus can discriminate prey odors from control odors from birth (Loop and Scoville, 1972; Burghardt, 1973). One omnivorous lizard, Gerrhosaurus validus (Gerrhosauridae, Cordyliformes), increased its tongue-flicking rate when exposed to cricket odor and romaine lettuce odor (their regular food), but not to odor of yellow squash (which they had consistently rejected earlier) (Cooper 2000).

Olfaction is not necessary in other lizard species. For instance, the ground-skink, *Scincella lateralis*, which feeds on insects in the ground litter, does not require chemical cues to attack prey (Nicoletto, 1985). Presumably, vision alone suffices for this purpose.

Amphisbaenids

The mostly ant- or termite-eating amphisbaenids use odors for foraging. The amphisbaenan *Blanus cinereus* increased its tongue flicking to the odor of ants presented on a cotton swab but responded little to cologne or water, and not at all to the odor of beetles (López and Salvadore, 1992).

Turtles

Turtles also use chemical cues for food, as shown for the marine loggerhead turtle, *Caretta caretta* (Grassman *et al.*, 1984). Among freshwater turtles, the European pond turtle, *Emys orbicularis*, locates food by chemoreception. This species moves chemical stimuli from the pond water to the chemoreceptors of the nasal and oral cavities by slow "jaw testing movements". The frequency of these movements increases in response to the amino acids alanine, arginine, and glutamine in a dose-dependent fashion (Manteifel and Goncharova, 1992; Manteifel *et al.*, 1992).

Crocodiles

Even crocodilians select prey by odor. If beef, nutria, or rattlesnake meat in perforated paper bags were presented on a pulley system over the water to American alligators, *Alligator mississippiensis*, the alligators contacted more bags with food than empty control bags, and contacted them sooner. They also removed more bags of those that contained meat. They did not distinguish between different types of meat, even though they had been fed nutria meat for years (Scott and Weldon, 1990). In a separate experiment, chloroform extracts of beef elicited more gular pumping behavior in American alligators than did chloroform alone. Gular pumping also increased in response to odors of crayfish, chicken, nutria, catfish, and alligator (Weldon *et al.*, 1990).

Birds

We now know that some bird taxa use their sense of smell in foraging. These include kiwis, vultures, seabirds, and honey guides. Others, such as seed-burying birds, and oilbirds, most likely use olfaction for finding food.

Kiwis

The kiwi (*Apteryx* sp.) has nostrils at the tip of its beak and feeds on earthworms, grubs, and small insects. When given a choice of containers covered with nylon screening and soil that contained either food or only soil, kiwis broke consistently into those with food, indicating that they use chemoreception to locate prey (Wenzel, 1968).

Raptors

Turkey vultures, *Cathartes aura*, find buried carcasses on the floor of tropical forests and will correctly detect meat experimentally hidden under leaves (Owre and Northington, 1961). Ethyl mercaptan released in the path of migrating turkey vultures attracted them to the general area. Once in the area, they seemed to search visually for the exact location of food. This is adaptive in forested and vegetated areas (Stager, 1967). The turkey vulture has the eighth largest olfactory bulb among the 108 bird species investigated by Bang and Cobb (1968). In an experiment at the Smithsonian Tropical Research Institute on Barro Colorado Island in Panama, chicken carcasses were placed 200 or 400 m apart on the floor of the tropical forest and covered with dry leaves. The turkey vultures efficiently located 1-day-old carcasses. However, they did not find

animals that had died very recently as easily and they rejected rotten meat. After the turkey vultures, black vultures, *Coragyps atratus*, arrived at the food. This species does not smell the carcasses but visually follows the turkey vultures. By itself, the black vulture forages in open country, and nowadays frequents garbage dumps. Both species eventually removed 90% of the provided food (Houston, 1987).

The large king vulture, *Sarcorhamphus papa*, of South America soars at 300 to 500 m, where presumably little or no odor can be detected. Like the black vulture, the king vulture probably first observes the turkey vultures and two species of yellow-headed vultures (*Cathartes melambrotus* and *Cathartes burrovianus*), which also can smell carcasses. The king vulture appears to use the sense of smell of the other three species for its benefit. In return, the king vulture aids the other vultures by tearing and feeding on the skin of larger animals, making them available to these smaller species. This highlights the interdependence of the members of the community of American forest vultures (Houston, 1984). By comparison, Old World vultures do not forage by smell, stay in more open areas, and do not scavenge in the forest. Using the sense of smell has opened up a new niche to New World vultures.

The turkey vulture's sense of smell has even been recruited to detect leaks in natural gas pipelines. In 1938, Union Oil Co. injected ethanethiol into gas lines and watched for turkey vultures to appear over leaks (Stager, 1964). The vultures were attracted from up to 61 m altitude and 183 m downwind. To achieve this attraction, a detection threshold of 1×10^{-12} to 1×10^{-13} mol/l was necessary. Later, the vulture's detection thresholds were measured and found to be much higher even than that (Smith and Paselk, 1986; see p. 115). It still is not clear what compounds turkey vultures use to detect carcasses and at what concentrations.

Seabirds

Among the Procellariiformes (shearwaters [Puffinus spp.] and petrels), Wilson's storm-petrels (Oceanites oceanicus), Leach's storm-petrels (Oceanodroma leucorhoa), and greater shearwater (Puffinus gravis) approached cod liver oil-soaked sponges over water, while sooty shearwaters (Puffinus griseus) did not (Grubb, 1972). A variety of other Procellariiformes respond to food odors on the ocean surface. These include black-footed albatrosses (Diomedea nigripes), shearwaters (Puffinus creatopus, P. puffinus, P. bulleri, and P. tenuirostris) and northern fulmars (Fulmarus glacialis). Tuna oil, a tuna fraction, and squid homogenate attracted these birds, while bacon fat, vegetable oil, mineral oil, and petroleum oil did not. They were attracted to slicks contained in plastic pools and to odor-saturated wicks attached to vertical poles on floating inner tubes. Other seabirds such as gulls, terns, cormorants, or pelicans were not attracted to the odors (Hutchison and Wenzel, 1980). In field tests off the California coast, northern fulmars,

E. glacialis, and sooty shearwaters, P. griseus, arrived sooner and more reliably at an experimental fraction of cod liver oil than at the whole oil. This may be because of the higher volatility of the fraction. Shearwaters also approached squid and krill from downwind. Seawater or heptane controls did not attract these birds. Non-Procellariids did not differ in their behaviors with respect to stimuli or wind directions (Hutchison et al., 1984). The "fishy odor" of krill is mainly a result of pyrazines and N,N-dimethyl-2-phenylethyl amine (Kubota and Kobayashi, 1988). Leach's storm petrel, Oceanodroma leucorhoa, was more sensitive to amines than to carboxylic acids from krill. These amines are more volatile than carboxylic acids. However, the carboxylic acid fraction attracted petrels more than did a "fishy" fraction. The more-volatile amines may be a longer-distance attractant while less-volatile carboxylic acids could be responsible for more concentrated searching in a restricted active space. The detection threshold for volatile carboxylic acids is also higher than that of amines (Clark and Shah, 1992).

The anatomy of the northern fulmar's olfactory bulbs is better suited for powerful odor *detection* than for keen odor discrimination. The periglomerular and external tufted cells are relatively sparse. These cells are important for superior odor *discrimination* in macrosmatic mammals. The interior granule cells are also loosely organized (Meisami and Wenzel, 1987).

Several Antarctic petrels distinguished floating sponges soaked with cod liver oil from those soaked with mineral oil. These included Wilson's storm petrel (*O. oceanicus*), snow petrel (*Pagodroma nivea*), and the pintado petrel (*Daption capense*) (Jouventin and Robin, 1984). The snow petrel has the most highly developed olfactory bulbs (Bang, 1965). Non-breeding snow petrels found herring hidden under perforated cups and even pecked at the hand that had handled the fish and not at the other hand. They also found pieces of fish buried in snow. By contrast, South Polar skuas, *Catharacta maccormicki*, never found a piece of fish unless they had seen it buried by the experimenter (Jouventin, 1977).

Sub-Antarctic seabirds were examined in the Crozet archipelago. With 26 species, it boasts the world's highest concentration of Procellariiforms. A sponge treated with cod liver oil was placed in a box on a raft and presented to free-ranging seabirds. A seawater sponge served as control. Wilson's storm petrel, *O. oceanicus*, and the black-bellied storm petrel (*Fregetta tropica*) responded most strongly to the odor. Cape petrels, *D. capense*, white-chinned petrel, *Procellaria aequinoctalis*, and giant petrels, *Macronectes* sp., were also significantly attracted. Albatrosses, prions, and diving petrels showed no interest. In general, only surface feeders, not diving species, responded to the food odor (Lequette *et al.*, 1989).

Marine microalgae such as *Phaeocystis pouchetti* produce dimethyl sulfide and release it into the air, where it can persist for hours or days. Dimethyl Sulfide is

formed enzymically from dimethylsulfoniopropionate, particularly upon grazing by zooplankton such as Antarctic krill (*Euphausia superba*). Krill, in turn, is the prey of many seabirds. Experiments have shown that procellariiforms, especially Wilson's and black-bellied storm petrels, white-chinned petrels, and prions are attracted to dimethyl sulfide if it is in the nanomoles per cubic meter range on an oil slick. Dimethyl sulfide is as attractive as cod liver oil to these birds but did not attract albatrosses (black-browed, gray-headed, and wandering) and cape petrels (Nevitt *et al.*, 1995). In the presence of a DMS aerosol, white-chinned petrels also turned more often in the odor plume than they did to a water control, while black-browed albatrosses did not discriminate in this way (Nevitt, 1999).

Another experiment used cod liver oil, crude krill extract, and the krill odor components trimethylamine and pyrazine, with phenylethanol (rose odor) as control. Several procellariiform species were attracted to krill extract. The krill odor components attracted giant petrels, cape petrels, blue petrels, Antarctic petrels, Kerguelen petrels, and black-browed albatrosses. Cape petrels were more attracted to trimethylamine than to pyrazine and cod liver oil but blue petrels responded most to cod liver oil (Nevitt, 1999).

Shorebirds use their sense of taste when probing sand for food. The purple sandpiper, *Calidris maritima*, and the knot, *Calidris canutus*, forage much longer in jars that contain food buried in sand, or sand with an extract of food, than in jars with plain sand (Gerritsen *et al.*, 1983). Table 12.3 lists the responses of various seabirds to prey odors.

Honey guides: of bees and birds, and the badger

Some species of honey guides (Indicatoridae), sparrow-like birds in Africa and Southeast Asia, guide wild mammals to wild honeybee colonies. They are attracted to wax, and not the honey. The tropical African species Indicator indicator interacts with the honey badger or ratel, Mellivora capensis. The bird smells the wax that contains honey and gives characteristic calls in the presence of the ratel. The badger follows the bird to the wild bees' nest and opens up the hive with its powerful claws. The badger eats the honey, the honey guide the wax. A Portuguese missionary in East Africa reported in 1569 that honey guides flew into his mission, were attracted to the burning altar candles, and ate the beeswax. Honey guides could be kept alive for 32 days on a pure beeswax diet. A 100% beeswax candle put in a tree attracted honey guides only when lit: within 35 minutes, six honey guides appeared (Friedmann, 1955). Honey guides have a large olfactory apparatus (Stager, 1967). Honey guides (Indicator variegatus and Indicator exilis) have been attracted to mist nets by burning honey comb (Archer and Glen, 1969). The odor of fresh beeswax plays a pheromonal role in the beehive: it stimulates food hoarding in honeybees (Blum et al., 1988). Honey

 Table 12.3 Responses of seabirds to food odors

Seabird species	Cod liver oil	Tuna oil, tuna fraction, squid homogenate		Buried fish	Dimethyl sulfide (from phytoplankton)	Reference
Shearwaters						
(Puffinus spp.)						
P. griseus	_	+				1-5
P. gravis	+					1
P. creatopus		+				3
P. puffinus		+				3
P. bulleri		+				3
P. tenuirostris		+				3
Storm petrels						
Wilson's	+				+	1,4,6–9
Leach's	+		$+^a$			1–7
Black-bellied	+				+	4–9
Petrels						
Snow	+			+		2,4
Cape (pintado)	+		+		_	4, 6, 8, 9
White-chinned	+				+	6–9
Giant	+		+			6–8
Blue		+	+			8
Antarctic			+			8
Kerguelen			+			8
Albatrosses	_					6
Black-footed		+				3
Black-browed		'	+		_	8,9
Grayheaded			·		_	9
Wandering					_	9
Prions	_				+	6–9
Diving petrels	_					6
Fulmars northern		+				5
Gulls, terns,		1				3,5
cormorants,						J,J
pelicans						
South Polar skua				_		
- Cathi i Olai Situa						

^a Amines, carboxylic acids.

^{1.} Grubb, 1972; 2. Jouventin, 1977; 3. Hutchison and Wenzel, 1980; 4. Jouventin and Robin, 1984;

^{5.} Hutchison et al., 1984; 6. Lequette et al., 1989; 7. Clark and Shah, 1992; 8. Nevitt 1995, 1999;

^{9.} Nevitt et al., 1995.

guides probably have secondarily cued in to the pheromone, which, therefore, also qualifies as a kairomone. Archer and Glen (1969) caught all their honey guide specimens within 50 yards of wild bee colonies, even as long as 2 weeks after bees had been absent from two of the tree hives. The wax of bees plays a vital role in the social organization of honey guides: males of the orange-rumped honeyguide of Asia defend a colony of the bee *Apis dorsata* year round. The wax is a critical resource: females are attracted to these polygynous males on the basis of the resource they defend (Cronin and Sherman, 1976).

Finding food caches

Birds such as nuthatches, nutcrackers, and jays store food by burying seeds such as acoms, beechnuts, and pine seeds in the ground and find it later very well. A single Clark's nutcracker, *Nucifraga columbiana*, buries as many as 33 000 piñon seeds in up to 3750 different caches (Vander Wall, 1982). Experiments have shown that these nutcrackers use memory (Tomback, 1980), visual cues (Vander Wall, 1982), and probably little olfaction to locate buried food caches. Other birds use olfaction. Black-billed magpies, *Pica pica*, discovered buried suet or raisins better when the cache was scented with cod liver oil (Buitron and Nuechterlein, 1985).

The oilbird

The oilbird or guácharo, *Steatornis caripensis*, of South America and Trinidad – a relative of the nighthawks – has large olfactory bulbs (Stager, 1967) and probably uses olfaction for finding the aromatic, spicy fruits of the oil palm during its nocturnal foraging. These birds fly up to 75 km from their cave to feed. Oilbirds were first made famous by Alexander von Humboldt's visit to the Guácharo Cave in Venezuela in 1799. and locals have traditionally turned the fat of young birds into "guácharo butter" for cooking and lighting.

Other senses in birds

Birds do not necessarily distinguish palatable and unpalatable prey by odor. Free-ranging European birds such as chaffinch, house sparrow, robin, starling, blackbird, and song thrush recognized bread pieces treated with quinine and mustard powder only by their size, but not other visual cues or smell. This has a bearing on model–mimic relationships. Batesian mimics are often smaller than their unpalatable models (Marples, 1993).

In ultraviolet light (320–400 nm) birds can see, rather than smell, urine and feces on vole trails. In the laboratory, kestrels, *Falco tinnunculus*, spent more time near ultraviolet-illuminated, artificial urine- and feces-soaked trails of voles, *Microtus agrestis*, than near such trails in visible light, and clean trails under both

types of illumination. Likewise, in the field, kestrels hunted, perched, and used nestboxes more often near urine- and feces-soaked artificial vole trails than near artificial trails without urine or feces, and areas with no vole trails (Viitala *et al.*, 1995). Whether and under what conditions kestrels and possibly other birds use the fluorescence caused by the ultraviolet portion of sunlight while hunting remains unclear.

Mammals

Mammals use all senses for foraging. A species' natural history will determine which sensory modality predominates. Nocturnally active species and those living in dense vegetation or underground are expected to be more olfactorily oriented. Predators can be attracted to one odor of a prey species more than to others of the same species. For instance, foxes, ferrets, and cats seek out blood odors of *stressed* Mongolian gerbils, *Meriones unguiculatus*. Cats are attracted to food pellets that are contaminated with a few pellets soaked in stress odorcontaining blood of gerbils (Thiessen and Cocke, 1986). Hunting by smell is the hallmark of carnivores. In the following, insectivores, canids, felids, mustelids, and ursids will be discussed.

Insectivores

Shrews (*Sorex cinerus cinereus* and *Blarina brevicauda talpoides*) use olfactory cues to find buried prepupae of the European pine sawfly, *Neodiprion sertifer*. Specifically, these cues play a role in digging, removing, and opening this prey, while taste stimuli guide the eating behavior (Holling, 1958).

Canids

Wolves, *Canis lupus*, detect prey from 300 (27m) yards or even up to 1.5 miles (2.5 km) downwind by "direct scenting," i.e. smelling the air. They may also track initially, followed by direct scenting at close range (Mech, 1970). The same individual may be guided by different senses according to circumstances. For instance, a coyote, *Canis latrans*, uses mostly vision when finding a rabbit in an enclosed room, and olfaction is least important. Outdoors, however, olfaction becomes more important and the coyote approaches from downwind approximately 84% of the time. In addition, wind may interfere with the sounds of the prey outdoors (Bekoff and Wells, 1980).

Felids

Predators of open country, such as Serengeti lions, *Panthera leo*, do not necessarily approach their quarry from downwind. In 300 hunts, Schaller (1972)

observed lions approach prey equally often (28% each) from up- and downwind, and in the remainder of cases with wind from the side. This, random approach direction with regard to wind (and odor) suggests that lions primarily use vision in hunting.

Mustelids

Small predators such as mustelids extensively use their sense of smell to locate prey. Moreover, they respond more to familiar prey and *learn* certain odor preferences early in life during a *sensitive period*. In one experiment, polecats, *Putorius putorius*, first lived on a diet of domestic chicks and, starting at different ages, later received dead mice and rats in addition. Subsequently, up to 1 year of age, they were tested for prey odor preferences. The later they had been exposed to mouse and rat carcasses, the less they responded to mouse odor. It was concluded that prey odors are learned around the second or third month of life (Apfelbach, 1973). This can be important in captive breeding of rare animals. For instance, captive black-footed ferrets need prairie dog meat while raising their young. They should be exposed to this type of meat early in life. Otherwise, the reintroduction of these captive-raised young might fail.

Ursids

Bears have a keen sense of smell, even though we lack formal and thorough studies. According to popular television programs, polar bears can smell a ringed seal pup buried in snow from distances exceeding 1 mile (1.6 km). The exact distances for the bears prey detection is not yet clear (I. Stirling, personal communication) and only 1 in 20 hunts is successful.

Rodents

Many birds and mammals bury seeds in soil for storage. Mammals that cache food subterraneously include tree and ground squirrels, and deer mice, *P. maniculatus*. Underground storage provides many benefits: competing surface foragers such as birds, deer, and turkeys cannot reach the food and the seeds may loose undesirable secondary plant compounds such as tannins in the moist soil, amounting to food processing. Small mammals use olfaction, spatial memory, and visual landmarks to locate buried seeds. Spatial memory favors the owner of the buried cache, while olfaction permits other individuals to pilfer stored food. For underground hoarding to remain a viable behavior, the animal that caches the food must enjoy a greater chance of finding it again than do competitors.

Deer mice, *P. maniculatus*, readily find buried seeds. In forest reseeding projects, deer mice often remove 70 to 100% of the planted conifer seeds. In the laboratory, deer mice found the aromatic seeds of sugar, Jeffrey, and Ponderosa

pine buried in 5 cm peat moss more easily than seeds of wheat, barley, or oats. If these cereal seeds were treated with safflower oil or lecithin-mineral oil, the mice found them more easily. This suggests that deer mice use olfactory cues to find buried seeds (Howard and Cole, 1967; Howard *et al.*, 1968). Deer mice also use olfactory stimuli to find buried prepupae of the European pine sawfly, *N. sertifer*. They distinguish between parasitized and healthy larvae, and between males and females (Holling, 1958).

Yellow pine chipmunks, *Tamias amoenus*, and deer mice of the Sierra Nevada scatter-hoard seeds of Jeffrey pine, *Pinus jeffreyi*, and other trees and shrubs. Knowledgeable foragers found the buried seeds whether the soil was dry or wet. Naive foragers found caches only when the soil was wet and found seeds buried by either species equally well. Few seeds were discovered when the soil was dry regardless of the species of the caching individual or the forager. Wet seeds release organic molecules, which the rodents spy on; these odors are now functioning as kairomones. Under dry conditions, spatial memory gives the owner of the cache the edge over competitors. As the soil moisture rises, owners and competitors vie increasingly more for the seeds and pilfering increases (Vander Wall, 2000).

Primates

Lower primates hunt by smell. To the nocturnal African prosimians *Galagoides demidovii* and *Galago alleni*, the odor of a concealed insect is more important than a visible, but odorless animal. *G. alleni* even distinguishes the odors of the front and rear end of an insect. Aqueous extracts applied to the "wrong" end of the body can trick the galago (Molez-Verriere, 1988).

Marine mammals

Even marine mammals may use airborne odors for food detection. Baleen whales (Mysticeti) feed on krill near the ocean's surface. Oldtime Antarctic whalers noted a "krill odor" near large schools of krill. Cruising at the surface and inhaling periodically, baleen whales may detect krill odor. Anatomically, they have a well-developed olfactory organ (Cave, 1988), in contrast to toothed whales (Kusnetzov, 1988).

12.1.2 Predator odors used by prey

Potential prey species can chemically assess predation risk from a distance and/or from the safety of their refuge by evaluating predator odors in the area (Kats and Dill, 1998). Such odors emanate from the predator itself or its

Table 12.4 Respones o	f prey fish to	predator chemicals
-----------------------	----------------	--------------------

Prey species	Predator species	Stimulus	Response	Reference
Coho salmon Oncorhynchus kisutch	Human, bear, dog	L-Serine, L-alanine	Retreat to lower water	Idler <i>et al.</i> , 1956; Rehnberg and Schreck, 1986
	Northern squawfish Ptychocheilus oregonensis	Body rinse, broken skin extract	Avoidance, plasma cortisone and glucose rise	Rehnberg and Schreck, 1987
Arctic charr Salvelinus alpinus	Human	L-serine	Avoidance	Jones and Hara, 1982
Mosquito fish Gambusia patruelis	Chain pickerel Esox niger; redfin pickerel Esox americanus	Mucus coat of pickerel	Swim to upper water levels	George, 1960
Shiners Notropis texanus and Notropis venustusi and chubs Hybopsis aestivalis	Pikes, Esox sp.; sunfish Lepomis macrochirus; bass Micropterus punctulatus	Rinse from piscivores of North and South America	Swim to bottom of tank; no response to human hand rinse	Reed, 1969
Threadfin shad Dorosoma petenense	Large mouth bass Micropterus salmonides	Bass rinse	No response (fish move faster than chemicals)	McMahon and Tasch, 1979

"sign" such as tracks, droppings, urine or other scent marks, food remains, or disturbed soil or plants.

Fish

Several fish species, notably salmon, respond to the odors of their predators in an adaptive manner (Table 12.4). Coho and spring salmon retreated to the lower parts of a fish ladder when rinses of human hand, bear paw, dog meat, or sea lion meat were added to the water upstream. Even the odor of deer feet had the same effect (Brett and MacKinnon, 1954). L-Serine was identified as the active compound in mammalian skin (Idler *et al.*, 1956). Arctic char, *Salvelinus alpinus*, also responds to L-serine from human hand wash, while D-serine and several other amino acids were not active. L-Serine is less active than the whole hand wash, suggesting that two or more compounds are required for the full response (Jones and Hara, 1982).

Coho salmon, *Oncorhynchus kisutch*, tested in a two-choice Y-trough, avoided whole body rinses and broken-skin extracts of the predatory northern squawfish, *Ptychocheilus oregonensis*, but not those of the non-predatory largescale sucker,

Catostomus macrocheilus. However, both rinses induced elevated levels of plasma cortisol and glucose, commonly referred to as a stress response. This experiment demonstrates that the behavioral and physiological responses are not necessarily coupled (Rehnberg and Schreck, 1987).

The mosquitofish, *Gambusia patruelis*, avoids its predators, the chain pickerel, *Esox niger*, and redfin pickerel, *Esox americanus*, by swimming to the upper water levels. It also responds to odor from the pickerel mucus coat. This odor survives passage through filter paper and hours of bubbling but loses activity if heated or passed over charcoal (George, 1960).

Shiners (*Notropis texanus* and *N. venustus*) and chubs (*Hybopsis aestivalis*) avoid the water from piscivorous fish, such as pikes (*Esox* sp.), sunfish (*Lepomis macrochirus*), bass (*Micropterus punctulatus*), and two South American cichlids (*Astronotus ocellatus* and *Cichlasoma severum*). They respond by swimming to the bottom of the experimental tank and remaining motionless in clusters. Plain water or human hand rinse do not trigger this response. In this case, predator and prey do not have to occur sympatrically for the avoidance response to occur (Reed, 1969).

Fish avoid more vigorously the odor of predators that have fed on members of their species than that of those on different diets. For example, young Arctic charr avoid water from brown trout fed on Arctic charr and are less wary of that from pellet-fed trout (Hirvonen *et al.*, 2000). Prey fish also reduce their "predator inspection" behavior vis-à-vis predators that have eaten members of their own species. For instance, finescale dace, *Phoxinus neogaeus*, dash toward predators such as yellow perch, *Perca flavescens*, and withdraw. Dace inspect perch models less often if the model is accompanied by water from perch that had eaten dace than if accompanied by water from perch on a swordtail, *Xiphophorus helleri*, diet. Dace produce alarm pheromone, while swordtails do not. The Central American swordtails do not cooccur with finescale dace (Brown *et al.*, 2001).

A chemical cue from piscivorous fish may be responsible for the size and shape change in crucian carp, *Carassius carassius*, in response to the presence of the carp-eating pike, *Esox lucius*. The carp increases its bulk and becomes more difficult to catch by the pike. The chemical cue could emanate from the predator, or from injured or frightened carps (Bronmark and Miner, 1992). Crucian carp (or bronze carp) form schools. Crucian carp show stronger alarm responses to unfamiliar predatory fish such as pike or perch. Where the carp coexist with pike, their alarm behavior is attenuated. Larger predator individuals triggered stronger responses than smaller ones. The diet of the predatory fish affected the responses of crucian carp: after eating prey fish that produce alarm substance, large pike induced more alarm behavior in their prey than small pike. Crucian carp use cues from the predators as well as the alarm substance of their prey. They also distinguish large and small predators and habituate to sympatric predators.

In summary, growing a deep body in response to sympatric predators appears to reduce the need for behavioral alarm behavior (Pettersson *et al.*, 2000).

Other fish species do not respond to predator odors. The threadfin shad, *Dorosoma petenense*, is strongly attracted to odors of its prey such as brine shrimp (*Artemia*) or *Daphnia* spp. but does not respond to those of its predator, the largemouth bass, *M. salmonides*, or conspecifics. Both shad and bass swim faster than chemicals travel in water, which may explain this behavior difference (McMahon and Tash, 1979).

In minnows, taste is not sufficient for predator recognition. Anosmic fathead minnows, *P. promelas*, did not show the flight reaction to the odor of northern pike, *Esox lucius* (Chivers and Smith, 1993). Naive European minnows, *Phoxinus phoxinus*, do not exhibit a fright reaction when first exposed to a predator odor, such as that of pike, *E. lucius*. They develop a conditioned fright response only after experiencing the predator odor in dangerous circumstances, such as when accompanied by schreckstoff (alarm pheromone) of conspecifics. Responses to the odor of non-piscivorous fishes such as tilapia, *Tilapia mariae*, can also be conditioned in this fashion but the responses are much weaker (Magurran, 1989).

Predator density, and hence probability of attack, and the cost to the prey species alter responses to predator cues, including chemical ones, by the same prey species. For instance, in a laboratory tank, Trinidadian guppies, *Peocilia reticulata*, from predator-dense downstream sections stay at greater distances from a hungry predatory largemouth bass, *M. salmonides*, than do guppies from headwater streams that have few predators. Both populations discriminated a hungry bass from a sated one (Licht 1989).

Amphibia

Salamanders

Salamanders and their larvae detect predators by odor cues (Table 12.5). They take refuge and are also distasteful to predators. The salamander *Eurycea bislineata* and tadpoles of the tree frog, *Hyla chrysoscelis*, avoid the odor of the predatory green sunfish, *Lepomis cyanellus*. They do not respond to the odors of the green frog, *Rana clamitans*, and brine shrimp (*Artemia* sp.) used as controls (Petranka *et al.*, 1987). Tadpoles of the small-mouthed salamander, *Ambystoma texanum*, hide in refuges when the odor of their predator, the sunfish *L. cyanellus*, is added to the water. They do not respond to the odors of snapping turtle (*Chelydra serpentina*), water snakes (*Nerodia sipedon*), crayfish (*Oronectus rusticus*), or odorless water. Only larvae from water bodies that have fish do actually respond to fish cues (Kats, 1988). Why do these larvae not respond to the odors of sympatric predaceous crayfish, turtles, and snakes? If further studies confirm these

Prey species	Predator species	Chemical cue	Response	Reference
Small-mouthed salamander Ambystoma texanum: tadpoles	Green sunfish Lepomys cyanellus	Rinse	Hide in refuges	Kats, 1988
Salamander Eurycea bislineata and tree frog Hyla chrysocelis: tadpoles	Green sunfish	Rinse	Avoidance	Petranka <i>et al.</i> , 1987
Salamander Plethodon richmondi	Ringneck snake Diadophis punctata	Traces on substrate	Avoidance	Cupp, 1988
Tadpoles of frogs Rana lessonae and Rana esculenta	Pike Esox luteus	Rinse	Swim, rest, edge use	Stauffer and Semlitch, 1993

Table 12.5 Responses of amphibian prey to predator chemicals

laboratory results, one might expect these predators to have a more recent association with small-mouthed salamanders (Kats, 1988).

On land, snakes prey on salamanders and their eggs. The ringneck snake (*Diadophis punctata*) and its prey, the salamanders *Plethodon richmondi* and *Plethodon dorsalis* provide an example. Salamanders were given a choice between a paper towel that had had a ringneck snake on it for 48 hours, and one with clean water. The salamanders of both species avoided the substrate with the odor of the ringneck snake (Cupp, 1988).

In their predator avoidance, salamanders use complex odors that combine chemicals from both predator and prey. In the laboratory, red-backed salamanders, *P. cinereus*, avoid filter papers soaked with water extracts from garter snakes that had been preying on salamanders, while earthworm-fed snakes lacked this effect. Exudations from unfed snakes and extracts from homogenized salamanders had no such alarming effect (Madison *et al.*, 2002).

Frogs and toads

Tadpoles of the two closely related frog species *Rana lessonae* and *Rana esculenta* respond more to chemical cues of their predator, the pike *E. lucius*, than to visual and tactile ones. The strongest swimming, resting, and edge-use behaviors – all considered antipredator responses – occurred to a combination of

chemical and tactile cues (Stauffer and Semlitch, 1993). Rana temporaria tadpoles became less active when exposed to chemical cues from their predators, perch, *Perca fluviatilis*, and dragonfly larvae, *Aeschna juncea*. Tadpoles of different parentages differed in their responses, suggesting genetic factors. Overall, their behavior appeared to be rather plastic (Laurila, 2000). In one experiment in Sweden, tadpoles of the frog *R. temporaria* avoided water-borne cues from their predator the rainbow trout, *O. mykiss*, by hiding in refuges, while toad tadpoles, *Bufo bufo*, remained unresponsive. The behavior of the frog tadpoles affected lower trophic levels: their reduced grazing behavior resulted in more surviving plant (periphyton) mass (Nyström and Abjörnsson, 2000).

After metamorphosis, juvenile toads generally avoid predators by visual cues, whereas juvenile Great Plains toads, *Bufo cognatus*, and southwestern toads, *Bufo microscaphus*, detect and avoid odors of their respective predators, the eastern plains garter snake, *T. radix*, and the wandering garter snake, *T. elegans*. This was demonstrated by presenting the odors on paper towels on which the snakes had been living for 24 hours. Lizard odor had no effect (Flowers and Graves, 1997).

Ecological and evolutionary aspects

Ephemeral ponds and streams harbor different species of larval amphibians to those in permanent bodies of water. Several factors may be responsible for this, although predation appears to play an important role. In permanent aquatic habitats, predator densities are higher, and fish, including predators, are usually restricted to such permanent bodies of water. Chemical recognition of predators and chemical defense compounds are two ways in which amphibian larvae counteract predation. In one experiment, predator-naive larvae of different amphibian species were exposed to water from tanks with a predatory green fish, Lepomis cyanellus. Amphibians from temporary ponds and streams who rarely encounter fish did not avoid the fish odor by hiding. These species are also palatable to predators. By contrast, larvae from permanent ponds often encounter fish and so would usually avoid the fish odor regardless of whether they were palatable or unpalatable. Therefore, the response of amphibian larvae to the odor of a predatory fish correlated the with the probability of fish encounters (i.e. predation risk). Closely related taxa differed in palatability and their responses to predator chemicals. For instance, green frog, R. clamitans, and bullfrog, Rana catesbeiana, were unpalatable and encounter fish often, while wood frog, Rana sylvatica, was palatable and rarely encountered fish. Variation also occurs within one species: small-mouth salamander, Ambystoma texanum, from streams with fish avoided the sunfish odor, while salamanders from fishless ponds did not. Thus, natural selection rather than phylogeny appears best to explain the observed differences in antipredator defenses (Kats *et al.*, 1988). Investigators have found that amphibian larvae from permanent bodies of water had at least one of the two chemosensory defenses, while those from ephemeral habitats consistently lacked such defenses. Chemical detection of predatory fish appears to be the major defense of palatable amphibian larvae that coexist with fish predators. These studies imply interaction by chemical cues as a proximate mechanism operating where predators organize animal communities.

Life history shifts

Chemical cues from predators can change the rate of development in amphibians. For example, red-legged frogs, *Rana aurora*, of western North America were raised either in the presence of chemicals from one of their predators, the rough-skinned newt, *Taricha granulosa* (which had eaten red-legged frogs) or in the presence of chemicals from injured conspecifics. In both cases, the tadpoles metamorphosed earlier and at a smaller size than usual (Kiesecker *et al.*, 2002). Similarly, tadpoles of the western toad, *Bufo borealis*, metamorphosed faster when exposed to visual and chemical cues from a predator such as backswimmers (*Notonecta* sp.), or chemical alarm cues from injured tadpoles of their own species (Chivers *et al.*, 1999; Fig. 12.1)

Conservation implications

The decline of amphibian populations has been linked to habitat destruction, increased pathogens, global warming, and ultraviolet radiation, but it may also be linked to failed chemical predator recognition. For example, the California newt, *Taricha torosa*, has decreased or disappeared from streams inhabitated by the introduced crayfish, *Procambarus clarki*, and mosquitofish, *Gambusia affinis*. In laboratory and field experiments, the newts, their eggs, and their larvae were successfully attacked and/or eaten by the introduced predators. The newt's chemical predator recognition and its defense by tetrodotoxin do not appear to cope with the introduced species (Gamradt and Kats, 1996).

Reptiles

In the terrestrial environment, olfactory cues have several advantages for detecting and avoiding predators: long-distance propagation by air currents; detection of signals in the dark; reception despite obstacles, such as vegetation; slow fade-out of signals; and possible deception by spatial separation of animal and its scent.

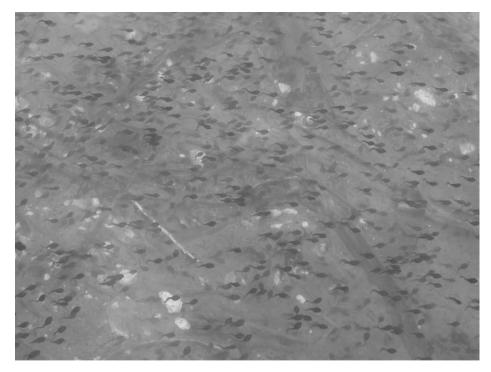


FIGURE 12.1 Tadpoles of the western toad, *Bufo boreas*, metamorphose sooner when exposed to chemical and visual predator cues or alarm substances of conspecific tadpoles (Chivers *et al.*, 1999). (Photograph: D. Müller-Schwarze.)

Snakes

Snakes respond to odors of their predators (Table 12.6). Best known are their reactions to ophiophageous snakes. The odor of the California king snake, *Lampropeltis getulus californiae*, induces *body bridging* in rattlesnakes: the midbody is raised and undulated. Blindfolding does not change this reaction, while a stick rubbed on a king snake elicits the response. Dorsal skin material is most effective. North American rattlesnakes also responded to skin rubbings of the puffing snake *Pseustes sulphureus* from South America, which feeds on birds and lizards. Rubbings from non-ophiophagous snakes of the families of boids, crotalines, and colubrids were ineffective (Bogert, 1941). More than 20 species of crotaline snakes from North, Central and South America show body-bridging when exposed to methanol-soaked cotton balls that had been rubbed on the dorsal skin of ophiophagous snakes (Weldon and Burghardt, 1979).

In a number of species, the active predator odors originate on the dorsal skin. Neonate pygmy rattlesnakes, *Sistrurus miliarius*, and timber rattlesnakes, *C. horridus*, respond to dorsal skin chemicals of the ophiophagous king snakes and indigo snakes, *Drymarchon corais*, but not to those from ventral skin or skin

Prey species	Predator species	Cue	Response	Reference
Rattlesnakes	California king snake Lampropeltis getulus californiae	Skin rubbings	Body bridging	Bogert, 1941
Rattlesnakes	King snake, human	Air or skin extract, human breath	Increased heart rate	Cowles and Phelan, 1958
Neonate pygmy (Sistrurus miliarius) and timber (Crotalis horridus) rattlesnakes	King and indigo (<i>Drymarchon corais</i>) snakes	Dorsal skin odor	Body-bridging	Marchisin, 1980
Garter snakes, Thamnophis sp.	Racer, <i>Coluber</i> <i>constrictor</i> , and king snake	Swabs or air	Increased tongue flicking	Weldon, 1982
Pine snake Pituophis melanoleucus: hatchlings	King snake	Body odor in Y-maze	Avoidance	Burger, 1989
Lizard Lacerta vivipara	Vipera berus, smooth snake Coronella	Soiled cage	Increased tongue flicking	Thoen <i>et al.</i> , 1986

Table 12.6 Responses of reptiles to predator chemicals

austriaca

glands. Furthermore, the rattlesnake *Crotalis atricaudatus* body-bridged to odor of the water snake, *Nerodia fasciata*, which feeds on amphibia and fish (Marchisin, 1980).

Garter snakes (*Thamnophis* sp.) increase their rate of tongue flicking when exposed to swabs or air from snake-eating snakes such as racer, *Coluber constrictor*, and king snake, *L. getulus*, to air that has passed over a king snake, or to skin extract from a king snake. Human breath is also active (Cowles and Phelan, 1958). Odor from the non-predatory hognose snake, *Heterodon nasicus*, does not affect tongue flicking (Weldon, 1982). Hatchling pine snakes, *P. melanoleucus*, also avoided the odor of king snakes in a Y-maze, while they were attracted to conspecific odor (Burger, 1989).

Lizards

Lizards respond to odors of predatory snakes with increased tongue flicking. The common lizard, *Lacerta vivipara*, tongue flicks more when placed in a cage that had held the predator *Vipera berus* or the smooth snake, *Coronella austriaca*, but only slightly more to the odor of the grass snake, *Natrix natrix*, which does not feed on snakes. The behavior also changed to a slow locomotion when

in a cage with predaceous snake odor (Thoen *et al.*, 1986). Gekkonid lizards also responded to odors of predatory snakes (Dial *et al.*, 1989).

Lizards that prey on snakes but are also eaten by larger snakes discriminate skin chemicals of snakes very well. Monitor lizards, *Varanus albigularis*, fall in this group. Hatchling monitors attacked harmless snakes but avoided venomous species. However, they accepted meat of all snakes if carefully skinned. The hatchlings tongue flicked to invertebrate prey covered with skin from venomous snakes and rejected these samples (Phillips and Alberts, 1992).

Mammals

Insectivores

Hedgehogs, *Erinaceus europaeus*, avoided areas in an enclosure that were tainted with feces of badger, *Meles meles*, which prey on hedgehogs. The response lasted about 2 days, but free-ranging hedgehogs on golf courses and farmland reduced their feeding near badger odor only for minutes or hours (Ward *et al.*, 1997).

Rodents

Many decades ago, Griffith (1919, 1920) described the white (laboratory) rat's response to cats. From the age of 3 weeks, rats huddle in corners, freeze, and crouch when presented with a cat. They tremble, twitch their muscles, whine, and cease to feed and nurse. A cloth with cat scent or an arena with cat scent sufficed to trigger these responses. Anosmic rats or those confronted with a cat in a glass jar remained unaffected. Hence, the critical cue proved to be odor (Griffith, 1919). Cat feces, urine, heart, or other tissues did not elicit these "fright reactions" (Griffith, 1920).

In settings that permit the recipient to move more freely, many different avoidance responses to predator odors have been observed (Table 12.7). Meadow voles, *Microtus pennsylvanicus*, avoid areas of enclosures that previously had been occupied by short-tailed shrews, *Blarina brevicauda* (Fulk, 1971). This shrew is a voracious predator with venomous saliva that feeds mostly on invertebrates but occasionally takes small mice, voles, or shrews. Free-ranging European field voles, *M. agrestis*, and field mice, *Apodemus sylvaticus*, avoid traps that are scented with anal secretion of weasel, *Mustela nivalis*, or urine from tiger, *Panthera tigris*, and jaguar, *Panthera onca* (Stoddart, 1976). Bank voles avoid areas with weasel odor as much as areas that contain live weasels (Jedrzejewska and Jedrzejewski, 1990).

In Australia, house mice living on islands without predators did not avoid traps treated with predator odors. In areas where the introduced red fox or house cat occur, or the native western quoll, *Dasyurus geoffroyii*, the mice avoided

Prey species	Predator species	Cue	Response	Reference
Rat Rattus norvegicus	Red fox Vulpes vulpes	Trimethylthiazoline	"Stress," raised corticosterone	Vernet-Maury et al., 1984
Meadow vole Microtus pennsylvanicus	Short-tailed shrew Blarina brevicauda	Soiled enclosure	Avoidance	Fulk, 1971
Field voles Microtus agrestis; field mice Apodemus sylvaticus	Weasel Mustela nivalis	Soiled trap	Avoidance of traps	Stoddart, 1976
Field mice and Clethrionomys glareolus	Mink Mustela vison	2, 2-Dimethylthietane from anal gland	Avoidance of traps	Robinson, 1990
Black-tailed deer Odocoileus hemionus columbianus	Coyote Canis latrans; puma, Felis concolor	Fecal odor	Inhibition of feeding	Müller-Schwarze, 1972

Table 12.7 Examples of responses of mammals of predator chemicals

traps with predator odors. They also used dense vegetation on moonlit nights, while mice on the predator-free islands showed no such preference for denser vegetation. Finally, predator-naive and predator-experienced mice, transferred to areas with cats and foxes, differed in their mortality: over twice the number of predator-experienced mice survived compared with naive ones (Dickman, 1992).

Free-ranging North American beaver, *Castor canadensis*, feed less on experimental aspen sticks that have been treated with extracts from predator excrement or urine. Odors from the sympatric coyote and river otter, and extirpated lynx, were most effective, while those from allopatric lion and extirpated wolf odor were less active. However, these response differences between species were small (Fig. 12.2; Engelhart and Müller-Schwarze 1995).

Urine and feces odors of mink applied to soil and vegetation in outdoor enclosures had no effect on gray-tailed voles, *Microtus canicaudus*, in Oregon. The voles did not seek taller vegetation for cover, and their reproduction was not affected. Specifically, reproductive rates, time to sexual maturation, juvenile recruitment, and activity did not change after exposure to mink odors (Wolff and Davis-Born, 1997).

The odor of ferret, *Mustela putorius furo*, urine causes male outbred laboratory *Mus musculus*, to reduce their overmarking of rival urine marks. These mice reduce predation risk at the price of tolerating more intrasexual competition (Roberts *et al.*, 2001).

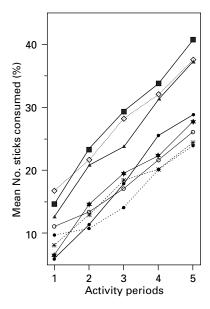


FIGURE 12.2 Responses of beavers to predator chemicals applied to aspen sticks. Activity periods were consecutive 5 days of experiment in two areas in New York and the percentages of sticks consumed is the mean of several replications of the experiment. All treated sticks were punctured to improve the uptake of chemicals; intact sticks were untreated (\triangle), punctured but not treated (\square), punctured and treated with the solvent methanol (\diamondsuit) (the three controls), or treated with extracts from other animals. Treatments that inhibited consumption most were excrement extracts from lynx (*) and coyote ($\bullet \dots \bullet$), both sympatric predators (lynx now extirpated). Beavers accepted most readily the three control sets. Other chemicals were from otter (\circ), wolf ($\bullet \bullet \bullet$), lion (*). (From Engelhart and Müller-Schwarze, 1995.)

Ungulates

In black-tailed deer, *Odocoileus hemionus columbianus*, fecal odors of sympatric predators (coyote, *C. latrans*, and mountain lion, *Felis concolor*) in vials next to food pellets inhibited feeding, while those of allopatric predators (lion, *Felis leo*, snow leopard, *Uncia uncia*) do not, or very little (Müller-Schwarze, 1972; Fig. 12.3). Note that mammals discriminate between the odors of sym- and allopatric predators, while fish and rattlesnakes do not (pp. 359 and 364). Freeranging adult female wapiti, *Cervus elaphus canadensis*, respond to the odors of dog urine, and cougar and wolf feces (presented as water slurry) with increased heart rates. It was concluded that the main effect of predator odors may be for assessing the risk of predation (Chabot *et al.*, 1996).

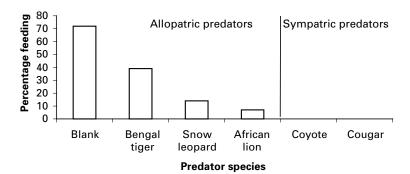


FIGURE 12.3 Responses of black-tailed deer fawns to predator odors.

Primates

Among primates, red-bellied tamarins (Saguinus labiatus) sniff and avoid fecal extracts of jaguar (P. onca), jaguarundi (Herpailurus yagouaroundi), and margay (Felis wiedi) more than those from non-predatory mammals. In this case, it is not clear whether these predators actually prey on tamarins (Caine and Weldon, 1989).

Mammals also respond to odors of predatory snakes. Female laboratory mice deposited more fecal boli in a section of an arena that was lined with paper from the cage of rat snakes, *Elaphe obsoleta*, which preys on rodents. Paper from the cage of the rough earth snakes, *Virginia striatulata*, a worm eater, had no effect. The females also ate less from snake-scented food pellets. Males did not respond to these odors (Weldon *et al.*, 1987). Kangaroo rats, *Dipodomys merriami*, avoided sidewinder rattlesnakes, but not after olfactory bulbectomy. Blinded or deafened individuals still avoided the snakes (Webster, 1973). Snake odors alarm even bears: brown bears retreat drooling from the odor of dead or live rat snakes. A rope treated with snake odor still proved active (Kano, 1976).

Chemistry of predator odors

Fox feces contain trimethylthiazoline (Fig. 3.1 p. 37), which by itself induces in rats even stronger "fear" responses than whole feces (Vernet-Maury, 1980). Of 12 compounds from fox feces tested, seven elicited "fear" responses, confirmed by postmortem corticosterone measurements. Some structure–activity relationships emerged. The thiol group is essential, while the keto group is not; molecular size can vary; mercaptoketones can be aliphatic or alicyclic; the thiol can be secondary or tertiary; and the thiol can be alpha or beta to the keto group (Vernet-Maury *et al.*, 1984).

Dickman and Doncaster (1984) suggested that feces and urine of different carnivore species may share chemicals that elicit avoidance responses in small prey mammals. This might explain why prey species avoid chemicals from allopatric or extirpated sympatric predators. Nolte *et al.* (1994b) have suggested that herbivores are repelled by odors from the excreta of various predators because these have in common sulfur compounds resulting from protein digestion. Sulfur compounds from anal gland secretions of several mustelids have been shown to be deterrents for herbivorous prey mammals such as gophers, voles, snowshoe hares, and black-tailed deer (Sullivan and Crump, 1984, 1986a; Sullivan *et al.*, 1988). These effects and the active compounds are further discussed in Chapter 13.

The European rodents *A. sylvaticus* and *Clethrionomys glareolus* tend to avoid traps scented with 2,2-dimethylthietane from the anal sacs of the mink, *Mustela vison*, while it suppresses feeding in European wild rabbits, *Oryctolagus cuniculus* (Robinson, 1990).

Effects of predator diet

When given a choice, mountain beaver, guinea pigs, house mice, and deer mice consumed less food from containers near urine from coyotes on a meat diet than from containers near urine from coyotes on a fruit diet (Nolte *et al.*, 1994b).

Multisensory control of responses

Visual *and* olfactory stimuli contribute to anti-snake responses in mammals: California ground squirrels, *Spermophilus beecheyi*, flag their tail and kick sand at a rattlesnake, *C. viridis*, more often than at a gopher snake, *P. melanoleucus*. The squirrels kicked sand at and approached a snake in a perforated transparent bag more frequently than one in an intact bag. Visual and chemical cues are important, but the latter seem to be the primary releasers (Henessy and Owings, 1979).

Tenrecs from Madagascar respond to urine, feces, or scent gland material from viverrids by showing distinct behavior patterns. The long-tailed tenrec, *Microgale dobsoni*, displays an open-mouth threat to scents of the Madagascar ringtailed mongoose, *Galidia elegans*. The streaked tenrec, *Hemicentetes semispinosus*, erected its quills and bucked while running toward a cloth impregnated with odor from *G. elegans* or the fanaloka, *Fossa fossa*. The tenrec, *Tenrec ecaudatus*, hissed and erected its hair when exposed to viverrid odors. These tenrecs respond similarly to dogs or humans (Eisenberg and Gould, 1970).

Responses to predator odors raise several questions:

Is the odor detected?
Is it recognized as predator odor?
Is it important information that is used in adaptive behavior?
Is it important energetically (i.e. does it save the animal locomotory energy)?
Is it useful for application as a repellent?
If so, is it an area repellent, feeding inhibitor, or activity depressor?
Can land and wildlife managers use it in assessing predator pressure?

The last three questions will be addressed in Ch. 13.

12.2 Host odors used by parasites

12.2.1 Vertebrates as parasites

Adult sea lampreys, *Petromyzon marinus*, parasitize other fish species. They find brook trout, *Salvelinus fario fontinalis*, and brown trout, *Salvelinus fario trutta*, by odor and can detect 0.014 mg trout extract in water. Trout odor activates sea lampreys even during the rest phase of their 24 hour cycle. When rendered anosmic by nasal plugs, the response disappeared. Even sea lampreys reared in the laboratory in absence of any other potential host fish responded to trout odor, indicating the genetically programmed nature of the response. Of two active amines, isoleucine methyl ester proved to be the most active (Kleerekoper and Mogensen, 1963).

Turning to a *mammalian parasite*, the vampire bat, *Desmodus rotundus*, is very sensitive to butyric acid, a common compound given off by mammals. It detects 0.0039–0.00784 % (vol.) butyric acid. This is lower than the human threshold (Schmidt and Greenhall, 1971).

12.2.2 Vertebrates as hosts

Salmon lice

Salmon lice, *Lepeophtheirus salmonis* (Copepoda) feed on mucus, epithelium, and blood of salmon. The resulting wounds lead to anemia, osmoregulation problems, infections, and disfigured heads. Salmon lice also carry pathogenic viruses and bacteria. Such damage harms the salmon farming industry. Chemical baiting of traps could be a way to control salmon lice. To this end, these copepods were exposed to salmon-conditioned water and extracts from salmon skin, mucus, and flesh, and a number of control substances. Water conditioned by Atlantic salmon, *S. salar*, and turbot, *Scophthalmus maximus*, excited the

salmon lice, but they swam in a directed way only in response to salmon stimuli (Hull, 1997; Devine *et al.*, 2000). 6-Methyl-5-hepten-2-one attracts the larvae of salmon lice to their host (Genna *et al.*, 2004). Furthermore, male salmon lice find females on the salmon host by small lipophilic organic molecules (Mordue-Luntz *et al.*, 2004).

Cattle and the tsetse fly

Many arthropod parasites use butyric acid and/or carbon dioxide emanating from mammals as kairomones. The well-studied tsetse fly, Glossina spp., may serve as an example. Tsetse flies transmit *Trypanosoma* spp., the flagellates that cause sleeping sickness in people, and Nagana sickness in cattle. The flies are attracted to oxen even when these are invisible in underground pits, indicating the chemical nature of the decisive cue (Vale, 1974). The larger the ox, the more flies it attracts (Hargrove and Vale, 1978). Ox odor contains 1-octen-3-ol, acetone, and carbon dioxide. A mixture of these three compounds in their natural ratios is almost as attractive as the total odor from an ox (Vale and Hall, 1985). The effects of carbon dioxide and acetone are additive (Vale, 1980). The three compounds together attract tsetse flies from a long distance, while carbon dioxide alone attracts them into a trap at short range (Vale and Hall, 1985). In a wind tunnel, 1-octen-3-ol triggers upwind flight (Bursell, 1984; Hall et al., 1984). Traps already baited with acetone and 1-octen-3-ol catch more flies if cattle urine is added (Vale et al., 1986). Ox urine also attracts tsetse flies by itself and is used by Kenya tribesmen as bait (Owaga, 1984; Brightwell et al., 1987). The phenolic fraction of cattle urine is particularly active. It contains phenol, 3-methyl phenol, 4-methyl phenol, 3-ethyl phenol, 4-ethyl phenol, 3-propyl phenol, 4-propyl phenol, and 2-methoxy phenol (Bursell et al., 1988). A mixture of 4-methyl phenol and 3-methyl phenol is as attractive to tsetse flies as the total mixture of all eight phenols (Owaga et al., 1988). The diet of livestock affects the active odor: starved oxen are less attractive to tsetse flies but become more so when a fattening diet resumes (Vale, 1981).

A recent study in Ethiopia found the most effective attractant for tsetse flies was acetone, octenol, and cow urine (Belete *et al.*, 2004). An American company markets Mosquito Magnet, a device that purportedly attracts mosquitoes with 1-octen-3-ol, heat, carbon dioxide, and water vapor (Enserink, 2002). Simpler traps are being used in East Africa. A black-and-blue cloth, impregnated with insecticide, attracts mosquitoes with acetone and octenol, or alternatively, buffalo urine (Enserink, 2002).

Urine of a wild host bovid, the African buffalo, *Syncerus caffer*, also attracts tsetse flies. Up to eight phenols are active (Hassanali *et al.*, 1986). The phenols are the

same as those in cattle urine described above (Bursell *et al.*, 1988). The phenolics in urine of cattle and buffalo are formed by microbes over several days. Assuming that the hosts move on, tsetse cannot find their hosts by means of (fresh) urine. Urine aged in vessels without soil attracts more tsetse than if soil is present. It is thought that glandular secretions, "processed" by microbes on the body of the host mammals, attract tsetse flies (Madubunyi *et al.*, 1996).

Rodents and fleas

Fleas (Siphonaptera) typically ectoparasitize small to medium-sized burrowing mammals. They find their hosts by odor. The fleas employ behavior that is adapted to the ecology and social behavior of the particular rodent species. For example, in the Negev highlands of Israel, the gerbil *Gerbillus dasyurus* is almost exclusively parasitized by the flea *Xenopsylla dipodilli*, while the spiny mouse, *Acomys cahirinus*, harbors almost exclusively the flea species *Parapulex chephrenis*. In laboratory experiments, odors of whole animals attracted each flea species to its specific host. Moreover, *X. dipodilli* started to approach its gerbil host sooner, and all animals responded, while *P. chephrenis* took longer to start moving, and only 55% of the animals chose an *Acomys* host. This reflects differences in host ecology: the gerbil is solitary and as many as 50% of the animals do not have their own home range and burrow. Therefore, *X. dipodilli* must search for and approach its host more actively. By contrast, the spiny mouse largely nests communally and most animals reside in their burrow for a long time. Accordingly, *P. chephrenis*, its parasite, follows a sit-and-wait strategy (Krasnov *et al.*, 2002).

Humans and mosquitoes

Heat, moisture, and carbon dioxide attract mosquitoes to warm-blooded animals. Mosquitoes that single out humans, such as *Anopheles gambiae* and *Aedes aegypti*, need more specific compounds for host specificity. Incidentally, *A. gambiae* is also attracted by Limburger cheese. Fatty acids are common to cheese and human foot odors, and bacteria of the genus *Brevibacterium* produce fatty acids in both cheese and between human toes. Mosquitoes vary in their preferences: ammonia attracts *A. gambiae*, but *A. aegypti* relies more on lactic acid (Enserink, 2002).

Humans and blood flukes (Trematoda)

Cercariae of the blood fluke (or schistosome) *Schistosoma mansoni* are attracted to humans first by temperature and light. The first signal that

facilitates brief attachment to human skin is arginine. Arginine is present in trace amounts on human skin. At the same time, the cercaria's tail is rich in arginine. The cercaria discards its tail and leaves it on the skin surface. So this amino acid may originate from human skin or other cercariae, being kairomone and/or pheromone. The second and third signals are two different mixtures of skin lipids. The third signal is required for penetrating the skin (perceived as "swimmer's itch"), shedding the tail, and transformation from a free-living freshwater form to an endoparasite. Cercariae are even attracted to a dab of human skin lipids applied to the wall of a glass dish. They try to penetrate the glass and actually shed their tails. They even perform this "suicidal" behavior to the lipid cue dispersed in the water (e.g. Feiler and Haas, 1988). This has led Haas to propose that cercaria-infested waters should be sprayed with lipids. The cercariae would then self-destroy.

Humans and leeches

The leech *Hirudo medicinalis* responds to human blood or plasma by a sequence of probing, attachment to skin, biting, and ingestion of blood. Spring water at 38 °C releases probing and transient attachment, but not ingestion. Sodium chloride plus arginine is an effective feeding stimulus, sodium chloride alone triggers probing only, and no feeding takes place. Likewise, arginine alone does not lead to feeding. The ions in the salt can be substituted. Lithium ions are as affective as sodium ions but only bromide was as effective as chloride ions. The amino acid is highly specific: D-arginine does not lead to ingestion even in concentrations up to 1000 times the threshold for L-arginine. Other common L-amino acids such as histidine or lysine are not active. Finally, of the arginine analogues, only homoarginine and canavanine were effective feeding stimuli. These two compounds have all three functional groups of arginine intact (Elliot, 1986).

12.3 Eavesdropping

12.3.1 Pheromones as kairomones

Vertebrates using vertebrate signals

In laboratory tests, least weasels, *Mustela nivalis*, were more attracted to estrous than diestrous urine of prairie deer mice, *Peromyscus maniculatus bairdii*. This may increase the weasel's preying chances through the presence of pups in the case of postpartum estrus or estrous females may indicate a higher population density (Cushing, 1984).

FIGURE 12.4 Skatole, a prey odor used by snakes.

Canids such as wolves or dogs who track their prey most likely eavesdrop on intraspecific signals, such as deposits from interdigital glands in cervids and bovids, or urine marks of rodents.

Finally, the responses of grizzly bear, *Ursus arctos* and polar bear, *Ursus maritimus*, to human menstrual odors during attacks on campers (Cushing, 1983) appears to be a form of eavesdropping, even though the intraspecific function of the odor in humans is not yet understood.

Vertebrates using invertebrate signals

Blind snakes find their prey by using the prey's pheromones. The worm-like Texas blind snake, *Leptotyphlops dulcis*, of the southwestern United States leads a subterranean life and feeds on termites and ant brood. It finds ants by following their pheromone trails (Gehlbach *et al.*, 1968). Other blind snakes such as the American blindsnake, *Typhlops pusillus* (Gehlbach *et al.*, 1971), and the Australian blindsnake, *Ramphotyphlops nigrescens* (Webb and Shine, 1992), also follow odor trails of ants, their prey. Texas blind snakes are attracted to the simple alkaloid *skatole* (methyl indole; Fig. 12.4), an amine with an unpleasant odor from the ant *Neivamyrmex* sp. (Watkins *et al.*, 1969).

The wax pheromones of honeybees that honey guides use as feeding stimuli have been discussed above (p. 352).

Invertebrates using vertebrate signals

The klipspringer, *Oreotragus oreotragus*, is an antelope of the rocky hills (*kopjes*) of eastern and southern Africa, where it is the ecological equivalent of the ibex or chamois. This animal marks its home range by applying secretion from the preorbital glands to blades of grass. Rain washes scent material down the stem, creating a gradient along an odor track. The tick *Ixodes mentzii* (or *I. matopi*) crawls up the stem, following this odor gradient, and then waits at the tip of a grass blade. Now the parasite is ready to transfer to another klipspringer that is

attracted to the scent mark and touches the stem while marking in turn (Spickett et al., 1981).

12.3.2 Hormones as kairomones

Host hormones may act as kairomones and attract or otherwise benefit parasites. The female rabbit flea, *Spilopsyllus cuniculi*, has to eat corticoidladen blood from a pregnant rabbit before its own eggs can mature (Rothschild, 1965).

12.4 The self-anointed: chemical mimicking

Various reptiles, birds, and mammals treat their own bodies with odoriferous or toxic secretions and excretions from other species (Table 12.8). The benefits of these acquired odors are not well understood. In small prey animals, such as rodents or insectivores, an antipredator function seems plausible. Predators may self-anoint to cover up their own odor to improve their ability to approach prey. A second function for carnivores may be to transfer information about food to members of their own group. For this to be likely, the behavior should be more prevalent in social species. Self-anointing is also thought to deter ectoparasites and/or microbial pathogens. Passive anointing is mere exposure to chemicals of other species, while in active anointing an animal actively applies scents to its own body. Using such acquired odors represents an "extended phenotype" (Weldon 2004). No direct interaction between the odor donor and the target animal, such as a predator or parasite, is necessary. Indeed, as Weldon (2004) pointed out, the donor and the target may be allopatric, not even occurring in the same area.

Young Komodo monitors, *Varanus komodoensis*, roll themselves in the feces of prey animals. This may protect them from predation by larger conspecifics, as these avoid gut contents of their prey by shaking them out before eating the prey (Auffenberg, 1981; Ciofi, 1999).

Among the canids, dogs, wolves, coyotes, and foxes roll in dung and carcasses and other materials such as scent glands of rattlesnakes (Weldon and Fagre, 1989).

The chipmunk, *Eutamias sibiricus asiaticus*, approaches a dead snake, gnaws its skin, chews the gnawed bits, and finally applies them to its own fur. Snake urine, feces, the rectum end section, cloaca, and cloacal sacs release the same behavior. Snakes eat dead mice treated externally with urine from the Japanese rat snake, *Elaphe climacophora*, less readily than untreated controls. This suggests that chipmunks conceal their own odor by self-anointing, reducing predation (Kobayashi and Watanabe, 1986). Chipmunks may rub off the acquired odor on their burrow

Species	Material	From species	Context/function	Reference
Hedgehogs Erinaceus europaeus, Atelerix pruneri, Hemiechinus auritus	Venom	Toad	Self-defense	Brodie, 1977
Siberian chipmunk Eutamias sibiricus asiaticus	Carcass	Snake	Self-defense	Kobayashi and Watanabe, 1986
Rice rat Rattus rattoides	Anal gland secretion	Weasel Mustela sibirica	Self-defense	Xu et al., 1995
Dog	Dung, carcass	Various: cattle	Unclear	Common observation
Capuchin monkey cebus sp.	Benzoquinones	Millipede	Mosquito repellent	Valderrama et al., 2000; Weldon et al., 2003

Table 12.8 Self-anointing in mammals

entrance. Thus the material passes from snake to chipmunk to burrow to predators – snake or polecat – who may come along.

The rice rat, *Rattus rattoides*, from southern China also self-anoints, at least in the laboratory. Presented with filter paper carrying anal gland secretion from the weasel *Mustela sibirica*, rice rats chewed the paper and rubbed the pieces against both of their flanks. Even laboratory-born naive rice rats anointed themselves in this fashion. Weasel or fox urine did not release anointing behavior (Xu *et al.*, 1995).

Among insectivores, at least five genera of hedgehogs anoint themselves with toad toxins. Although the exact adaptive function of this behavior is not known, it has been observed that the anointed spines hurt humans more than clean ones (Brodie, 1977).

Capuchin monkeys (*Cebus* spp.) in Venezuela self-anoint with a benzoquinone-secreting millipede. Two of the compounds by themselves also release self-anointing. These are 2-methyl-1,4-benzoquinone and 2-methoxy-3-methyl-1,4-benzoquinone. These compounds repel mosquitoes (Valderrama *et al.*, 2000; Weldon *et al.*, 2003).

12.5 Evolutionary considerations

12.5.1 Ghosts from predation past: history of association of prey and predator

Responses of prey to predator cues depend on predator-prey association in evolutionary time. Nevertheless, some rodents avoid the odor of a predator

Prey species	Predator species	Location	Reference
Orkney vole Microtus arvalis orcadensis	Red fox Vulpes vulpes	Orkney Islands	Calder and Gorman, 1991
Lizard Podarcis hispanica atrata	Stoat Mustela erminea Viper Vipera latastei	Orkney Islands Spain: Isla Columbreta Grande	Gorman, 1984 van Damme and Castilla, 1996

Table 12.9 Ghosts of predation past: atavistic responses to chemicals of former predators

even if the two species have not been sympatric for many generations. For instance, the lizard *Podarcis hispanica atrata* from the Spanish island Columbreta Grande has not experienced its predator, the viper *Vipera latastei* since the snake's eradication in the 1880s. The lizard responds still to odors of the viper as much as another subspecies, *Podarcis hispanica*, from the mainland where vipers occur today (van Damme and Castilla, 1996).

Deer mice, *P. maniculatus*, on an island (Moresby Island, Gulf Islands, British Columbia) that now lacks their usual predator, the short-tailed weasel, *M. erminea*, still respond to its odor. However, they only show the more delayed and prolonged stress-type, opioid-sensitive behavior. By contrast, this island population has lost its fear and flight responses, which are benzodiazepine sensitive and more immediate. Mainland deer mice that are sympatric with weasels show both types of response (Kavaliers, 1990).

Similarly, on an island in the Orkneys, the vole *Microtus arvalis orcadensis*, introduced there by neolithic settlers between 3700 and 3400 BC, still avoids the anal gland secretion of the stoat *M. erminea*, even though the island has lacked predators for 5000 years (Gorman, 1984). Likewise, the Orkney vole avoids fecal odors of red fox, *Vulpes vulpes*, in live traps in the field and the laboratoy. Moreover, the vole almost never eats bark from Scots pine (*Pinus sylvestris*) saplings treated with fox feces extract (Calder and Gorman, 1991). Avoidance of red fox odor by the Orkney vole is an example of a "ghost from predation past" (Peckarsky and Penton, 1988) (Table 12.9).

12.6 Plant chemicals used by vertebrates

12.6.1 Aromatic plants used by birds

Even bird species with smaller olfactory bulbs respond to odors more than previously assumed. Starlings, *Sturnus vulgaris*, for instance, olfactorily discriminate species of aromatic plants they place in their nests. Three lines of

evidence support this First, six plant species evoked strong multiunit electrophysiological responses in the olfactory nerves of adult birds. Second, if plant odors were paired with gastrointestinal malaise caused by the bird repellent methiocarb, a conditioned avoidance was produced. Third, birds with sectioned olfactory nerves did not develop the avoidance (Clark and Mason, 1987). Experiments showed that aromatic herbs in the nest do not reduce the ectoparasite load in starlings but may stimulate those parts of the immune system that mitigate effects of ectoparasites (Gwinner *et al.*, 2002).

Corsica blue tits, *Parus caeruleus ogliastrae*, select 6–10 species of aromatic plants out of about 200 species that occur in their habitat. They add them to their nests during the nestling stage. An ingenious experiment demonstrated that the blue tits maintain an aromatic environment around their nest. Containers filled with invisible aromatic herbs and attached to the nest box reduced the collecting of aromatic plants. Gas chromatographic analysis of volatiles from one plant species (*Achillea ligustica*) showed a rapid loss within 24 hours. Accordingly, the birds started to replenish their aromatic plants about 1 day after the experimenters had placed fresh aromatic plants near the nest. The birds selected aromatic plants that contain proven antibacterial, antiviral, fungicidal, insecticidal, and insect-repellent compounds. These include linalool, camphor, limonene, eucalyptol, myrcene, terpin-4-ol, pulegone, and piperotenone. Moreover, 40% of the plants involved possess activity against the human immunodeficiency virus (Petit *et al.*, 2002).

12.6.2 Regulators of mammal reproduction

Rodents in unpredictable environments such as deserts or alpine and polar habitats cannot easily rely on photoperiod for starting their annual breeding. Instead, many species rely on fresh vegetation to start reproducing. This, in turn, depends on rainfall in deserts and the spring thaw in the mountains and tundra. Reproduction in the montane vole, *Microtus montanus*, of western North America is triggered by (6-MBOA; 6-methoxybenzoxazolinone Fig. 12.5), a cyclic carbamate found in seedlings and growing tissues of grasses and some dicotyledonous plants (Berger *et al.*, 1981; Sanders *et al.*, 1981). The two separate effects of 6-MBOA on reproduction are cueing to time the onset of reproduction and affecting the quality of the reproductive effort. The latter is basically uterotropic in that it stimulates an increase in uterine mass. When given 6-MBOA, montane voles have larger litter sizes and more litters. If both sexes received 6-MBOA, 25% more females than males are produced (Berger *et al.*, 1987). Experimental administration of 6-MBOA to non-breeding winter populations stimulates breeding in montane voles (Berger *et al.*, 1981) and Townsend's

FIGURE 12.5 6-Methoxybenzoxazolinone. This compound occurs in young plants and stimulates reproduction in mammals.

vole, *Microtus townsendii* (Korn and Taitt, 1987). In the woodland vole, *Microtus pinetorum*, 6-MBOA increases uterine and ovarian mass and gonadotropin activity (Schadler *et al.*, 1988).

Ovarian weight is also increased in the laboratory mouse by 6-MBOA (Sanders *et al.*, 1981), and the laboratory rat shows similar responses to those of the woodland vole (Butterstein *et al.*, 1985). Even in predictable environments, such as salt marshes, 6-MBOA may regulate reproduction. The breeding cycles of North American microtine voles, which feed almost exclusively on salt grass, *Distichlis stricta*, parallel the growing season of that grass. It has high levels of 6-MBOA when growing, and little when senescing (Negus *et al.*, 1986).

Ord's kangaroo rat, *Dipodomys ordii* (Heteromyidae), also responds to 6-MBOA. This usually granivorous species of western North America can also consume up to 40–60% of green plants. It becomes reproductively active after rainfall, which nourishes desert annuals. Peritoneal injections of 6-MBOA increased uterus mass in this kangaroo rat. In the field, providing 6-MBOA-treated rolled barley near burrows resulted in more pregnant females. Mere excess food (untreated barley) did not affect reproduction. Litter size was not affected. This compound is but one factor of a complex of stimuli that regulate reproduction in this kangaroo rat (Rowsemitt and O'Connor, 1989). Uterus and ovary weights of Merriam's kangaroo rat, *Dipodomys merriami*, did not change after peritoneal injection of 6-MBOA. This desert animal of western North America may not rely on 6-MBOA as a reproduction trigger; the compound may not even be present in the desert vegetation (McClenaghan, 1987). Table 12.10 summarizes the effects of 6-MBOA in mammals.

Steroids or steroid-mimicking plant compounds affect mammalian reproduction. After giving birth, female muriqui monkeys, *Brachyteles arachnoides*, eat leaves that contain isoflavonoids, which structurally resemble estrogen (see Ch. 10). This might reduce fertility. When the females are ready to reproduce, they consume more "monkey ear," a legume containing a steroid that might boost fertility (Strier and Ziegler, 1994).

1		,	
Mammal species	Diet	Effect	Reference
Montane vole Microtus montanus	Grasses, some dicotylons	Timing reproduction; inceased uterine mass: larger and more litters	Berger et al., 1981; Sanders et al., 1981
Townsend's vole, <i>Microtus townsendi</i> and montane vole	6-MBOA on rations	Winter breeding	Berger <i>et al.</i> , 1981; Korn and Taitt, 1987
Woodland vole Microtus pinetorum	6-MBOA	Increased uterine and ovarian weight; more gonadotropin	Schadler <i>et al.</i> , 1988
Laboratory mouse	6-MBOA	Increased ovarian weight	Nelson and Shiber, 1990
Laboratory rat	6-MBOA	Increased uterine and ovarian mass; more gonadotropin	Butterstein et al., 1985
Ord's kangaroo rat Dipodomys ordii	Green plants	Reproduces after rainfall	Rowsemitt and O'Connor, 1989
	Injected 6-MBOA	Increased uterus mass	
	6-MBOA on rolled	More pregnant females	

Table 12.10 Reproductive effects of 6-methoxybenzoxazolinone in rodents

6-MBOA, 6- methoxybenzoxazolinone.

barley in field

12.6.3 Self-medication

Chimpanzees, *Pan troglodytes*, take up leaves of certain plants and massage them in their mouth between the tongue and the buccal surface. The leaves are swallowed whole without chewing. Whole leaves have been found in chimpanzee feces. The plants include several species of *Aspilia* (Compositae: sunflower family), *Lippia plicata*, *Commelina* spp., and *Ficus exasperata*. They are eaten early in the morning, while most of the regular feeding occurs in the afternoon. The selected plants are widely used in medicine (reviewed by Newton and Nishida, 1990). *Aspilia mossambicensis* contains thiarubrine A, an antifungal, antibacterial, and antihelminthic compound with the appearance of a red oil (Wrangham and Nishida, 1983; Rodriguez *et al.*, 1985). The two diterpenes kaurenoic acid and grandiflorenic acid from the same plant species are strongly uterotonic, and also antibacterial and antihepatotoxic (Page *et al.*, 1992). Chimpanzees carry a heavy load of nematodes, and this plant may control these. An anorexic adult

female chimpanzee with a presumed gastrointestinal disorder sucked out the bitter juice from the piths of shoots of the composite Vernonia amygdalina, a plant usually not eaten by chimpanzees. This is used in tropical medicine to treat parasitoses and gastrointestinal disorders in humans and domestic animals (Huffman and Seifu, 1989). Chimps in the Mahale Mountains National Park in Tanzania use this plant. Four known sesquiterpene lactones have been isolated, plus seven new steroid glucosides, and two aglycones of these glucosides. In vitro, the sesquiterpene lactones are antischistosomal, plasmodicidal, and leishmanicidal. The major steroid glucoside, vernonioside B₁, was also antischistosomal. The glycosides inhibit schistosome movement and egg laying most when the sugar moiety is eliminated. Vernodalin was the most potent antiparasitic compound in vitro, but proved toxic in vivo to cercariae-infected mice. The pith, chewed by chimps, contains steroid-related compounds, but vernodalin was found only in bark and leaves. Vernodalin and other sesquiterpene lactones may be too toxic for chimps. During the rainy season (mid-October to December) parasite diseases are more prevalent, and levels of vernonioside B increase in stems and leaves yet chimps avoid the leaves (Ohigashi et al., 1994). Chimps possibly practice preventive medicine: they eat more medicinal plants during the rainy season when pneumonia threatens. The chimps may use the forest as a medicine cabinet. After these observations, a new subdiscipline, zoopharmacog*nosy* was born. It held its first symposium in 1992.

Capuchin monkeys rub millipedes on their fur and skin to repel insects. Benzoquinones are the active principle. These compounds deter mosquitoes from feeding on human blood in the laboratory. Even captive capuchin monkeys seek benzoquinone-soaked tissues when these are offered by experimenters (Weldon *et al.*, 2003; see also p. 268 and p. 377).

12.6.4 Food preservatives

Pikas, Ochotona princeps, of the North American Rocky Mountains store winter food in "hay piles." An individual collects about 28 kg of material. One plant species, Geum (Acomastylis) rossi, constitutes up to 75% in some piles. It is almost never eaten in summer and contains considerable levels of phenolics. Geum extracts are bacteristatic. Experimental hay piles of this plant retained their biomass better than controls. The pika perhaps uses the phenolics to preserve its food for the mountain winter, lasting several months. Over time, the food loses most of the phenolics and becomes palatable (Dearing, 1997).

12.7 Animal chemicals benefiting plants

12.7.1 Saliva

Saliva of herbivorous mammals and insects (specifically grasshoppers) can stimulate growth of clipped vegetation. Grazers influence pastures not only mechanically but also biochemically via thiamine in saliva. Thiamine in saliva stimulates growth in higher plants when temperatures are low and ascorbic acid is present (Vittoria and Rendina, 1960). Jameson (1964) proposed that thiamine should be applied to partially defoliated plants. In later greenhouse experiments, clipped plants of the grass sideoats grama, Bouteloua curtipendula, were treated with bovine saliva, collected from the mouth. This treatment increased forage yields, root yields, and basal area. Thiamine applied to the soil increased the forage yield by as much as 232%, and root yields by 398%. In the field, the increases were less pronounced. These results suggested that the native rangelands may require grazers for vigor and production (Reardon et al., 1972). Sideoats grama grazed by cattle, sheep, or goats regrew significantly more (72.5 cm) than clipped plants (55 cm). Thiamine added to the soil increased forage yield by as much as 42%, but bovine saliva did not affect growth (Reardon et al., 1974).

Saliva and especially urine contain epidermal growth factor, a small protein, which may be very important to stimulate plant growth (Dyer, 1980). Epidermal growth factor from mouse urine stimulates growth in sorghum seedlings and the effect is proportional to the amount used.

Other studies used saliva after it had reached the rumen. This saliva had no effect on plant growth (Johnson and Bailey, 1972; Detling *et al.*, 1980). The effect of thiamine depends on the extent of "grazing": thiamine stimulated growth in clipped grasses as long as they retained 25% of their height (McNaughton, 1985).

Whole saliva from the North American bison, *Bison bison*, stimulated elongation of *Avena* coleoptiles at high pH. Sodium bicarbonate from saliva enhanced growth in a dose-dependent fashion and, therefore, inorganic ions were thought to be the main growth-promoting factor in bison saliva rather than organic compounds (Detling *et al.*, 1981).

12.8 Synomones

Chemicals mediating relationships where both partners benefit are termed synomones.

Tryptamine

FIGURE 12.6 Synomones attracting anemone fish to its sea anemone partner. Amphikuemin attracts the anemone fish *Amphiprion perideraion* to its sea anemone partner *Radianthus kuekenthali*. Tyramine and tryptamine attract the anemone fish *Amphiprion ocellaris* to its anemone partner *Stoichactis kenti*.

12.8.1 Fish

Anemone fish

The relationship between anemone fish, *Amphiprion* sp., and their sea anemone partners are regulated by alkaloids from the sea anemone. *Amphiprion perideraion* is attracted to the sea anemone *Radianthus kuekenthali* by the simple lysine-derived alkaloid amphikuemin (Fig. 12.6; Murata *et al.*, 1986). Other simple alkaloids, aplysinopsin and dihydroaplysinopsin, also attract *A. perideraion*, regulate its swimming rate and induce their species-specific partnership. A related anemone fish, *Amphiprion ocellaris*, is attracted to the sea anemone *Stoichactis kenti* by tyramine (Fig. 12.6) and tryptamine induces searching by the fish (Murata *et al.*, 1986).

12.8.2 Reptiles, birds and mammals

Fruit flavor, consumption and seed dispersal by birds

Many fruits attract birds by colors, odors, and taste. After consuming the fruit, the birds will disperse the seeds to new habitat. Preference orders exist, and some fruits are rarely eaten by birds (see p. 306). For instance, the flavor of buckthorn, *Rhamnus cathartica*, ranked lowest of 11 fruit extracts tested in blackbirds, *Turdus merula*, and a song thrush, *Turdus philomelus* (Sorensen, 1983). Least preferred fruits contain toxins that deter birds, perhaps to avoid seeds being dispersed to unfavorable habitat. For instance, it is disadvantageous for forest

plants to be dispersed by open-country birds to unsuited open habitat (Janzen, 1978; Sorensen, 1983).

Passage of seeds through animals

Fruit- and seed-eating birds and mammals not only disperse seeds but can also mechanically and chemically weaken or break the seed coat. The chemical alteration of seed coats by animal digestive tracts can affect germination rates and patterns, often benefiting the plant in the process. Since this chemical plant—animal interaction takes place in the digestive system, the chemical senses are not directly involved, although the first step, ingestion, is largely guided by olfaction and taste.

Earlier it was thought that plants and animals might have coevolved, the plant depending on seed conditioning by seed-eating birds or mammals. A case in point was the idea that the Tambalacoque tree, *Calvaria major*, on Mauritius, which might have depended on the now extinct dodo for germination. The last dodo died in 1681 and no trees seemed to have germinated from seed during the intervening 300 years (Temple, 1977). To test whether a large bird can enhance germination, Temple force-fed turkeys with 17 of the 5cm diameter seeds of *Calvaria*. Of the 10 seeds that survived the digestive tract, three germinated. Temple claims that "these may well have been the first *Calvaria* seeds to germinate in more than 300 years." However, more trees have been found since, and the dodo's elevational range was probably not identical with that of *Calvaria* (Owadally, 1979).

Perhaps not by coincidence, another island provides a striking example. On Galapagos, less than 1% of the seeds of the tomato *Lycopersicon esculentum* var. *minor* germinate by themselves. By contrast, the germination rate of tomato seeds fed to two Galapagos tortoises from Indefatigable Island rose to 60–81%, after a 14 to 18 day passage through their digestive tract (Rick and Bowman, 1961).

Even Fish affect the germination rate of seeds that they eat; for example, only 0.5% of untreated seeds of beaked tasselweed, *Ruppia maritima*, and Spiny naiad, *Najas marina*, germinate. Passage through grass carp, *Ctenopharyngodon idella*, increased germination to 5.7% and 10.5%, respectively, and passage through tilapia, *Oreochromis* sp., to 12% and 16% respectively (Agami and Waisel, 1988).

Passage through birds or mammals conditions seeds in several ways. Passage time varies from 30 minutes in some songbirds to up to 60 days in horses (Table 12.11). Seeds may not necessarily be affected, especially during short passage times, and at the other extreme, they may be completely destroyed, as by the

Class	Species	Passage time	Reference
Fish	Tilapia; grass carp	30–60 hours	Agami and Waisel, 1988
Reptiles	Galapagos tortoise	11–12, up to 20 days	Rick and Bowman, 1961
Birds	Turkey	Up to 6 days	Temple, 1977
	Hermit thrush	30 minutes	Krefting and Roe, 1949
	Phainopepla	12–45 minutes	Walsberg, 1975
Mammals	Cattle	2–3, up to 10 days	Peinetti <i>et al.</i> , 1993
	Horse	14 (75%), up to 60 days	Janzen, 1981

Table 12.11 Time of seed passage through vertebrate digestive tracts

gizzards of birds, notably gallinaceous birds. Between these extremes lies a continuum. For some plants, only a few seeds remain viable after passing through digestive tracts. For instance, only 7 of 40025 ingested seeds from various plants germinated after passage through the California linnet (Roessler, 1936). About 5 to 10% of the ingested seeds remained viable in several studies.

Birds

Seeds of Tatarian honeysuckle, *Lonicera tatarica*, germinate in 33 days after passing through robins, *Turdus migratorius*, while untreated seeds take 43 days and those passed through the catbird, *Dumetella carolinensis*, take 47 days (Krefting and Roe, 1949). Table 12.12 lists how seven eastern Mediterranean scrubland birds (Izhaki and Safiel, 1990) and pheasants (Swank, 1944) conditioned seeds of various Mediterranean plants. Germination probability rose, dormancy was shortened, and germination time pattern of the seeds became variable.

Mammals

In mammals, gazelles, cattle, and grizzly bear increased germination probability of some seeds (Table 12.13). Elephants enhance germination and seedling survival of the marula plum tree, *Sclerocarya caffra* (Anacardiaceae) (Lewis, 1987). Certain tropical seeds germinate only if passed through the digestive system of monkeys (Hladick and Hladik, 1969).

Many investigators agree that mammalian, and bird, digestive tracts condition seeds to different degrees, especially if several animal species prey on the seeds. In unpredictable environments, such as deserts with little rain, there will always be some seeds ready to germinate. To conditioning must be added the requirement for dispersal, common to all surviving seeds, regardless of any mechanical and chemical abrasion of the seeds. In mainland ecosystems, many plant and animal species interact, so that coevolution between plants and

Table 12.12 How avian frugivores affect seed germination

Reference	chemical 1990 decomposition of seed coat	u, uigotive owann, 1944
Process	Abrasion and chemical decompositi	juices"
Temporal pattern Process of germination	Variable, bird-dependent in 4 spp.	
Probability of Length of dormancy germination	Decreased in all 5 spp. Variable, to 5–90 days bird-dep in 4 spp.	germinated in 14–35 days (control: 38 days)
Probability of germination	Increased in 4 of 5 spp.	(control: 20%)
Plant species	Five Mediterranean Increased in 4 Species of 5 spp.	nightshade, clover, black locust
Bird species	Seven eastern Mediterranean scrubland frugivores ^a	colchicus

ND, No differences.

^a Blackbird (Turdus merula), song thrush (Turdus philomelos), bulbul (Pycnonotus barbatus), blackcap (Sylvia atricapilla), orphean warbler (Sylvia hortensis), lesser whitethroat (Sylvia curruca) and redstart (Phoenicurus phoenicurus).

 Table 12.13 How mammalian frugivores affect seed germination

Mammal species Plant species	Plant species	Germination probability	Length of dormancy	Process	Function	Reference
Bat Rousettus aegyptiacus	Fig Ficus carica	ND	ND	Inhibitors stay in feces, chemical and physical gut processes	† Asynchronous G spreads risk (adaptation to desert rain)	Izhaki et al., 1995
	Mulberry <i>Morus nigra</i>	ND	↑G delay			
	Arbutus Arbutus andrachna	ND	↑G delay			
Dorcas gazelle	Acacia (3 spp.)	† to 13% (C: 1%); † to 21% (C: 4%)	<pre>\ Acceleration: 7% / 5days; 13% / 10 days</pre>	Bruchid beetle opens seed	Dispersal; adaptation to desert rain pattern	Or and Ward, 2003
Cattle	Caldén Prosopis caldenia	† during first month	↑ G delay	Seed scarification	↑G under varying environmental and site conditions	Peinetti <i>et al.</i> , 1993
Grizzly bear	Cow parsnip Heracleum lanatum	↑ to 65% (C: 51%); frozen: ↑ to 85% (C: 69%)	NT	NT	Dispersal	Applegate et al., 1979

ND, no difference. C, control. G, germination; \uparrow Increase; \downarrow , Decrease; NT, not tested.

seedeaters is less likely than on islands. Relationships among several trophic levels can grow complex. For example, the numbers of agouti and paca on Barro Colorado Island in Panama are over 10 times greater than in a comparable area in Amazonian Peru. Barro Colorado Island is too small (16 km²) to support large predators such as pumas or jaguars. The lack of predators increased the abundance of some plants, while that of others decreased. Seed predators such as peccaris, pacas, and agoutis will particularly affect numbers of seedlings of large-seeded plants (Terborgh, 1988). In general, passage through vertebrate guts enhances seed germination more in temperate zone plant species than tropical ones, and more often in trees than in herbs and shrubs (Traveset, 1998).

Animals can even affect germination without passing seeds through the digestive system. Heteromyid rodents select certain types of seed, hull them, and collect them in their external, hairy pouches; in *Oryzopsis hymenoides*, a perennial grass of western North America, 31% of seeds germinated after hulling by two *Dipodomys* species whereas only 7% of intact seeds germinated (McAdoo *et al.*, 1983).

The pig and the truffle

The highly priced hypogeous Périgord truffle, Tuber melanosporum, occurs in Spain, France, Italy, and Greece. It is commercially grown there in oak and hazelnut woods. The fungus produces an odor that humans can detect as it emanates through cracks in the ground. Traditionally, dogs and female pigs, and sometimes goats, have been used to find the truffles. The truffle produces the steroid androstenol (5α -androst-16-en- 3α -ol) (Claus et al., 1981). The same compound occurs in the saliva of the domestic boar and stimulates mating behavior in the sow. Truffles have had a reputation as an aphrodisiac for many years (Harris, 1987). In short, the truffle attracts the female pig with an odor that mimics the sex pheromone of the boar. Since the animal uses the "prey" odor to its advantage, a kairomone is involved. Undoubtedly, the truffle also benefits, probably by spore dispersal. This would make the steroid a synomone. Insects also respond and serve as indicators: Some human truffle hunters find truffles by lying on the ground, and observing the black-and-yellow truffle flies that hover over the invisible truffles, enter the ground, and lay their eggs in mature truffles. Truffles also produce chemicals that kill plants above them, resulting in a patch of "scorched earth," terre brûlée in French. It, too, helps in detecting these prized fungi.

Bats, flowers and fruits

Bat-pollinated flowers have "musky" odors, resembling that of bats or fermentation. These flowers contain butyric acid, which smells to humans similar to bat gland odors (Kulzer, 1961). The flower aroma may possibly resemble a bat pheromone, and the bat may be sexually attracted to the flower and pollinate it in the process (Baker, 1963). Fruit bats (Macrochiroptera) use odors to locate and discriminate food. Egyptian fruit bats, *Rousettus aegyptiacus*, discriminated boxes containing 100 mg banana from empty boxes. They also distinguish natural from synthetic banana odor. Frugivorous bats were attracted to odors in a field experiment in Costa Rica's Corcovado National Park. Mist nets were baited with bananas wrapped in plastic, unwrapped, or smeared on leaves. Fourteen species of phyllostomid bats were caught. Smeared banana attracted more bats of the genus *Artibeus* and the family Stenodermatinae. In rain, bats did not discriminate. It was concluded that "olfactory ability may provide a mechanistic link between plant dispersal strategies and chiropteran foraging strategies" (Rieger and Jakob, 1988).

The short-tailed fruit bat, *Carollia perspicillata*, discriminates the odor of ripe from that of green bananas. Odor cocktails of banana odor components were distinguished from the whole odor of banana (Laska, 1990). The two most common esters of banana odor are isopentyl acetate and isobutyl acetate. Nearly all fruit odors contain ethyl acetate. Isopropyl acetate and 2-pentanone are characteristic of putrefying fruits.

Mammals and fungal spores

Small mammals often smell like the fungi they eat. They probably benefit the fungi by dispersing their spores. A strong coevolution between the fungi and these mammalian fungivores has been suggested (Maser *et al.*, 1978).

Practical applications of semiochemicals

Give me an ounce of civet, good apothecary, to sweeten my imagination SHAKESPEARE: King Lear Act IV, sc. 6, 1.132

Smelly secretions as repellents: "The devil can be completely undone if you manage to fart into his nostrils." (beating the devil with his own weapons).

MARTIN LUTHER (Erik Erikson: Young Man Luther, p. 61-62)

Chemical cues hold considerable promise for manipulating behavior in vertebrates, provided we understand an animal's natural history, biology, and behavior well. However, the development of chemical attractants, stimulants, inhibitors, and repellents for vertebrates has progressed rather slowly for several reasons. First, chemical stimulus and behavior are not connected as rigidly as in insects, for example. Second, the same stimulus may elicit different behaviors, depending on the state of the recipient and the context. Third, chemical cues often are rather complex mixtures of compounds. Fourth, learning, especially early experience plays a major role in vertebrate, notably mammalian behavior. Finally, many behaviors are modulated by several sensory modalities so that chemical stimuli alone trigger only incomplete responses at best.

13.1 Fish

The desire for improved techniques in fishing, aquaculture, and limnological management motivates much of the current basic work in the chemical ecology of fish.

13.1.1 Chemical imprinting

Hatcheries raise fish and release them in lakes and rivers on a large scale. Some species such as salmon imprint early in life on the odor of their home stream. Coho salmon imprint when just over a year old (smolt stage), and sockeye salmon even earlier, right after hatching. Coho salmon prepare to migrate from rivers to the ocean as smolt. Proper imprinting ensures their return as adults to the home stream in order to spawn. Morpholine-imprinted Coho salmon have been successfully attracted to the Mad River in California by adding morpholine to the water (Hassler and Kucas, 1988). Sockeye salmon leave their streams around lakes just a few months after hatching and develop further in a lake where they smolt before they leave for the sea.

The goal is to imprint fish to be released properly so that they do not scatter into other, wild populations and interfere with wild gene pools. Now it is possible to assess whether imprinting has taken place. Fish imprinted on 2-phenylethanol (phenethyl alcohol) have a higher percentage of neurons that respond to this compound in their noses, and these neurons are more sensitive to this compound. Thus, learning leaves its traces in the nose as well as the brain. The sensitive neurons make more cyclic GMP, an intracellular messenger. Changes such as these may be a suitable basis for assays to ascertain whether hatchery-raised fish have been properly imprinted before they are released into the wild (Barinaga, 1999).

13.1.2 Chemical lures, pheromones and feeding attractants and stimulants

Chemical lures, based on feeding stimuli, are now on the market as odorimpregnated artificial bait. They are specific for certain predatory fish and are extremely effective (Schisler and Bergesen, 1996). Efforts to understand improving of feeding attractants and stimulants for economically sound *mariculture* started early (Bardach and Villars, 1974). Another obvious application of fish odors is the use of waterborne male and female pheromones to improve spawning in fish in aquaculture.

13.1.3 Acidification of Water

Acid precipitation and the resulting higher acidity of freshwater bodies may result in severe disruption of feeding and migration, since both behaviors depend on water chemistry. Atlantic salmon (Salmo salar) alter responses to amino acids with changes in pH (Royce-Malmgren and Watson, 1987). In the

laboratory, the fish are attracted to glycine, but avoid L-alanine. When the pH is lowered from 7.6 to 5.1, the salmon become indifferent to glycine but are attracted to L-alanine. Acidic water also disrupts fish responses to alarm pheromones. At pH 6.0 fish do not respond to the alarm pheromone hypoxanthine 3N-oxide. This occurs because the N-oxide group on the molecule is lost through an irreversible covalent change. At pH 8.0 this does not happen (Brown et al., 2000), water pollution by copper affects responses to alarm pheromone in fish such as rainbow trout, chinook salmon, and pikeminnow, *Ptychocheilus lucius* (e.g. Beyers and Farmer, 2001; Scott et al., 2003).

13.1.4 Mosquito control

Mosquito fish, *Gambusia affinis*, are being released into freshwater bodies to control mosquitoes. It appears that chemicals from the fish "warn" the mosquitoes and they avoid laying their eggs in ponds with these fish. The number of mosquito larvae reduced as more fish chemicals (2 liter/day from pools with mosquito fish) were added to outdoor pools in North Carolina. The problem is that if mosquitoes avoid stocked pools, the efforts to control these insects with fish may fail (Angelon and Petranka, 2002).

13.2 Reptiles

13.2.1 Habitat Imprinting

Efforts to apply semiochemicals to manipulate behavior in reptiles are still young. One example is habitat imprinting in sea turtles. Almost all of the endangered Kemp's ridley sea turtles, Lepidochylus kempi used to nest on a 15 km beach near Rancho Nuevo, Mexico. In the 1940s, approximately 40000 females came ashore to lay their eggs. By 1978 there were only 924. Since 1978 a second nesting population 400 km north at Padre Island National Seashore in Texas has slowly been established. The task was to imprint hatchlings to the water of Padre Island so that they return to this new "home" to lay eggs when adult. From 1978 to 1988, 22507 eggs were collected at Rancho Nuevo and placed in sand and water from Padre Island. They were then brought to Padre Island and hatched there. The juveniles were raised for 9-12 months and then released at the beach where they enter the surf. In multiple-choice experiments, these juveniles spent more time in seawater and samples from Padre Island than samples from other areas. For years, none of the released turtles returned to Padre Island and many were found dead at beaches and in fishing nets. Methodological problems, such as not tagging palm-sized hatchlings serving as controls, also

occurred (Tauber, 1992). In 1996, this effort at olfactory imprinting and "headstarting" bore fruit: two tagged turtles, 10 and 13 years old, landed at the Padre Island beach. Both laid eggs (a total of 590), and 369 hatchlings were released into the ocean. In 1999, 16 nests were found in South Texas. In 2002, 23 of the 40 nests in the USA were found at Padre Island National Seashore. In 2003, 24 sea turtle nests were reported. Of these, 19 were Kemp's ridleys. The hatching success was 90–95%, and 1426 hatchlings were released. In 2004, 3928 eggs were laid in 42 nests and 2608 hatchings were released. That year saw the first *arribada* nesting of more than three females in 1 day. In the 2005 season, 56 clutches had been found by mid-July (www.nps.gov/pis/home). The turtles appear to have established themselves; the headstarting effort has paid off.

13.2.2 Snake control

The brown snake, Boiga irregularis, was accidentally introduced on Guam during or shortly after the Second World War. There are now 16–50 snakes per hectare (or as many as 30000 per square mile) on that island. They have caused extinction of native birds, reduced bat and lizard populations, are feeding on chickens and caged birds, and cause frequent power outages. Dispersal to other Pacific islands, notably Hawaii, by commercial traffic is a serious danger. Some snakes have already found their way to Hawaii. To trap brown snakes, the potential of both feeding stimuli from prey (Shivik, 1998) and blood and its components (Chiszar *et al.*, 1988b, 1992, 1993a) and pheromones (Mason and Greene, 2001) are being studied. In the meantime, strict inspections of airplanes from Guam try to prevent accidental spread of the snake.

13.3 Birds

13.3.1 Antifeedants: methyl anthranilate

Feeding repellents for pest birds are the most important application of chemical stimuli to manipulate bird behavior. Methyl anthranilate (Fig. 13.1) and dimethyl anthranilate, esters of benzoic acid, are found in concord grapes and are used as artificial flavorings. Starlings, Sturnus vulgaris, have an aversion to methyl anthranilate, which irritates the trigeminal nerve, and they feed less on food flavored with a variety of anthranilates. They avoid the more volatile anthranilates most. The odor is partly responsible for the effect; contact is not necessary. In one particular experiment, only volatile compounds were aversive (Mason and Clark, 1987). If only anthranilate-treated food is offered, the birds will accept more of the flavored food than they do if they offered a choice between

FIGURE 13.1 Methyl anthranilate deters birds from feeding and stimulates feeding in mammals. Capsaicin deters mammals from feeding but stimulates birds.

treated and untreated food (Mason *et al.*, 1989c). This underscores the need for alternatives for a repellent to work.

Commercially, methyl anthranilate (trade name ReJeX-iT AG-36) is used to repel red-winged blackbirds, *Agelaius phoeniceus*, from feeding on rice seed (Avery *et al.*, 1995). This species avoided methiocarb-poisoned apples; additional chemical (methyl anthranilate) and visual (calcium carbonate) cues enhanced the avoidance response. This way the amount of the toxin methiocarb could be reduced and yet still be equally effective and at lower cost (Mason, 1989). Canada geese (*Branta canadensis*) also feed less on grass treated with methyl anthranilate (Cummings *et al.*, 1995).

13.3.2 Antifeedants: compounds from unripe fruit

Unripe fruits of many plants contain compounds such as quinines that impart an aversive taste and discourage fruit eaters from consuming them before they are ripe and their seeds are mature and ready for dispersal. Unripe fruit of some *Rhamnus* species contain emodin, an anthraquinone that deters birds and mammals from feeding. Anthraquinone reduced red-winged blackbird feeding on rice by 84%, and 71% in boat-tailed grackles (Avery *et al.*, 1997).

13.3.3 Exploitation of automimicry (Batesian mimicry)

Brown-headed cowbirds, *Molothrus ater*, and red-winged blackbirds, *A. phoeniceus*, removed 95–98% less rice seeds sown in flight pens if the seeds had been treated with methiocarb. The effect was the same whether 100% or only 50% of the seeds had been treated. This shows that an entire prey population can be protected even if only a portion is unpalatable. Such automimicry may permit partial treatment of crops with repellents, which would be effective, environmentally sound, and economical (Avery, 1989). Batesian mimicry offers advantages over food aversion learning (Reidinger and Mason, 1983).

Birds are more sensitive to repellents in drinking water than on food. The active molecules in aqueous solution may have better access to the chemoreceptors (Clark *et al.*, 1991).

13.3.4 Food flavor preferences in domestic birds

Domestic birds rely more on odors than initially thought. Chickens are attracted to a sample of familiar odor (soiled substrate, or familiar substrate with orange oil) in an otherwise novel and probably frightening environment (Jones and Gentle, 1985). Chickens reared on mash with a few drops of water added were presented with mash treated with a few drops of orange oil. They showed neophobia (i.e. their latency to feed was longer and they fed less than from the water mash), but only initially. Conversely, chicks raised on orange-scented mash accepted water mash readily. This suggests that diet changes in poultry operations may have little impact on the animals' food intake or well-being (Jones, 1987). Still, farmers in the future may be able to imprint chicks to certain food or coop odors while the chicks are still *in ovo* (Section 9.4).

13.3.5 Non-lethal contact repellents

In the search for environmentally benign chemicals that might deter birds such as starlings, crows, or pigeons from roosting *en masse*, spices and herbs such as rosemary, cumin, and thyme look promising. In some experiments, the birds' feet were immersed in oil extracts of the spices. Starlings also avoided perches treated with starch mixes of (R)-limonene, (S)-limonene, β -pinene, or methiocarb. The first three occur in rosemary, cumin, and thyme (Clark, 1997).

13.3.6 Bird olfaction and conservation biology

Given how sensitive many birds are to odors, breeders of endangered bird species may be able to use food and environmental odors to imprint young birds on relevant cues of their future habitat (Nevitt, in Malakoff, 1999).

13.3.7 Differences between birds and mammals

Birds and mammals differ in their responses to some flavor compounds

- birds are repelled by methyl anthranilate but accept capsaicin
- mammals are repelled by capsaicin and are accepting of methyl anthranilate.

Mammals avoid capsaicin (Fig. 13.1) from "hot" jalapeño peppers, widely used as dog repellent, but birds eat it and it is used as parrot food (Reidinger and Mason, 1983). Capsaicin is used in commercial feeding repellents for squirrels at bird feeders. Conversely, birds reject methyl anthranilate (Kare and Pick, 1960), but mammals such as livestock and obviously humans, accept it. However, five known bird repellents given in water also reduced drinking in mice. The most active was orthoaminoacetophenone, which eliminated drinking but methyl anthranilate, 2-amino-4',5'-methoxyacetophenone, 2-methoxyacetophenone, and veratryl amine were also effective (Nolte *et al.*, 1993). Different capsaicin-related compounds repel both starlings and rats. These responses involve the trigeminal nerve. Repellency in birds is enhanced by electron richness of the phenyl ring and basicity and reduced by acidic functionalities. The reverse is true for mammals. Compounds with long side chains (lipophilicity), electron-poor phenyl ring, and acidity repel mammals (Mason *et al.*, 1991; Table 13.1).

Birds may have a trigeminal receptor for *o*-aminoacetophenone and methyl anthranilate, analogous to the capsaicin receptor in mammals (Clark and Shah, 1994).

13.4 Non-human mammals

The bulk of the applications of semiochemicals in vertebrates are aimed at mammals. However, it must be remembered that mammalian behavior is complex and management by chemical cues is more challenging than in insects, for example.

13.4.1 Control of rodent and other herbivore damage

Area repellents

Predator odors

The search for environmentally benign pest repellents has spawned many experiments with excretions and gland secretions from felids, canids, and mustelids.

A predator odor affected a pocket gopher, *Thomomys talpoides*, population in western Canada. First live-trapping removed the gophers from plots of 4ha area in an orchard. Then synthetic sulfur compounds from stoat, *Mustela erminea* anal glands (1:1 mixture of 2-propylthietane and 3-propyl-1,2-dithiolane) were

Table 13.1 Structure–function relationships in repellents: opposite effects on a bird and a mammal

Compound	Structure	Repell	lent effect
		Starling	Rat
Capsaicin	N C ₉ H ₁₇	None	Strongest
Methyl capsaicin	H_3 CO C_9 H $_{17}$	None	Second strongest
Vanillyl acetamide	HO CH ₃	None	Medium
Veratryl acetamide	H_3 CO OCH_3 H CH_3	Yes	Lowest
Veratryl amine	NH ₂ OCH ₃	Yes	Second lowest

Factor	Species	Effect	Reference
Cover	Townsend's vole Microtus townsendii	Avoid R in open; ignore R in cover	Merkens et al., 1991
Available choice of food plant species	Many species	R avoided only when food choice is wide	Common experience
Palatability of plant	White-tailed deer Odocoileus virginianus	R protects less-preferred plants better	Swihart <i>et al.</i> , 1991
Established feeding habits	White-tailed deer	R deters early in growing season but not in summer after having fed in gardens	Müller-Schwarze, 1983
Hunger	Elk Cervus canadensis	R ineffective in starving animals	Andelt et al., 1992

Table 13.2 Some factors that affect chemical repellent efficacy in mammals

R. repellent.

applied on clay pellets inserted into burrows in November. In the following spring (April), recolonization was found to be reduced. The population fell by 50% as judged by the numbers of fresh mounds and did not recover fully, as would happen under usual circumstances (Sullivan *et al.*, 1990).

Predator odors are also effective area repellents for lagomorphs. A rabbit warren sprayed with an extract from lion feces had as many as 80% fewer animals than before the treatment and also fewer than a control warren. Adult rabbits stayed away from the treated warren longer than young ones. The effect lasted up to 5 months (Boag, 1991; Boag and Mlotkiewicz, 1994).

For odors to work, certain environmental conditions have to be met. Townsend's voles, *Microtus townsendii*, avoid a repellent odor if no *cover* is available. With cover present, they feed whether or not the area is scented. Therefore, for effective area repellents, the "pest rodent" should be able to retreat to unscented areas in preferred habitat such as sufficient cover (Merkens *et al.*, 1991; Table 13.2).

Non-target rodent species in the same forests may escape effects of mustelid sulfur compounds. In British Columbia, densities, survival rates, and reproduction of deer mice, *Peromyscus maniculatus*, were little affected when these compounds were used against long-tail voles (*Microtus longicaudus*), meadow voles (*Microtus pennsylvanicus*), and boreal redback voles (*Clethrionomys gapperi*) (Zimmerling and Sullivan, 1994).

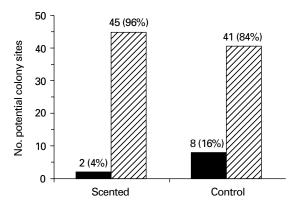


FIGURE 13.2 Beavers colonized only 2 of 47 (4%) experimentally scented potential colony sites (left), but 8 of 49 (16%) unscented control sites (right). Black indicates colonized sites; cross-hatched, sites left vacant. Fulton County Experiment. (From Müller-Schwarze 1990.)

Conspecific odors

Secretions and excretions from the same species can be used as repellents as well as attractants to traps or bait. Repellents are based on territorial exclusion and same-sex competition, while attractants exploit general gregariousness and preference for the opposite sex.

Territorial rodents typically mark their breeding territory with scent gland secretions, urine, feces, or combinations of these. It is possible to mimic occupancy of a vacant area by artificial scenting. North American beaver, *Castor canadensis*, are less likely to colonize available "potential beaver sites" along streams if a mixture of male and female castoreum (from castor sacs) and anal gland secretion is placed as scent marks at "strategic places." These are points of entry along a stream and areas near dam(s) and lodge (Welsh and Müller-Schwarze, 1989; Müller-Schwarze Fig. 13.2).

Adult muskrats, *Ondatra zibethica*, in the dense population at the Montezuma Wildlife Refuge in New York avoided traps scented with musk, the secretion of the preputial glands of this species, whereas juveniles entered clean traps and traps with *Ondatra* musk or a control odor (phenylacetic acid) equally often. This suggests that adults avoid others' territories and their odors, while immature muskrats are tolerated (van den Berk and Müller-Schwarze, 1984). In simultaneous experiments in the Netherlands, however, adult muskrats, especially males, were attracted to musk-scented traps. Here the experimental animals belonged to a colonizing population that was in the process of invading along waterways and were not yet entrenched territorially (Ritter *et al.*, 1982). Both of these responses were noted in a field study of wild red-backed voles,

Clethrionomys clareolus, and wood mice, Apodemus sylvaticus, near Munich. Immature animals entered male-scented, female-scented, and control traps equally often, while male odor attracted adult males and female odor attracted adult females (Bäumler and Hock, 1987).

Scents from coyotes, foxes, and other carnivores traditionally have served to attract these animals to traps. Trappers also use conspecific secretions to attract beavers, raccoons, and other fur bearers to traps. The use of scent lures in predator control is discussed below.

Area attractants

The odor of a trap soiled by a previous occupant of the same or different species can affect subsequent trapping success and, therefore, deduced population estimates. Whether rodents prefer dirty to clean traps, the odor of their own species to others, their own odor to other conspecifics, the same or opposite sex, varies between species and studies (reviewed by Gurnell and Little, 1992). For instance, adult California voles, *M. californicus*, prefer volescented traps over clean ones only during the breeding season. Juveniles show no preference, whether during the breeding or non-breeding season (Heske, 1987). Short-tailed voles, *M. agrestis*, are also affected by trap odor (Stoddart, 1982) and often the sexes can differ in their responses. In California ground squirrels, *S. b. douglasii*, males are more attracted to male anal gland secretion on traps, but females showed no preferences (Salmon and Marsh, 1989). These responses resemble those by prairie deer mice, *P. m. bairdii*, and house mice, *M. musculus* (Wuensch, 1982).

Wood mice, *A. sylvaticus*, and bank voles, *Clethrionomys glareolus*, differ in their responses to soiled traps. While both species are attracted to conspecific odor, wood mice preferred odor of the opposite sex, while bank voles did not, and wood mice avoided bank vole odor. This study suggests caution when using live-trapping for population censuses of rodents (Tew *et al.*, 1994).

However, several European rodent and shrew species (*A. sylvaticus*, *C. glareolus*, *M. agrestis*, and *Sorex araneus*) entered cleaned (with boiled water or detergent) traps equally often as dirty ones (only scraped after use). This was true for each species and sex. This suggests it may not be necessary to clean traps between uses (Tew, 1987).

Antifeedants

Predator secretions and excretions inhibit feeding in a variety of herbivore species.

Gland secretions

Mountain beaver, *Aplodontia rufa*, a primitive rodent from northwestern North America, fed less from food bowls if the rim was scented with mink anal gland secretion, or urine from mink, bobcat, coyote, or dog. The control odors butyric acid or guinea pig urine had no effect (Epple *et al.*, 1993). Mountain beaver did not respond significantly to mustelid sulfur compounds though, yet these deter other small mammals (Epple *et al.*, 1993).

Excretions

Black-tailed deer, *Odocoileus hemionus columbianus*, feed less on salal leaves (*Gaultheria shallon*), red cedar (*Thuja plicata*), and Douglas fir (*Pseudotsuga menziesii*) in the presence of urine, or feces of large cats and canids (Sullivan *et al.*, 1985b). Browsing by black-tailed deer on salal was most reduced by bobcat feces (by 51%), followed by mountain lion, *Felis concolor* (by 27%), wolf, *Canis lupus*, and coyote, *Canis latrans* (by 8%) (Melchiors and Leslie, 1985). Coyote urine also reduced feeding by tame mule deer on pelleted rations (Andelt *et al.*, 1991).

Predator-derived repellents deterred white-tailed deer, *Odocoileus virginianus*, from gardens after an application in May, the beginning of the growing season, but a second application in August was not effective once the deer had had ample opportunity to sample and utilize the gardens (Müller-Schwarze, 1983). White-tailed deer feed less on Eastern hemlock (*Tsuga canadensis*) and Japanese yew (*Taxus cuspidata*) when these are treated with urine from coyote, *C. latrans*, or bobcat, *Lynx rufus* (Swihart *et al.*, 1991).

The repellent effect depends on hunger, as in female elk (Andelt *et al.*, 1992) and less-preferred plants are better protected by predator urine odors (Swihart *et al.*, 1991).

Dog feces are very effective as feeding repellents for sheep, roedeer, *Cervus capreolus*, and red deer, *Cervus cervus*, while droppings from omnivorous bears had very little repulsive effect (Arnould and Signoret, 1993; Arnould *et al.*, 1993). In Australia, dog urine significantly reduced browsing damage to eucalypt and pine plantations by swamp wallabies, *Wallabia bicolor*, and rabbits (Montague *et al.*, 1990).

The Orkney vole, *Microtus arvalis orcadensis*, removes almost no bark from seedlings of Scots pine, *Pinus sylvestris*, if they are treated with an extract of feces of red fox, *Vulpes vulpes*, even though the two species have been separated for approximately 5500 years (Calder and Gorman, 1991).

Synthetic compounds

The odors of predatory mustelids can be very effective in inhibiting feeding on plants by rodents, lagomorphs, and deer, and, therefore, hold promise

	_		_
Compound	Source animal and secretion	Affected animal	Reference
3,3-Dimethyl-1,2-dithiolane	Ferret AGS Red fox feces	Rat	Vernet-Maury et al., 1984
2-Propylthietane	Stoat AGS	Montane vole	Sullivan et al., 1988
3-Propyl-1,2-dithiolane	Stoat AGS	Montane vole	Sullivan et al., 1988
2,5-Dihydro-2,4,5- trimethylthiazoline	Red fox urine	Montane and meadow vole	Sullivan et al., 1988
3-Methyl-3-butenyl- methyl sulfide	Red fox urine	Snowshoe hare	Sullivan and Crump, 1986a
Trimethylthiazoline	Red fox feces	Rat, mouse	Vernet-Maury <i>et al.</i> , 1984; Coulston <i>et al.</i> , 1993;

Table 13.3 Predator sulfur compounds active as rodent repellents

AGS, anal gland secretion

for protecting plantations. Sulfur compounds from mustelid anal gland secretions suppress feeding by snowshoe hares, *Lepus americanus*. The most active compounds were 3-propyl-1,2-dithiolane from the stoat, *M. erminea*, and 2,2-dimethylthietane from stoat and mink, *M. vison*. The effect lasted for up to 38 days (Sullivan and Crump, 1984). Urine, feces and anal gland secretions from various mustelids, canids, and felids suppressed feeding by snowshoe hares (Sullivan *et al.*, 1985a). The most effective constituent of anal gland secretions from the red fox, *V. vulpes*, for reducing feeding on lodgepole pine seedlings was 3-methyl-3-butenylmethyl sulfide (Table 13.3). Other sulfur compounds such as 2-phenylethylmethyl sulfide and 3-methylbutylmethyl sulfide were not active (Sullivan and Crump, 1986b). Odors from wolverines also deter snowshoe hares from feeding (Sullivan, 1986). Sulfur compounds from mustelid anal glands inhibited feeding in a number of rodents such as pocket gophers, *Thomomys talpoides* (Sullivan and Crump, 1986a) and montane, *Microtus montanus*, and meadow, *M. pennsylvanicus*, voles (Sullivan *et al.*, 1988).

A mixture of five sulfur compounds that occur in lion (Felis leo) dung proved to be a feeding inhibitor in captive red deer, Cervus elaphus. The five compounds are dimethyl disulfide, dimethyl trisulfide, dimethyl tetrasulfide, 3-mercaptopropan-1-ol, and 3-mercaptothiopropan-1-ol (Abbott et al., 1990). This odor also deters rabbits from feeding on carrots for up to 1 month (Boag and Mlotkiewicz, 1994). Sulfur compounds, generated during protein digestion, could be the "general theme" underlying the repellency of predator odors for rodents and other herbivores (Nolte et al., 1994b). Mule deer Odocoileus hemionus hemionus, avoided tree seedlings treated with proteins that are high in

methionine, a sulfur-containing amino acid, although methionine alone was not effective (Kimball and Nolte, 2004).

Experimentation with sulfur-containing compounds has also utilized egg extract. Such extract, with acrylic adhesive and a red dye, proved effective in repelling sheep from feeding on seedlings of Monterey pine, *Pinus radiata*, in New Zealand. Thiram (tetramethylthiuram disulfide, a fungicide), with or without egg, was not effective (Knowles and Tahan, 1979). Crozier (1991) recommended egg-based repellents, which had proved effective against rabbits, hares, and possums in New Zealand, because they are readily available, easy to apply, and nontoxic to humans. Mixed with acrylic paint adhesives, such repellents stay on tree seedlings for months. A commercial deer and elk repellent containing putrescent whole egg solids has been tested as a spray to change the uneven distribution of steers on their range. The repellent effect in putrefied egg concoctions results from aldehydes and possibly fatty acids (Tiedman *et al.*, 1976).

Plant compounds

"Natural resistance in trees to damage by animals remains essentially unrecognized and unexploited" (Dimock *et al.*, 1976). Despite great interest in chemical plant defenses since the mid-1970s, we still are just beginning to understand these systems well enough to apply them successfully. (Well-defended trees in non-glaciated "resistance centers" and from areas with large wildfires were discussed in Ch. 11, p. 336 and p. 337.)

Over evolutionary time, chemical warfare between plants and herbivores has produced many effective antifeedants that have significant effects on herbivores (Ch. 11). When seeking applications, we can avoid exaggerated expectations by remembering that the vast majority of plants do not totally eliminate herbivory. Instead, they manage to keep it at acceptable levels. It is not surprising that some plant toxins have been used in rodent control. Scilliroside (red squill) is a rodenticidal glycoside from the Mediterranean plant *Urginea maritima*. Quebracho, a condensed tannin, reduces *Microtus* spp. damage in orchards (Lindroth and Batzli, 1984). Intense efforts are underway to harness natural antiherbivore compounds as non-toxic repellents to protect crops and the potential is enormous.

Pine oil is a by-product of the pulp industry. This oily liquid contains a mixture of terpene alcohols and monoterpenes and is available under the trade name Norpine-65. Snowshoe hare, *L. americanus*, and voles, *M. townsendii*, consume less laboratory chow, apples, or carrots if they are treated with pine oil. This response did not habituate within the time of the experiment. When considering the use of pine oil as a herbivore repellent, we have to keep in mind that it is phytotoxic to seedlings if applied topically (Bell and Harestad, 1987). Red deer, *C.*

elaphus, calves avoid monoterpenes from Sitka spruce, *Picea sitchensis*, and lodgepole pine, *Pinus contorta*, even without prior experience. Females are more sensitive than males. Monoterpenes with higher vapor pressure were avoided more, raising the question whether the deer respond to the strength of an odor or to its quality (Elliot and Loudon, 1987). This avoidance is adaptive as monoterpenes inhibit rumen microorganisms (Schwartz *et al.*, 1980). In the search for new and effective deer repellents to protect Douglas fir, *P. menziesii*, fresh leaves of wild ginger, *Asarum caudatum*, deterred black-tailed deer, *O. h. columbianus*, effectively. The deer ate only 2% of a food mix containing 5% shredded wild ginger leaves (Campbell and Bullard, 1972). Poisonous, foxglove, *Digitalis purpurea*, also quickly conditions herbivores to avoid this plant (Campbell, 1987).

In the first practical application of plant antifeedants for forest crop protection and wildlife management, snowshoe hare, *L. americanus*, fed less on coniferous tree seedlings after they had been sprayed with pinosylvin or pinosylvin methyl ether (Sullivan *et al.*, 1992).

Conditioned flavor aversions

"Flavor" consists of taste and odor combined. Animals often avoid tastes after only one experience if delayed illness follows it, but odors are not avoided under such circumstances. If an odor and a taste are presented together before an illness is induced experimentally (e.g. with lithium chloride), animals can learn to avoid the odor after just one such experience. When the odor is intensified in such a combined presentation, only the odor aversion increases, not the taste aversion. If the taste is made more intense, both odor and taste aversions increase. As a rule, the odor aversion strength depends on the taste experience, but not vice versa (Garcia *et al.*, 1986).

Mountain beavers, *A. rufa*, and black-tailed deer, *O. h. columbianus*, severely damage seedlings of forest trees in western North America. These two species learnt to avoid Douglas fir, *P. menziesii*, when mountain beaver were provided with Douglas fir cuttings treated with a putrescent egg preparation, and black-tailed deer were exposed to treated plastic ribbons on seedlings (Campbell, 1987). Damage to Douglas-fir seedlings by mountain beaver can also be reduced if the animals are conditioned first. Sulfur compounds such as the fungicide thiram, dimethyl dithiocarbamate compound are feeding repellents suited for aversion conditioning. Thiram-treated seedlings were placed into burrows and treated and untreated seedlings planted in front of the burrow. After 8 months (note the long-term effect), feeding on treated and untreated seedlings was reduced to 17.4% and 16.0%, respectively. On sites without thiram conditioning, 94.4% of untreated seedlings were cut outside untreated burrows (Campbell and Evans, 1989).

Modifying plants

Lowering or increasing the concentrations of plant compounds that affect palatibilty by selective breeding or bioengineering remains an important goal. Douglas fir, *P. menziesii*, is an example where an increase in secondary compounds would be desirable to render it more resistant to browsing by black-tailed deer, *O. h. columbianus* (Dimock *et al.*, 1976). Sagebrush, *Artemisia tridentata*, poses the opposite problem: selective breeding for *reduced* monoterpene content is needed so that mule deer, *O. h. hemionus*, can better utilize it (Welch and McArthur, 1981).

The levels of antifeedants can be raised safely only in plants such as forest trees or fiber crops, not in food plants for humans or livestock. Such problems have arisen inadvertently. For instance, a new insect-resistant potato cultivar had to be withdrawn from the market because it contained high levels of the carcinogens solanine and chaconine (Renwick *et al.*, 1984). In another example, an insect-resistant celery had 10 times the usual concentration of the carcinogen 8-methoxypsoralen (and related psoralens), which caused dermatitis in produce handlers (Seligman *et al.*, 1987).

Watering the foliage of seedlings with quadrivalent selenium reduced deer browsing of Douglas fir. This produces a garlic-like odor that might be responsible for the effect (Allan *et al.*, 1984).

Feeding stimulants

Conspecific secretions and excretions may stimulate pest rodents to consume bait. An example is the metad, or soft-furred rat, *Rattus meltada pallidior*, of the grasslands and crop fields of India. In a laboratory test, 1% conspecific urine from males or females added to the diet of millet increased food consumption by 32 to 70%. The metads also stayed longer in the half of the cage with the urine-treated food than in that with untreated food, and scent marked more there. The males marked with sebum, the females with urine. Male urine tended to be more effective, and males responded more than females (Soni and Prakash, 1987).

Rats are initially reluctant to take new bait. This bait shyness can be overcome by applying urine of estrous females to attract males, and male urine to attract females, unless pregnant. Males consume more of a feed if female urine odor is present (Gao, 1991). Bank voles are attracted to traps scented with $16-5\alpha$ -androsten- 3α -ol (Bäumler and Haag, 1989).

A novel odor for attracting rats or mice to bait is carbon disulfide. It is found in the breath of rats, serves in communication about food, attracts rodents to bait, and increases bait consumption (Bean *et al.*, 1988). Use of carbon disulfide and carbonyl sulfide in baits attracts mice and rats and prevents bait shyness from

developing. It has been patented for that purpose (US Patent No. 4,861,585 to J. R. Mason and B. G. Galef).

13.4.2 Domestic Animals

Predator-derived odor repellents

Alpine goats, *Capra hircus*, fed considerably less from buckets with food (cottonseed hull, corn, and alfalfa) covered with shredded filter paper soaked with extracts of predator feces. The predators were Bengal tiger (*Panthera tigris tigris*), Siberian tiger (*Panthera tigris altaica*), African lion (*Panthera leo*), and brown bear (*Ursus arctos*) (Weldon *et al.*, 1993). Dog odor is also active. A water slurry of dog feces repels sheep, roe-deer, and possibly red deer from feeding on grass, yoke-elm, *Carpinus betulus*, and oak, *Quercus* spp. (Arnould *et al.*, 1993).

Predator odors do not necessarily keep herbivores away from forage. Sheep and cattle, presented with odors from coyote, fox (*V. vulpes*), cougar (*Felis concolor*), or bear (*Ursus americanus*) near their feed rations spent less time feeding there, but they did not stay away from the treated feed (Pfister *et al.*, 1990b).

Similarly, two sulfur compounds from predators, 3,3-dimethyl-1,2-dithiolane and (*E*)-(*Z*)-2,4,5-trimethyl- Δ^3 -thiazoline had no effect on wild roof rats, *Rattus rattus*, in a Hawaiian macadamia orchard (Burwash *et al.*, 1998).

Laboratory and domestic animals may be poor models for avoidance of predator odors. For example, in one experiment, chickpeas were painted with the sulfur compounds *n*-propyldithiolane and *n*-propylthiolane from stoat anal gland secretion and 2,4,5-trimethylthiazoline (Fig. 3.1, p. 37) from fox feces. The chickpeas were planted and wild mice and house mice were tested to see if they would dig up and eat the peas. Wild mice remembered the predator odors better after odor exposure for 1 or 4 weeks and, consequently, may be better than laboratory mice at risk assessment (Coulston *et al.*, 1993).

Conspecific herbivore odors as feeding deterrents

Cattle avoid grazing near cow pats for at least 30 days. The aversive stimulus emanates from the dung and not the plants: cattle feces, 0–35 days old, placed in a feeding trough inhibit feeding. Volatiles extracted from cow dung also reduce feeding. A neutral fraction of a diethyl ether extract was active in one experiment (Dohi *et al.*, 1991). Likewise, horses will not feed near their own droppings (Ödberg and Francis-Smith, 1977). House mice, *M. musculus*, differ. They consume the same amount of food whether it contains fecal pellets or not (Pennycuick and Cowan, 1990).

Plant compounds and food consumption

Secondary plant metabolites can reduce food consumption by livestock. Sheep refuse hay if it contains coumarin, gramine, tannic acid, malonic acid, or glycine. However, butyric acid or amyl acetate stimulate hay consumption. Anosmic and agustatory sheep show the same responses so sensory stimuli are not involved. Tannic acid and gramine lower *in vitro* digestibility, coumarin does so slightly, and the other compounds did not affect digestibility. In general, the compounds affect food intake only if there is an effect on the rumen (Arnold *et al.*, 1980). Some plants are outright toxic to livestock, presenting the chemical ecologist with the twin problems of the effects of plant compounds on herbivores and the need to develop effective repellents to steer grazing animals away from stands of toxic plants. Larkspur (*Delphinium* sp.) contains diterpenoid alkaloids and poses the most serious poisonous plant problem for cattle of the mountain rangeland in the western United States. Cattle are more affected than sheep. During the years 1913–1916, more than 5000 cattle died annually in the national forests of the west from larkspur poisoning (Ralphs *et al.*, 1988).

Olfactory imprinting

Since time immemorial, farmers, breeders, and zookeepers have had to cope with cross-fostering problems. They either have to save orphaned livestock newborns or attach a young of a different species to a lactating female for later interbreeding to produce desirable hybrids. Recent studies have confirmed that in cows, for example, odor transfer between young is an inexpensive, rapid and safe technique to help postpartum beef cows accept alien calves (Dunn *et al.*, 1987). The alien calf is wrapped in a stockinette that the cow's own calf had worn for 48–96 hours and this makes the alien calf acceptable to 9 out of 10 cows. By comparison, only 1 of 12 alien calves was accepted if they wore a jacket with their own odor. Acceptance was defined as lack of aggression and suckling periods lasting at least 20 seconds. Four days after fostering, 100% of the alien calves with the jacket of the cow's own calf were accepted, but only 33% with their own jacket. The highest acceptance rate for this second group was 50% on day 5. All cows accepted calves with transferred jackets, while two (17%) did not accept young with an alien calf's jacket (Dunn *et al.*, 1987).

In sheep too, a ewe whose lamb has died soon after birth will accept an alien lamb if it carries the odor of her own young. Several methods of scenting the new lamb have been compared. First, Hessian coats were fitted on the lambs when they died and left there for 2 to 18 hours; they were then put on the foster lambs. When presented to the ewes whose lambs had died, 73% accepted the new lambs.

Second, dead lambs were smeared with neatsfoot oil (an oil obtained by boiling feet and leg bones of cattle), left with the mother for 2–18 hours and then substituted by neatsfoot oil-treated foster lambs. Here 91% were accepted. The best acceptance rate (100%), however, was achieved by draping the skin of the dead lamb over the alien one. This is the method of choice because it is not only most effective but also saves labor as no treatment is necessary (Alexander *et al.*, 1987). Ewes accept alien lambs if they are rubbed with non-polar oils such as white soft paraffin or liquid paraffin. Polar materials such as polyethylene glycol, glycerol silicone, or diisooctyl phthalate have no effect. Ewes appear to use non-polar volatiles for discrimination (Alexander *et al.*, 1989).

When ewes and lambs become separated during sheep management operations, mothers and lambs first bleat. After approaching one another, the ewe sniffs a lamb at close range. This olfactory test decides whether the mother lets the lamb nurse, or rejects it, possibly by head-butting (Fig. 13.3).

Amniotic fluid

Along with eating the afterbirth, female ungulates lick amniotic fluid during and after giving birth. For cross-fostering experiments, it is important to know whether acceptance of amniotic fluid is species specific. In one experiment, ewes accepted food treated with ovine amniotic fluid, also caprine amniotic fluid (albeit less so) at and after parturition, but always rejected bovine amniotic fluid (Arnould *et al.*, 1991).

Priming pheromones

Priming pheromones hold great promise as means to accelerate puberty or to synchronize and regularize estrus cycles in livestock.

Puberty acceleration

Oronasal application of bull urine accelerated puberty in beef heifers; 67% of the animals reached puberty within 8 weeks of treatment, compared with 32% in water-treated controls. Urine-treated heifers that reached puberty during the experiment calved earlier than water-treated heifers. The calving season also was shorter for urine-treated animals (Izard and Vandenbergh, 1982a). Puberty in female pigs can be accelerated by 1 month if a boar is present (Pearce and Paterson, 1992).

Estrus synchronization

Dairy heifers, after being brought into their follicular phase by injection of prostaglandin $F_{2\alpha}$, were treated with cervical mucus or urine placed on





FIGURE 13.3 Ewes recognize their lambs by first calling and then sniffing lambs after separation during herding operations in paddocks. (Photographs: D. Müller-Schwarze.)

the oronasal area. Cervical mucus had the greatest effect: all heifers in this group were in estrus within 72 hours and the degree of synchrony was high (Izard and Vandenbergh, 1982b).

Signaling pheromones

The boar sex pheromone, consisting of androstenol and androstenone, from the submaxillary salivary gland has been used commercially for artificial insemination of pigs. It was available as a spray under the trade name Suidor (formerly Jeyes Boar Mate) in Europe (Glei *et al.*, 1989). Sows who did not assume the

mating stance are sprayed in the face. After this treatment, 60% will assume the mating stance. In addition, fighting among young pigs was reduced by introducing the odor of an adult boar (McGlone, 2002).

13.4.3 Breeding of rare species

Declining species are sometimes hard to breed in captivity. Two techniques have been used to enhance breeding behavior in the harvest mouse, *Micromys minutus*. First, males rejected by females were given scent marks from preferred mates. This stimulated their mating behavior and made them more attractive to females. Second, females were given male scent marks to prefamiliarize them with a male. The females preferred the male odor donors and were less aggressive towards them (Roberts and Gosling, 2004).

13.4.4 Predator control

Gland secretions and urine as scent lures in predator control

Many odor lures have been tried for coyotes, *C. latrans*. The behaviors of coyotes at chemical lures include approach, sniffing, digging, scratching, licking, chewing, biting, rolling, urinating, and defecating. Fagre $et\ al.$ (1983) have listed 23 behavioral and chemical characteristics of an optimal coyote lure. Some effective coyote lures such as a synthetic fermented egg formulation comprise four odor classes: fruity (esters), sulfurous (organosulfurs), sweaty (volatile fatty acids), and fishy (amines) (Bullard $et\ al.$, 1983). Trimethylamine occurs in anal sacs of coyotes, dogs, and red foxes, valeric acid in coyote urine. In combination with trimethylammonium valerate, trimethylamine is attractive to coyotes. The combination slowly releases trimethylamine and lasts longer than the latter alone (Fagre $et\ al.$, 1983). Even more attractive is trimethylammonium decanoate, a salt of a C_{10} fatty acid (Fagre $et\ al.$, 1983).

Both male and female coyotes, *C. latrans*, are attracted to volatile aldehydes from sheep liver and estrous urine of female coyotes. The most active compounds were octanal, nonanal, decanal, and undecanal. The aldehydes released sniffing and rub-rolling, but little lick-chewing and biting. Therefore, these compounds are better suited to attract coyotes to traps than to toxicant-delivery systems that rely on the latter behaviors (Scrivner *et al.*, 1984).

The introduced ferret, *Mustela furo*, in New Zealand destroys native fauna and has to be controlled. For trapping, lures based on conspecific scent gland secretions or urine compounds can be as effective as food bait. Eight constituents of the anal sac secretion have been synthesized and tested for their ability to attract

ferrets to traps. Ferrets were most attracted to a mixture of 2-propylthietane and *trans*- and *cis*-2,3-dimethylthietane (Clapperton *et al.*, 1989).

Numbers of visits to scent stations baited with fatty acid tablets or prey odor such as shellfish oil allows estimation of population sizes of free-living red and gray foxes, raccoons, striped skunks, river otter, bobcats, cottontail rabbits, and oppossums. However, Smith *et al.* (1994) showed that the numbers of raccoon visits to scent stations did not reflect the size of their population.

Coyote scent (urine) stations also attracted cottontail rabbits *Sylvilagus floridanus*. In Texas, this behavior has aided rabbit censuses. The scent station count is more accurate than the traditional "headlight count" but more labor intensive (Drew *et al.*, 1988).

Predator repellents

Of many compounds tested, *trans*-cinnamaldehyde was most effective in delaying the approach to food by coyotes (Jankowsky *et al.*, 1974). Other studies found β -chloroacetyl chloride, a strong irritant and lachrymator, an effective olfactory repellent, while cinnamaldehyde was somewhat active in delaying approach to a visible food source (Lehner *et al.*, 1976).

Wildlife managers and conservationists often need carnivore repellents. As an example, in Lothringia, kites' nests had to be moved from barley fields. Red foxes followed the human tracks and preyed on the nestlings (A. Engelhart, personal communication). Measures that discouraged the foxes included chemical repellents, covering the human tracks, and aversive conditioning of the foxes.

Aversive conditioning of predators

To discourage birds and mammals from preying on eggs of the greater sandhill crane, *Grus canadensis*, Nicolaus (1987) injected surrogate crane eggs (turkey eggs) with 2,3,5- and 3,4,5-trimethylphenylmethyl carbamate (UC 27867) and placed them in breeding territories of ravens. Untreated chicken eggs, painted blue–green, were presented in a second experimental nest. At additional control sites, neither the surrogate eggs nor the painted chicken eggs were injected. Significantly more treated eggs (70%) survived intact than did uninjected blue–green eggs (38%). At the control sites, 37% of untreated surrogate eggs and 41% of blue–green chicken eggs survived.

A long line of investigations has tried aversion conditioning for mammalian predators, particularly coyotes. Traditionally, lithium chloride is either injected

into the predator after it has consumed a certain food, or it is worked into the bait. A study of aversive conditioning to rabbit meat used three methods: lithium chloride was injected into a coyote after feeding, used to lace rabbit meat wrapped in rabbit fur as a bait package, or used to lace rabbit carcasses. The aversion tended to be specific: the coyotes avoided the bait packages but continued hunting live rabbits (Horn, 1983).

Mongooses, Herpestes auropunctata, have been introduced on many tropical islands to fight rats in sugar cane fields. Unfortunately, they also damage or destroy native populations of mammals, birds, reptiles, or amphibians. Mongooses are particularly destructive where they prey on endangered species, such as the eggs of endangered hawksbill, green, and leatherback sea turtles. In a study in the field and in captivity on St. Thomas, American Virgin Islands, mongooses were conditioned with carbachol (carbamyl choline chloride) to avoid eggs. Carbachol acts as a cholinesterase blocker. Unlike the more often used trimethacarb, it is water soluble and it is less detectable by taste or odor. Both are harder to detect than lithium chloride, which is salty-tasting, and they are, therefore, thought to be more suitable for producing conditioned taste aversions. In a T-maze pre-test, the mongooses were exposed to an egg treated with anise extract or one treated with vanilla extract, both without carbachol. Next, during the acquisition phase, each end of the T-maze held an egg treated with anise extract and laced with 12 mg carbachol. Finally, in the post-testing phase, the two eggs smelled as they did in the pretest but the anise egg was injected with 6 mg carbachol. The toxic eggs were not avoided at a distance nor after tasting the outer shell. In the field, the mongoose behaved differently. Here they avoided toxic eggs sprayed with vanilla or mint odor from a distance and 39% of the clutches were left alone (Nicolaus and Nellis, 1987).

Aversive conditioning to deter mammalian predators from preying on water-fowl nests remains elusive. In one experiment, nests with chicken eggs injected with lithium chloride were placed along transects in the Sand Lake National Wildlife Refuge in South Dakota. Later, there was no difference in nest success between treated and control plots. The predators in the area were red fox (V. vulpes), raccoon (Procyon lotor), striped skunk (M. mephitis), mink (M. vison), and Franklin's ground squirrel (Spermophilus franklinii). The waterfowl nesting in the area were blue-winged teal (Anas discors), mallard (Anas platyrhynchos), common pintail (Anas acuta), gadwall (Anas strepera), and northern shoveler (Anas clypeata) (Sheaffer and Drobney, 1986).

In a similar study, chicken eggs were injected with emetine dihydrochloride (20–25 mg) and placed along three transects, 0.7 to 1.0 km long. Raccoons, opossums, *Didelphis virginiana*, and striped skunks consumed 75% less treated eggs than untreated controls if the animals were first conditioned and untreated eggs

were offered later. Simultaneously offered eggs of both types resulted in both eaten equally often. This suggests that predators should be conditioned before the nesting season, not after egg laying has begun (Conover, 1990).

Aversive conditioning of beeyard-raiding black bears, *Ursus americanus*, using lithium chloride wrapped in honey or brood comb has been inconclusive. Emetics also proved to be "overspecific": bears avoided only the particular foods that had been used as bait. In Yosemite National Park, black bears were successfully discouraged from raiding the foodsacks of backcountry campers by "aversion sacks" that contained balloons filled with ammonium hydroxide (Hastings and Gilbert 1980).

Social learning can modify conditioned aversions. Spotted hyenas conditioned by lithium chloride treatment lost their aversion in the presence of naive conspecifics (Yoerg, 1991). The reverse occurs in rats: a conditioned aversion is transmitted to naive animals (Strupp and Levitsky, 1984).

13.4.5 Use of mammal noses

While the performance of dogs in rescue and police work is legendary, other mammals have been used to detect fungi, explosives, illegal drugs or diseases.

Tracking by dogs

The domestic dog provides a convenient model for wild canids. Like wild dogs, domestic breeds find prey by tracking with their nose near the ground, or by air scenting, picking up odor from the source directly with nose held high. In air scenting, practitioners speak of a "scent cone" spreading from a point source. The dog quarters in a zigzag pattern – like insects – toward the odor source. The age-old practice of training dogs for hunting and finding lost persons acquired a scientific basis during the first three decades of the twentieth century. For training and testing a dog's performance, Löhner (1926) introduced the method of letting the dog fetch a scented piece of wood of standard size. He found that dogs use human sweat, sebum, and dandruff to identify an individual. Dogs are very responsive to body odor components such as fatty acids up to C_8 – formic, acetic, propionic, valeric, caproic, and caprylic acids – plus nitrogen compounds such as ammonia and skatole, and also sulfur compounds. Contact for only 1–2s between skin and wood left enough scent on the wood for the dog to identify it. Contact with an area as small as a fingertip sufficed. The dogs distinguish whether the odor side on the wood is placed up, sideways, or down when the wood is put on the ground.

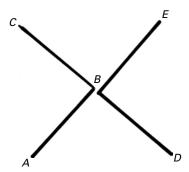


FIGURE 13.4 The track cross was used to test the ability of dogs to follow a track (A to C) that angles back while a second, diversionary track (D to E) leads straight on at point B. (Redrawn from Most and Brückner, 1936.)

Schmid (1935) continued that work. He found that odor from individuals can drift up to 60 m from the actual trail in lateral winds. When confronted with diversionary tracks, dogs circle at the intersection with the real track. They are not distracted by diversionary tracks and still can follow individual horse or dog tracks. Most and Brückner (1936) used a "track cross" (Fig. 13.4) to assess trackfollowing ability. The test track veers off at a right angle to the left or right, while a diversionary track is laid as a straight continuation of the test track. A dog will follow the straight line (i.e. the wrong track) while a specially trained dog would double back and was not distracted by the straight diversionary track. The age of the track matters: trained police dogs discriminated tracks that differed only by 3 minutes in age.

Most and Brückner (1936) found human tracks to be complex. A track contains a human species odor and an individual odor, both extruded through boots, but also the odors of crushed plants, disturbed soil, and leather or other shoe material. Dogs still tracked correctly after removal of one or several of these components. To separate the track components, the authors built a chair lift and a track wheel (Fig. 13.5). The rim of the metal wheel (approximately 2m in diameter) carried raised replicas of shoes at stride intervals. Pulled over the terrain, the wheel makes a track of crushed plants and disturbed soil without human scent. The lift consists of a chair suspended on a steel cable about 1.5 m above the ground. Although a person riding this chair leaves no foot imprints or disturbed soil, he/she still sheds odorous "rafts" of skin cells. A trained dog uses one or the other of these track components, as terrain and surface change.

As discussed in Ch. 5, dogs are extremely sensitive to odors (Neuhaus, 1953, 1956a,b; Neuhaus and Müller, 1954). A dog can identify a human odor against background odors such as bergamot, clove or oregano oil, tobacco smoke,

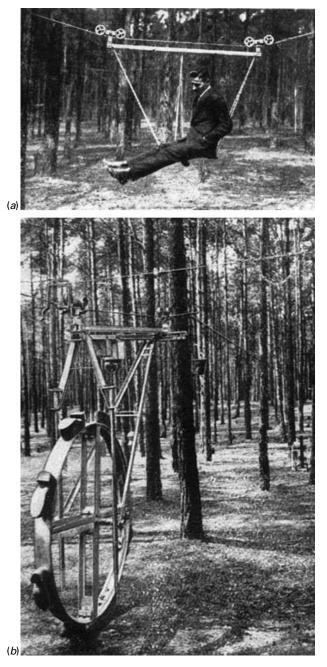


FIGURE 13.5 Analysis of tracking components. (a) A chairlift. (b) The track wheel was used to separate a human track into a person's olfactory footprint and odors emanating from crushed soil and vegetation. The track wheel has wooden footpads on a metal rim, spaced at an average human stride. The wheel produces a track that lacks human scent and consists entirely of odors from disturbed soil and vegetation. (From Most and Brückner, 1936.)

or hay or barn odor. Odor differences between body regions of one individual can be greater than differences between individuals.

Identical twins provide the ultimate test for a dog's prowess in distinguishing individual odors. In his 1875 Anthropological Miscellany: the History of Twins and the Criterion of their Relevant Powers of Nature and Nurture, Francis Galton suggested "it would be an interesting experiment for twins who were closely alike to try how far dogs can distinguish between them by scent." H. Kalmus of the Galton Laboratory in London tested seven trained police dogs and two show dogs skilled at retrieving for their ability to discriminate identical twins by smell. He challenged them to single out a handkerchief scented by a particular person, or to follow a person's track among four other confusing tracks. A 3-year old Alsatian bitch was unable to distinguish handkerchiefs scented by two identical twin sisters if the handkerchiefs were laid out in a row, to be smelled one at a time. Tracking experiments, however, permitted the dog to compare the two tracks of the identical twins simultaneously. In this test, a male Alsatian followed the "correct" track of identical twin A among confusing tracks laid by twin B after sniffing a handkerchief scented by A (Kalmus, 1955). Of these two experiments, the tracking test is more relevant to the question of a predator following a prey odor (i.e. a kairomone).

Can dogs, and by extension any mammalian predator, determine the direction of a track? Thesen *et al.* (1993) found that dogs that approached a track of human footprints at a right angle, found the "right" direction after sniffing only two to five footprints in 3–5 seconds. In these experiments, the footprints were 3 or 20 minutes old and had been laid 1s apart. The dogs apparently were able to use differences in scent concentration in the air above the footprints (Thesen *et al.*, 1993).

Rodents such as gerbils or rats are able to sniff out drugs or explosives. In efforts to find land mines in Mozambique, the Gambian giant pouched rat, *Cricetomys gambianus*, performed well in sniffing out buried TNT scents. (The rats find their buried food stashes by smell.) APOPO, a Belgian organization, trains these giant rats in Tanzania. Trainers condition the animals, harnessed and running along a taut line, for scratching soil at the correct spot to respond to a metallic double click that signals a food reward. Upon this signal, the rat runs back to the handler and receives a food such as banana. These rats found all 20 live mines in a 400 m² area. Rats are well suited for this work. They are abundant, cheap, and small enough for easy transport, too light to detonate mines accidentally, and tire less than dogs. Furthermore, human mine specialists are rare, and metal detectors give many false alarms (Mott, 2004; Cox *et al.*, 2004; Wines, 2004).

The mammalian olfactory sense is also being exploited to detect disease. African giant pouched rats can detect tuberculosis in sputum (Holden, 2004)

and dogs can discriminate urine from patients with bladder cancel from normal urine, but not consistently (Willis *et al.*, 2004).

13.4.6 Odors, earthquakes, and animals

Wild and domestic birds and mammals, even snakes, might be sensitive to cues preceding earthquakes (Deshpande 1986; Tributsch 1988). If odors are among the pertinent stimuli, sulfurous volatiles are the most likely candidates, as sulfur dioxide is commonly released in earthquakes, along with methane and radon. Methane from the earth migrates to the surface through weak zones. In cold zones it reaches the surface unchanged and could possibly be detected by animals, while in hot volcanic lava it will be oxidized to carbon dioxide (Gold and Soter, 1980). The San Francisco earthquake of October 1989 gave reason to examine anew whether the olfactory sense of animals is reliable enough to predict imminent earthquakes.

13.5 Humans

Odors affect human behavior more than we realize. They are now appreciated as important in human health and disease. Above all, the powerful role of learning is impressive. Odors become associated with pleasant and unpleasant experiences and can retain their hedonic value lifelong. This applies to food, to social and sexual relationships, and to environments such as houses, workplaces, or landscapes. Writers rather than scientists have described such anecdotes. In *Remembrances of Things Past*, Marcel Proust evoked a flood of childhood memories by the taste of a madeleine dipped in lime-blossom tea. Jean-Paul Sartre tells in his autobiography *Les Mots* how the halitosis of his grade-school teacher became to him the odor of authority.

New, powerful techniques in chemistry, odor formulations, bioassays, and olfactometry have supplied us with deeper as well as fresh insights into olfactory effects on our behavior. Medicine, psychology, environmental design, occupational safety, air-quality control, marketing, and advertising now consider and contribute to human chemical ecology.

13.5.1 Diet odors

Odors are important for food discrimination, including detection of spoiled or toxic food, and perhaps surprisingly for stimulating food intake. People who become anosmic in the wake of head injuries often loose their appetite. To them food no longer appears desirable; conversely odors stimulate

food intake in obese people (Schecter and Henkin 1974). Reducing stimulating food odors would suggest itself as one measure in treatment of obesity.

Food aversions

Most of us have one or the other food aversion. These aversions develop when we overindulge, are poisoned by a food, or ate the particular food while an illness was developing. It follows that "the time to give children junk food and candy is when they are sick, certainly not when they are healthy and hungry" (Cain, 1982). Drug tolerance and addiction may be acquired in a similar fashion (Siegel, 1979). Aversion therapy for overeating, alcoholism, and drug abuse should couple the odor of food, alcoholic beverages, or drugs, respectively, with the condition of being sick or feeling miserable. So far, these efforts have not been successful. The subjects habituate to the odors if no further negative consequences occur. To counteract habituation, several or many different negative odors are needed. Repeated exposure makes pleasant odors less so and unpleasant odors more pleasant. This applies to employees in the perfume industry and candy stores as well as workers in chemical factories, rendering plants, or pigsties.

Unfortunate food aversions can develop in people who receive chemotherapy. Even though they are nauseous, children have to eat regular meals and, subsequently, they may dislike the food because of its association with feeling sick. Even anorexia nervosa may be the result.

13.5.2 Social and sexual odors

Removing or covering body odors is an age-old preoccupation and the basis of lucrative toiletries and perfume industries. Obviously, odors – good and bad – and their control are extremely important to humans. Yet the often-posed question of *human pheromones* has not been answered satisfactorily. It probably never will if we apply standards and definitions that were developed originally for insects – not even for non-human vertebrates. Stories of potent and irresistible sex attractants have remained the province of imaginative writers, as in *Perfume* by Patrick Süsskind, *Come Out To Play* by Alex Comfort, or *Switch Bitch* by Roald Dahl. The much more meager results scientists have reaped so far probably reflect the real biological state of affairs. First, female odors may be important as *primers* that drive – among other stimuli – the ovarian cycles of associating women (McClintock, 1971; Preti *et al.*, 1986). Second, daily nasal application of a chemical analogue of luteinizing hormone-releasing hormone inhibited ovulation in 25 of 27 women (Bergquist *et al.*, 1979). This is the opposite effect to its usual one of increasing fertility. Third, the body odors of men and women may

signal attraction or repulsion, based on prior pleasant or negative experiences with certain person(s) and/or odors. Finally, recognition of offspring, mate, sibling, or parent may be aided by body odors.

The influence of odors such as perfumes and fragrances on human behavior is assumed to be acquired, and the responses elicited depend on the often complex previous social experiences. The response will be altered if a laboratory experiment eliminates contextual stimuli (Kirk-Smith and Booth, 1987). Social odors include those of the well-known security blankets in toddlers, familiarly scented bed sheets in new surroundings, and treating insomnia with mother's axillary odor on handkerchief. Removal of bad body odors (diet, metabolism defects) that disrupt interpersonal harmony appears to be universal.

Human odors have been used for sinister purposes. In East Germany, the Stasi (state security police) kept a library of dissidents' smells. Samples of their scarves or dirty underwear were kept in several hundred glass jars. Trained dogs were able to compare the odors of antigovernment materials with those of the standards in this smell library (Rosenberg, 1995). At other times, the persecuted have tried to protect themselves with olfactory countermeasures. During the second World War, the Danish inventor Ernst Morch thwarted the bloodhounds of the Gestapo. The dogs searched for Jews hidden in false bottoms of fishing boats on their way to safety in Sweden. Morch perfected a mixture of rabbit's blood and cocaine that disabled the dogs' noses (Thomas, 1996).

13.5.3 Health and safety hazards

Environmental pollution

The chemical ecology of *Homo sapiens* increasingly includes unpleasant odors from chemical and other factories, oil refineries, rendering plants, sewage plants, car and truck exhausts, lignite fuel burning, dairy or chicken farms, feedlots, silage, liquid manure, or open landfills. These odors may present more than mere esthetic problems by physically irritating or even signaling danger from toxins. Discharges from sulfate cellulose plants (Berglund, 1974) and malodors such as hydrogen sulfide from combustion toilets (Lindvall and Svensson, 1974) are well-investigated examples. Many American cities, such as Jacksonville, Florida or Berlin, New Hampshire have struggled with obnoxious industrial odors, mostly from paper mills (the *Aroma of Tacoma*; Pesce, 1990).

Industrial effluents contain sulfur compounds, with their burnt, pungent quality, and fishy or ruinous-smelling nitrogen compounds. Humans are very sensitive to these odors: hydrogen sulfide has an extremely low recognition threshold of 0.00047 ppm (vol.) (Cain, 1978).

Attempts to label dangerous substances with either characteristic or outright unpleasant odors have not been successful with children, the primary victims of accidental poisonings by toxic household products. Children tolerate odors that adults find unpleasant, such as that of butyric acid. The range between the most pleasant and unpleasant odors is much narrower for children around 4 years of age than for adults, and also much narrower than for taste stimuli. This means that olfactory cues are not suited to produce aversive responses in children (Engen, 1974b, Cain, 1978).

Toxic pollutants may affect the chemical ecology of humans indirectly by altering steroid metabolism and other functions, as animal models indicate. The cancer-causing dioxins found in herbicides, for example, may mimic steroid hormones, altering chemical communication. Hormones affect the expression, development, and functioning of both scent production in glands and chemoreception. Fish, amphibians, and mammals provide examples. In Atlantic salmon, S. salar, males lower their response to a priming pheromone from females after exposure to the pesticide cypermethrin (Moore and Waring, 2001). Female newts, Notophthalmus viridescens, suffer impaired sexual pheromone communication and lower mating success after exposure to the pesticide endosulfan (Park et al., 2001). Lastly, low doses of pesticides even affect the next generation: male mice scent-marked their territories more if they had been exposed in utero to dichlorodiphenyltrichloroethane (DDT) and methoxychlor ingested by their pregnant mothers (vom Saal et al., 1995). Zala and Penn (2004) and Clotfelter et al. (2004) have reviewed the effects of endocrine-disrupting chemicals on animal behavior.

13.5.4 Odors in advertising

Much has been expected from odors in advertising and sales, as with fresh bread odor, new car odor, or fragrance samples in magazines. But the last, particularly are controversial: readers of magazines and libraries object to the scent strips that permeate the entire issue. In response to "a couple of hundred letters," the *New Yorker* discontinued running fragrance strips in October 1992 after publishing them for over a year. The media continue to report on novel attempts at using odors to sell products. Popcorn smell in movie theaters is said to lift popcorn sales by 5–15%. There are claims that people spent 45% more money at scented gambling machines. Even the "odor of an honest car salesman" has been tried. Hotels are using floral-citrus fragrances in their lobbies, trying to induce guests to stay longer. Real estate agents advise home sellers to scent the premises with floral-spicy, inviting kitchen odors. In one experiment, people spent more time in textile department stores when odors (at near threshold

levels) were added to the fresh air supply; however, sales increased only slightly (Kleinfield, 1992). *Public Citizen* and other research and lobbying groups reject such manipulation. Because the odors being used are kept secret, replication and scientific scrutiny are impossible. Some companies scent invoices with the urinary odor of androstenone. The unpleasant odor is supposed to induce the recipient to "pay fast to get rid of the stink" (Kleinfield, 1992).

Odor and other senses: movies with scent

Visual stimuli may influence the perceived odor intensity. An odorous and an odorless solution that were slightly tinted with yellow were more often considered odorous than the corresponding clear solutions with and without odor (Engen, 1972). Based on these findings, Cain (1978) pointed out that in the realm of air pollution the sight of black smoke from a factory may drastically increase the number of *odor* complaints whether or not the odor is indeed more intense.

Motion pictures still lack the odor dimension. Experimental scent movies are improving. The Science City in Paris features a small theater that screens films with an odor component. Scent cans on a carousel rotate in position, get activated, and an air stream blows odors toward the viewer at the appropriate scene. A more recent computer-controlled mixing apparatus can provide many more odor combinations.

GLOSSARY

- active space three-dimensional zone near an odor source where the odor concentration is above the biologically relevant threshold of conspecifics; given air movements, the active space typically extends downwind
- aggression aggressive Behaviors such as threat, attack, fighting, or chasing that result in dominance or spacing; injury or death may ensue, but are not the typical outcome (Latin: aggredior, aggressus sum, to get ready, to assert oneself)
- **agonistic** interactions between animals that include attack, fighting, defense, or flight (Greek: *agonistes*, combatant)
- **alarm pheromone** animal-produced odors that arouse conspecifics and/or cause them to flee, attack, or defend in the face of danger such as predators
- **alert odor** Animal-produced odor that causes conspecifics to interrupt ongoing behavior and prepare them for evasive action in the face of disturbance or danger
- **alkaloid** A heterogeneous class of compounds that contain nitrogen and often taste bitter (Arabic: *Al-qaliy*: ashes of saltwort *Salsola kali*)
- allelochemics Interspecific chemical signals (Greek: allel-, mutual)
- **allomone** a chemical substance produced or acquired by an organism, which evokes in the receiver a behavioral or physiological reaction adaptively favorable to the emitter (Greek: *allos*, different)
- **anadromous (fish)** migrating upstream (Greek an-, up, upwards; *dromos*, a running, from *dramein*, to run)
- anosmia inability to perceive all or certain specific odors
- antifeedants chemicals that inhibit food ingestion, usually by herbivores
- **apocrine gland** a modified sweat gland that produces an aqueous secretion and discharges it into the pilosebaceous canal around a hair shaft; milk glands are a type of apocrine gland (Greek: *apo*, away from, off)
- **aposematism, aposematic** an animal signals distastefulness or other dangerous attributes by visual, olfactory, or auditory cues (e.g. warning colors) to would-be predators (Greek *aposemanein*, to announce by sign)
- aversion, primary a non-learned aversion, usually to food
- **avomic** unable to detect chemicals by the vomeronasal organ (Graves and Duvall 1985)
- **bioassay** determination of active compounds by experimental presentation to animals in the field or the laboratory; fractions and compounds to be tested depend on prior responses by the experimental animals (response-guided bioassay)

- **biotransformation** conversion of xenobiotics (compounds ingested with food, but not needed by the organism and potentially harmful) into non-toxic, excretable compounds, usually in the liver
- **body odor** a term used in lieu of pheromone when no clear and compulsory function has been established
- **chemical ecology** the study of interactions among and between organisms and between organisms and their abiotic environment by means of chemical stimuli
- **chemical signal** compound that serves as a messenger between organisms, is processed by the chemical senses, and is usually part of an evolved communication system
- **communication.** To give or exchange information, signals, or messages in any meaningful way; many varied definitions exist depending on the context (Latin: *communicare*, to impart or share)
- **conditioned aversion** avoidance of an originally neutral or attractive food (or other stimulus) by an animal as a consequence of becoming sick (nausea), experiencing pain, or other negative effects after consuming the food; a single experience can have lasting effect
- **coumarin** white, vanilla-flavored crystalline substance for baking, perfumes, etc.; the tree coumarou, *Dipteryx odorata*, yields the sweet-scented tonka bean (native of French Guiana) (French *coumaron*, tonka bean)

detoxication see biotransformation

- **eccrine sweat glands** these produce an aqueous secretion, open directly to the skin surface (not associated with hairs), and occur over most of the body in humans being most numerous on the sole of the foot and the palm of the hand (Greek *ex,* out, and *krinein*, to separate; referring to the production of a liquid secretion without removing other cell contents)
- **exocrine gland** a gland secreting to the surface of the body, in contrast to *endocrine* glands that send secretions to other tissues *within* the body
- **fear substance** (**schreckstoff**) compound(s) released by an animal when encountering a danger, such as a predator or rival, or in the laboratory electric shock or other noxious stimuli; also known as *alarm substance*
- **feeding stimulant** chemicals from plants or animals that facilitate ingestion **flavor** the combination of odor and taste (of a foodstuff)
- **holocrine gland** sebaceous glands, producing a fatty secretion and opening into the pilosebaceous canal of the hair follicle. From Greek *holos*, whole, and *krinein*, to separate, after the mechanism of secreting whereby the entire cell dissolves and forms the secretions)
- **home range** area utilized, but not necessarily defended, by a resident animal, pair, family, or group of animals; it maybe as large as, or larger than, the defended territory and includes all points the resident(s) visit during the non-migratory (especially breeding) phase of the annual cycle
- **homing** an animal's returning to its site of residence after migration, feeding trips, or artificial displacement; the term does not imply any particular mechanism of reaching the target

- **information** in animal communication and cybernetics, this is the property of a signal or through which message something unpredictable and meaningful to a receiver is conveyed; usually measured in bits
- **kairomones** chemical messengers between species where the adaptive benefit falls on the recipient rather than the emitter; can be non-adaptive or maladaptive to the transmitter (Greek: *kairos*, fitness, opportunity, [by stealth, opportunistic)

macrosmatic endowed with keen sense of smell

message information that is transmitted from one individual to another during com-

MHC molecules all-surface glycoprotien encoded by the major histocompatibility complex of genes; they are involved in immunity and cell recognition

mimicry, Batesian resemblance of one animal (the mimic) to another (the model) to the benefit of the mimic, as when the model is dangerous or inedible (described by Henry Walter Bates 1825–1892)

mimicry, Müllerian the resemblance of two species to their mutual advantage, e.g. if two or more distasteful or toxic species share the same visual or chemical pattern or behavior, a predator needs to learn to avoid only one pattern rather than two or more, which reduces predation on the several prey species in question (described by Fritz Müller, 1821–1897)

navigation finding a target by means other than landmarks, usually over large distances **neophobia** avoidance of new food or unfamiliar places.

odor image a biologically relevant odor signal consisting of many chemical compounds, all of which are necessary; reducing the number of components results in loss of activity

odor pattern two or more compounds in certain proportions that are characteristic for a secretion from a particular individual, sex, subspecies, etc.; it is usually used for analytical findings, but the pattern is also thought to be the relevant discrimination cue for the animals themselves

odor profile similar to odor pattern

odor trail chemicals deposited by one individual as it moves along the substrate; it is used as cue by conspecifics, but predators and parasites also cue in on it

odortype genetically based individual odor, as in the urine odors in the laboratory mouse controlled by the major histocompatibility complex

olfaction reception of volatile cues and processing of information about them by means of sensory neurons and higher centers; includes the *main olfactory system* and the *vomeronasal organ*

orientation reaching a target area by means of landmarks

osmetrichia modified hairs involved in chemical communication (scent hairs); they store secretion, release odor, and serve in applying scents to the environment

palatability likelihood that a food will be ingested under specific conditions

phenolics compounds with a benzene ring and a hydroxyl group (French *phene*, for a contaminant that showed up in gas for illumination; Greek: *phenain*: appear, show)

- **pheromone** a chemical released (usually in minute amounts) by one organism into the external environment that is detected by conspecifics resulting in behavioral or physiological changes; these are likely to benefit both individuals (Greek: *pherein*, to carry)
- **pilosebaceous canal** the space around a hair follicle; it receives secretions from sebaceous and apocrine glands and conveys them to the surface of the skin
- **poison** term refers to any substance (of biological or synthetic origin) that is ingested, inhaled, or absorbed and has detrimental systemic effects on respiratory enzymes, neural transmission, or cell-membrane processes; in biology, it is often used for toxic compounds present in potential prey species and it may be associated with visual signals such as "warning colors," as in tropical frogs, or other cues to signal danger to predators (aposematism)
- preference a higher likelihood that an animal approaches or stays near one of two or more stimuli (odors or animals), or eats more of a food than from others, under specific circumstances and given a peculiar set of choices; it is a relative measure and characteristic neither of the stimulus nor of the animal
- **pregnancy block** termination of pregnancy upon exposure to an adult male (or his odor) different from the stud male; it occurs typically up to the blastula stage
- primary aversion the (usually taste) aversion is not learned
- **primer (priming) pheromones** intraspecific chemical cues that modulate physiological processes, such as growth, development, sexual maturation, estrus cycles, or pregnancy
- **releaser pheromones** intraspecific chemical signals that trigger behavioral responses immediately (i.e. within seconds or minutes of exposure); the original notion of a rigid response being triggered is not easily applicable to mammals
- **repellent** natural repellents are secretions that irritate, are malodorous, or are distasteful to other animals, notably predators, and can be ejected or sprayed; synthetic repellents are compounds or concoctions that reduce damage (by consuming or soiling) inflicted by animals to humans, livestock, crops, forests, stored foods, materials, buildings or any other assets
- scent marking depositing glandular secretions, urine, feces, saliva, or their mixtures on the substrate (e.g. soil, rocks, vegetation, or other, often conspicuous features of the environment) or other, conspecifics; the behavior is olfactorily guided as evidenced by sniffing before and after marking and subsequent sniffing by conspecifics and specific, often rigidly elaborated motor patterns are used. The scent mark conveys information on species, sex, age, kinship, reproductive state, social status, or individuality; it facilitates reproductive behavior and regulates social encounters, thereby increasing the inclusive fitness of both marker and addressee
- **scent matching** comparing a scent mark or airborne odor with the odor of a territory owner; this is essential in territorial defense (coined by Gosling, 1981)
- **semiochemicals** chemical compounds or mixtures of compounds that serve as signals in communication. (Greek: *semeion* sign, flag)
- **sex attractant** chemical produced by one sex that causes members of the other sex of the same species to move toward the sender

- **signal** something that conveys information and occasions a certain action or response; it may be a behavior, a chemical, or other communication and it may or may not serve other functions as well
- **Signaling pheromone** provides information to the recipient that may or may not lead to a change in the recipient's behavior; also called *informer pheromone*
- **skin gland** a composite glandular area (macrogland) on the surface of a vertebrate body; it consists mostly of varying proportions of sebaceous and apocrine glands, along with hair, muscles, and blood vessels
- **stress odor** odor emanating from an animal that experiences stress, trauma or danger; in the laboratory, stress odors are stimulated by predators, their odor, or electric foot shock; the odor may transmit information to conspecifics
- **synergism** two or more compounds act together to achieve or enhance a specific biological effect: the total effect is greater than the sum of the component effects and, individually, the compounds may even have no effect
- **terpene** a large class of compounds with the 5-carbon unit isopentane as bulding block; they are classified by multiples of 5-carbon units: monoterpenes (C_{10}), diterpenes (C_{20}), etc. (word derived from *turpentine*)
- **territory** an area defended by an individual, pair, family, clan, or group of animals against conspecifics, and used for breeding, and/or feeding; it should be distinguished from *home range*
- **threshold** smallest amount of a compound detected by a given organism; *detection* threshold and *recognition* threshold may lie at different concentrations
- toxin any poison derived from a plant, animal, or microorganism; toxins can cause damage to organisms that feed on, or come in contact with, the producing organism and they are usually considered to be adaptive for the producer as defensive compounds (often also used for specific chemical compounds present in a venom)
- **trail pheromone** chemical(s) deposited on the substrate that conspecifics follow; common in insects but also occurs in snakes and deer
- **venom** toxins produced by animals and *injected* by stings, spines, or specialized mouth parts such as fangs in snakes; an animal may use venom for preying as well as antipredator defense, as in venomous snakes; venoms have a complex composition and often are potent enzymes
- **volatility** ability of compounds to evaporate and disperse through air, thus suited for distance communication between animals (or even plants); a small molecular size is required
- vomerolfaction the vomeronasal sense; used analogous to olfaction or gustation vomodor the largely non-volatile chemical stimuli received by the vomeronasal organ xenobiotics natural or synthetic compounds that animals ingest but do not need; they are often harmful and have to be detoxified, usually in the liver

REFERENCES

- Abbott, D. H., Baines, D. A., Faukes, C. G., *et al.* (1990). A natural deer repellent: chemistry and behavior. In *Chemical Signals in Vertebrates*, vol. 5, ed. D. W. Macdonald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 599–609. Oxford: Oxford University Press.
- Adams, M. A. and Johnsen, P. B. (1986). Chemical control of feeding in herbivorous and carnivorous fish. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 45–61. New York: Plenum.
- Adams, M. A., Teeter, J. H., Katz, Y., and Johnsen, P. B. (1987). Sex pheromones of the sea lamprey, *Petromyzon marinus*: steroid studies. *Journal of Chemical Ecology* **13**, 387–395.
- Adams, R., Johnson, J. R., and Wilcox, C. F. (1970). *Laboratory Experiments in Organic Chemistry*, 6th edn. London: MacMillan.
- Adron, J. W. and Mackie, A. M. (1978). Studies on the chemical nature of feeding stimulants for rainbow trout, *Salmo gairdneri* Richardson. *Journal of Fish Biology* **12**, 303–310.
- Agami, M. and Waisel, Y. (1988). The role of fish in distribution and germination of seeds of the submerged macrophytes *Najas marina* and *Ruppia maritima*. *Oecologia* **76**, 83–88.
- Ågren, G., Zhou, Q., and Zhong, W. (1989). Ecology and social behavior of Mongolian gerbils, *Meriones unguiculatus*, at Xilinhot, Inner Mongolia, China. *Animal Behaviour* 37, 11–27.
- Ahn, Y.-J., Lee, S.-B., Okubo, T., and Kim, M. (1995). Antignawing factor of crude oil derived from *Thujopsis dolabrata* S. and Z. var. *hondai* sawdust against mice. *Journal of Chemical Ecology* **21**, 263–271.
- Albers, H. E. and Rowland, C. M. (1989). Ovarian hormones influence odor stimulated flank marking behavior in the hamster, *Mesocricetus auratus*. *Physiology and Behavior* **45**, 113–118.
- Alberts, A. C. (1989). Ultraviolet visual sensitivity in desert iguanas: implications for pheromone detection. *Animal Behaviour* **38**, 129–137.
 - (1992a). Constraints on the design of chemical communication systems in terrestrial vertebrates. *American Naturalist* **139**, 562–589.
 - (1992b). Density-dependent scent gland activity in desert iguanas. *Animal Behaviour* **44**, 774–776.
 - (1992c). Pheromonal self-recognition in desert iguanas. *Copeia* 1, 229–231.
 - (1993). Chemical and behavioral studies of femoral gland secrections in iguanid lizards. *Brain Behavior and Evolution* **41**, 255–260.

- Alberts, A. C. and Werner, D. I. (1993). Chemical recognition of unfamiliar conspecifics by green iguanas: functional significance of different signal components. *Animal Behaviour* **46**, 197–199.
- Alberts, A. C., Sharp, T. R., Werner, D. I., and Weldon, P. J. (1992). Seasonal variation of lipids in femoral gland secretions of male green iguanas (*Iguana iguana*). *Journal of Chemical Ecology* **18**, 703–712.
- Alberts, A. C., Jackintell, L. A., and Phillips, J. A. (1994). Effects of chemical and visual exposure to adults on growth, hormones, and behavior of juvenile green iguanas. *Physiology and Behavior* **55**, 987–992.
- Albone, E. S. (1984). *Mammalian Semiochemistry*. The Investigation of Chemical Signals between Mammals. Chichester: Wiley.
- Albone, E. S., Blazquez, N. B., French, J., Long, S. E., and Perry, G. C. (1986). Mammalian semiochemistry: issues and futures, with some examples from a study of chemical signalling in cattle. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 27–36. New York: Plenum.
- Albonetti, B. D. and D'Udine, B. (1986). Social experience occuring during adult life: its effects on socio-sexual olfactory preferences in inbred mice, *Mus musculus*. *Animal Behaviour* **34**, 1844–1847.
- Aleksiuk, M. (1968). Scent mound communication, territoriality, and population regulation in the beaver. *Journal of Mammalogy* **49**, 759–762.
- Alexander, G., Stevens, D., and Bradley, L. R. (1987). Fostering in sheep V. Use of unguens to foster an additional lamb onto a ewe with a single lamb. *Applied Animal Behaviour Science* 17, 95–108.
- Alexander, G., Goodrich, B. S., Stevens, D., and Bradley, L. R. (1989). Maternal interest in lambs smeared with polar and nonpolar substances. *Australian Journal of Experimental Agriculture* **29**, 513–516.
- Ali, M. A. (ed.) (1978). Sensory Ecology. New York: Plenum Press.
- Allan, G. G., Gustafson, D. I., Mikels, R. A., Miller, J. M., and Neogi, S. (1984). Reduction of deer browsing of Douglas fir (*Pseudotsuga menziesii*) seedlings by quadrivalent selenium. *Forest Ecology and Management* 7, 161–181.
- Allison, M. J. (1978). The role of ruminal microbes in the metabolism of toxic constituents from plants. In *Effects of Poisonous Plants on Livestock*, ed. R. F. Keeler, K. R. Van Kampen, and L. J. James, pp. 101–118. New York: Academic Press.
- Alyan, S. H. and Jander, R. (1994). Short-range learning in the house mouse, *Mus musculus*: stages in the learning of directions. *Animal Behaviour* **48**, 285–298.
 - (1997). Interplay of directional navigation mechanisms as a function of near-goal distance: experiments with the house mouse. *Behavioural Processes* **41**, 245–255.
- American Meteorological Association (2000). Glossary of Meterorology, 2nd edn.
- Ames, B. N. and Gold, L. S. (1990). Too many rodent carcinogens: mitogenesis increases mutagenesis. *Science* **249**, 970–971.
- Ames, B. N., Profet, M., and Gold, L. S. (1990). Nature's chemicals and synthetic chemicals: comparative toxicology. *Proceedings of the National Academy of Sciences of the USA* 87, 7782–7786.

- Amoore, J. E., Pelosi, P., and Forrester, L. J. (1977). Specific anosmias to 5α -androst-16-en-3-one and ω -pentadecalactone: the urinous and musky primary odors. *Chemical Senses and Flavour* **2**, 401–425.
- Andelt, W. F., Burnham, K. P., and Manning, J. A. (1991). Relative effectiveness of repellents for reducing mule deer damage. *Journal of Wildlife Management* **55**, 341–347.
- Andelt, W. F., Baker, D. L., and Burnham, K. P. (1992). Relative preference of captive cow elk for repellent-treated diets. *Journal of Wildlife Management* **56**, 164–173.
- Andersen, K. K. and Bernstein, D. T. (1975). Some chemical constituents of the scent of the striped skunk (*Mephitis mephitis*). *Journal of Chemical Ecology* **1**, 493–499.
- Andren, C. (1982). The role of the vomeronasal organs in the reproductive behavior of the adder *Viper berus. Copeia* **1982**, 148–157.
- Angelon, K. A. and Petranka, J. W. (2002). Chemicals of predatory mosquitofish (*Gambusia affinis*) influence selection of oviposition site by *Culex* mosquitoes. *Journal of Chemical Ecology* **28**, 797–806.
- Apfelbach, R. (1973). Olfactory sign stimulus for prey selection in polecats. *Zeitschrift für Tierpsychologie* **33**, 270–273.
- Apfelbach, V. R., Schutz, S., and Slotnick, B. (1990). Eine verhaltensphysiologische Untersuchung zur Ermittlung olfaktorischer Schwellenwerte bei männlichen Ratten. *Zeitschrift für Säugetierkunde* 55, 407–412.
- Applebaum, S. W. and Birk, Y. (1979). Saponins. In *Herbivores: Their Interaction with Secondary Plant Metabolites*, ed. G. A. Rosenthal and D. H. Janzen, pp. 539–566. New York: Academic Press.
- Applegate, R. D., Rogers, L. L., Casteel, D. A., and Novak, J. M. (1979). Germination of cow parsnip seeds from grizzly bear feces. *Journal of Mammalogy* **60**, 655.
- Apps, P., Viljoen, H. W., Richardson, P. R. K., and Pretorius, V. (1989). Volatile components of anal gland secretion of aardwolf (*Proteles cristatus*). *Journal of Chemical Ecology* **15**, 1681–1688.
- Archer, A. T. and Glen, R. M. (1969). Observations on the behavior of two species of honey-guides *Indicator variegatus* (Lesson) and *Indicator exilia* (Cassin). *Los Angeles County Museum Contributions in Science* **160**, 1–6.
- Archibald, J. D. (1996). Fossil evidence for a late Cretaceous origin of 'hoofed" mammals. *Science* **272**, 1150–1152.
- Arnold, G. W. (1966). The special senses in grazing animals. II. Smell, taste, and touch and dietary habits in sheep. *Australian Journal of Agricultural Research* 17, 531–542.
- Arnold, G. W. and Hill, J. L. (1972). Chemical factors affecting selection of food plants by ruminants. In *Phytochemical Ecology*, ed. J. B. Harborne, pp. 71–101. London: Academic Press.
- Arnold, G. W., de Boer, E. S., and Boundy, C. A. P. (1980). The influence of odour and taste on the food preferences and food intake of sheep. *Australian Journal of Agricultural Research* 31, 571–587.
- Arnold, S. J. (1978). Some effects of early experience on feeding responses in the common garter snake, *Thamnophis sirtalis*. *Animal Behaviour* **26**, 455-462.

- (1981a). Behavioral variation in natural populations. I. Phenotypic, genetic and environmental correlations between chemoreceptive responses to prey in the garter snake, *Thamnophis elegans*. *Evolution* **35**, 489–509.
- (1981b). Behavioral variation in natural populations. II. The inheritance of a feeding response in crosses between geographic races of the garter snake, *Thamnophis elegans*. *Evolution* **35**, 510–515.
- Arnould, C. and Signoret, J. P. (1993). Sheep food repellents: efficacy of various products, habituation, and social facilitation. *Journal of Chemical Ecology* **19**, 225–236.
- Arnould, C., Piketty, V., and Levy, F. (1991). Behavior of ewes at parturition toward amniotic fluids from sheep, cows and goats. *Applied Animal Behaviour Science* **32**, 191–196.
- Arnould, C., Orgeur, P., Sempéré, A., and Signoret, J.-P. (1993). Repulsion alimentaire chez trois espèces d'ongulés en situation de pâturage: effet des excréments de chien. *Revue Ecologique (Terre Vie)* **48**, 121–132.
- Arnould, C., Malosse, C., Signoret, J.-P., and Descoins, C. (1998). Which chemical constituents from dog feces are involved in its food repellent effect in sheep? *Journal of Chemical Ecology* **24**, 559–576.
- Arnts, R. R., Peterson, W. B., Seila, R. L., and Gay, B. W., Jr. (1982). Estimates of alphapinene emissions from a loblolly pine forest using an atmospheric diffusion model. *Atmospheric Environment* **16**, 2127–2137.
- Asa, C. S. (1993). Relative contributions of urine and anal sac secretion in scent marks of large felids. *American Zoologist* **33**, 167–172.
- Asa, C. S., Peterson, E. K., Seal, U. S., and Mech, D. L. (1985). Deposition of anal-sac secretions by captive wolves (*Canis lupus*). *Journal of Mammalogy* **66**, 89–93.
- Asa, C. S., Mech, L. D., and Seal, U. S. (1990). The influence of social and endocrine factors on urine-marking by captive wolves (*Canis lupis*). *Hormones and Behavior* **24**, 497–509.
- Ashkenazy, D., Kashman, Y., Nyska, A., and Friedman, J. (1985). Furocoumarins in shoots of *Pituranthos triradiatus* (Umbelliferae) as protectants against grazing by hyrax (*Procavia capensis syriaca*). *Journal of Chemical Ecology* **11**, 231–239.
- Atema, J. (1986). Review of sexual selection and chemical communication in the lobster, Homarus americanus. Canadian Journal of Fisheries and Aquatic Sciences 43, 2283–2290.
- (1988). Distribution of chemical stimuli. In *Sensory Biology of Aquatic Animals*, ed. J. Atema, A. N. Popper, R. R. Fay, and W. H. Tavolga, pp. 29–56. Heidelberg: Springer-Verlag.
- Atema, J., Holland, K., and Ikehara, W. (1980). Olfactory responses of yellow fin tuna (*Thunnus albacares*) to prey odors: chemical search image. *Journal of Chemical Ecology* **6**, 457–465.
- Auffenberg, W. (1981). The Behavioral Ecology of the Komodo Monitor. Gainesville, FL: University Press of Florida.
- Augner, M. and Bernays, E. A. (1998). Plant defence signals and Batesian mimicry. *Evolutionary Ecology* **12**, 667–680.
- Austin, P. J., Suchar, L. A., Robbins, C. T., and Hagerman, A. E. (1989). Tannin-binding proteins in saliva of deer and their absence in saliva of sheep and cattle. *Journal of Chemical Ecology* **15**, 1335–1347.

- Avery, M. L. (1989). Experimental evaluation of partial repellent treatment for reducing bird damage to crops. *Journal of Applied Ecology* **26**, 433–439.
- Avery, M. L., Decker, D. G., and Nelms, C. O. (1992). Use of a trigeminal irritant for wildlife management. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 319–322. New York: Plenum.
- Avery, M. L., Decker, D. G., Humphrey, J. S., et al. (1995). Methyl anthranilate as a rice seed treatment to deter birds. *Journal of Wildlife Management* **59**, 50–56.
- Avery, M. L., Humphrey, J. S., and Decker, D. G. (1997). Feeding deterrence of anthraquinone, anthracene, and anthrone to rice-eating birds. *Journal of Wildlife Management* **61**, 1359–1365.
- Aylor, D. E. (1976). Estimating peak concentrations of pheromones in the forest. In *Perspectives in Forest Entomology*, ed. J. F. Anderson and H. K. Kaya, pp. 177–188. New York: Academic Press.
- Badcock, J. (1986). Aspects of the reproductive biology of *Gonostoma bathyphilus*, Gonostomatidae. *Journal of Fish Biology* **29**, 589–604.
- Bailey, K. (1978). Flehmen in the ring-tailed lemur (*Lemur catta*). Behavior LXV, 309–319.
- Baker, H. G. (1963). Evolutionary mechanisms in pollination biology. *Science* **139**, 877–883.
- Balogh, R. D. and Porter, R. H. (1986). Olfactory preferences resulting from mere exposure in human neonates. *Infant Behavior and Development* **9**, 395–402.
- Balthazart, J. and Schoffeniels, E. (1979). Pheromones are involved in the control of sexual behavior in birds. *Naturwissenschaften* **66**, 55–56.
- Bang, B. G. (1965). Anatomical adaptations for olfaction in the snow petrel. *Nature* **205**, 513–515.
 - (1971). Functional Anatomy of the Olfactory System in Orders of Birds. Basel: Karger.
- Bang, B. G. and Cobb, S. (1968). The size of the olfactory bulb in 108 species of birds. *Auk* **85**, 55–61.
- Bardach, J. E. and Villars, T. (1974). The chemical senses of fishes. In *Chemoreception by Marine Organisms*, ed. P. T. Grant and A. M. Mackie, pp. 49–104. New York: Academic Press.
- Bardach, J. E., Todd, J. D., and Crickmes, R. (1967). Orientation by taste in fish of the genus *Ictalurus. Science* **155**, 1276–1278.
- Barinaga, M. (1999). Salmon follow watery odors home. Science 286, 705-706.
- Barnard, C. J. and Fitzsimmons, J. (1988). Kin recognition and mate choice in mice: the effects of kinship, familiarity and social interference on intersexual interaction. *Animal Behaviour* **36**, 1078–1090.
- Barrette, C. (1977). The social behavior of captive muntjacs (*Muntiacus reevesi*). Zeitschrift für Tierpsychologie **43**, 188–213.
- Barron, S., Gagnon, W. A., Mattson, S. N., *et al.* (1988). The effects of prenatal alcohol exposure on odor associative learning in rats. *Neurotoxicology and Teratology* **10**, 333–340.
- Barrows, E. M. (1995). Animal Behavior Desk Reference. Boca Raton, FL: CRC Press.
- Barthalmus, G. T. and Zielinski, W. J. (1988). *Xenopus* skin mucus induces oral dyskinesias that promote escape from snakes. *Pharmacology, Biochemistry and Behavior* **30**, 957–959.

- Bastakov, V. A. (1986). Preference by young-of-the-year of the edible frog (*Rana esculenta* complex) for their own reservoir ground smell. *Zoologicheskii Zhurnal* **65**, 1864–1868.
- Bate-Smith, E. C. (1972). Attractants and repellents in higher animals. *Proceedings of the Phytochemistry Society* **8**, 45–56.
- Batzli, G. O. and Pitelka, F. A. (1971). Condition and diet of cycling populations of the California vole, *Microtus californicus*. *Journal of Mammalogy* **52**, 141–163.
- Batzli, G. O., Getz, L. L., and Hurley, S. S. (1977). Suppression of growth and reproduction of microtine rodents by social factors. *Journal of Mammalogy* **58**, 583–591.
- Baudinette, R. V., Wheldrake, J. F., Hewitt, S., and Hawke, D. (1980). The metabolism of [14C] phenol by native Australian rodents and marsupials. *Australian Journal of Zoology* **28**, 511–520.
- Baumann, T. W. and Gabriel, H. (1984). Metabolism and excretion of caffeine during germination of *Coffea arabica*. L. *Plant Cell Physiology* **25**, 1431–1436.
- Bäumler, W. and Haag, I. (1989). Sekrete und Delta-16-steroide als Lockmittel für die Rötelmaus (Clethrionomys glareolus). Anzeiger für Schädlingskunde, Pflanzenschutz und Umweltschutz 62, 55–59.
- Bäumler, W. and Hock, V. (1987). Über den Einfluss von Duftstoffen und Sekreten auf Verhalten und Entwicklung der Rötelmaus (*Clethrionomys clareolus*) und sympatrischer Arten. Anzeiger für Schädlingskunde, Pflanzenschutz und Umweltschutz **60**, 105–109.
- Bauwens, D., Nuijten, K., van Wezel, H., and Verheyen, R. F. (1987). Sex recognition by males of the lizard *Lacerta vivipara*: an introductory study. *Amphibia-Reptilia* **8**, 49–58.
- Beaman, A. L. (1988). A novel approach to estimating the odor concentration distribution in the community. *Atmospheric Environment* **22**, 561–568.
- Bean, N. J. (1982). Modulation of agonistic behavior by the dual olfactory system in male mice. *Physiology and Behavior* **29**, 433–437.
- Bean, N. J., Galef, B. G., Jr., and Mason, J. R. (1988). The effect of carbon disulfide on food consumption by house mice. *Journal of Wildlife Management* **52**, 502–507.
- Bearder, S. W. and Randall, R. M. (1978). The use of fecal marking sites by spotted hyenas and civets. *Carnivore* 1, 32–48.
- Beauchamp, G. K. (1976). Diet influences attractiveness of urine in guinea pigs. *Nature* **263**, 587–588.
- Beauchamp, G. K. and Berüter, J. (1973). Source and stability of attractive components in guinea pig (*Cavia porcellus*) urine. *Behavioral Biology* **9**, 43–47.
- Beauchamp, G. K., Doty, R. L., Moulton, D. G., and Mugford, R. A. (1976). The pheromone concept in mammalian chemical communication: a critique. In *Mammalian Olfaction, Reproductive Processes and Behavior*, ed. R. L. Doty, pp. 144–160. New York: Academic Press.
- Beauchamp, G. K., Wysocki, C. J., and Wellington, J. L. (1985). Extinction of response to urine odor as a consequence of vomeronasal organ removal in male guinea pigs. *Neuroscience* **99**, 950–955.
- Beauchamp, G. K., Yamazaki, K., Bard, J., and Boyse, E. A. (1988). Preweaning experience in the control of mating preferences by genes in the major histocompatibility

- complex of the mouse. [Symposium of the American Society of Zoology on Kin Recognition in Animals: Empirical Evidence and Conceptual Issues, New Orleans, December, 1987.] Behavioral Genetics 18, 537–548.
- Beauchamp, G. K., Curran, M., and Yamazaki, K. (2000). MHC-mediated fetal odourtypes expressed by pregnant females influence male associative behavior. *Animal Behaviour* **60**, 289–295.
- Bechler, D. L. (1986). Pheromone and tactile communication in the subterranean salamander, *Typhlomolge rathbuni*. *Proceedings of the Ninth International Congress of Speleology*, Barcelona, Spain, vol. 2, pp. 120–122.
- Becker, J. and van Raden, H. (1986). Meteorologische Gesichtspunkte zur olfaktorischen Navigationshypothese. *Journal of Ornithology* **127**, 1–8.
- Begg, C. M., Begg, K. S., Du Toit, J. T., and Mills, M. G. L. (2003). Scent-marking behaviour of the honey badger, *Mellivora capensis* (Mustelidae), in the southern Kalahari. *Animal Behaviour* **66**, 917–929.
- Bekoff, M. and Wells, M. C. (1980). The social ecology of coyotes. *Scientific American* **242**, 9–30.
- Belcher, A., Epple, G., Küderling, I., and Smith, A. B. III. (1988). Volatile components of scent material from cotton-top tamarin (*Saguinus o. oedipus*): a chemical and behavioral study. *Journal of Chemical Ecology* **14**, 1367–1384.
- Belcher, A. M., Epple, G., Greenfield, K. L., *et al.* (1990). Proteins: biologically relevant components of the scent marks of a primate (*Saguinus fuscicollis*). *Chemical Senses* **15**, 431–446.
- Belete, H., Tikubet, G., Petros, B., Oyibu, W. A., and Otigbuo, I. N. (2004). Control of human African trypanosomiasis: trap and odour preference of tsetse flies (*Glossina morsitans submorsitans*) in the upper Didessa River Valley of Ethiopia. *Tropical Medicine and International Health* **9**, 710–714.
- Bell, C. M. and Harestad, A. S. (1987). Efficacy of pine oil as repellent to wildlife. *Journal of Chemical Ecology* **13**, 1409–1417.
- Bell, D. J. (1986). Social effects on physiology in the European rabbit. *Mammal Review* **16**, 131–138.
- Bellairs, A. (1970). The Life of Reptiles. New York: Universe Books.
- Bellringer, J. F., Pratt, H. P. M., and Keverne, E. B. (1980). Involvement of the vomeronasal organ and prolactin in pheromonal induction of delayed implantation in mice. *Journal of Reproduction and Fertility* **59**, 223–228.
- Belvedere, P., Colombo, L., Giacoma, C., Malacarne, G., and Andreoletti, G. E. (1988). Comparative ethological and biochemical aspects of courtship pheromones in European newts. *Monitore Zoologica Italiano (Italian Journal of Zoology)* 22, 397–403.
- Benjamini, L. (1987). Primer pheromones in the Levant vole (*Microtus guentheri*): activation of reproduction in the female by male-related stimuli. *Phytoparasitica* **14**, 3–14.
- Bennett, M. H. (1968). The role of the anterior limb of the anterior commissure in olfaction. *Physiology and Behavior* **3**, 507–515.

- Bennett, N. and Jarvis, J. U. M. (1988). The social structure and reproductive biology of colonies of the mole-rat, *Cryptomys damareasis* (Rodentia, Bathyersidae). *Journal of Mammalogy* **69**, 293–302.
- Benson, J. M., Seiber, J. N., Bagley, C. V., et al. (1979). Effects on sheep of the milkweeds Asclepias eriocarpa and A. labriformis and of cardiac glycoside-containing derivative materials. Toxicon 17, 155–165.
- Bent, A. C. (1964). Life Histories of North American Nuthatches, Wrens, Thrashers, and Their Allies. New York: Dover.
- Benvenuti, S., Fiaschi, V., Fiore, L., and Papi, F. (1973). Homing performance of inexperienced and directionally trained pigeons subjected to olfactory nerve section. *Journal of Comparative Physiology* **83**, 81–92.
- Benvenuti, S., Ioalé, P., and Papi, F. (1992). The olfactory map of homing pigeons. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 429–434. New York: Plenum.
- Berejikian, B. A., Smith, R, J. F., Tezak, E. B., Schultz, W., and Knudsen, C. M. (1999). Chemical alarm signals and complex hatchery rearing habitats affect antipredator behavior and survival of Chinook salmon (*Oncorhynchus tshawytscha*) juveniles. *Canadian Journal of Fisheries and Aquatic Sciences* **56**, 830–838.
- Berenbaum, M. (1981). Patterns of furanocoumarin production and insect herbivory in a population of wild parsnip (*Pastinaca sativa* L.). *Oecologia* **49**, 236–244.
- Berenbaum, M. R. (1991). Coumarins. In *Herbivores: Their Interactions with Secondary Plant Metabolites*, ed. G. A. Rosenthal and M. R. Berenbaum, pp. 221-249. San Diego, CA: Academic Press.
- Berger, P. J., Sanders, E. H., Gardner, P. D., and Negus, N. C. (1977). Phenolic plant compounds functioning as reproductive inhibitors in *Microtus montanus*. *Science* **195**, 575–577.
- Berger, P. J., Negus, N. C., Sanders, E. H., and Gardner, P. D. (1981). Chemical triggering of reproduction in *Microtus montanus*. *Science* **214**, 69–70.
- Berger, P. J., Negus, N. C., and Rowsemitt, C. N. (1987). Effect of 6-methoxybenzoazolinone on sex ratio and breeding performance in *Microtus montanus*. *Biology of Reproduction* **36**, 255–260.
- Bergeron, J. M., Jodoin, L., and Jean, Y. (1987). Pathology of voles (*Microtus pennsylvanicus*) fed with plant extracts. *Journal of Mammalogy* **68**, 73–79.
- Berglund, B. (1974). Quantitative and qualitative analysis of industrial odors with human observers. *Annals of the New York Academy of Sciences* **237**, 35–51.
- Bergquist, C., Nillius, S. J., and Wide, L. (1979). Inhibition of ovulation in women by intranasal treatment with a luteinizing hormone-releasing agonist. *Contraception* **19**, 497–506.
- Berti, R., Thinès, G., and Lefevre, B. (1982). Effets des informations chimiques provenant d'un milieu habité par des congéneres su l'orientation topographique du poisson cavernicole *Phreatichthys andruzzii* Vinciguerra (Pisces, Cyprinidae). *International Journal of Speleology* **12**, 103–117.

- Beyers, D. W. and Farmer, M. S. (2001). Effects of copper on olfaction of Colorado pikeminnow. *Environmental Toxicology and Chemistry* **20**, 907–912.
- Beynon, R. G., Robertson, D. H. L., Hubbard, S. J., Gaskell, S. J., and Hurst, J. L. (1999). The role of protein binding in chemical communication. In *Advances in Chemical Signals in Vertebrates*, vol. 8, ed. R. E. Johnston, D. Müller-Schwarze, and P. W. Sorensen, pp. 137–147. New York: Kluwer Academic/Plenum.
- Beynon, R. G., Veggerby, C., Payne, C. E., *et al.* (2002). Polymorphism in major urinary proteins: molecular heterogeneity in a wild mouse population. *Journal of Chemical Ecology* **28**: 1429–1446.
- Bierich, J. R. (1981). Puberty. [in German.] Klinische Wochenschrift 59, 985-994.
- Billard, R., Bieniarz, K., Popek, W., Epler, P., and Saad, A. (1989). Observations on a possible pheromonal stimulation of milt production in carp (*Cyprinus carpio* L.). *Aquaculture* 77, 387–392.
- Billewicz, W. Z., Fellowes, H. M., and Thomson, A. M. (1981). Menarche in Newcastle upon Tyne, England, UK, girls. *Annals of Human Biology* **8**, 313–320.
- Birch, E. J., Knight, T. W., and Shaw, G. J. (1989). Separation of male goat pheromones responsible for stimulating ovulatory activity in ewes. *New Zealand Agricultural Research* 32, 337–342.
- Bird, S. and Gower, D. B. (1982). Axillary 5α -androst-11-en-3-one, cholesterol and squalene in men: preliminary evidence for 5α -androst-16-en-3-one being a product of bacterial action. *Journal of Steroid Biochemistry* **17**, 517–522.
- Birke, L. I. A. and Sadler, D. (1987a). Differences in maternal behavior of rats and the socio-sexual development of the offspring. *Developmental Psychobiology.* **20**, 85–100.
 - (1987b). Effects of odor familiarity on the development of systematic exploration in the spiny mouse, *Acomys cahirinus*. *Developmental Psychobiology* **20**, 627–640.
- Bjerselius, R. and Olsen, K. H. (1993). A study of the olfactory sensitivity of the crucian carp (*Carassius carassius*) and goldfish (*Carassius auratus*) to 17α ,20 β -dihydroxy-4-pregnen-3-one. *Chemical Senses* **18**, 427–436.
- Bjerselius, R., Li, W., Teeter, J. H., *et al.* (2000). Direct behavioral evidence that unique bile acids released by larval sea lamprey (*Petromyzon marinus*) function as a migratory pheromone. *Canadian Journal of Fisheries and Aquatic Science* **57**, 557–569.
- Black, G. A. and Dempson, J. B. (1986). A test of the hypothesis of pheromone attraction in salmon migration. *Environmental Biology of Fishes* **15**, 229–235.
- Bland, K. P. and Jubilan, B. M. (1987). Correlation of flehmen by male sheep with female behavior and oestrus. *Animal Behaviour* **35**, 735–738.
- Blasquez, N. B., Long, S. E., Perry, G. C., and Matson, E. D. (1987). Effect of estradiol- 17β on perineal and neck skin glands in heifer calves. *Journal of Endocrinology* **115**, 43–46.
- Blass, E. M. and Teicher, M. H. (1980). Suckling. Science 210, 15-22.
- Blass, E. M., Shuleikina-Turpaeva, K., and Luschiekin, V. (1988). Sensory determinants of nipple-attachment behavior in 2- to 4-day-old kittens. *Developmental Psychobiology* **21**, 365–370.
- Blaustein, A. R. (1981). Sexual selection and mammalian olfaction. *American Naturalist* 117, 1001–1010.

- Blaustein, A. R. and O'Hara, R. K. (1981). Genetic control for sibling recognition? *Nature* **290**, 246–248.
- Blaustein, A. R. and Waldman, B. (1992). Kin recogition in anuran amphibians. *Animal Behaviour* **44**, 207–221.
- Block, M., Volpe, V., and Hayes, M. (1981). Saliva as a chemical cue in the development of social behavior. *Science* **211**, 1062–1064.
- Bloom, S. J. (1975). Olfaction in children one to five years of age. B.A. Thesis, Brown University, Providence, Rhode Island, USA.
- Blum, M. S., Murray, S., Jones, T. H., Rinderer, T. E., and Sylvester, H. A. (1988). Oxygenated compounds in beeswax: identification and possible significance. *Biochemical and Molecular Biology* **91**B, 581–583.
- Blytt, H. J., Guscar, T. K., and Butler, L. G. (1988). Antinutritional effects and ecological significance of dietary condensed tannins may not be due to binding and inhibiting digestive enzymes. *Journal of Chemical Ecology* **14**, 1455.
- Boag, B. (1991). Evaluation of an odour derived from lion faeces on the behavior of wild rabbits. *Annals of Applied Biology* **118** (Suppl. 12).
- Boag, B. and Mlotkiewicz, J. A. (1994). Effect of odor derived from lion feces on behavior of wild rabbits. *Journal of Chemical Ecology* **20**, 631–637.
- Böcskei, Z., Groom, C. R., Flower, D. R., *et al.* (1992). Pheromone binding to two rodent urinary proteins revealed by X-ray crystallography. *Nature* **360**, 186–188.
- Bogert, C. M. (1941). Sensory cues used by rattlesnakes in their recognition of ophidian enemies. *Annals of the New York Academy of Sciences* **41**, 329–343.
- Bolhuis, J. J., Strijkstra, A. M., Moore, E., and van der Lende, K. (1988). Preferences for odors of conspecific non-siblings in the common vole, *Microtus arvalis*. *Animal Behaviour* **36**, 1551–1553.
- Bollinger, K. S. (1980). Scent marking behavior of beaver (*Castor canadensis*) M. Sc. Thesis, University of Massachusetts, Amherst.
- Bonadonna, F. and Bretagnolle, V. (2002). Smelling home: a good solution for burrow-finding in nocturnal petrels? *Journal of Experimental Biology* **205**, 2519–2523.
- Bonadonna, F. and Nevitt, G. A. (2004). Partner-specific odor recognition in an Antarctic seabird. *Science* **306**, 835.
- Bonadonna, F., Villafane, M., Bajzak, C., and Jouventin, P. (2004). Recognition of burrow's olfactory signature in blue petrels, *Halobaena caerulea*: an efficient discrimination mechanism in the dark. *Animal Behaviour* **67**, 893.
- Booth, W. D. (1984). A note on the significance of boar salivary pheromones to the male-effect puberty attainment in gilts. *Animal Production* **39**, 149–152.
 - (1987). Factors affecting the pheromone composition of voided boar saliva. *Journal of Reproduction and Fertility* **81**, 427–432.
 - (1989). Boar pheromones: a model for the integration of endocrinology and exocrinology. *Journal of Endocrinology* **123** (Suppl.), 16.
- Bossert, W. H. and Wilson, E. O. (1963). The analysis of olfactory communication in animals. *Journal of Theoretical Biology* **5**, 443-469.

- Bounds, D. and Pope, G. S. (1960). Light-absorption and chemical properties of mirestrol, the estrogenic substance of *Pueraria mirafica*. *Journal of the Chemical Society* 3696–3705.
- Bouvet, J. F., Godinot, F., Delaleu, J. C., and Holley, A. (1989). Interaction between the olfactory and the trigeminal systems in the frog. *Chemical Senses* **14**, 200.
- Bowen, W. D. (1978). Social organization of the coyote in relation to prey size, Ph.D. Thesis, University of British Columbia.
- Bowers, J. M. and Alexander, B. K. (1967). Mice: individual recognition by olfactory cues. *Science* **158**, 1208–1210.
- Bowers, M. D. and Farley, S. (1990). The behaviour of grey jays, Perisoreus canadensis towards palatable and unpalatable Lepidoptera. *Animal Behaviour* **39**, 699–705.
- Bowers, M. D. and Larin, Z. (1989). Acquired chemical defense in the lycaenid butterfly, *Eumaeus atala. Journal of Chemical Ecology* **15**, 1133–1146.
- Boyd, S. K. and Blaustein, A. R. (1985). Familiarity and inbreeding avoidance in the gray-tailed vole (*Microtus canicaudus*). *Journal of Mammalogy* **66**, 348–352.
- Boyle, R. (1999). Folivorous specialization: adaptations in the detoxification of the dietary terpene, *p*-cymene, in Australian marsupial folivores. *American Zoologist* **39**, 102A.
- Boyle, R. R. and McLean, S. (2004). Constraint of feeding by chronic ingestion of 1,8-cineole in the brushtail possum (*Trichosurus vulpecula*). *Journal of Chemical Ecology* **30**, 757–775.
- Bradbury, J. W. and Vehrencamp, S. L. (1977). Social organization and foraging in emballonurid bats. III–IV. *Behavioral Ecology* 2, 1–29.
- Brannon, E. L. and Quinn, T. B. (1990). Field test of the pheromone hypothesis for homing by Pacific salmon. *Journal of Chemical Ecology* **16**, 603–609.
- Braun, J. J. and Marcus, J. (1969). Stimulus generalization among odorants by rats. *Physiology and Behavior* **4**, 245–248.
- Bray, R. O., Wamboldt, C. L., and Kelsey, R. G. (1991). Influence of sagebrush terpenoids on mule deer preference. *Journal of Chemical Ecology* 17, 2053–2062.
- Brennan, P., Kaba, H., and Keverne, E. B. (1990). Olfactory recognition: a simple memory system. *Science* **250**, 1223–1226.
- Brett, J. R. and MacKinnon, D. (1954). Some aspects of olfactory perception in migrating adult coho and spring salmon. *Journal of the Fisheries Research Board of Canada* 11, 310–318.
- Brett, L. P., Hankins, W. G., and Garcia, J. (1976). Prey-lithium aversions III: Buteo hawks. *Behavioral Biology* **17**, 87–98.
- Brightsmith, D. J. (2002). *The Tambopata Macaw Project*. Washington, DC: Earthwatch Institute.
- Brightwell, R., Dransfield, R. D., Kyorku, C., Goldes, T. K., and Tarimo, S. A. R. (1987). A new trap for *Glossina pallidipes*. *Tropical Pest Management* **33**, 151–159.
- Brinck, C., Erlinge, S., and Sandell, M. (1983). Anal sac secretion in mustelids: a comparison. *Journal of Chemical Ecology* **9**, 727–746.
- Brock, O. G. and Meyers, S. N. (1979). Responses of ingestively native *Lampropeltis getulus* (Reptilia, Serpentes, Colubridae) to prey extracts. *Journal of Herpetology* **13**, 209–212.

- Brodie, E. D., Jr. (1977). Hedgehogs use toad venom in their own defense. *Nature* **268**, 627–628.
- Brodie, E. D., Jr. and Brodie, E. D. III. (1980). Differential avoidance of mimetic salamanders by free-ranging birds. *Science* **208**, 181–182.
- Brodie, E. D. Jr., Hensel, J. L., and Johnson, J. A. (1974). Toxicity of the urodele amphibians *Taricha*, *Notophthalmus*, *Gynops* and *Paramesotriton*. *Copeia* **2**, 506–511.
- Brodie, E. D. Jr., Nowak, R. T., and Harvey, W. R. (1979). The effectivness of anti-predator secrections and behavior of selected salamanders against shrews. *Copeia* 2, 270–274.
- Brodie, E. D., Jr., Ridenhour, B. J., and Brodie, E. D., III. (2002). The evolutionary response of predators to dangerous prey: hotspots and coldspots in the geographic mosaic of coevolution between garter snakes and newts. *Evolution* **56**, 2067–2082.
- Bronmark, C. and Miner, J. G. (1992). Predator-induced phenotypical change in body morphology in Crucian carp. *Science* **258**, 1348.
- Bronson, F. H. (1979). The reproductive ecology of the house mouse. *Quarterly Review of Biology* **54**, 265–299.
- Bronson, F. H. and Coquelin, A. (1980). The modulation of reproduction by priming pheromones in house micee: Speculatons on adaptive function. In *Chemical Signals: Vertebrates and Aquatic Invertebrates*, ed. D. Müller-Schwarze and R. M. Silverstein, pp. 243–265. New York: Plenum.
- Brooksbank, B. W. L. (1970). Labelling of steroids in axillary sweat after administration of ³H-5-pregnenolone and ¹⁴C-progesterone to a healthy man. *Experientia* **26**, 1012–1014.
- Brooksbank, B. W. L., Brown, R., and Gustafsson, J. A. (1974). The detection of 5α -androst-16-en- 3α -ol in human male axillary sweat. *Experientia* **30**, 864–865.
- Brouette-Lahlou, I., Amouroux, R., Chastrette, F., et al. (1991). Dodecyl propionate, attractant from rat preputial gland: characterization and identification. *Journal of Chemical Ecology* 17, 1343–1345.
- Brower, L. P. (1969). Ecological chemistry. Scientific American, 220, 22-29.
- Brower, L. P. and Brower, J. V. Z. (1964). Birds, butterflies, and plant poisons: a study in ecological chemistry. *Zoologica* **49**, 137–159.
- Brower, L. P., Nelson, C. J., Seiber, J. N., Fink, L. S. and Bond, C. (1988). Exaptation as an alternative to coevolution in the cardenolide-based chemical defense of monarch butterflies (*Danaus plexippus* L.) against avian predators. In *Chemical Mediation of Coevolution*, ed. K. C. Spencer, pp. 447–475. San Diego, CA: Academic Press/Harcourt Brace.
- Brown, G. E. and Brown, J. A. (1993). Social dynamics in salmonid fishes: do kin make better neighbours? *Animal Behaviour* **45**, 863–871.
- Brown, G. E. and Godin, G. J. (1999). Chemical alarm signals in wild Trinidadian guppies (*Poecilia reticulata*). *Canadian Journal of Zoology* 77, 562–570.
- Brown, G. E., Brown, J., and Crosbie, A. (1993). Phenotype matching in juvenile rainbow trout. *Animal Behaviour* **46**, 1223–1225.
- Brown, G. E., Adrian, J. C., Smyth, E., Leet, H., and Brennan, S. (2000). Ostariophysan alarm pheromones: laboratory and field tests of the functional significance of nitrogen oxides. *Journal of Chemical Ecology* **26**, 139–154.

- Brown, G. E., Golub, J. L., and Plata, D. L. (2001). Attack cone avoidance during predator inspection visits by wild finescale dace (*Phoxinus neogaeus*): the effect of predator diet. *Journal of Chemical Ecology* **27**, 1657–1666.
- Brown, G. E., Adrian, J. C., Jr., Naderi, N. T., Harvey, M. C., and Kelly, J. M. (2003). Nitrogen oxides elicit antipredator responses in juvenile channel catfish, but not in convict cichlids or rainbow trout: conservation of the ostariophysan alarm pheromone. *Journal of Chemical Ecology* **29**, 1781–1796.
- Brown, R. E. (1988). Individual odors of rats are discriminable independently of changes in gonadal hormone levels. *Physiology and Behavior* **43**, 359–364.
- Brown, R. E. and MacDonald, D. (1985). *Social Odours in Mammals*, vol. 2, p. 635. Oxford: Clarendon Press.
- Brown, R. E., Singh, P. B., and Roser, B. (1987a). Both class I and class II regions of the major histocompatibility complex influence the distinctive urinary odors of congenic rat. *American Zoologist* 27, 47A.
- (1987b). The major histocompatibility complex and the chemosensory recognition of individuality in rats. *Physiology and Behavior* **40**, 65–74.
- Brown, R. E., Hauschild, M., Holman, S. D., and Hutchison, J. B. (1988). Mate recognition by urine odors in the Mongolian gerbil, *Meriones unguiculatus*. *Behavioral and Neural Biology* **49**, 174–183.
- Brown, W. L., Jr., Eisner, T., and Whittaker, R. H. (1970). Allomones and kairomones: transspecific chemical messengers. *BioScience* **20**, 21–22.
- Brown, W. S. and MacLean, F. M. (1983). Conspecific scent-trailing by newborn timber rattlesnakes, *Crotalus horridus*. *Herpetologica* **39**, 430–436.
- Brownlee, R. G., Silverstein, R. M. Müller-Schwarze, D., and Singer, A. G. (1969). Isolation, identification and function of the chief component of the male tarsal scent in black-tailed deer. *Nature* **221**, 284–285.
- Bruce, H. M. (1959). An exteroceptive block to pregnancy in the mouse. *Nature* **184**, 105.
- Brundin, A., Andersson, G., Andersson, K., Mossing, T., and Källquist, L. (1978). Short-chain aliphatic acids in the interdigital gland secretion of reindeer (*Rangifer tarandus* L.), and their discrimination by reindeer. *Journal of Chemical Ecology* **4**, 613–622.
- Bryant, B. P. and Atema, J. (1987). Diet manipulation affects social behavior of catfish: importance of body odor. *Journal of Chemical Ecology* **13**, 1645–1662.
- Bryant, J. P. (1981). Phytochemical deterrence of snowshoe hare browsing by adventitious shoots of four Alaskan trees. *Science* **213**, 889–890.
- Bryant, J. P. and Kuropat, P. J. (1980). Selection of winter forage by subarctic browsing vertebrates: the role of plant chemistry. *Annual Review of Ecology* **11**, 261–285.
- Bryant, J. P., Wieland, G. D., Reichardt, P. B., Lewis, V. E., and McCarthy, M. C. (1983). Pinosylvin methyl ether deters snowshoe hare feeding on green alder. *Science* 222, 1023–1025.
- Bryant, J. P., Tahvanainen, J., Sulkinoja, M. *et al.* (1989). Biogeographic evidence for the evolution of chemical defense by boreal birch and willow against mammalian browsing. *American Naturalist* **134**, 20–34.

- Bryant, J. P., Kuropat, P. J., Reichardt, P. B., and Clausen, T. P. (1991). Controls over the allocation of resources by woody plants to chemical antiherbivore defense. In *Plant Defenses Against Mammalian Herbivory*, ed. R. T. Palo and C. T. Robbins, pp. 83–102. Boca Raton, FL: CRC Press.
- Buchsbaum, R., Valiela, I., and Swan, A. (1984). The role of phenolic acids and other plant constituents in feeding of Canada geese in a coastal marsh. *Oecologia* **63**, 343–349.
- Buck, L. (1996). Information coding in the vertebrate olfactory system. *Annual Review of Neuroscience* **19**, 517–544.
- Buck, L. and Axel, R. (1991). A novel multigene family may encode odorant receptors: a molecular basis for odor recognition. *Cell* **65**, 175–187.
- Bucyanayandi, J. D., Bergeron, J.-M., and Menard, H. (1990). Preference of meadow voles (*Microtus pennsylvaticus*) for conifer seedlings: chemical components and nutritional quality of bark of damaged and undamaged trees. *Journal of Chemical Ecology* **16**, 2569–2579.
- Buesching, C. D., Waterhouse, J. S., and MacDonald, D. W. (2002a). Gaschromatographic analyses of the subcaudal gland secretion of the European badger (*Meles meles*). Part I: chemical differences related to individual parameters. *Journal of Chemical Ecology* 28, 41–56.
 - (2002b). Gaschromatographic analyses of the subcaudal gland secretion of the European badger (*Meles meles*). Part II: time-related variation in the individual-specific composition. *Journal of Chemical Ecology* **28**, 57–69.
- Bugloss, A. J., Darling, F. M. C., and Waterhouse, J. S. (1990). Analysis of the anal sac secretion of Hyaenidae. In *Chemical Signals in Vertebrates*, vol. 5, ed. D. W. MacDonald, D. Müller–Schwarze, and S. T. Natynczuk, pp. 65–69. Oxford: Oxford University Press.
- Buitron, D. and Nuechterlein, G. L. (1985). Olfactory detection of food caches by black-billed magpies. *Condor* **87**, 92–95.
- Bullard, R. W., Turkowski, F. J., and Kilburn, S. R. (1983). Responses of free-ranging coyotes to lures and their modifications. *Journal of Chemical Ecology* **9**, 877–888.
- Burger, B. V., Pretorius, P. J., Spies, H. S. C., Bigalke, R. C., and Geierson, G. R. (1990). Mammalian phenomones VIII: chemical characterization of preorbital gland secretion of grey duiker, *Sylvicapra grimmia* (Artiodactyla: Bovidae). *Journal of Chemical Ecology* 16, 397–416.
- Burger, J. (1989). Following of conspecific and avoidance of predator chemical cues by pine snakes (*Pituophis melanoleucus*). *Journal of Chemical Ecology* **15**, 799–806.
 - (1991). Response to prey chemical cues by hatchling pine snakes (*Pituophis melanoleucus*). Effects of incubation temperature and experience. *Journal of Chemical Ecology* **17**, 1069–1078.
- Burger, J. and Gochfeld, M. (1985). A hypothesis on the role of pheromones on age of menarche. *Medical Hypotheses* 17, 39–46.
- Burger, J., Boarman, W., Kurzava, L. and Gochfeld, M. (1991). Effect of experience with pine (*Pituophis melanoleucus*) and king (*Lampropeltis getulus*) snake odors on Y-maze behavior of pine snake hatchlings. *Journal of Chemical Ecology* 17, 79–87.

- Burghardt, G. M. (1966). Stimulus control of the prey attack response in naive garter snakes. *Psychonomic Science* **4**, 37–38.
 - (1967). Chemical-cue preferences of inexperienced snakes: comparative aspects. *Science* **157**, 718–721.
 - (1970). Intraspecific geographical variation in chemical food cue preferences of newborn garter snakes (*Thamnophia sirtalis*). *Behaviour* **36**, 246–257.
 - (1971). Chemical-cue preferences of newborn snakes: influence of prenatal maternal experience. *Science* **171**, 921–923.
 - (1973). Chemical release of prey attack: extension to naive newly hatched lizards. *Journal of Herpetology* **15**, 77–81.
 - (1975). Chemical prey preference polymorphism in newborn garter snakes *Thamnophis sirtalis* M. *Behavior* **52**, 202–225.
 - (1977). The ontogeny, evolution, and stimulus control of feeding in humans and reptiles. *The Chemical Senses and Nutrition*, ed. M. R. Kare and O. Maller, pp. 253–275. In New York: Academic Press.
 - (1983). Aggregation and species discrimination in newborn snakes. Zeitschrift für Tierpsychologie **61**, 89–101.
- (1992). Prior exposure to prey cues influences chemical prey preference and prey choice in neonatal garter snakes. *Animal Behaviour* **44**, 787–789.
- Burghardt, G. M. and Abeshaheen, J. P. (1971). Responses to chemical stimuli of prey in newly hatched snakes of the genus *Elaphe*. *Animal Behaviour* **19**, 486–489.
- Burghardt, G. M. and Hess, E. H. (1966). Food imprinting in the snapping turtle, *Chelydra serpentina*. *Science* **151**, 108–109.
- Burghardt, G. M., Wilcoxon, H. C., and Czaplicki, J. A. (1973). Conditioning in garter snakes: aversion to palatable prey induced by delayed illness. *Animal Learning and Behavior* 1, 317–320.
- Burghardt, G. M., Goss, S. E., and Schell, F. M. (1988). Comparison of earthworm- and fish-derived chemicals eliciting prey attack by garter snakes (*Thamnophis*). *Journal of Chemical Ecology* **14**, 855–881.
- Burritt, E. A. and Provenza, F. D. (2000). Role of toxins in intake of varied diets by sheep. *Journal of Chemical Ecology* **26**, 1991–2005.
- Bursell, E. (1984). Effects of host odour on the behavior of tsetse. *Insect Science and its Application* **5**, 345–349.
- Bursell, E., Gough, A. J. E., Beevor, P. S., *et al.* (1988). Identification of components of cattle urine attractive to tsetse flies, *Glossina* spp. (Diptera: Glossinidae). *Bulletin of Entomological Research* **78**, 281–291.
- Burwash, M. D., Tobin, M. E., Woolhouse, A. D., and Sullivan, T. P. (1998). Field testing synthetic predator odors for roof rats (*Rattus rattus*) in Hawaiin macadamia nut orchards. *Journal of Chemical Ecology* **24**, 603–630.
- Butterstein, G. M., Schadler, M. H., Lysogorski, E., Robin, L., and Sipperly, S. (1985). A naturally occurring compound, 6-methoxybenzoxyzolinone, stimulates reproductive responses in rats. *Biology of Reproduction* **32**, 1018–1023.
- Buzzell, G. R., Menendez-Pelaez, A., Chlumecky, V., and Reiter, R. J. (1991). Gender differences and time course of castration-induced changes in porphyrins, indoles, and

- proteins in the Harderian gland of the Syrian hamster. *Canadian Journal of Physiology and Pharmacology* **69**, 1814–1818.
- Caicedo, A. and Roper, S. D. (2001). Taste receptor cells that discriminate between bitter stimuli. *Science* **291**, 1557–1560.
- Cain, W. S. (1978). The odoriferous environment and the application of olfactory research. In *Handbook of Perception*, vol. VIA: *Tasting and Smelling*, ed. E. C. Carterette, and M. P. Friedman, pp. 277–304. London: Academic Press.
 - (1982). Odor identification by males and females: predictions vs. performance. *Chemical Senses* 7, 129–142.
- Caine, N. G. and Weldon, P. J. (1989). Responses by red-bellied tamarins (*Saguinus labiatus*) to fecal scents of predatory and non-predatory Neotropical mammals. *Biotropica* **21**, 186–189.
- Calder, C. J. and Gorman, M. L. (1991). The effects of red fox *Vulpes vulpes* faecal odours on the feeding behavior of Orkney voles *Microtus arvalis*. *Journal of Zoology* **224**, 599–606.
- Caldwell, J. P. (1996). The evolution of myrmecophagy and its correlates in poison frogs (family Dendrobatidae). *Journal of Zoology* **240**, 75–101.
- Camazine, S. M. (1983). Mushroom chemical defense: food aversion learning induced by a hallucinogenic toxin, muscimol. *Journal of Chemical Ecology* **9**, 1473–1481.
 - (1985). Olfactory aposematism: association of food toxicity with naturally occurring odor. *Journal of Chemical Ecology* **11**, 1289–1295.
- Camazine, S. M., Resch, J. F., Eisner, T., and Meinwald, J. (1983). Mushroom chemical defense: pungent sesquiterpenoid dialdehyde antifeedant to opossum. *Journal of Chemical Ecology* **10**, 1439–1447.
- Campbell, D. L. (1987). Potential for aversive conditioning in forest animal damage control. In *Proceedings of a Symposium on Animal Damage Management in Pacific Northwest Forests*, March, 1987, pp. 117–118.
- Campbell, D. L. and Bullard, R. W. (1972). A preference-testing system for evaluating repellents for black-tailed deer. *Proceedings of the Vertebrate Pest Conference* **5**, 56–63.
- Campbell, D. L. and Evans, J. (1989). Aversive conditioning with thiram to reduce mountain beaver damage to Douglas-fir seedlings. *Northwest Science* **63**, 70.
- Cantoni, D. and Rivier, L. (1992). Analysis of the secretions from the flank glands of three shrew species and their possible function in a social context. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 99–106. New York: Plenum.
- Carolsfeld, J., Tester, M., Kreiberg, H., and Sherwood, N. M. (1997a). Pheromone-induced spawning of Pacific herring. 1. Behavioral characterization. *Hormones and Behavior* **31**, 256–268.
- Carolsfeld, J., Scott, A. P., and Sherwood, N. M. (1997b). Pheromone-induced spawning of Pacific herring. 2. Plasma steroids distinctive to fish responsive to spawning pheromone. *Hormones and Behavior* **31**, 269–276.
- Carr, W. E. S. (1976). Chemoreception and feeding behavior in the pigfish, *Orthopristis chrysopterus*: characterization and identification of stimulatory substances in a shrimp extract. *Comparative Biochemical Physiology* **55A**, 153–157.

- Carr, W. and Chaney, T. B. (1976). Chemical stimulation of feeding behavior in the pinfish, *Lagodon rhomboides*: characterization and identification of stimulatory substances extracted from shrimp. *Comparative Biochemistry and Physiology* **54A**, 437–441.
- Carr, W., Gondeck, A. R., and Delanoy, R. L. (1976). Chemical stimulation of feeding behavior in the pinfish, *Lagodon rhomboides*: a new approach to an old problem. *Comparative Biochemistry and Physiology* **54A**, 161–166.
- Carter, C. S. and Roberts, R, L. (1997). The psychobiology of cooperative breeding in rodents. In *Cooperative Breeding in Mammals*, ed. N. G. Solomon and J. French, pp. 231–266. New York: Cambridge University Press.
- Carter, C. S., Getz, L. L., Gavish, L., McDermott, J. L., and Arnold, P. (1980). Male-related pheromones and the activation of reproduction in the prairie vole (*Microtus ochrogaster*). *Biology of Reproduction* **23**, 1038–1045.
- Carter, C. S., Getz, L. L., and Cohen-Parsons, M. (1986). Relationships between social organization and behavioral endocriology in a monogramous mammal. *Advances in the Study of Behavior* **16**, 109–146.
- Cave, A. J. E. (1988). Note on olfactory activity in mysticetes. *Proceedings of the Zoological Society of London*, **214**, 307–311.
- Cernoch, J. M. and Porter, R. H. (1985). Recognition of maternal axillary odors by infants. *Child Development* **56**, 1593–1598.
- Chabot, D., Gagnon, P., and Dixon, E. A. (1996). Effect of predator odors on heart rate and metabolic rate of wapiti (*Cervus elaphus canadensis*). *Journal of Chemical Ecology* **22**, 839.
- Champlin, A. K. (1971). Suppression of oestrus in grouped mice: the effect of various densities and the possible nature of the stimulus. *Journal of Reproduction and Fertility* 27, 233–241.
- Chanin, P. (1985). The Natural History of Otters. London: Croom Helm.
- Chapple, R. S. and Wodzicka-Tomaszewska, M. (1987). The learning behavior of sheep when introduced to wheat. II. Social transmission of wheat feeding and the role of the senses. *Applied Animal Behaviour Science* **18**, 163–172.
- Chelazzi, G. and Delfino, G. (1986). A field test on the use of olfaction in homing by *Testudo hermanni* (Reptilia: Testudinidae). *Journal of Herpetology* **20**, 451–455.
- Chen, L. and Martinich, R. L. (1975). Pheromonal stimulation and metabolite inhibition of ovulation in the zebrafish, *Brachydanio rerio*. *NOAA Fisheries Bulletin* **73**, 889–894.
- Chernayeva, G. N., Dolgodvorova, S., and Peryshkina, Y. (1982). Seasonal dynamics of tannin content in European white birch bark. *Rastitielnii Resurci* **18**, 63–66.
- Chien, A. K. (1973). Reproductive behavior of the angelfish *Pterophyllum scalare* (Pisces: Cichlidae) II. Influence of male stimuli upon the spawning rate of females. *Animal Behaviour* **21**, 457–463.
- Chipman, R. K. and Fox, K. A. (1966). Oestrus synchronization and pregnancy blocking in wild house mice (*Mus musculus*). *Journal of Reproduction and Fertility* **12**, 233–236.
- Chiszar, D. (1986). Motor patterns dedicated to sensory function, *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 37–44. New York: Plenum.

- Chiszar, D. and Scudder, K. M. (1980). Chemosensory searching by rattlesnakes during predatory episodes. In *Chemical Signals: Vertebrates and Aquatic Invertebrates*, ed. D. Müller-Schwarze and R. M. Silverstein, pp. 125–139. New York: Plenum.
- Chiszar, D., Radcliff, C. W., Scudder, R. M., and Duvall, D. (1983). Strike-induced chemosensory searching by rattlesnakes: the role of envenomation-related chemical cues in the post-strike environment. In *Chemical Signals in Vertebrates* vol. 3, ed. D.Müller-Schwarze and R. M. Silverstein, pp. 1–24. New York: Plenum.
- Chiszar, D., Nelson, P., and Smith, H. M. (1988a). Analysis of the behavioral sequence emitted by rattlesnakes during feeding episodes III: strike-induced chemosensory searching and location of rodent carcasses. *Bulletin of the Maryland Herpetological Society* **24**, 99–108.
- Chiszar, D., Kandler, K., Lee, R., and Smith, H. M. (1988b). Stimulus control of predatory attack in the brown tree snake, *Boiga irregularis*. 2. Use of chemical cues during foraging. *Amphibia-Reptilia* **9**, 77–88.
- Chiszar, D., Melner, T., and Lee, R. (1990). Chemical cues used by prairie rattlesnakes (*Crotalus viridis*) to follow trails of rodent prey. *Journal of Chemical Ecology* **16**, 79–86.
- Chiszar, D., Fox, K., and Smith, H. M. (1992). Stimulus control of predator behavior in the brown tree snake (*Boiga irregularis*). Behavioral and Neural Biology **57**, 167–169.
- Chiszar, D., Dunn, T. M., and Smith, H. M. (1993a). Response of brown tree snakes (*Boiga irregularis*) to human blood. *Journal of Chemical Ecology* **19**, 91–96.
- Chiszar, D., Grant, H., and Hobart, M. (1993b). Prairie rattlesnakes (*Crotalus viridis*) respond to rodent blood with chemosensory searching. *Brain, Behavior and Evolution* **41**, 229–233.
- Chivers, D. P. and Smith, R. J. F. (1993). The role of olfaction in chemosensory-based predator recognition in the fathead minnow, *Pimephales promelas*. *Journal of Chemical Ecology* **19**, 623–633.
- (1994). The role of experience and chemical alarm signaling in predator recognition by fathead minnows, *Pimephales promelas. Journal of Fish Biology* **44**, 273–285.
- (1998). Chemical alarm signaling in aquatic predator-prey systems: a review and prospectus. *Ecoscience* **5**, 338-352.
- Chivers, D. P., Kiesecker, J. M., Anderson, M. T., and Wildy, B. A. R. (1996). Avoidance response of a terrestrial salamander (*Ambystoma macrodactylum*) alarm cues. *Journal of Chemical Ecology* 22, 1709–1716.
- Chivers, D. P., Kiesecker, J. M., Marco, A., Wildy, E. L., and Blaustein, A. R. (1999). Shifts in life history as a response to predation in Western toads (*Bufo boreas*). *Journal of Chemical Ecology* **25**, 2455-2463.
- Chow, Y. S. and Lin, Y. M. (1986). Actinidine, a defensive secretion of stick insect, *Megacrania alpheus*, Westwood (Orthoptera: Phasmatidae). *Journal of Entomological Science* 21, 97-101.
- Ciofi, C. (1999). The Komodo dragon. Scientific American 280, 84–91.
- Cipollini, M. L. and Levey, D. J. (1997). Antifungal activity of *Solanum* fruit glycoalkaloids: implications for frugivory and seed dispersal. *Ecology* **78**, 799–809.

- Clancy, A. N., Coquelin, A., Macrides, F., Gorski, R. A., and Nobel, E. P. (1984). Sexual behavior and aggression in male mice: involvement of the vomeronasal system. *Journal of Neuroscience* 4, 2222–2229.
- Clapperton, B. K., Minot, E. D., and Crump, D. R. (1988). An olfactory recognition system in the ferret *Mustela furo L.* (Carnivora: Mustelidae). *Animal Behaviour* **36**, 541–553.
 - (1989). Scent lures from anal sac secretions of the ferret, *Mustela furo* L. *Journal of Chemical Ecology* **15**, 291–308.
- Clark, L. (1991). Odor detection thresholds in tree swallows and cedar waxwings. *Auk* **108**, 177–180.
 - (1997). Dermal contact repellents for starlings: foot exposure to natural plant products. *Journal of Wildlife Management* **61**, 1352–1358.
- Clark, L. and Mason, J. R. (1985). Use of nest material as insecticidal and anti-pathogenic agents by the European starling. *Oecologia* **67**, 169–176.
 - (1987). Olfactory discrimination of plant volatiles by the European starling. *Animal Behaviour* **35**, 227–235.
 - (1989). Sensitivity of brown-headed cowbirds to volatiles. Condor 91, 922-932.
- Clark, L. and Shah, P. S. (1992). Information content of prey odor plumes: what do foraging Leach's storm petrels know? In *Chemical Signals in Vertebrates* vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 421–427. New York: Plenum.
 - (1994). Tests and refinements of a general structure–activity model for avian repellents. *Journal of Chemical Ecology* **20**, 321–339.
- Clark, L. and Smeraski, C. A. (1990). Seasonal shifts in odor acuity by starlings. *Journal of Experimental Zoology* **255**, 22–29.
- Clark, L., Shah, P. S. and Mason, J. R. (1991). Chemical repellency in birds: relationship between structure of anthranilate and benzoic acid derivatives and avoidance response. *Journal of Experimental Zoology* **269**, 310–322.
- Clark, M. M., Whiskin, E. E., and Galef, B. G. (2003). Mongolian gerbil fathers avoid newborn male pups, but not newborn female pups: olfactory control of early paternal behavior. *Animal Behaviour* **66**, 441–447.
- Clark, R. W. 2004. Timber rattlesnakes (*Crotalus horridus*) use chemical cues to select ambush sites. *Journal of Chemical Ecology* **30**, 607–617.
- Claus, R. and Hoppen, H. O. (1979). The boar pheromone identified in vegetables. *Experientia* **35**, 1674–1675.
- Claus, R., Hoppen, H. O., and Karg, H. (1981). The secret of truffles: a steroidal pheromone? *Experientia* 37, 1178–1179.
- Claus, R., Dehnhard, M., Götz, U., and Lacorn, M. (2001). The pheromone of the male goat: function, sources, androgen dependency and partial chemical characterization. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 133–140. New York: Kluwer Academic/Plenum.
- Clausen, T. P., Reichardt, P. B., and Bryant, J. P. (1986). Pinosylvin and pinosylvin methyl ether as feeding deterrents in green alder. *Journal of Chemical Ecology* **12**, 2117–2131.

- Clausen, T. P., Bryant, J. P., and Swihart, R. K. (2004). Has browsing by mammals caused continent-scale variation in the chemical defenses of woody plants? In *Annual Meeting of International Society of Chemical Ecology*, July 2004, Ottawa, Canada.
- Clotfelter, E. D., Bell, A. M., and Levering, K. R. (2004). The role of animal behaviour in the study of endocrine-disrupting chemicals. *Animal Behaviour* **68**, 665–676.
- Clulow, F. V. and Baddaloo, E. G. Y. (1987). Influence of odors of male organ homogenates on maturation of young female meadow voles, *Microtus pennsylvanicus*. *Behavioral Processes* 14, 225–228.
- Clulow, F. V. and Clarke, J. R. (1968). Pregnancy-block in *Microtus agrestis*, an induced ovulator. *Nature* **219**, 511.
- Clulow, F. V. and Langford, P. E. (1971). Pregnancy-block in the meadow vole, *Microtus pennsylvanicus*. *Journal of Reproduction and Fertility* **24**, 275–277.
- Cole, K. S. and Smith, R. J. F. (1987). Release of chemicals by prostaglandin-treated female fathead minnows, *Pimephalus promelas*, that stimulate male courtship. *Hormones and Behavior* **21**, 440–456.
 - (1992). Attraction of female fathead minnows, *Pimephales promelas*, to chemical stimuli from breeding males. *Journal of Chemical Ecology* **18**, 1269–1284.
- Coley, P. D., Bryant, J. P., and Chapin, F. S., III (1985). Resource availability and plant antiherbivore defense. *Science* **230**, 895–899.
- Colombo, L., Marconato, A., Belvedere, P. C., and Friso, C. (1980). Endocrinology of teleost reproduction: a testicular steroid pheromone in the black gobi, *Gobius jozo* L. *Bolletino di Zoologia* 47: 355–364.
- Commetto-Muñiz, J. E. and Cain, W. S. (1993). Detection thresholds for an olfactory mixture and its three constituent compounds. *Chemical Senses* **18**, 723–734.
- Conover, M. R. (1990). Reducing mammalian predation on eggs by using a conditioned taste aversion to deceive predators. *Journal of Wildlife Management* **54**, 360–365.
- Cooper, S. M. and Owen-Smith, N. (1985). Condensed tannins deter feeding by browsing ruminants in a South African savanna. *Oecologia* **67**, 142–146.
- Cooper, W. E., Jr. (1989a). Strike-induced chemosensory searching occurs in lizards. *Journal of Chemical Ecology* **15**, 1311–1320.
 - (1989b). Prey odor discrimination in the varanoid lizards: *Heloderma suspectum* and *Varanus exanthematicus*. *Ethology* **81**, 250–258.
 - (1992). Elevation in tongue-flick rate after biting prey in the broad-headed skink, *Eumeces laticeps*. *Journal of Chemical Ecology* **18**, 455–467.
 - (1994). Chemical discrimination by tongue-flicking in lizards: a review with hypotheses on its origin and its ecological and phylogenetic relationships. *Journal of Chemical Ecology* **20**, 439–487.
 - (2000). Responses to chemical cues from plant and animal food by an omnivorous lizard, *Gerrhosaurus validus. Journal of Herpetology* **34**, 614–617.
- Cooper, W. E., Jr. and Alberts, A. C. (1991). Tongue-flicking and biting in response to chemical food stimuli by an iguanid lizard (*Dipsosaurus dorsalis*) having sealed vomeronasal ducts: vomerolfaction may mediate these behavioral processes. *Journal of Chemical Ecology* 17, 135–146.

- Cooper, W. E., Jr. and Garstka, W. (1987). Discrimination of male conspecific from male heterospecific odors by male scincid lizards, Eumeces laticeps. Journal of Experimental Zoology 241, 253–256.
- Cooper, W. E. and Trauth, S. E. (1992). Discrimination of conspecific male and female cloacal chemical stimuli by males and possession of a probable pheromone gland by females in a cordylid lizard, *Gerrhosaurus nigrolineatus*. *Herpetologia* **48**, 229–232.
- Cooper-Driver, G. A. and Swain, T. (1976). Cyanogenic polymorphism in bracken (*Pteridium*) in relation to herbivore predation (*Schistocerca gregaria*). *Nature* **260**, 604.
- Coopersmith, C. B. and Banks, E. M. (1983). Effects of olfactory cues on sexual behavior in the brown lemming, *Lemmus trimucronatus*. *Journal of Comparative Psychology* **97**, 120–126.
- Coopersmith, R. and Leon, M. (1986). Neurobehavioral analysis of odor preference development in rodents. In *Ontogeny of Olfaction: Principles of Olfactory Maturation in Vertebrates*, ed. W. Breiphol, pp. 237–242. Irvine, CA: Department of Psychobiology.
- Coppola, D. M. (1986). The puberty delaying pheromone of the house mouse: field data and a new evolutionary perspective, In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 457–462 New York: Plenum.
 - (2001). The role of the main and accessory olfactory systems in prenatal olfaction. In *Chemical Signals in Vertebrates 9*, ed. A. Marchlewska-Koj, J. J. Lepri and D. Müller-Schwarze, pp. 189–196. New York: Kluwer Academic/Plenum.
- Coppola, D. M. and Coltrane, J. A. (1994). Retronasal or internasal olfaction can mediate odor-guided behaviors in newborn mice. *Physiology and Behavior* **56**, 729–736.
- Coppola, D. M. and O'Connell, R. J. (1988). Behavioral responses of peripubertal female mice towards puberty-accelerating and puberty-delaying chemical signals. *Chemical Senses* 13, 407–424.
- Coppola, D. M. and Vandenbergh, J. G. (1985). Effect of density, duration of grouping and age of urine stimulus on the puberty delay pheromone in female mice. *Journal of Reproduction and Fertility* 73, 517.
- (1987). Induction of a puberty-regulating chemosignal in wild mouse populations. *Journal of Mammalogy* **68**, 86–91.
- Coquelin, A., Clancy, A. N., Macrides, F., Nobel, E. P. and Gorski, R. A. (1984). Pheromonally induced release of luteinizing hormone in male mice: involvement of the vomeronasal system. *Journal of Neuroscience* 4, 2230–2236.
- Cornwell-Jones, C. A., Velasquez, P. Wright, E. L. and McGaugh, J. L. (1988). Early experience influences adult retention of aversively motivated tasks in normal but not DSP4-treated rats. *Developmental Psychobiology* **21**, 177–185.
- Costanzo, J. P. (1989). Conspecific scent trailing by garter snakes (*Thamnophis sirtalis*) during autumn. *Journal of Chemical Ecology* **15**, 2531–2538.
- Cott, H. B. (1947). The edibility of birds. *Proceedings of the Zoology Society of London* **116**, 371–542.
 - (1948). Edibility of the eggs of birds. *Nature* **161**, 8–11.
 - (1952). The palatability of the eggs of birds. *Proceedings of the Zoological Society of London* **122**, 1–54.

- (1953). The palatability of the eggs of birds: illustrated by experiments on the food preferences of the ferret (*Putorius furo*) and cat (*Felis cato*); with notes on other egg-eating carnivores. *Proceedings of the Zoological Society of London* **123**, 123–141.
- (1955). The palatability of the eggs of birds: mainly based upon observations of an egg panel. *Proceedings of the Zoological Society of London* **124**, 335–463.
- Cott, H. B. and Benson, J. M. (1969). The palatability of birds, mainly based upon observations of a tasting panel in Zambia. *Ostrich* **8**, 357–384.
- Coulston, S., Stoddart, D. M., and Crump, D. R. (1993). Use of predator odors to protect chick-peas from predation by laboratory and wild mice. *Journal of Chemical Ecology* **19**, 607–612.
- Coureaud, G., Schaal, B., Langlois, D., and Perriers, G. (2001). Orientation response of newborn rabbits to odours of lactating females: relative effectiveness of surface and milk cues. *Animal Behaviour* **61**:153–162.
- Courtney, A. J. and Masel, J. M. (1997). Spawning stock dynamics of two penaeid prawns, *Metapenaeus bennettae* and *Penaeus esculentus*, in Moreton Bay, Queensland, Australia. *Marine Ecology Progress Series* **148**, 37–47.
- Cowles, R. B. and Phelan, R. L. (1958). Olfaction in rattlesnakes. Copeia 1958, 77-83.
- Cowley, J. J. and Wise, D. R. (1972). Some effects of mouse urine on neonatal growth and reproduction. *Animal Behaviour* **20**, 499–506.
- Cox, C., Weetjens, B., Machangu, R., Billet, M., and Verhagen, R. (2004). Rats for demining: an overview of the APOPO program. www.apopo.org.
- Cox, T. P. (1989). Odor-based discrimination between noncontiguous demes of wild *Mus. Journal of Mammalogy* **70**, 549–556.
- Cox, T. P. and Sacks, O. W. (2002). Cycad neurotoxins, consumption of flying foxes, and ALS-PDC diseases in Guam. *Neurology* **26**, 1664–1665.
- Creigh, S. L. and Terman, C. R. (1988). Reproductive recovery of inhibited male prairie deer mice (*Peromyscus maniculatus bairdii*) from laboratory populations by contact with females or their urine. *Journal of Mammalogy* **69**, 603–607.
- Cronin, E. W., Jr. and Sherman, P. W. (1976). A resource-based mating system: the orange rumped honeyguide. *Living Bird* **15**, 5–32.
- Crozier, E. R. (1991). Practical animal repellents for tree seedlings: a success story. *Forest Research Institute Bulletin* **156**, 172–177.
- Cruz, E., Gibson, S., Kandler, K., Sanchez, G., and Chiszar, D. (1987). Strike-induced chemosensory searching in rattlesnakes: a rodent specialist (*Crotalus viridis*) differs from a lizard specialist (*Crotalus pricei*). *Bulletin of the Psychonomic Society* 25, 136–138.
- Cummings, J. L., Pochop, D. A., Davis, J. E., and Krupa, H. W. (1995). Evaluation of ReJeXiT AG-36 as a Canada goose grazing repellent. *Journal of Wildlife Management* **59**, 47–50.
- Cummins, K. A. and Myers, L. J. (1988). Effect of visual and olfactory alterations on social behavior in lactating dairy cows. *Journal of Dairy Science* **71**(Suppl. 1), 189.
- Cupp, P. V., Jr. (1988). Avoidance of predators by salamanders through the detection of chemical odors. *American Zoologist* **28**, 156A.

- Cushing, B. S. (1983). Responses of polar bears to human menstrual odors. In *International Conference on Bear Research and Management*, pp. 270–274.
 - (1984). A selective preference by least weasels for oestrus versus dioestrous urine of prairie deer mice. *Animal Behaviour* **32**, 1263–1265.
- Cutler, W. B., Preti, G., Huggins, G. R., Garcia, C. R., and Lawley, H. J. (1986). Human axillary secretions influence women's menstrual cycles: the role of donor extract from men. *Hormones and Behavior* **20**, 463–473.
- Dagg, A. I. and Windsor, D. E. (1971). Olfactory discrimination limits in gerbils. *Canadian Journal of Zoology* **49**, 283–285.
- Daltry, J. C., Wüster, W., and Thorpe, R. S. (1996). Diet and snake venom evolution. *Nature* **379**, 537–540.
- Daly, J. W. and Meyers, C. W. (1967). Toxicity of Panamanian poison frogs (*Dendrobates*): some biological and chemical aspects. *Science* **156**, 970–973.
- Daly, J. W. and Spande, T. F. (1986). Amphibian alkaloids: chemistry, pharmacology and biology, In *Alkaloids: Chemical and Biological Perspectives*, vol. 4, ed. S. W. Pelletier, pp. 4–274. New York: Wiley.
- Daly, J. W., Meyers, C. W., Warnick, J. E., and Albuquerque, E. X. (1980). Levels of bactrachotoxin and lack of sensitivity to its action in poison-dart frogs (*Phyllobates*). *Science* **208**, 1383–1385.
- Daly, J. W., Secunda, S. I., Garaffo, H. M., *et al.* (1992). Variability in alkaloid profiles in neotropical poison frogs (Dendrobatidae): genetic versus environmental determinants. *Toxicon* **30**, 887–898.
- Daly, J. W., Garraffo, H. M., Spande, T. F., Jaramillo, C., and Stanley, R. A. (1994). Dietary source for skin alkaloids of poison dart frogs (*Dendrobatidae*)? *Journal of Chemical Ecology* **20**, 943–955.
- Daly, J. W., Garaffo, H. M., Jain, P., *et al.* (2000). Arthropod–frog connection: decahydro-quinoline and pyrrolizidine alkaloids common to microsympatric myrmicine ants and dendrobatid frogs. *Journal of Chemical Ecology* **26**, 73–85.
- Daly, M. and Daly, S. (1975). Behavior of *Psammomys obesus* (Rodentia: Gerbillinae) in the Algerian Sahara. *Zeitschrift für Tierpsychologie* **37**, 298–321.
- Daly, R. P. (1988). Status of the Arabian oryx in Oman. In *Proceedings of the 63rd Meeting of IUCN Species Survival Commission* San Jose, Costa Rica, January, 1988.
- D'Amato, F. and Cabib, S. (1987). Chronic exposure to a novel odor increases pups' vocalizations, maternal care, and alters dopaminergic functions in developing mice. *Behavioral and Neural Biology* **48**, 197–205.
- Danell, K., Huss-Danell, K., and Bergström, R. (1985). Interactions between browsing moose and two species of birch in Sweden. *Ecology* **66**, 1867–1878.
- Danell, K., Gref, F., and Reza, Y. (1990). Effects of mono- and diterpenes in Scots pine needles on moose browsing. *Scandinavian Journal of Forest Research* **5**, 535–539.
- Daranzo, J. P., Sydow, M., and Garris, D. R. (1983). Influence of isolation and training on fighting in mice with olfactory-bulb lesions. *Physiology and Behavior* **31**, 857–860.
- Daumae, M. and Kimura, T. (1986). Analysis of urination pattern of male-male counter marking in mice. *Zoological Society of Japan* (Tokyo), **3**, 1103.

- David, C. T., Kennedy, J. S., Ludlow, A. R., Perry, J. N., and Wall, C. (1982). A re-appraisal of insect flight towards a distant, point source of wind-borne odor. *Journal of Chemical Ecology* **9**, 1207–1215.
- Davis, W. (1996). *One River. Explorations and Discoveries in the Amazon Rain Forest*. New York: Simon and Schuster.
- Dawkins, R. and Krebs, J. R. (1979). Arms races between and within species. *Proceedings of the Royal Society of London Series B* **205**, 489–511.
- Dawley, E. M. (1984). Recognition of individual, sex and species odours by salamanders of the *Plethodon glutinosus–P. jordani* complex. *Animal Behaviour* **32**, 353–361.
 - (1987). Salamander vomeronasal systems: why plethodontids smell well. *American Zoologist* **27**, 166A.
 - (1992). Correlation of salamander vomeronasal and main olfactory system anatomy with habitat and sex: behavioral interpretations. In *Chemical Signals in Verte-brates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 403–409. New York: Plenum.
- Dawson, T. M., Arriza, J. L., Jaworsky. D. E., *et al.* (1993). β -Adrenergic receptor kinase-2 and β -arrestin-2 as mediators of odorant-induced desensitization. *Science* **259**, 825–829.
- Dearing, M. D. (1997). The manipulation of plant toxins by a food-hoarding herbivore, *Ochotona princeps. Ecology* **78**, 774–781.
- de Boer, J. N. (1977). The age of olfactory cues function in chemo-communication among male domestic cats. *Behavioral Processes* **2**, 209–225.
- De Fanis, E. and Jones, G. (1995). The role of odour in the discrimination of conspecifics by pipistrelle bats. *Animal Behaviour* **49**, 835–839.
- De Fraipont, M. (1987). Chemical detection in *Astyanax mexicanus*, Teleostei, Characidae, (cave-dwelling form) as a function of group density. *Annales de la Societé Royale Zoologique de Belgique* **117**, 63–67.
- De Fraipont, M. and Thines, G. (1986). Responses of the cavefish *Astyanax mexicanus* (*Anoptichthys antrobius*) to the odor of known or unknown conspecifics. *Experientia* (Basel) **42**, 1053–1054.
- De Fraipont, M. and Sorensen, P. W. (1993). Exposure to the pheromone $17\alpha,20$ β -dihydroxy-4-pregnen-3-one enhances the behavioral spawning success, sperm production and sperm motility of male goldfish. *Animal Behaviour* **46**, 245–256.
- Dehnhard, M. and Claus, R. (1988). Reliability criteria of a bioassay using rats trained to detect estrus-specific odor in cow urine. *Theriogenology* **30**, 1127–1138.
- De Lorgeril, M., Salen, P., Paiilard, F., *et al.* (2002). Mediterranean diet and the French paradox: two distinct biogeographic concepts for one consolidated scientific theory on the role of nutrition in coronary heart disease. *Cardiovascular Research* 3, 503–515.
- Del Punta, K., Leinders-Zufall, T., Rodriguez, I., *et al.* (2002). Deficient pheromone responses in mice lacking a cluster of vomeronasal receptor genes. *Nature* **419**, 70–74.
- DeMonte, M. and Roeder, J. J. (1990). Responses to inter- and intraspecific scent marks in pine martens (*Martes martes*). *Journal of Chemical Ecology* **16**, 611–618.

- Demski, L. S. and Northcutt, R. G. (1983). The terminal nerve: a new chemosensory system in vertebrates? *Science* **220**, 435–437.
- Denmead, O. T., Simpson, J. R., and Freney, J. R. (1974). Ammonia flux into the atmosphere from a grazed pasture. *Science* **185**, 609–610.
- Deshpande, B. G. (1986). Earthquakes, animals and man: Chapter III: Animal response to earthquakes. *Proceedings of the Indian National Science Academy, Part B, Biological Sciences* **52**, 585–618.
- Desjardins, C., Maruniak, J. A., and Bronson, F. H. (1973). Social rank in house mice: differentiation revealed by ultraviolet visualization of urinary marking patterns. *Science* **182**: 939–941.
- De Souza, L. L., Ferrari, S. F., Da Costa, M. L., and Kern, D. C. (2002). Geophagy as a correlate of folivory in red-handed howler monkeys (*Alouatta belzebul*) from eastern Brazilian Amazonia. *Journal of Chemical Ecology* **28**, 1613–1621.
- Detling, J. K., Dyer, M. I., Procter-Greeg, C., and Winn, D. T. (1980). Plant-herbivore interactions: examination of potential effects of bison saliva on regrowth of *Bouteloua gracilis* (H. B. K.) Lag. *Oecologia* **45**, 26–31.
- Detling, J. K., Ross, C. W., Walmsley, M. H., *et al* (1981). Examination of North American bison saliva for potential plant growth regulators. *Journal of Chemical Ecology* **7**, 239–246.
- Deutsch, J. D. and Nefdt, R. J. C. (1992). Olfactory cues influence female choice in two lekbreeding antelopes. *Nature* **356**, 596–598.
- Devine, G. J., Ingvarsdottir, A., Mordue, W., et al. (2000). Salmon lice, Lepeophtheirus salmonis, exibit specific chemotactic responses to semiochemicals originating from the salmonid, Salmo salar. Journal of Chemical Ecology 26, 1833–1847.
- Devine, M. C. (1977). Copulatory plugs, restricted mating opportunities and reproductive competition among male garter snakes. *Nature* **267**, 345–346.
- Dewsbury, D. A. (1981). Effects of novelty on copulatory behavior. *Psychological Bulletin* **89**, 464–482.
- Dial, B. E., Weldon, P. J., and Curtis, B. (1989). Chemosensory identification of snake predators (*Phyllorhynchus decurtatus*) by banded geckos (*Coleonyx variegatus*). *Journal of Herpetology* **23**, 224–229.
- Dickinson, C. and Keverne, E. B. (1988). Importance of noradrenergic mechanisms in the olfactory bulbs for the maternal behavior of mice. *Physiology and Behavior* **43**, 313–316.
- Dickman, C. R. (1992). Predation and habitat shift in the house mouse, *Mus domesticus*. *Ecology* **73**, 313–322.
- Dickman, C. R. and Doncaster, C. P. (1984). Responses of small mammals to red fox (*Vulpes vulpes*) odour. *Journal of Zoology* (London) **204**, 521–531.
- Dieterlen, F. (1959). Das Verhalten des Syrischen Goldhamsters. *Zeitschrift für Tierpsychologie* **16**, 47–103.
- Dietz, B. A., Hagerman, A. E., and Barrett, G. W. (1994). Role of condensed tannin on salivary tannin-binding proteins, bioenergetics and nitrogen digestibility in *Microtus pennsylvanicus*. *Journal of Mammalogy* **75**, 880–889.

- Dimock, E. J., Silen, R. E., and Allen, V. E. (1976). Genetic resistance on Douglas-fir to damage by snowshoe hare and black-tailed deer. *Forest Science* **22**, 106–121.
- Distel, R. A. and Provenza, F. D. (1991). Experience early in life affects voluntary intake of beachbrush by goats. *Journal of Chemical Ecology* **17**, 431–450.
- Dixon, M. D., Johnson, W. C., and Adkisson, C. S. (1997). Effects of caching on acorn tannin levels and Blue Jay dietary performance. *Condor* **99**, 756–764.
- Dluzen, D. E., Ramirez, V. D., Carter, C. S., and Getz, C. C. (1981). Male vole urine changes luteinizing hormone-releasing hormone and norepinephrine in female olfactory bulb. *Science* **212**, 573–575.
- Dmitrieva, T. M. and Ostroumov, V. A. (1986). Role of chemocommunication in the organization of the spawning behavior of the bullhead, *Cottocomephorus grewingki* (Dyb.). *Biologicheskie Nauki* (Moscow) **10**, 38–42.
- Do, J. T., Sullivan, R. M., and Leon, M. (1987). Differential respiration during training is not required for early olfactory learning in infant rats. *Society of Neuroscience Abstracts* 13, 1402.
- Dohi, H., Yamada, A. and Entsu, S. (1991). Cattle feeding deterrents emitted from cattle feces. *Journal of Chemical Ecology* **17**, 1197–1203.
- Dominic, C. J. (1964). Source of the male odour causing pregnancy-block in mice. *Journal of Reproduction and Fertility* **8**, 266–267.
- Dorries, K. M., Adkins-Regan, E., and Halpern, B. P. (1991). Sex difference in olfactory sensitivity to the boar chemosignal, androstenone, in the domestic pig. *Animal Behaviour* **42**, 403–411.
- Dorries, K. M., Adkins-Regan, E., and Halpern, B. P. (1997). Sensitivity and behavioral responses to the pheromone androstenone are not mediated by the vomeronasal organ in domestic pigs. *Brain, Behavior and Evolution* **49**, 53–62.
- Doty, R. L. (1972). Odor preferences of female *Peromyscus maniculatus bairdii* for male mouse odors of *P. m. bairdii* and *P. leucopus noveboracensis* as a function of estrous state. *Journal of Comparative and Physiological Psychology* **81**, 191–197.
 - (1981). Human olfaction. Chemical Senses 6, 351-376.
 - (1986). Ontogeny of human olfactory function. In *Ontogeny of Olfaction*. Principles of Olfactory Maturation in Vertebrates, ed. W. Breipohl, pp. 3–17. Berlin: Springer-Verlag.
- Doty, R. L. and Ferguson-Segall, M. (1989). Influence of adult castration on the olfactory sensitivity of the male rat: a signal detection analysis. *Behavioral Neuroscience* **103**, 691–694.
- Doty, R. L., Green, P. A., Ram, C., and Yankell, S. L. (1982). Communication of gender from human breath odors: relationship to perceived intensity and pleasantness. *Hormones and Behavior* **16**, 13–22.
- Doty, R. L., Shaman, P., Applebaum, S. L., *et al.* (1984). Smell identification ability: changes with age. *Science* **226**, 1441–1443.
- Douglas, H. D., III, Co, J. E., Jones, T. H., and Conner, W. E. (2004). Interspecific differences in *Aethia* spp. auklet odorants and evidence for chemical defense against ectoparasites. *Journal of Chemical Ecology* **30**, 1921–1935.

- Døving, K. B. and Pinching, A. J. (1973). Selective degeneration of neurons in the olfactory bulb following prolonged odour exposure. *Brain Research* **52**, 115–129.
- Døving, K. B. and Trotier, D. (1998). Structure and function of the vomeronasal organ. *Journal of Experimental Biology*, **201**, 2913–2925.
- Døving, K., Selset, B., and Thommsen, R. (1980). Olfactory sensitivity to bile acids in salmonid fishes. *Acta Physiologica Scandinavica* **108**, 123–131.
- Døving, K. B., Westerberg, H., and Johnsen, P. B. (1985). Role of olfaction in the behavior and neuronal responses of Atlantic salmon, *Salmo salar*, to hydrographic stratification. *Caradian Journal of Fisheries and Aquatic Science* **42**, 1658–1667.
- Døving, K. B., Trotier, D., Rosin, J. F., and Holley, A. (1993). Functional architecture of the vomeronasal organ of the frog (genus *Rana*). *Acta Zoologica* **74**, 173–180.
- Downum, K. R., Villegas, S., Rodriguez, E., and Keil, D. J. (1989). Plant photosensitizers: a survey of their occurrence in arid and semiarid plants from North America. *Journal of Chemical Ecology* **15**, 345–355.
- Drea, C. M., Vignieri, S. N., Cunningham, S. B., and Glickman, S. E. (2002). Responses to olfactory stimuli in spotted hyenas (*Crocuta crocuta*): I. Investigation of environmental odors and the function of rolling. *Journal of Comparative Psycholology* **116**, 331–341.
- Drew, G. S., Fagre, D. B., and Martin, D. J. (1988). Scent-station surveys for cottontail rabbit populations. *Wildlife Society Bulletin* **16**, 396–398.
- Drickamer, L. C. (1977). Delay of sexual maturation in female housemice by exposure to grouped females or urine from grouped females. *Journal of Reproduction and Fertility* **51**, 77–81.
 - (1979). Acceleration and delay of first estrus in wild Mus musculus. Journal of Mammology **60**, 215–216.
 - (1982). Acceleration and delay of sexual maturation in female house mice by urinary cues: dose levels and mixing uring from different sources. *Animal Behaviour* **30**, 456–460.
 - (1984a). Acceleration of puberty by a urinary chemosignal from pregnant and lactating *Mus musculus. Journal of Mammalogy* **65**, 697–699.
 - (1984b). Effects of very small doses of urine on acceleration and delay of sexual maturation in female house mice. *Journal of Reproduction and Fertility* **71**, 475–477.
 - (1986). Behavioral aspects of rodent urinary chemosignals. American Zoologist 26, 118A.
 - (1989a). Patterns of deposition of urine containing chemosignals that affect puberty and reproduction by wild stock male and female house mice (*Mus domesticus*). *Journal of Chemical Ecology* **15**, 1407–1421.
 - (1989b). Odor preferences of wild stock female house mice (*Mus domesticus*) tested at three ages using urine and other cues from conspecific males and females. *Journal of Chemical Ecology* **15**, 1971–1987.
 - (1989c). Pregnancy block in wild stock house mice, *Mus domesticus*: olfactory preferences of females during gestation. *Animal Behaviour* **37**, 690–698.
 - (1992). Oestrous female house mice discriminate dominant from subordinate males and sons of dominant from sons of subordinate males by odour cues. *Animal Behaviour* **43**, 868–870.

- (2001a). Ecological aspects of house mouse urinary chemosignals. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri and D. Müller-Schwarze, pp. 35–41. New York: Kluwer Academic/Plenum.
- (2001b). Intrauterine position effects on rodent urinary chemosignals. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 211-216. New York: Kluwer Academic/Plenum.
- Drickamer, L. C. and Hoover, J. E. (1979). Effects of urine from pregnant and lactating female house mice on sexual maturation of juvenile females. *Developmental Psychobiology* **12**, 545–551.
- Drickamer, L. C. and Lenington, S. (1987). T-locus effects on the male urinary chemosignal that accelerates puberty in female mice. *Animal Behaviour* **35**, 1581–1582.
- Drickamer, L. D. and McIntosh, T. K. (1980). Effects of adrenal ectomy on the presence of a maturation-delaying pheromone in the urine of female mice. *Hormones and Behavior* **14**, 146–152.
- Drickamer, L. C. and Mikesic, D. G. (1990). Urinary chemosignals, reproduction, and population size for house mice (*Mus domesticus*) living in field enclosures. *Journal of Chemical Ecology* **16**, 2955–2968.
- Drummond, H. (1985). The role of vision in the predatory behavior of natricine snakes. *Animal Behaviour* **33**, 206–215.
- Drummond, H. and Garcia, M. (1995). Congenital responsiveness of garter snakes to a dangerous prey abolished by learning. *Animal Behaviour* **49**, 891–900.
- Dubost, G. and Feer, F. (1981). The behavior of the male *Antilope cervicapra L.*, its development and social rank. *Behaviour* **76**, 62–127.
- Ducey, P. K. and Ritsema, P. (1988). Intraspecific aggression and responses to marked substrates in *Ambystoma maculatum*, (Caudata: Ambystomatidae). *Copeia* 1988, 1008–1013.
- Duchamp-Viret, P., Chaput, M. A. and Duchamp, A. (1999). Odor response properties of rat olfactory receptor neurons. *Science* **284**, 2171–2174.
- Duellman, W. E. and Trueb, L. (1986). Biology of Amphibians. New York: McGraw Hill.
- Dulka, J. G., Stacey N. E., and Sorenson, P. W. (1986). A gonadal sex steroid: $17\alpha-20\beta$ -dihydroxy-4-pregnen-3-one, acts as a pheromone to rapidly increase gonadotropin and milt volume in male goldfish *Carassius auratus*. *Society for Neuroscience Abstracts* **12**, 1413.
- Dulka, J. G., Stacey, N. E., Sorensen, P. W. and van der Kraak, G. J. (1987). A steroid sex pheromone synchronizes male–female spawning readiness in goldfish. *Nature* **325**, 251–253.
- Dumbacher, J. P. (2003). Natural history of chemical defense in New Guinea birds. In *Proceedings of the 10th International Symposium on Chemical Signals in Vertebrates*, July, 2003, Oregon State University, Corvallis.
- Dumbacher, J. P., Beehler, B. M., Spande, T. F., Garraffo, H. M., and Daly, W. (1992). Homobatrachotoxin in the genus *Pitohui*: chemical defense in birds? *Science* **258**, 799–801.

- Dumbacher, J. P., Spande, T. F., and Daly, J. W. (2000). Batrachotoxin alkaloids from passerine birds: a second toxic bird genus (*Ifrita kowaldi*) from New Guinea. *Proceedings of the National Academy of Sciences of the USA* **97**, 12970–12975.
- Dunbar, I. F., Ranson, E., and Buehler, M. (1981). Pup retrieval and maternal attraction to canine amniotic fluids. *Behavioral Processes* **6**, 249–260.
- Duncan, M. W. (1991). Role of the cycad neurotoxin BMAA in the amyotrophic lateral sclerosis–parkinsonism dementia complex of the western Pacific. *Advances in Neurology* **56**, 301–310.
- Dunn, G. C., Price, E. O., and Katz, L. S. (1987). Fostering calves by odor transfer. *Applied Animal Behaviour Science* 17, 33–39.
- Dusenberry, D. B. 1992. Sensory Ecology. New York: W. H. Freeman.
- Duvall, D. (1986). Snake, rattle and roll. Natural History 95, 66-73.
- Duvall, D. and Chiszar, D. (1990). Behavior and chemical ecology of vernal migration and pre-and poststrike predatory activity in prairie rattlesnakes: field and laboratory experiments. In *Chemical Signals in Vertebrates*, vol. 5, ed. D. W. Macdonald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 539–554 New York: Plenum.
- Duvall, D., Scudder, K. M., and Chiszar, D. (1980). Rattlesnake predatory Behavior: mediation of prey discrimination and release of swallowing by cues arising from envenomated mice. *Animal Behaviour* **28**, 674–683.
- Duvall, D., Graves, B. M., and Carpenter, G. C. (1987). Visual and chemical composite signaling effects of *Sceloporus* lizard fecal boli. *Copeia* **1987**, 1028–1031.
- Dyer, M. I. (1980). Mammalian epidermal growth factor promotes plant growth. *Proceedings of the National Academy of Sciences, USA* 77, 4836–4837.
- Eberhard, I. H., McNamara, J., Pearse, R. N., and Southwell, I. A. (1975). Ingestion and excretion of *Eucalyptus punctata* D. C. and its essential oil by the koala, *Phascolarctas cinereus* (Goldfuss). *Australian Journal of Zoology* **23**, 169–179.
- Ebling, F. J. (1977). Hormonal control of mammalian skin glands. In *Chemical Signals in Vertebrates*, vol. 1, ed. D. Müller-Schwarze and M. M. Mozell, pp. 17–33. New York: Plenum.
- Egid, K. and Brown, J. L. (1989). The major histocompatibility complex and female mating preference in mice. *Animal Behaviour* **38**, 548–549.
- Egid, K. and Lenington, S. (1985). Responses of male mice to odor of females: effects of T- and H2-locus genotype. *Behavior and Genetics* **15**, 287–295.
- Ehrenfeld, J. G. and Ehrenfeld, D. W. (1973). Externally secreting glands of freshwater and sea turtles. *Copeia* **1973**, 305–314.
- Ehrlich, P. R. and Raven, P. H. (1964). Butterflies and plants: a study in coevolution. *Evolution* **18**, 586–608.
- Eibl-Eibesfeldt, I. (1949). Über das Vorkommen von Schreckstoffen bei Erdkrötenquappen. Experientia 5, 236.
- Eidelman, A., Katz, M., Good, A., and Roken, A. (1987). Mothers' recognition of their newborns by olfactory cues. *Pediatric Research* 21, 180A.
- Eisenberg, J. F. and Gould, E. (1970). *Smithsonian Contributions to Zoology*, No. 27: The *Tenrecs: A Study in Mammalian Behavior and Evolution*. Washington, DC: Smithsonian Institution Press.

- Eisner, T. (1989). Prospecting for nature's chemical riches. *Issues in Science and Technology* **6**, 31-34.
- Eisner, T. and Grant, R. P. (1981). Toxicity, odor aversion and "olfactory aposematism." *Science* **213**, 213–476.
- Eisner, T., Conner, M. E., Hicks, K., *et al.* (1977). Stink of stinkpot turtle identified: ω -phenylalkanoic acids. *Science* **196**, 1347–1349.
- Eleftheriou, B. E., Bronson, F. H., and Zarrow, M. X. (1962). Interaction of olfactory and other environmental stimuli on implantation in the deer mouse. *Science* **137**, 764.
- Elkinton, J. S., Cardé, R. T., and Mason, C. J. (1984). Evaluation of time-average dispersion models for estimating pheromone concentrations in a deciduous forest. *Journal of Chemical Ecology* **10**, 1081–1108.
- Ellin, R. I., Farrand, R. L., Oberst, F. W., *et al.* (1974). An apparatus for the detection and quantitation of volatile human effluents. *Journal of Chromatography* **100**, 137–152.
- Ellingsen, O. F. and Døving, K. B. (1986). Chemical fractionation of shrimp extracts inducing bottom food search behavior in cod (*Gadus morhua L.*). *Journal of Chemical Ecology* **12**, 155–168.
- Elliot, E. J. (1986). Chemosensory stimuli in feeding behavior of the leech *Hirudo medicinalis*. *Journal of Comparative Physiology* **159**, 391–401.
- Elliot, S. and Loudon, A. (1987). Effects of monoterpene odors on food selection by red deer calves (*Cervus elaphus*). *Journal of Chemical Ecology* **13**, 1343–1349.
- Ellis, H. H. (1920). Sexual Selection in Man. Philadelphia, PA: F. A. Davis.
- El-Sayed, S. Z. (1988). Fragile life under the ozone hole. Natural History 97, 72.
- Emlen, S. 1969. Homing ability and orientation in the painted turtle *Chrysemys picta marginata*. Behaviour **33**, 58–76.
- Endler, J. A. and McLellan, T. (1988). The processes of evolution: toward a newer synthesis. *Annual Review of Ecology and Systematics* 395–421.
- Engelhart, A. and Müller-Schwarze, D. (1995). Responses of beavers (*Castor canadensis* Kuhl) to predator chemicals. *Journal of Chemical Ecology* **21**, 1349–1364.
- Engen, T. (1972). The Effect of Expectation on Judgements of Odor. [Research Report ICRL-RR-70-11.] Washington, DC: Injury Control Research Laboratory, US Department of Health, Education and Welfare.
 - (1974a). Method and theory in the study of odor preferences. In *Human Responses to Environmental Odors*, ed. A. Turk, J. Johnston, and D. Moulton, pp. 121–141. New York: Academic Press.
 - (1974b). The potential usefulness of sensations of odor and taste in keeping children away from harmful substances. *Annals of New York Academy of Sciences* **237**, 224–228.
 - (1986). The combined effect of carbon monoxide and alcohol on odor sensitivity. *Environment International* **12**, 207–210.
- Enns, H. P. and Hornung, D. E. (1988). Comparisons of the estimate of smell, taste and overall intensity in young and elderly people. *Chemical Senses* **13**, 131–140.
- Enomoto, S., Shoji, T., Taniguchi, M., and Kurihara, K. (1992). Role of lipids of receptor membranes in odor reception. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 55–58. New York: Plenum.
- Ense rink, M. (2002). What mosquitoes want: secrets of host attraction. Science 298, 90-92.

- Epple, G., Belcher, A. M., and Smith, A. B., III. (1986). Chemical signals in callitrichid monkeys: a comparative review. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 653–672. New York: Plenum.
- Epple, G., Küderling, I. and Belcher, A. (1988). Some communicatory functions of scent marking in the cotton-top tamarin (*Saguinus oedipus oedipus*). *Journal of Chemical Ecology* **14**, 503–515.
- Epple, G., Mason, J. R., Nolte, D. L., and Campbell, D. L. (1993). Effects of predator odors on feeding in the mountain beaver (*Aplodontia rufa*). *Journal of Mammology* **74**, 715–722.
- Erlinge, S., Sandell, M., and Brinck, C. (1982). Scent-marking and its territorial significance in stoats, *Mustela erminea*. *Animal Behaviour* **30**, 811–818.
- Estes, R. D. (1972). The role of the vomeronasal organ in mammalian reproduction. *Mammalia* **36**, 315–342.
- Evans, C. M., Mackintosh, J. H., Kennedy, J. T., and Robertson, S. M. (1978). Attempts to characterise and isolate aggression-reducing olfactory signals from the urine of female mice *Mus musculus*. *Physiology and Behavior* **20**, 129–134.
- Eyck, G. T. and Halpern, M. (1988). Aggregation in infant corn snakes (*Elaphe guttata*) and garter snakes (*Thamnophis radix*). *Chemical Senses* 13, 740.
- Faak, M. (ed.) (1990). Die amerikanische Reise. A. v. Humboldt's Travel Log, vol. 2. Berlin Akademie-Verlag.
- Faeth, S. H. (1992). Do defoliation and subsequent phytochemical responses reduce future herbivory on oak trees? *Journal of Chemical Ecology* **18**, 915–925.
- Fagre, D. B., Howard, W. E., Barnum, D. A., et al. (1983). Criteria for the development of coyote lures. In *Proceeding of the 4th Symposium of the ASTM STP 812 on Vertebrate Pest Control and Management Materials*, ed. D. E. Kaukeinen, pp. 265–277. Philadelphia, PA: American Society for Testing and Materials.
- Farentinos, R. C., Capretta, P. J., Kepner, R. E., and Littlefield, V. M. (1981). Selective herbivory in tassel-eared squirrels: role of monoterpenes in ponderosa pines chosen as feeding trees. *Science* **213**, 1273–1275.
- Fares, Y., Sharpe, P. J., and Magnuson, C. E. (1980). Pheromone dispersion in forests. *Journal of Theoretical Biology* **84**, 335–359.
- Faulkes, C. G. and Abbott, D. H. (1993). Evidence that primer pheromones do not cause social supression of reproduction in naked mole rats (*Heterocephalus glaber*). *Journal of Reproduction and Fertility* **99**, 225–230.
- Feiler, W. and Haas, W. (1988). Trichobilharzia ocellata: chemical stimuli of duck skin for cercarial attachment. *Parasitology* **96**, 507–517.
- Feist, J. D. and McCullough, D. (1976). Behavior patterns and communication in feral horses. *Journal of Reproduction and Fertility* **23** (Suppl.), 337–371.
- Fergusson, B., Bradshaw, S. D., and Cannon, J. R. (1985). Hormonal control of femoral gland secretion in the lizard, *Amphibolurus ornatus*. *General and Comparative Endocrinology* **57**, 371–376.
- Ferkin, M. H. (1988). The effect of familiarity on social interactions in meadow voles, *Microtus pennsylvanicus*: a laboratory and field study. *Animal Behaviour* **36**, 1816–1822.

- Ferkin, M. H. and Johnston, R. E. (1995a). Meadow voles, *Microtus pennsylvaticus*, use multiple sources of scent for sex recognition. *Animal Behaviour* **49**, 37–44.
 - (1995b). Effects of pregnancy, lactation and postpartum oestrus on odor signals and the attraction to odours in female meadow voles, *Microtus pennsylvaticus*. *Animal Behaviour* **49**, 1211–1217.
- Ferkin, M. H. and Seamon, J. O. (1987). Odor preference and social behavior in meadow voles, *Microtus pennsylvanicus*: seasonal differences. *Canadian Journal of Zoology* **65**, 2931–2937.
- Ferkin, M. H., Sorokin, E. S., Johnston, R. E., and Lee, C. J. (1997). Attractiveness of scents varies with protein content of the diet in meadow voles. *Animal Behaviour* **53**, 133–141.
- Ferreira, A., Dahlof, L. G., and Hansen, S. (1987). Olfactory mechanisms in the control of maternal aggression, appetite, and fearfulness: effects of lesions to olfactory receptors, mediodorsal thalamic nucleus, and insular prefrontal cortex. *Behavioral Neuroscience* **101**, 709–717.
- Ferris, C. G., Axelson, J. F., Shinto, L. H., and Albers, H. E. (1987). Scent marking and the maintenance of dominant subordinate status in male golden hamsters. *Physiological Behavior* **40**, 661–664.
- Filho, O. G. and Mazzafera, P. (2000). Caffeine does not protect coffee against the leaf miner *Perileucoptera coffeella*. *Journal of Chemical Ecology* **26**:1447–1464.
- Fillion, T. J. and Blass, E. M. (1986). Infantile experience with suckling odors determines adult sexual behavior in male rats. *Science* **231**, 729–731.
- Fine, J. M. and Sorensen, P. W. (2004). Bioassay-guided fractionation demonstrates that the sea lamprey migratory pheromone is a mixture of at least three sulfated steroids. In *Annual Meeting of International Society of Chemical Ecology*, July 2004, Ottawa, Canada.
- Fine, J. M., Vrieze, L. A., and Sorensen, P. W. (2004). Evidence that petromyzontid lampreys employ a common migratory pheromone that is partially comprised of bile acids. *Journal of Chemical Ecology* **30**, 2091–2110.
- Flannery, T. (1998). Throwing Way Leg: Tree-kangaroos, Possums, and Penis Gourds: On the track of Unknown Mammals in Wildest New Guinea. New York: Atlantic Monthly Press.
- Fleming, A. S., Vaccarino, F., Tambosso, L., and Chee, P. (1979). Vomeronasal and olfactory system modulation of maternal behavior in the rat. *Science* **203**, 372–374.
- Fletcher, K. J. C. and Michener, D. C. (1987). Kin Recognition in Animals. New York: Wiley.
- Fletcher, T. C. and Lindsay, D. R. (1968). Sensory involvement in the mating behavior of domestic sheep. *Animal Behaviour* **16**, 410–416.
- Flier, J. M., Edwards, W., Daly, J. W., and Meyers, C. (1980). Widespread occurrence in frogs and toads of skin compounds interacting with the ovabain site of Na⁺, K⁺-ATPase. *Science* **208**, 503–505.
- Flood, P. F., Abrams, S. R., Muir, G. D., and Rowell, J. E. (1989). Odor of the muskox: a preliminary investigation. *Journal of Chemical Ecology* **15**, 2207–2217.
- Flowers, M. A. and Graves, B. M. (1997). Juvenile toads avoid chemical cues from snake predators. *Animal Behaviour* **53**, 641–646.

- Fogel, R. and Trappe, J. M. (1978). Fungus consumption (mycophagy) by small animals. *Northwest Science* **52**, 1–31.
- Foley, W. J., McLean, S., and Cork, S. J. (1995). Consequences of biotransformation of plant secondary metabolites on acid-base metabolism in mammals: a final common pathway? *Journal of Chemical Ecology* **21**, 721–743.
- Fombon, A. M. and Polak, E. H. (1987). Odor similarity between stress-inducing odorants in Wistar rats. *Journal of Chemical Ecology* **13**, 153–166.
- Ford, N. B. (1978). Evidence for the species specificity of pheromone trails in two sympatric garter smakes, (*Thamnophis*). *Herpetological Review* **9**, 10.
 - (1981). Seasonality of pheromone trailing behavior in two species of garter snake, *Thamnophis* (Colubridae). *Southwestern Naturalist* **26**, 385–388.
 - (1982). Species specificity of sex pheromone trails of sympatric and allopatric garter snakes (*Thamnophis*). *Copeia* **1**, 10–13.
- Ford, N. B. and Low, J. R. (1984). Sex pheromone source location by garter snakes: a mechanism for detection of direction in nonvolatile trails. *Journal of Chemical Ecology* **10**, 1193–1199.
- Forester, D. C. and Wisnieski, A. (1991). The significance of airborne olfactory cues to the recognition of home area by the dart-poison frog *Dendrobates pumilio*. *Journal of Herpetology* **25**, 502–504.
- Formanowicz, D. R. and Brodie, E. D. (1982). Relative palatabilities of members of a larval amphibian community. *Copeia* **1982**, 91–97.
- Fornasieri, I. and Roeder, J. J. (1992). Behavioral responses to own and other species' scent marks in *Lemur fulvus* and *Lemur macaco*. *Journal of Chemical Ecology* **18**, 2069–2082.
- Fowler, M. E. (1992). Veterinary Zootoxicology. Boca Raton, FL: CRC Press.
- Fraenkel, G. S. (1959). The raison d'etre of secondary plant substances. *Science* **129**, 1466–1470.
- Frank, F. (1954). Beiträge zur Biologie der Feldmaus, *Microtus arvalis* (Pallas). Teil I. Gehegeversuche. *Zoologische Jahrbücher* **82**, 354–404.
- Frank, R. A. and Byram, J. (1988). Taste–smell interactions are tastant and odorant dependent. *Chemical Senses* 13, 445–455.
- Franklin, W. L. (1983). Contrasting socioecologies of South America's wild camelids: the vicuña and the guanaco. *American Society of Mammologists, Special Publication* 7, 573–631.
- Freeland, W. J. (1974). Vole cycles: another hypothesis. *American Naturalist* **108**, 238–245. (1991). Plant secondary metabolites, biochemical coevolution with herbivores. In *Plant Defenses Against Mammalian Herbivory*, ed. R. T. Palo and C. T. Robbins, pp. 61–81. Boca Raton, FL: CRC Press.
- Freeland, W. J. and Janzen, D. H. (1974). Strategies in herbivory by mammals: the role of plant secondary compounds. *American Naturalist* **108**, 269–289.
- French, J. A., Abbot, D. H., and Snowdon, C. T. (1984). The effect of social environment on estrogen excretion, scent marking, and sociosexual behavior in tamarins (*Saguinus oedipus*). *American Journal of Primatology* **6**, 155–167.

- Frey, R. and Hofmann, R. R. (1997). Skull, proboscis musculature and preorbital gland in the saiga antelope and Guenther's dikdik (Mammalia, Artiodactyla, Bovidae). *Zoologischer Anzeiger* **235**, 183–199.
- Frid, L. and Turkington, R. (2001). The influence of herbivores and neighboring plants on risk of browsing: a case study using arctic lupine (*Lupinus arcticus*) and arctic ground squirrels (*Spermophilus parryi plesius*). *Canadian Journal of Zoology* **79**, 874–880.
- Friedmann, A. (1955). The honey guides. *Bulletin of the United States National Museum* **208**, 1–292.
- Friedman, L. and Miller, J. G. (1971). Odor incongruity and chirality. *Science* **172**, 1044–1046.
- Frischknecht, P. M., Ulmer-Dufek, J., and Baumann, T. W. (1986). Purine alkaloid formation in buds and developing leaflets of *Coffea arabica*: expression of an optimal defence strategy? *Phytochemistry* **25**, 613–616.
- Fry, B. G., Vidal, N., Norman, J. A., *et al.* (2005). Early evolution of the venom system in lizards and snakes. *Nature* advance online publication, 16 November 2005. (doi.10.1038/nature04328).
- Fuchs, J. L. and Burghardt, G. M. (1971). Effects of early feeding experience on the responses of garter snakes to food chemicals. *Learning and Motivation* **2**, 271–279.
- Fulk, G. W. (1971). The Behavioral Interactions of the Short-tailed Shrew and the Meadow Vole. Kingston, RI: University of Rhode Island.
- Gagliardo, A., Faschi, V., and Benvenuti, S. (1988). Pigeon homing: olfactory experiments with young inexperienced birds. *Naturwissenschaften* **75**, 211–213.
- Galef, B. (1982). Acquisition and waning of exposure-induced attraction to a non-natural odor in rat pups. *Developmental Psychobiology* **15**, 479–490.
- Galef, B. G. and Kaner, H.C. (1980). Establishment and maintenance of preference for natural and artificial olfactory stimuli in juvenile rats. *Journal of Comparative and Physiological Psychology* **94**, 588–595.
- Galef, B. G. and Stein, M. (1985). Demonstrator influence on observer diet preference: analysis of critical social interactions and olfactory signals. *Animal Learning and Behavior* 13, 31–38.
- Galef, B. G., Kennett, D. J., and Stein, M. (1985). Demonstrator influence on observer diet preference: effects of simple exposure and the presence of a demonstrator. *Animal Learning and Behavior* 13, 25–30.
- Galef, B. G., Mason, J. R., Prety, G., and Bean, N. J. (1988). Carbon disulfide: a semiochemical mediating socially-induced diet choice in rats. *Physiology and Behavior* **42**, 119–124.
- Gamradt, S. C. and Kats, L. B. (1996). Effect of introduced crayfish and mosquitofish on California newts. *Conservation Biology* **10**, 1155–1162.
- Gangrade, B. K. and Dominic, C. J. (1986). Effect of storage and lyophilization on estrusinducing capacity of male mouse urine. *Indian Journal of Experimental Biology* **24**, 728–729.
- Gao, Y. (1991). Behavioral responses of rats to the smell of urine from conspecifics. *Animal Behaviour* **42**, 506–508.

- Garcia, J. and Rusiniak, K. W. (1980). What the nose learns from the mouth. In *Chemical Signals in Vertebrates*, vol. 2, ed. D. Müller-Schwarze, and R. M. Silverstein, pp. 141–156. New York: Plenum.
- Garcia, J., Holder, M. D., and Yirmiya, R. (1986). Taste and odor interactions in conditioned flavor aversions. *Appetite* 7, 259.
- Garstka, W. R. and Crews, D. (1986). Pheromones and reproduction in garter snakes. In *Chemical Signals in Vertebrates*, vol 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 243–260. New York: Plenum.
- Garton, J. D. and Mushinsky, H. R. (1979). Integumentary toxicity and unpalatability as an antipredator mechanism in the narrow mouthed toad, *Gastrophryne carolinensis*. *Canadian Journal of Zoology* **57**, 1965–1973.
- Gause, G. F. (1934). The Struggle for Existence. New York: Hafner.
- Gauthier, G. and Hughes, R. J. (1995). The palatability of Arctic willow for greater snow geese: the role of nutrients and deterring factors. *Oecologia* **103**, 390–392.
- Gauthier-Pilters, H. (1974). The behavior and ecology of camels in the Sahara, with special reference to nomadism and water management. In *The Behavior of Ungulates and its Relation to Management*, ed. V. Geist and F. Walther, pp. 542–551. Morges, Switzerland: International Union for Conservation of Nature and Natural Resources.
- Gavish, L., Hofmann, J. E., and Getz, L. L. (1984). Sibling recognition in the prairie vole, *Microtus ochrogaster. Animal Behaviour* **32**, 362–366.
- Gazdewich, K. J. and Chivers, D. P. (2002). Acquired predator recognition by fathead minnows: influence of habitat characteristics on survival. *Journal of Chemical Ecology* **28**, 439–445.
- Gehlbach, F. R., Watkins, J. F., and Reno, H. W. (1968). Blindsnake defensive behavior elicited by ant attacks. *BioScience*. **18**, 784–785.
- Gehlbach, F. R., Watkins, J. F., and Kroll, J. C. (1971). Pheromone trail following studies of typhlopid, leptotyphlopid, and colubrid snakes. *Behaviour* **40**, 282–294.
- Geiger, R. (1965). The Climate Near the Ground. Cambridge, MA: Harvard University Press.
- Genna, R. L., Mordue, W., Pike, A. W., and Mordue (Luntz) A. J. (2004). Identification of semiochemicals involved in sea lice host location, and their potential use in pest control. In *Annual Meeting of the International Society of Chemical Ecology*, July 2004, Ottawa, Canada.
- George, C. J. W. (1960). Behavioral interaction of the pickerel (*Esox niger* LeSueur and *Esox americanus* LeSueur) and the mosquitofish (*Gambusia patruellis* Bairdad Girard). Ph.D. Thesis, Harvard University Massachussetts.
- Gerhart, D. J., Bondura, M. E., and Commito, J. A. (1991). Inhibition of sunfish feeding by defensive steroids from aquatic beetles: structure–activity relationships. *Journal of Chemical Ecology* 17, 1363–1370.
- Gerritsen, A. F. C., van Heezik, Y. M., and Swennen, C. (1983). Chemoreception in two further *Calidris* species (*C. maritima and C. canutus*) with a comparison of the relative importance of chemoreception during foraging in *Calidris* species. *Netherlands Journal of Zoology* **33**, 485–496.

- Gervais, R., Holley, A., and Keverne, B. (1988). The importance of central noradrenergic influences on the olfactory bulb in the processing of learned olfactory cues. *Chemical Senses* **13**, 3–12.
- Gheusi, G., Goodall, G., and Dantzer, R. (1997). Individually distinctive odors represent individual conspecifics in rats. *Animal Behaviour* **53**, 935–944.
- Gilardi, J. D., Duffey, S. S., Munn, C. A., and Tell, L. A. (1999). Biochemical functions of geophagy in parrots: detoxification of dietary toxins and cytoprotective effects. *Journal of Chemical Ecology* **25**, 897–922.
- Gilbert, B. K. (1973). Scent marking and territoriality in pronghorn (*Antilocapra americana*) in Yellowstone National Park. *Mammalia* 37, 25–33.
- Gilder, P. M. and Slater, P. J. B. (1978). Interest of mice in conspecific male odours is influenced by degree of kinship. *Nature* **274**, 364–365.
- Gleeson, R. A. (1978). Functional adaptation in chemosensory systems. In *Sensory Ecology*, ed. M. A. Ali, pp. 291–317. New York: Plenum.
- Glei, M., Schlegel, W., Straube, D., and Blankenger, J. (1989). Untersuchungen zur Beeinflussung des Pubertätseintritts von Jungsauen mittels maskuliner Stimuli. *Archiv für Tierzucht* **32**, 173–179.
- Glendinning, J. F., Brower, L. P., and Montgomery, C. A. (1990). Responses of three mouse species to deterrent chemicals in the Monarch butterfly. I. Taste and toxicity tests using artificial diets laced with digitoxin or monocrotaline. *Chemoecology* 1, 114–123.
- Godfrey, J. (1958). The origin of sexual isolation between bank voles. *Proceedings of the Royal Physiological Society Edinburgh* **27**, 47–55.
- Golan, L., Radcliffe, C. W., Miller, T., O'Connell, R. J., and Chiszar, D. (1982). Prey trailing by the prairie rattlesnake (*Crotalus v. viridis*). *Journal of Herpetology* **16**, 287–293.
- Gold, T. and Soter, S. (1980). The deep-earth-gas hypothesis. *Scientific American* **242**, 154–161.
- Goldstein, W. S. and Spencer, K. C. (1985). Inhibition of cyanogenesis by tannins. *Journal of Chemical Ecology* **11**, 847–858.
- Goodrich, B. S. and Mykytowycz, R. (1972). Individual and sex differences in the chemical composition of pheromone-like substances from the skin glands of the rabbit, *Oryctolagus cuniculus*. *Journal of Mammalogy* **53**, 540–548.
- Goodrich, B. S., Gambale, S., Pennycuik, P. R., and Redhead, T. D. (1990). Volatiles from feces of wild male house mice. Chemistry and effects on behavior and heart rate. *Journal of Chemical Ecology* **16**, 2091–2106.
- Gorman, M. L. (1976). A mechanism for individual recognition by odour in *Herpestes auro- punctatus* (Carnivora: Viverridae). *Animal Behaviour* **24**, 141–145.
 - (1984). The response of prey to stoat (*Mustela erminea*) scent. *Journal of Zoology* (London) **202**, 419–423.
- Gorman, M. L. and Mills, M. G. L. (1984). Scent marking strategies in hyaenas (Mammalia). *Journal of Zoology* (London) **202**, 535–547.
- Gorman, M. L., Jenkins, D., and Cooper, R. J. (1978). The anal scent sacs of the otter (*Lutra lutra*). *Journal of Zoology* **186**, 463–474.

- Gosden, P. E. and Ware, G. C. (1976). The aerobic bacterial flora of the anal sac of the red fox. *Journal of Applied Bacteriology* **41**, 271–275.
- Gosling, L. M. (1981). Demarkation in a gerenuk territory: an economic approach. *Zeitschrift für Tierpsychologie* **56**, 305–322.
 - (1982). A reassessment of the function of scent marking in territories. *Zeitschrift für Tierpsychologie* **60**, 89–118.
 - (1986). Economic consequences of scent marking in mammalian territoriality. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 385–395. New York: Plenum.
- Gould, S. J. and Vrba, E. S. (1982). Exaptation: a missing term in the science of form. *Paleobiology* **8**, 4–15.
- Gove, D. and Burghardt, G. M. (1975). Responses of ecologically dissimilar populations of the water snake, *Natrix s. sipedon*, to chemical cues from prey. *Journal of Chemical Ecology* **1**, 25–40.
- Gower, D. B. (1989). Pheromones in the human. Journal of Endocrinology. 123(Suppl.), 13.
- Gower, D. B. and Booth, V. (1986). Ontogeny of Olfaction: Principles of Olfactory Maturation in Vertebrates, Berlin: Springer-Verlag.
- Gower, D. B. and Ruparelia, B. A. (1993). Olfaction in humans with special reference to odourous 16-androstenes: their occurrence, perception and possible social, psychological, and sexual impact. *Journal of Endocrinology* **137**, 167–187.
- Gower, D. B., Watkins, J., Mallett, A. I., Rennie, P. J., and Holland, K. T. (1989). Transformations of odorous 16-androstene steroids by human axillary coryneform bacteria. *Chemical Senses* 14, 208.
- Graham, C. A. and McGrew, W. C. (1980). Menstrual synchrony in female undergraduates living in a co-educational campus. *Psychoneuroendocrinology* **5**, 245–252.
- Graham, T., Georges, A., and Mcelhinney, N. (1996). Terrestrial orientation by the eastern longnecked turtle, *Chelodina longicollis*, from Australia. *Journal of Herpetology* **30**, 467–477.
- Grajal, A. (1995). Structure and function of the digestive tract of the hoatzin (*Opisthocomus hoazin*): a folivorous bird with foregut fermentation. *Auk* **112**, 20–28.
- Grajal, A., Strahl, S. D., Parra, R., Dominquez, M. G., and Neher, A. (1989). Foregut fermentation in the hoatzin, a neotropical leaf-eating bird. *Science* **245**, 1236–1238.
- Grant, D., Andersen, O., and Twitty, V. (1968). Homing orientation by olfaction in newts (*Taricha rivularis*). *Science* **160**, 1354–1356.
- Grassman, M. and Owens, D. (1987). Chemosensory imprinting in juvenile green sea turtles, *Chelonia mydas*. *Animal Behaviour* **35**, 929–931.
- Grassman, M. A., Owens, D. W., McVey, J. P., and Marquez, M. R. (1984). Olfactory-based orientation in artificially imprinted sea turtles. *Science* **224**, 83–84.
- Grau, H. J. (1982). Kin recognition in the white-footed deer mice (*Peromyscus leucopus*). *Animal Behaviour* **30**, 497–505.
- Graves, B. M. and Duvall, D. (1985). Avomic prairie rattlesnakes (*Crotalus viridis*) fail to attack rodent prey. *Zeitschrift für Tierpsychologie* **67**, 161–166.

- (1988). Evidence of an alarm pheromone from the cloacal sacs of prairie rattlesnakes. *Southwestern Naturalist* **33**, 339–345.
- Graves, B. M. and Halpern, M. (1990). Roles of vomeronasal chemoreception in tongueflicking, exploratory and feeding behavior of the lizard, *Chalcides ocellatus*. *Animal Behaviour* **39**, 692–698.
 - (1991). Discrimination of self from conspecific chemical cues in *Tiliqua scincoides* (Sauria: Scincidae). *Journal of Herpetology* **25**, 125–126.
- Graves, B. M., Carpenter, G. C. and Duvall, D. (1987). Chemosensory behaviors of neonate prairie rattlesnakes, *Crotalus viridis*. *Southwest Naturalist* **32**, 515–517.
- Graves, B. M., Halpern, M., and Gillingham, J. C. (1993). Effects of vomeronasal system deafferentation on home range use in a natural population of eastern garter snakes, *Thamnophis sirtalis*. *Animal Behaviour* **45**, 307–311.
- Graziadei, P. P. C. (1977). Functional anatomy of the mammalian chemoreceptor system. In *Chemical Signals in Vertebrates*, vol. 1, ed. R. T. Mason, M. P. LeMaster, and D. Müller-Schwarze, pp. 435–454. New York: Plenum.
- Green, G. A. (1988). Living on borrowed turf. Natural History 97, 58-64.
- Greenberg, B. (1943). Social behavior of the western banded gecko, *Coleonyx variegatus* Baird. *Physiological Zoology* **16**, 110–122.
- Greene, M. J. and Mason, R. T. (2000). Courtship, mating and male combat of the brown tree snake, *Boiga irregularis*. Herpetologica **56** 166–175.
 - (2003). Pheromonal inhibition of male courtship behavior in the brown tree snake, *Boiga irregularis*: a mechanism for the rejection of potential mates. *Animal Behaviour* **65**, 905–910.
- Greene, M. J., Stark, S. L., and Mason, R. T. (2002). Predatory response of brown tree snakes to chemical stimuli from human skin. *Journal of Chemical Ecology* **28**: 2465–2473.
- Greenwood, D. R., Comeskey, D., Hunt, M. B., and Rasmussen, L. E. L. (2005). Chirality in elephant pheromones. *Nature* **438**, 1097–1098.
- Gregory, M. J. and Cameron, G. N. (1989). Scent communication and its association with dominance behavior in the Hispid cotton rat (*Sigmodon hispidus*). *Journal of Mammalogy* **70**, 10–17.
- Greig-Smith, P. W. (1988). Bullfinches and ash trees: assessing the role of plant chemicals in controlling damage by herbivores. *Journal of Chemical Ecology* **14**, 1889–1903.
- Griffin, R. W. and Beidler, L. M. (1984). Studies in canine olfaction, taste and feeding: a summing up and some comments on the academic–industrial relationship. *Neuroscience and BioBehavioral Reviews* **8**, 261–263.
- Griffith, C. R. (1919). A possible case of instinctive behavior in the white rat. *Science* **50**, 166–167.
 - (1920). The behavior of white rats in the presence of cats. *Psychobiology* **2**, 19–28.
- Grimmer, J. L. (1962). Strange little world of the hoatzin. *National Geographic* **122**, 390–401.

- Grønneberg, T. Ø. (1978–79). Analysis of a wax ester fraction from anal gland secretion of beaver (*Castor fiber*) by chemical ionization mass spectrometry. *Chemical Scripta* 13, 56–58.
- Grønneberg, T. Ø. and Lee, T. (1984). Lipids of the anal gland secretion of beaver (*Castor canadensis*). *Chemica Scripta* 24, 100-103.
- Grubb, J. C. (1973a). Olfactory orientation in breeding Mexican toads, *Bufo valliceps*. *Copeia* **1973**, 490–497.
 - (1973b). Olfactory orientation in Bufo woodhousei fowleri, Pseudacris clarki and P. streckeri. Animal Behaviour **21**, 726–732.
 - (1976). Maze orientation by Mexican toads, *Bufo valliceps*, using olfactory and configurational cues. *Journal of Herpetology* **10**, 97–104.
- Grubb, T. C., Jr. (1972). Smell and foraging in shearwaters and petrels. *Nature* **237**, 404–405.
- (1974). Olfactory navigation to the nesting burrow in Leach's petrel (*Oceanodroma leucorrhoa*). *Animal Behaviour* **22**, 192–202.
- Grundvig, J. L., Dustman, R. E., and Beck, E C. (1967). The relationship of olfactory receptor stimulation to stimulus environmental temperature. *Experimental Neurology*. **18**, 416–428.
- Gubernick, D. J. (1990). A maternal chemosignal maintains paternal behavior in the biparental California mouse, *Peromyscus californicus*. *Animal Behaviour* **39**, 936–942.
- Gubernick, D. J. and Klopfer, P. H. (1980). Parental Care in Mammals. New York: Plenum.
- Gubernick, D. J. and Nordby, J. C. (1992). Parental influences on female puberty in the monogamous California mouse, *Peromyscus californicus*. *Animal Behaviour* **44**, 259–267.
- Gunson, J. R. (1970). Dynamics of the beaver of Saskatchewan's northern forest. M. Sc. Thesis, University of Alberta, Edmonton, Alberta.
- Gurnell, J. and Little, J. (1992). The influence of trap residual odor on catching woodland rodents. *Animal Behaviour* **43**, 623–632.
- Gustin, M. K. and McCracken, G. F. (1987). Scent recognition between females and pups in the bat *Tadarida brasiliensis mexicana*. *Animal Behaviour* **35**, 13–19.
- Gwinner, H., Oltrogge, M., Trost, L., and Nienaber, U. (2002). Green plants in starling nests: effects on nestlings. *Animal Behaviour* **59**, 301–309.
- Habermehl, G. G. (1994). Gift-Tiere und ihre Waffen, 5th edn. Berlin: Springer-Verlag.
- Haftorn, S., Mehilum, F., and Bech, C. (1988). Navigation to nest site in the snow petrel, *Pagodroma nivea. Condor* **90**, 484–486.
- Hagelin, J. C., Jones, I. L., and Rasmussen, L. E. L. (2003). A tangerine-scented social odour in a monogamous seabird. *Proceedings of the Royal Society of London, Series B* **270**, 1323–1329.
- Haigh, G. R. (1987). Reproductive inhibition of female *Peromyscus leucopus*: female competition and behavioral regulation. *American Zoologist* 27, 867–868.
- Haim, A. and Fluxman, S. (1996). Daily rhythms of metabolic rates: role of chemical signals in coexistence of spiny mice of the genus *Acomys*. *Journal of Chemical Ecology* **22**, 223–231.

- Haldane, J. B. S. (1955). Animal communication and the origin of human language. *Science Progress* (London) **43**, 385–401.
- Hall, D. R., Beevor, P. S., Cork, A., Nesbitt, B. F., and Vale, G. A. (1984). 1-Octen-3-ol. A potent olfactory stimulant and attractant for tsetse isolated from cattle odors. *Insect Science and its Application* **5**, 335–339.
- Halpern, M. and Frumin, N. (1979). Roles of the vomeronasal and olfactory systems in prey attack and feeding in adult garter snakes. *Physiology and Behavior* **22**, 1183–1189.
- Halpern, M., Scribani, L., and Kubie, J. L. (1985). Vomeronasal stimuli can be reinforcing. *Chemical Senses* **10**, 422.
- Halpern, M., Schulman, N., and Kirschenbaum, D. M. (1986). Characteristics of earthworm washings detected by the vomeronasal system of snakes. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, Müller-Schwarze, D. and R. M. Silverstein, pp. 63–77, New York: Plenum.
- Halpin, Z. T. (1974). Individual difference in the biological odors of the Mongolian gerbil (*Meriones unguiculatus*). *Behavioral Biology* **11**, 253–259.
 - (1986). Individual odors among mammals: origins and functions. *Advances in the Study of Behavior* **16**, 39–70.
- Hanfin, C. T., Brodie, E. D., III, and Brodie, E. D., Jr. (2003). Tetrodotoxin levels in eggs of rough-skin newt, *Taricha granulosa*, are correlated with female toxicity. *Journal of Chemical Ecology* **29**, 1729–1739.
- Hansen, L. P., Døving, K.B., and Jonsson, B. (1987). Migration of farmed adult Atlantic salmon with and without olfactory sense, released on the Norwegian coast. *Journal of Fish Biology* **30**, 713–722.
- Hansson, L., Gref, R., and Theander, O. (1986). Susceptibility to vole attacks due to bark phenols and terpenes in *Pinus contorta* provenances introduced into Sweden. *Journal of Chemical Ecology* **12**, 1569–1578.
- Harada, K. (1989). Role of carboxylic group on feeding repellence of acidic amino acids and organic acids for oriental weatherfish. *Bulletin of the Japanese Society of Scientific Fisheries* **55**, 927.
- Harborne, J. B. (1993). *Introduction to Ecological Biochemistry*, 4th edn. London: Academic Press.
- Harden-Jones, F. R. (1968). Fish Migration. London: Edward Arnold.
- Hare, J. F. (1994). Group member discrimination by Columbian ground squirrels via familiarity with substrate-borne chemical cues. *Animal Behaviour* **47**, 803–813.
- Hargrove, J. W. and Vale, G. A. (1978). The effect of host odour concentration on catches of tsetse flies (Glossinidae) and other Diptera in the field. *Bulletin of Entomological Research* **68**, 607–612.
- Harju, A. (1996). Effect of birch (Betula pendula) bark and food protein level on root voles (Microtus oeconomus): II. Detoxification capacity. Journal of Chemical Ecology 22, 719–728.
- Harrington, F. H. (1981). Urine marking and caching behavior in the wolf. *Behavior* **61**, 82–105.

- Harris, B. (1987). Mushrooms and Truffles: Botany, Cultivation, and Utilization. Koenigstein: Koeltz Scientific.
- Harris, M. A. and Murie, J. O. (1982). Responses to oral gland scents from different males in Columbian ground squirrels. *Animal Behaviour* **30**, 140–148.
- Hart, B. L. (1983). Flehmen behavior and vomeronasal organ function. In *Chemical Signals in Vertebrates*, vol. 3, ed. D. Müller-Schwarze and R. M. Silverstein. New York: Plenum.
- Hart, B. L. and Leedy, M. G. (1987). Stimulus and hormonal determinants of flehmen behavior in cats. *Hormones and Behavior*. **21**, 44–52.
- Hart, B. L., Hart, L. A., and Maina, J. N. (1988). Alteration in vomeronasal system anatomy in alcelaphine antelopes: correlation with alteration in chemosensory investigation. *Physiology and Behavior* **42**, 155–162.
- Harvey, S., Jemiolo, B., and Novotny, M. (1989). Pattern of volatile compounds in dominant and subdominant male mouse urine. *Journal of Chemical Ecology* **15**, 2061–2072.
- Hasler, A. D. (1954). Odor perception and orientation in fishes. *Journal of the Fisheries Research Board of Canada* 11, 107–129.
- Hasler, A. D. and Scholz, A. T. (1983). Olfactory Imprinting and Homing in Salmon. New York: Springer-Verlag.
- Hasler, A. D. and Wisby, W. J. (1951). Discrimination of stream odors by fishes and relation to parent stream behavior. *American Naturalist.* **85**, 223–238.
- Hasler, A. D., Scholz, A. T., and Horral, R. M. (1978). Olfactory imprinting and homing in salmon. *American Scientist* **66**, 347–355.
- Hassanali, A., McDowell, P. G., Owaga, M. L. A., and Saini, R. K. (1986). Identification of tsetse attractants from excretory products of a wild host animal, *Syncerus caffer. Insect Science and its Application* 7, 5–9.
- Hassler, T. J. and Kucas, S. T. (1988). Returns of morpholine imprinted coho salmon to the Mad River, California. *North American Journal of Fisheries Management* **8**, 356–358.
- Hastings, B. C. and Gilbert, B. K. (1980). Aversive conditioning of black bears in the backcountry of Yosemite National Park. *Proceedings of the Second Conference on Scientific Research in National Parks*, vol. 7, pp. 294–303.
- Hatanaka, T. and Hanada, T. (1987). Structure of the vomeronasal system and the induced wave in the accessory olfactory bulb of red eared turtle. *Chemical Senses* **12**, 521.
- Hayashi, S. (1987). Female mice prefer odors of aggressive males. *Zoological Science* (Tokyo) 4, 1105.
- (1989). Male-mice: social-dominance influenced by strange male odors. *Aggressive Behavior*. **15**, 1–3.
- Hayes, R. A., Richardson, B. J., and Wyllie, S. G. (2001). Increased social dominance in rabbits, *Oryctalus cuniculus*, is associated with increased secretion of 2-phenoxyethanol from the chin gland. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koi, J. J. Lepri, and D. Müller-Schwarze, pp. 335–341. New York: Kluwer Academic/Plenum.
 - (2002). Semiochemicals and social signaling in the wild European rabbit in Australia: I. Scent profiles of chin gland secretion from the field. *Journal of Chemical Ecology* **28**, 363–384.

- Hébert, P. and Barrette, C. (1989). Experimental demonstration that scent marking can predict dominance in the woodchuck (*Marmota monax*). *Canadian Journal of Zoology* **67**, 575–578.
- Hefetz, A., Ben-Yaacov, R., and Yom-Tov, Y. (1984). Sex specificity in the anal gland secretion of the Egyptian mongoose *Herpestes ichneumon*. *Journal of Zoology* **203**, 205–209.
- Hendrichs, H. and Hendrichs, U. (1971). Dikdik und Elefanten: Ökologie und Soziologie zweier afrikanischer Huftiere. In *Studies in Ethology*, ed. W. Wickler. Munich: Piper.
- Henessy, F. F. and Owings, D. H. (1979). Snake species discrimination and the role of olfactory cues in the snake-directed behavior of the California ground squirrel. *Behaviour* **65**, 115–124.
- Henry, J. D. (1980). The urine marking behavior and movement patterns of red foxes (*Vulpes vulpes*) during a breeding and post-breeding period. In *Chemical Signals in Vertebrates*, vol. 2, ed. D. Müller-Schwarze and R. M. Silverstein, pp. 11–27. New York: Plenum.
- Henton, W. W., Smith, J. C., and Tucker, D. (1966). Odor discrimination in pigeons. *Science* **153**, 1138–1139.
- Hepper, P. G. (1983). Sibling recognition in the rat. Animal Behaviour 31, 1177–1191.
 - (1987). The amniotic fluid: an important priming role in kin recognition. *Animal Behaviour* **35**, 1343–1346.
 - (1988). Adaptive fetal learning, prenatal exposure to garlic affects postnatal preferences. *Animal Behaviour* **36**, 935–936.
 - (1990). Foetal olfaction. In *Chemical Signals in Vertebrates*, vol. 5, ed. D. W. MacDonald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 282–288. Oxford: Oxford University Press.
- Herb, R., Carroll, R., Yoshida, W. Y., Scheuer, P. J., and Paul, V. J. (1990). Polyalkylated cyclopentindoles: cytotoxic fish antifeedants from a sponge, *Axinella sp. Tetrahedron* **46**, 3089–3092.
- Herrera, E. A. (1992). Size of testes and scent glands in capybaras, *Hydrochaeris hydrochaeris* (Rodentia: Caviomorpha). *Journal of Mammalogy* **73**, 871–875.
- Herz, R. S. and Inzlicht, M. (2002). Sex differences in response to physical and social factors involved in human mate selection: the importance of smell for women. *Evolution and Human Behavior* **23**, 359–364.
- Heske, E. J. (1987). Responses of a population of California voles, *Microtus californicus*, to odor-baited traps. *Journal of Mammalogy* **68**, 64–72.
- Hews, D. K. (1988). Alarm response in larval western toads, *Bufo boreas*: release of chemicals by a natural predator and it's effect on predator capture efficiency. *Animal Behaviour*. **36**, 125–133.
- Hickey, M. B. C. and Fenton, M. B. (1987). Scent-dispersing hairs (Osmetrichia) in some Pteropodidae and Molossidae (Chiroptera). *Journal of Mammalogy* **68**, 381–384.
- Hill, J. O., Smith, G. L., III, Pavlik, E. H., Burghardt, G. M., and Coulson, P. B. (1976). Species characteristic responses to catnip by undomesticated felids. *Journal of Chemical Ecology* 2, 239–253.

- Hiller, A. and Wand, U. (1984). Radiocarbon dating of breeding places of petrels in the Antarctic. Academy of Sciences, German Democratic Republic. *Zentralinstitut für Isotopen-und Strahlenforschung ZFI-Mitteilungen* 89, 103–121.
- Hirvonen, H., Ranta E., Piironen, J., Laurila, A., and Peuhkuri, N. (2000). Behavioral responses of naive Arctic charr young to chemical cues from salmonid and non-salmonid fish. *Oikos* 88, 191–199.
- Hladick, A. and Hladik, C. M. (1969). Rapports trophiques entre vegetation et primates dans la forêt de Barro Colorado (Panama). *Terre Vie* **116**, 25–117.
- Ho, H.-Y. and Chow, Y. S. (1993). Chemical identification of defensive secretion of stick insect, *Megacrania tsudai* Shiraki. *Journal of Chemical Ecology* **19**, 39–46.
- Hofmann, J. E., Getz, L. L., and Gavish, L. (1987). Effect of multiple short-term exposures of pregnant *Microtus ochrogaster* to strange males. *Journal of Mammalogy* **68**, 166–169.
- Hold, B. and Schleidt, M. (1977). The importance of human odour in non-verbal communication. *Zeitschrift für Tierpsychologie* **43**, 225–238.
- Holden, C. (2001). Cyanide named as Kentucky foal killer. *Science* **292**, 1831. (2004). Ratting out tuberculosis. *Science* **303**, 166.
- Holland, K. (1978). Chemosensory orientation to food by a Hawaiian goatfish (*Parupeneus porphyreus*, Mullidae). *Journal of Chemical Ecology* **4**, 173–186.
- Holling, C. S. (1958). Sensory stimuli involved in the location and selection of sawfly cocoons by small mammals. *Canadian Journal of Zoology* **36**, 633–653.
- Holm, J. C. and Walther, B. T. (1988). Free amino acids in live freshwater zooplankton and dry feed: possible importance for first feeding in Atlantic salmon fry (*Salmo salar*). *Aquaculture* **71**, 223–234.
- Holmes, W. G. (1984). Sibling recognition in thirteen lined ground squirrels: effects of genetic relatedness, rearing association and olfaction. *Behavioral Ecology and Sociobiology* 14, 225.
 - (1986). Kin recognition by phenotype matching in female Belding's ground squirrels. *Animal Behaviour* **34**, 38–47.
 - (1992). Sternal odors as cues for social discrimination by female Virginia opossums, *Didelphis virginiana*. *Journal of Mammalogy* **73**, 286–291.
- Holy, T. E., Dulac, C., and Meister, M. (2000). Responses of vomeronasal organ to natural stimuli. *Science* **289**, 1569–1572.
- Horn, S. W. (1983). An evaluation of predatory suppression in coyotes using lithium chloride induced illness. *Journal of Wildlife Management* 47, 999–1009.
- Horne, E. A. and Jaeger, R. G. (1988). Territorial pheromones of female red-backed salamanders. *Ethology* **78**, 143.
- Houck, L. D. and Reagan, N. L. (1990). Male courtship pheromones increase female receptivity in a plethodontid salamander. *Animal Behaviour* **39**, 729–734.
- Houck, L. D., Bell, A. M., Reagan-Wallin, N. L., and Feldhoff, R. C. (1998). Effects of experimental delivery of male courtship pheromones on the timing of courtship in a terrestrial salamander, *Plethodon jordani* (Caudata: Plethodontidae). *Copeia* 1, 214–219.
- Houlihan, P. W. (1989). Scent mounding by beaver (*Castor canadensis*): functional and semiochemical aspects. M. Sc. Thesis, State University of New York, College of Environmental Science and Forestry, Syracuse, New York.

- Houston, D. C. (1984). Does the king vulture *Sarcorhamphus papa* use a sense of smell to locate food? *Ibis* **126**, 67–69.
 - (1987). Scavenging efficiency of turkey vultures in tropical forests. *Condor* **88**, 318–323.
- Howard, R. R. (1971). Avoidance learning of spotted salamanders by domestic chickens. *American Zoologist* **11**, 637.
- Howard, W. E. and Cole, R. E. (1967). Olfaction in seed detection by deer mice. *Animal Behaviour* **16**, 13–17.
- Howard, W. E., Marsh, R. E., and Cole, R. E. (1968). Food detection by deer mice using olfactory rather than visual cues. *Animal Behaviour* **16**, 13–27.
- Hradecky, P. (1989). Possible induction by estrous cows of pheromone production in penmates. *Journal of Chemical Ecology* 15, 1067-1076.
- Hubert, H. B., Fabsitz, R. R., Feinleib, M. and Brown, K. S. (1980). Olfactory sensitivity in humans: genetic versus environmental control. *Science* **208**, 607–609.
- Huck, U. W., Lisk, R. D., Kim, S., and Evans, A. B. (1989). Olfactory discrimination of estrous condition by the male golden hamster, *Mesocricetus auratus*. Behavioral and Neural Biology **51**, 1–10.
- Hudson, R. and Altbäcker, V. (1994). Development of feeding and food preference in the European rabbit: environmental and maturational determinants. In *Behavioral Aspects of Feeding: Basic and Applied Research in Mammals*, ed. B. G. Galef, M. Mainardi, and P. Valsecchi, pp. 125–145. Chur: Harwood Academic.
- Hudson, R. and Distel, H. (1983). Nipple location by newborn rabbits: behavioral evidence for pheromonal guidance. *Behaviour* **85**, 260–275.
 - (1986). Pheromonal release of suckling in rabbits does not depend on the vomeronasal organ. *Physiology and Behavior* 37, 123–128.
- Hudson, R. and Vodermayer, T. (1992). Spontaneous and odour-induced chin marking in domestic female rabbits. *Animal Behaviour* **43**, 329–336.
- Huffman, M. A. and Seifu, M. (1989). Observations on the illness of and consumption of a possibly medicinal plant, *Vernonia amygdalina* (Del.), by a wild chimpanzee in the Mahale Mountains National Park, Tanzania. *Primates* **30**, 51–63.
- Hugie, D. M. and Smith, R. J. F. (1987). Epidermal club cells are not linked with an alarm response in reedfish, *Erpetoichthys calabaricus*. *Canadian Journal of Zoology* **65**, 2057–2061.
- Hull, M. Q. (1997). The role of semiochemicals in the behavior and biology of *Lep-eophtheirus salmonis* (Krøyer 1837): Potential for control? Ph.D. Thesis, University of Aberdeen, Aberdeen, UK.
- Humphries, R. E., Robertson, D. H. L., Beynon, R. J., and Hurst, J. L. (1999). Unravelling the chemical basis of competitive scent marking in house mice. *Animal Behaviour* **58**, 1177–1190.
- Hurst, J. L. (1987). The functions of urine marking in a population of free-living house mice, *Mus domesticus* Rutty. *Animal Behaviour* **35**, 1433–1442.
 - (1989). The complex network of olfactory communicationin populations of wild house mice *Mus domesticus* Rutty: urine marking and investigation within family groups. *Animal Behaviour* **37**, 705–725.

- (1990a). Urine marking in populations of wild house mice *Mus domesticus* Rutty. I. Communication between males. *Animal Behaviour* **40**, 209–222.
- (1990b). Urine marking in populations of wild house mice *Mus domesticus* Rutty. II. Communication between females. *Animal Behaviour* **40**, 223–232.
- (1993). The priming effects of urine substrate marks on interactions between male house mice, *Mus musculus domesticus* Schwarz and Schwarz. *Animal Behaviour* **45**, 55–81.
- Hurst, J. L. and Rich, T. J. (1999). Scent marks as competitive signals of mate quality. In *Advances in Chemical Signals in Vertebrates*, ed. R. E. Johnston, D. Müller-Schwarze, and P. W. Sorensen, pp. 209–225. New York: Kluwer Academic/Plenum.
- Hurst, J. L. and Smith, J. (1995). Mus spretus Lataste: a hygienic house mouse? Animal Behaviour 49, 827–834.
- Hurst, J. L., Fang, J., and Barnard, C. J. (1993). The role of substrate odours in maintaining social tolerance between male house mice, *Mus musculus domesticus*. *Animal Behaviour* **45**, 997–1006.
- Hurst, J. L, Robertson, D. H. L., Tolladay, U., and Beynon, R. J. (1998). Proteins in urine scent marks of male house mice extend the longevity of olfactory signals. *Animal Behaviour* **55**, 1289–1298.
- Hurst, J. L., Payne, C. E., Nevison, C. M., *et al.* (2001). Recognition of mice mediated by major urinary protiens. *Nature* **414**, 631–634.
- Hutchison, L. V. and Wenzel, B. M. (1980). Olfactory guidance in foraging by procellariiforms. *Condor* **82**, 314–319.
- Hutchison, L. V., Wenzel, B. H., Stager, K. E. and Tedford, B. L. (1984). Further evidence for olfactory foraging by Sooty Shearwater and Northern Fulmars. In *Marine Birds: Their Feeding Ecology and Commercial Fisheries Relationships*, ed. D. N. Nettleship, G. A. Sanger, and P. F. Springer, pp. 78–89. Ottawa: Canadian Wildlife Service Special Publication.
- Hwang, C. J., Krasner, S. W., and McGuire, M. J. (1984). Determination of subnanogram per liter levels of earth-musty odorants in water by the salted closed-loop stripping method. *Environmental Science and Technology* **18**, 535–539.
- Iason, G. R. and Murray, A. H. (1996). The energy costs of ingestion of naturally occurring nontannin plant phenolics by sheep. *Physiological Zoology* **69**, 532–546.
- Iason, G. R. and Palo, R. T. (1991). The effects of birch phenolics on a grazing and a browsing mammal: a comparison. *Journal of Chemical Ecology* **17**, 1733–1743.
- Idler, D. E., Fagerland, V. H. M., and Mayoh, H. (1956). Olfactory perception in migrating salmon. I. L-Serine, a salmon repellent in mammalian skin. *Journal of General Physiology* **39**, 889–892.
- Idris, M. and Prakash, I. (1987). Scent marking activity in the Indian gerbil, *Tatera indica*, in relation to population density. *Animal Behaviour* **35**, 920–941.
- Ikeda, H. (1984). Raccoon dog scent marking by scats and its significance in social behavior. *Journal of Ethology* **2**, 77–84.
- Illius, A. W. and Jessop, N. S. (1995). Modeling metabolic costs of allelochemical ingestion by foraging herbivores. *Journal of Chemical Ecology* **21**, 693–719.

- Izard, M. K. and Vandenbergh, J. G. (1982a). The effects of bull urine on puberty and calving date in crossbred beef heifers. *Journal of Animal Science* **55**, 1160–1168.
 - (1982b). Priming pheromones from oestrous cows increase synchronization of oestrus in dairy heifers after PGC-Z injection. *Journal of Reproduction and Fertility* **66**, 189–196.
- Izhaki, I. and Safiel, U. N. (1990). The effect of some Mediterranean scrubland frugivores upon germination patterns. *Journal of Ecology* **78**, 56–65.
- Izhaki, I., Korine, C., and Arad, Z. (1995). The effect of bat (*Rousettus aegyptiacus*) dispersal on seed germination in Eastern Mediterranean habitats. *Oecologia* **101**, 335–342.
- Jachmann, H. (1989). Food selection by elephants in the "Miombo" dome, in relation to leaf chemistry. *Biochemical Systematics and Ecology* **17**, 15–24.
- Jackson, F. L. C. (1994). Bioanthropological impact of chronic exposure to sublethal cyanides from cassava in Africa. *Actae Horticulturae* **375**, 295–319.
- Jacob, J., Balthazart, J., and Schoffeniels, E. (1979). Sex differences in the chemical composition of uropygial gland waxes in domestic ducks. *Biochemical Systems Ecology* 7, 149–153.
- Jaeger, R. G. (1986). Pheromonal markers as territorial advertisement by terrestrial salamanders. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 191–203. New York: Plenum.
- Jaeger, R. G. and Gergits, W. F. (1979). Intra- and interspecific communication in salamanders through chemical signals on the substrate. *Animal Behaviour* **27**, 150–156.
- Jakubas, W. J. and Gullion, G. W. (1990). Coniferyl benzoate in quaking aspen: a ruffed grouse feeding deterrent. *Journal of Chemical Ecology* **16**, 1077–1087.
- Jakubas, W. J. and Mason, J. R. (1991). Role of avian trigeminal sensory system in detecting coniferyl benzoate, a plant allelochemical. *Journal of Chemical Ecology* 17, 2213–2221.
- Jakubas, W. J., Gullion, G. W., and Clausen, T. P. (1989). Ruffed grouse feeding behavior and its relationship to secondary metabolites of quaking aspen flower buds. *Journal of Chemical Ecology* **15**, 1899–1917.
- Jakubas, W. J., Wentworth, B. C., and Karasov, W. H. (1993). Physiological and behavioral effects of conifer benzoate on avian reproduction. *Journal of Chemical Ecology* **19**, 2353–2377.
- James, P. C. (1986). How do Manx shearwaters, *Puffinus puffinus*, find their burrows? *Ethology* 71, 287–294.
- Jameson, D. A. (1964). Forage plant physiology and soil-range relationships. Effect of defoliation on forage plant physiology. *American Society of Agronony, Special Publication* **5**, 67–80.
- Jankowsky, M. J., Swanson, V. B., and Cramer, D. A. (1974). Field trials of coyote repellents in western Colorado. *Proceedings of the Annual Meeting of the Western Section of the American Society of Animal Science* **25**, 74–76.
- Janzen, D. H. (1978). The ecology and evolutionary biology of seed chemistry as relates to seed predation. In *Biochemical Aspects of Plant and Animal Co-Evolution*, ed. J. B. Harborne, pp. 163–206. London: Academic Press.

- (1981). *Enterolobium cyclocarpum* seed passage rate and survival in horses, Costa Rican Pleistocene seed dispersal agents. *Ecology* **62**, 593–601.
- Jarvis, J. U. M. (1981). Eusociality in a mammal: cooperative breeding in naked mole-rat. *Science* **212**, 571–573.
- Jedrzejewska, B. and Jedrzejewski, W. (1990). Antipredator behaviour of bank voles and prey choice by weasels: enclosure experiments. *Annales Zoologici Fennici* **27**, 321–328.
- Jemiolo, B., Xie, T. M., Andreolini, F., Baker, A. E. M., and Novotny, M. (1991). The t complex of the mouse: chemical characterization by urinary volatile profiles. *Journal of Chemical Ecology* **17**, 353–367.
- Jobling, M. (1995). Environmental Biology of Fishes. London: Chapman and Hall.
- Jogia, M. K., Sinclair, A. R. E., and Andersen, R. J. (1989). An antifeedant in balsam poplar inhibits browsing by snowshoe hares. *Oecologia* **79**, 189–192.
- Johannesson, B. (1987). Observations related to the homing instinct of Atlantic salmon (*Salmo salar* L.). *Aquaculture* **64**, 339–341.
- Johanson, I. B. and Shapiro, E. G. (1986). Intake and behavioral responsiveness to taste stimuli in infant rats from 1 to 15 days of age. *Developmental Psychobiology* **19**, 593–606.
- Johns, M. A., Feder, H. H., Komisaruk, B. R., and Mayer, A. D. (1978). Urine-induced reflex ovulation in anovulatory rats may be a vomeronasal effect. *Nature* **272**, 446–447.
- Johns, T. (1986). The detoxification function of geophagy and the domestication of the potato. *Journal of Chemical Ecology* **12**, 635–646.
 - (1990). With Bitter Herbs They Shall Eat It: Chemical Ecology and Origins of Human Diet and Medicine. Tucson, AZ: University of Arizona Press.
- Johnsen, P. B., Zhou, H., and Adams, M. A. (1988). Olfactory sensitivity of the herbivorous grass carp, *Ctenopharyngodon idella*, to amino acids. *Journal of Fish Biology* **33**, 127–134.
- Johnson, A. and Bailey, C. B. (1972). Influence of bovine saliva on grass regrowth in the greenhouse. *Canadian Journal of Animal Sciences* **52**, 573–574.
- Johnson, B. A. and Leon, M. (2001). Spatial representations of odorant chemistry in the main olfactory bulb of the rat. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 85–91. New York: Kluwer Academic/Plenum.
- Johnston, R. E. (1983). Mechanisms of individual discrimination in hamsters. In *Chemical Signals in Vertebrates*, vol. 3, ed. D. Müller-Schwarze and R. M. Silverstein, pp. 245–258. New York: Plenum.
 - (1992). Olfactory and vomeronasal mechanisms of social communication in golden hamsters. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 515–522. New York: Plenum.
 - (1993). Memory for individual scents in hamsters (*Mesocricetus auratus*) as assessed by habituation methods. *Journal of Comprehensive Psychology* **107**, 201–207.
 - (2001). Neural mechanisms of communication: from pheromones to mosaic signals. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 61–67. New York: Kluwer Academic/Plenum.

- (2005). Communication by mosaic signals: Individual recognition and underlying neural mechanisms. In *Chemical Signals in Vertebrates*, vol. 10, ed. R. T. Mason, M. P. LeMaster, and D. Müller-Schwarze, pp. 269–282. New York: Springer.
- Johnston, R. E. and Jernignan, P. (1994). Golden hamsters recognize individuals, not just individual scents. *Animal Behaviour* **48**, 129–136.
- Johnston, R. E. and Schmidt, T. (1979). Responses of hamsters to scent marks of different ages. *Behavioral and Neural Biology* **26**, 64–75.
- Johnston, R. E., Derzie, A., Chiang, G., Jernigan, P., and Lee, H.-C. (1993). Individual scent signatures in golden hamsters: evidence for specialization of function. *Animal Behaviour* **45**, 1061–1070.
- Johnston, R. E., Munver, A., and Tung, C. (1995). Scent countermarks: selective memory for the top scent by golden hamsters. *Animal Behaviour* **49**, 1435–1442.
- Jones, A. S., Lamont, B. B., Fairbanks, M. M., and Rafferty, C. M. (2003). Kangaroos avoid eating seedlings with or near others with volatile essential oils. *Journal of Chemical Ecology* **29**, 2621–2635.
- Jones, D. A. (1998). Why are so many food plants cyanogenic? *Phytochemistry* 47, 155–162.
- Jones, I. L., Hagelin, J. C., Major, H. L., and Rasmussen, L. E. L. (2004). An experimental field study of the function of crested auklet feather odor. *Condor* **106**, 71–78.
- Jones, K. A. and Hara, T. J. (1982). Behavioral response by Arctic charr (*Salvelinus alpinus*) to taurocholic acid and L-serine, two putative semio-chemicals. *American Zoologist* 22, 925.
- Jones, R. B. (1987). Food neophobia and olfaction in domestic chicks. *Bird Behavior* 7, 78–81.
- Jones, R. B. and Gentle, M. J. (1985). Olfaction and behavioral modification in domestic chicks (*Gallus domesticus*). *Physiology and Behavior* **34**, 917–924.
- Jones, T. H., Gorman, J. S. T., Snelling, R. R., *et al.* (1999). Further alkaloids common to ants and frogs: decahydroquinilines and a quinolizidine. *Journal of Chemical Ecology* **25**, 1179–1193.
- Jorgenson, J. W., Novotny, M., Carmak, M., *et al.* (1978). Chemical scent constitutents in the urine of red fox (*Vulpes vulpes*) during the winter season. *Science* **199**, 796–798.
- Jouventin, P. (1977). Olfaction in snow petrels. Condor 79, 498-499.
- Jouventin, P. and Robin, J. P. (1984). Olfactory experiments on some Antarctic birds. *Emu* **84**, 46–48.
- Jung, H. G. and Batzli, G. O. (1981). Nutritional ecology of microtine rodents: effects of plant extracts on the growth of arctic microtines. *Journal of Mammalogy* **62**, 286–292.
- Kaba, H., Li, C.-S., Keverne, E. B., Saito, H., and Seto, K. (1992). Physiology and pharmacology of the accessory olfactory system. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 49–54. New York: Plenum.
- Kaiser, H. E. and Bartone, J. C. (1966). The carcinogenic activity of ordinary tea. *Journal of the National Medical Association* **58**, 361.
- Kaitz, M., Good, A., Korem, A. H., and Eidelman, A. I. (1987). Mothers' recognition of their newborns by olfactory cues. *Developmental Psychobiology* **20**, 587–592.

- Kalmus, H. (1955). The discrimination by the nose of the dog of individual human odours and in particular the odours of twins. *British Journal of Animal Behaviour* **5**, 25–31.
- Kano, N. (1976). Experiments on the avoidance by Japanese brown bear (*Ursus arctos*) in Noboribetsu Bear Park. *Higuma* 1, 16–17.
- Kare, M. R. and Pick, H. L. (1960). The influence of the sense of taste on feed and fluid consumption. *Poultry Science* **39**, 697–706.
- Kareem, A. M. and Barnard, C. J. (1982). The importance of kinship and familiarity in social interactions between mice. *Animal Behaviour* **30**, 594–601.
- Karlson, P. and Lüscher, M. (1959). "Pheromones": a new term for a class of biologically active substances. *Nature* **183**, 55–56.
- Kassil, V. G. and Gulina, L. K. (1987). Alteration of signal significance of olfactory stimuli in combining them with negative effects on the organism of puppies. *Fiziologicheskii Zhurnal SSSR* **73**, 246–253.
- Kasumyan, A. O. (1994). Olfactory sensitivity of the sturgeon to free amino acids. *Biophysics* **39**, 509–522.
- Kats, L. B. (1988). The detection of certain predators via olfaction by small-mouthed salamander larvae, *Ambystoma texanum*. *Behavioral and Neural Biology* **50**, 126–131.
- Kats, L. B. and Dill, L. M. (1998). The scent of death: chemosensory assessment of predation risk by prey animals. *Ecoscience* **5**, 361–394.
- Kats, L. B., Petranka, J. W., and Sih, A. (1988). Antipredator defenses and the persistence of amphibian larvae. *Ecology* **69**, 1865–1870.
- Katsel, P. L., Dmitrieva, T. M., Valeyer, R. B., and Kozlov, Y. P. (1992). Sex pheromones of male yellow fin Baikal sculpin (*Cottocomephorus grewingki*): Isolation and chemical studies. *Journal of Chemical Ecology* **18**, 2003–2010.
- Katsir, Z., and Crewe, R. M. (1980). Chemical communication in *Galago crassicaudatus*: investigation of the chest gland secrection. *South African Journal of Zoology* **15**, 249–254.
- Kavaliers, M. (1990). Responsiveness of deer mice to a predator, the short-tailed weasel: population differences and neuromodulatory mechanisms. *Physiological Zoology* **63**, 388–407.
- Kawamichi, T. and Kawamichi, M. (1979). Spatial organization and territory of tree shrews (*Tupaia glis*). *Animal Behaviour* 27, 381–393.
- Kaye, H., Mackintosh, N. J., Rothschild, M., and Moore, B. P. (1989). Odour of pyrazine potentiates an association between environmental cues and unpalatable taste. *Animal Behaviour* 37, 563–568.
- Keane, B. (1990). The effect of relatedness on reproductive success and mate choice in the white-footed mouse, *Peromyscus leucopus*. *Animal Behaviour* **39**, 264–273.
- Keefe, M. L. and Winn, H. E. (1991). Chemosensory attraction to the home stream water and conspecifics by native brook trout *Salvelinus fontinalis*, from two southern New England streams. *Canadian Journal of Fisheries and Aquatic Sciences* **48**, 938–944.
- Keeler, R. F. (1986). Teratology of steroidal alkaloids, In *Alkaloids: Chemical and Biological Perspectives*, ed. S. W. Pelletier, pp. 389–425. New York: Wiley.
- Keevin, T. M., Zuleyma, T. H., and McCurdy, N. (1981). Individual and sex specific odors in male and female eastern chipmunks (*Tamias striatus*). *Behavioral Biology* **6**, 329–338.

- Kelley, M. M. (1988). Chemosensory recognition of conspecifics by striped bass juveniles. *Pacific Science* **42**, 123–124.
- Kendall, W. A. and Leath, K. T. (1976). Effect of saponins on palatability of alfalfa to meadow voles. *Agronomy Journal* **68**, 473–476.
- Kendall, W. A. and Sherwood, R. T. (1975). Palatability of leaves of tall fescue and reed canarygrass and of some of their alkaloids to meadow voles. *Agronomy Journal* **67**, 667–671.
- Kendrick, K. M., Keverne, E. B., Chapman, C., and Baldwin, B. A. (1988). Microdialysis measurement of oxytoxcin, aspartate, gamma-aminobutryic acid and glutamate release from the olfactory bulb of sheep during vaginocervical stimulation. *Brain Research* **442**, 171–174.
- Kendrick, K. M., Levy, F., and Keverne, E. B. (1992). Changes in the sensory processing of olfactory signals induced by birth in sheep. *Science* **256**, 833–836.
- Kenney, A. M., Evans, R. L., and Dewsbury, D. A. (1977). Postimplantation pregnancy disruption in *Microtus ochrogaster*, M. pennsylvanicus and Peromyscus maniculatus. Journal of Reproduction and Fertility **49**, 365–367.
- Keverne, E. B. (1983). Chemical communication and reproductive behavior; primates. In *Pheromones and Reproduction in Mammals*, ed. J. G. Vandenbergh, pp. 79–92. New York: Academic Press.
 - (1999). The vomeronasal organ. Science 286, 716–720.
- Keverne, E. B. and de la Riva, C. (1982). Pheromones in mice: reciprocal interactions between the nose and brain. *Nature* **296**, 148–150.
- Keverne, E. B. and Rosser, A. E. (1986). The evolutionary significance of the olfactory block to pregnancy. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 433–439. New York: Plenum.
- Khan, T. Y. and Stoddart, D. M. (1986). Demonstration of an odorous intramale primer effect in short-tailed vole, *Microtus agrestis* L. *Journal of Chemical Ecology* **12**, 2097–2106.
- Khazenchdari, C., Buglass, A. J., and Waterhouse, J. S. (1996). Anal gland secretion of European mole: volatile constitutuents and significance in territorial maintenance. *Journal of Chemical Ecology* **22**, 383–392.
- Kiddy, C. A. and Mitchell, D. S. (1981). Estrus-related odors in cows: time of occurrence. *Journal of Dairy Science* **64**, 267–271.
- Kiddy, C. A., Mitchell, D. S., and Hawk, H. W. (1984). Estrus-related odors in body fluids of dairy cows. *Journal of Dairy Science* **67**, 388–391.
- Kiepenheuer, J. (1985). Can pigeons be fooled about the actual release site position by presenting them information from another site? *Behavioral Ecology* **18**, 75–82.
 - (1986). Are site specific airborne stimuli relevant for pigeon navigation only when matched by other release site information? *Naturwissenschaften* **73**, 42–43.
- Kiesecker, J. M., Chivers, D. P., Anderson, M., and Blaustein, A. R. (2002). Effect of predator diet on life history shifts of red-legged frogs, *Rana aurora*. *Journal of Chemical Ecology* **28**, 1007–1015.
- Kikuyama, S., Toyota, F., Ohmiya, Y., *et al.* (1995). Sodefrin: a female-attracting peptide pheromone in newt cloacal glands. *Science* **267**, 1643–1645.

- Kimball, B. A. and Nolte, D. L. (2004). That's disgusting: deer responses to seedlings treated with proteins. In *Proceedings of the Annual Meeting of International Society of Chemical Ecology*, July 2004, Ottawa, Canada.
- Kirk-Smith, M. D. and Booth, D. A. (1987). Chemoreception in human behavior: experimental analysis of the social effects of fragrances. *Chemical Senses* **12**, 159–166.
- Kirschenbaum, D. M. Schulman, N., and Halpern, M. (1986). Earthworms produce a collagen-like substance detected by the garter snake vomeronasal system. *Proceedings of the National Academy of Sciences, USA* **83**, 1213–1216.
- Kita, M., Nakamura, Y., Okumura, Y., et al. (2004). Blarinatoxin, a mammalian lethal venom from the short-tailed shrew, Blarina brevicauda: isolation and characterization. Proceedings of the National Academy of Science of the USA 101, 7542–7547.
- Kleerekoper, H. and Mogensen, J. (1963). Role of olfaction in the orientation of *Petromyzon marinus*. I. Response to a single amine in prey's body odor. *Physiological Zoology* **36**, 347–360.
- Kleiman, D. (1966). Scent marking in the Canidae. Symposium of the Zoological Society of London 18, 167–177.
- Kleinfield, N. R. (1992). The smell of money. *New York Times*, October 25, Section 9 (Styles), p. 1.
- Klemm, W. R., Hawkins, G. N., and De Los Santos, E. (1987). Identification of compounds in bovine cervico-vaginal mucus extracts that evoke male sexual behavior. *Chemical Senses* **12**, 77–88.
- Klingel, H. (1974). A comparison of the social behavior of the Equidae. In *New Series* 1 *Publication* 24: *The Behavior of Ungulates and Its Relation to Management*, ed. V. Geist and F. Walker, pp. 124–132. Morges, Switzerland: International Union for Conservation of Native and Natural Resources.
- Klopfer, P. H. and Gambale, J. (1966). Maternal imprinting in goats: the role of the chemical senses. *Zeitschrift für Tierpsychologie* **23**, 588–592.
- Knight, M., Glor, R., Smedley, S. R., et al. (1999). Firefly toxicosis in lizards. *Journal of Chemical Ecology* **25**, 1981–1986.
- Knight, T. W. and Lynch, P. R. (1980). Source of ram pheromones that stimulate ovulation in the ewe. *Animal Reproduction Science* **13**, 133–136.
- Knowles, R. I. and Tahan, F. (1979). A repellent to protect radiata pine seedlings from browsing by sheep. *New Zealand Journal of Forestry Science* **9**, 3–9.
- Kobal, G. S. van Toller, S., and Hummel, T. (1989). Is there directional smelling? *Experientia* (Basel) **45**, 130–132.
- Kobayashi, T. and Watanabe, M. (1986). An analysis of snake-scent application behavior in Siberian chipmunks (*Eutamias sibiricus asiaticus*). *Ethology* **72**, 40–52.
- Korn, H. and Taitt, M. J. (1987). Initiation of early breeding in a population of *Microtus townsendii* (Rodentia) with the secondary plant compound 6-MBOA. *Oecologia* **71**, 593–596.
- Krames, L. (1970). Responses of female rats to the individual body odors of male rats. *Psychonomic Science* **20**, 274–275.

- Krasnov, B., Khokhlova, I., and Shenbrot, G. (2002). The effect of host density on ectoparasite distribution: an example of a rodent parasitized by fleas. *Ecology* **83**, 164–175.
- Krause, J. (1993). The effect of "schreckstoff" on the shoaling behavior of the minnow: a test of Hamilton's selfish herd theory. *Animal Behaviour* **45**, 1019–1024.
- Krefting, L. W. and Roe, E. I. (1949). The role of some birds and mammals in seed germination. *Ecological Monographs* **19**, 269–286.
- Krestel, D., Passe, D., Smith, J. C., and Jonsson, L. (1984). Behavioral determination of olfactory thresholds to amyl acetate in dogs. *Neuroscience and Biobehavioral Reviews* 8, 169–174.
- Krieger, J., Schmitt, A., Loebel D., *et al.* (1999). Selective activation of G protein subtypes in the vomeronasal organ upon stimulation with urine-derived compounds. *Journal of Biological Chemistry* **274**, 4655–4662.
- Kristal, M. B., Whitney, J. F., and Peters, L. C. (1981). Placenta on pup's skin accelerates onset of maternal behavior in non-pregnant rats. *Animal Behaviour* **29**, 82–85.
- Kruse, K. C. and Stone, B. M. (1984). Largemouth bass (*Micropterus salmoides*) learn to avoid feeding on toad (*Bufo*) tadpoles. *Animal Behaviour* **32**, 1035–1039.
- Kruska, D. and Rohrs, M. (1974). Comparative-quantitative investigations on brains of feral pigs from the Galapagos Islands and of European domestic pigs. *Zeitschrift für Anatomie und Entwicklungsgeschichte* **144**, 61–73.
- Kruuk, H. (1972). *The Spotted Hyaena, a Study of Predation and Social Behavior*. Chicago, IL: University of Chicago Press.
 - (1978). Spatial organization and territorial behavior of the European badger *Meles meles*. *Journal of Zoology* **184**, 1–19.
 - (1992). Scent marking by otters (*Lutra lutra*): signaling the use of resources. *Behavioral Ecology* **3**, 133–140.
- Kruuk, H., Gorman, M., and Leitch, A. (1984). Scent-marking with the subcaudal gland by the European badger, *Meles meles L. Animal Behaviour* **32**, 899–907.
- Kubie, J. L. and Halpern, M. (1978). Garter snake trailing behavior: effects of varying preyextract concentration and mode of prey-extract presentation. *Journal of Comparative and Physiological Psychology* **93**, 362–373.
- Kubota, K. and Kobayashi, A. (1988). Identification of unknown methyl ketones in volatile flavor components from cooked small shrimp. *Journal of Agricultural and Food Chemistry* **36**, 121–123.
- Kucharski, D. and Hall, W. G. (1987). New routes to early memories. *Science* **238**, 786–788. Kulzer, E. (1961). Über die Biologie der Nil-Flughunde der Gattung (*Rousettus aegyptiacus*). *Natur und Volk* **91**, 219–228.
- Kumari, S. and Prakash, I. (1983). Seasonal variation in the dimension of scent-marking gland of three desert rodents and its possible relationship with their reproductive performance. *Proceedings of the Indian Academy of Sciences, Animal Science* **92**, 299–304.
- Kunz, T. H. (1982). Roosting ecology. In *Ecology of Bats*, ed. T. H. Kun, pp. 1–55. New York: Plenum.

- Kurland, L. T. (1972). An appraisal of the neurotoxicity of cycad and the etiology of amyotrophic lateral sclerosis on Guam. *Federation Proceedings* **31**, 1540–1542.
- Kusnetzov, V. B. (1988). Problem of olfaction reduction in Odontoceti toothed whales. *Zhurnal Obschei Biologii* **49**, 128–135.
- Kvitek, R. G. (1991). Sequestered paralytic shellfish poisoning toxins mediate glaucous-winged gull predation on bivalve prey. *Auk* **108**, 381–392.
- Labov, C. B. and Wysocki, C. J. (1989). Vomeronasal and social influences on urine marking by male mice. *Physiology and Behavior* **45**, 443–447.
- Lacher, T. E. J., Bouchardet de Fonseca, G. A., Alves, C., Jr., and Magalhaes-Castro, B. (1981). Exudate-eating, scent marking and territoriality in wild populations of marmosets. *Animal Behaviour* **29**, 306–307.
- Ladewig, J., Price, E. O., and Hart, B. L. (1980). Flehmen in male goats: role in sexual behavior. *Behavioral and Neural Biology* **30**, 312–322.
- Laffort, P. and Gortan, C. (1987). Olfactory properties of some gases in hyperbaric atmosphere. *Chemical Senses* **12**, 139–142.
- Laing, D. G. (1975). A comparative study of the olfactory sensitivity of humans and rats. *Chemical Senses and Flavour* **1**, 257–269.
- Langley, W. M. (1988). Spiny mouse's (*Acomys cahirinus*) use of its distance senses in prey localization. *Behavioral Processes* **16**, 67–73.
- Larson, J. K. and McCormick, M. I. (2005). The role of chemical alarm signals in facilitating learned recognition of novel cues in a coral reef fish. *Animal Behaviour* **69**, 51–57.
- Larson, R. A. and Berenbaum, M. R. (1988). Environmental phototoxicity. *Environmental Science and Technology* **22**, 354–360.
- Laska, M. (1990). Olfactory sensitivity to food odor components in the short-tail fruit bat, *Carollia perspicillata* (Phyllostomatidae, Chiroptera). *Journal of Comparative Physiology* **166**, 395–399.
- Laska, M. and Hudson, R. (1991). A comparison of the detection thresholds of odour mixtures and their components. *Chemical Senses* **16**, 651–662.
- Laska, M., Rother, G., and Schmidt, V. (1986). Die Beeinflussung des Riechvermögens durch die Luftfeuchte bei *Carollia perspicillata und Phyllostomus discolor* (Chiroptera). *Zeitschrift für Säugetierkunde* **51**, 74–79.
- Laska, M., Hudson, R., and Distel, H. (1990). Sensitivity to biologically relevant odours may exceed the sum of component thresholds. *Chemoecology* **1**, 139–141.
- Lauman, J., Pern, U., and Blum, V. (1974). Investigations on the function and hormonal regulation of the anal appendices in *Blennius payo* (Risso). *Journal of Experimental Zoology* **190**, 47–56.
- Launchbaugh, K. L., Provenza, F. D., and Burritt, E. A. (1993). How herbivores track variable environments: response to variability of phytotoxins. *Journal of Chemical Ecology* **19**, 1047–1056.
- Laurila, A. (2000). Responses to predator chemical cues and local variation in antipredator behaviour of *Rana temporaria* tadpoles. *Oikos* **88**, 159–168.
- Lavin-Murcio, P., Robinson, B. G., and Kardong, K. (1993). Cues involved in relocation of struck prey by rattlesnakes, *Crotalis viridis oreganus*. *Herpetologica* **49**, 463–469.

- Lawless, H. T. and Engen, T. (1977). Associations to odors: interference, mnemonics and verbal labeling. *Journal of Experimental Psychology H.* **3**, 52–59.
- Lawton, A. D. (1979). Inhibition of sexual maturation by a urinary pheromone in male prairie deer mice. *Hormones and Behavior* **13**, 128–138.
- Leffingwell, J. C. (2001). Olfaction. *Leffingwell Reports*, **1**, 1–24. www.Leffingwell.com/olfaction.htm.
- Lehner, P. N., Krumm, R., and Cringa, A. T. (1976). Tests for olfactory repellents for coyotes and dogs. *Journal of Wildlife Management* **40**, 145–150.
- Leinders-Zufall, T., Lane, A. P., Puche, A. C., *et al.* (2000). Ultrasensitive pheromone detection in the vomeronasal organ upon stimulation with urine-derived compounds. *Nature* **405**, 792–796.
- Leinders-Zufall, T., Brennan, P., Widmayer, P., *et al.* (2004). MHC class I peptides as chemosensory signals in the vomeronasal organ. *Science* **306**, 1033–1037.
- LeMaster, M. P. and Mason, R. T. (2002). Variation in a female sexual attractiveness pheromone controls male mate choice in garter snakes. *Journal of Chemical Ecology* **28**, 1269–1285.
 - (2003). Pheromonally mediated sexual isolation among denning populations of redsided garter snakes, *Thamnophis sirtalis parietalis*. *Journal of Chemical Ecology* **29**, 1027–1043.
- LeMaster, M. P., Moore, I. T., and Mason, R. T. (2001). Conspecific trailing behavior of redsided garter snakes, *Thamnophis sirtalis parietalis*, in the natural environment. *Animal Behaviour* **61**, 827–833.
- Lenington, S. and Egid, K. (1985). Female determination of male odor correlated with male genotype at the T-locus: A response to T-locus or H2 locus variability? *Behavior Genetics* **15**, 53–67.
- Leon, M. (1975). Dietary control of maternal pheromone in the learning rat. *Physiology and Behavior* **14**, 311–319.
- Leon, M. and Behse, J. H. (1977). Dissolution of the pheromonal bond: waning of approach response by weanling rats. *Physiology and Behavior* **18**, 393.
- Leopold, A. S., Erwin, M., Oh, J., and Browning, B. (1976). Phytoestrogens: adverse effects in reproduction in California quail. *Science* **191**, 98–99.
- Lepri, J. J. and Wysocki, C. J. (1987). Removal of the vomeronasal organ disrupts the activation of reproduction in female voles. *Physiology and Behavior* **40**, 349–355.
- Lepri, J. J., Wysocki, C. J., and Vandenbergh, J. G. (1985). Mouse vomeronasal organ: effects on pheromone production and maternal behavior. *Physiology and Behavior* **35**, 809–814.
- Lequette, B., Verheyden, C., and Jouventin, P. (1989). Olfaction in subantarctic sea-birds: its phylogenetic and ecological significance. *Condor* **91**, 732–735.
- Levin, D. A. (1971). Plant phenolics: an ecological perspective. *American Naturalist* **105**, 157–181.
- Levy, F. and Poindron, P. (1987). The importance of amniotic fluids for the establishment of maternal behavior in experienced and inexperienced ewes. *Animal Behaviour* **35**, 1188–1192.

- Lewis, D. M. (1987). Fruiting patterns, seed germination, and distribution of *Sclerocarya* caffra in an elephant-inhabited woodland. *Biotropica* **19**, 50–56.
- Leyden, J. J., McGinley, K. J., Hoelzle, E., Labows, J. N., and Kligman, A. M. (1981). The microbiology of the human axillae and its relation to axillary odors. *Journal of Investigative Dermatology* 77, 413–416.
- Li, W., Scott, A. P., Siefkes, M. J., *et al.* (2002). Bile acid secreted by male sea lamprey that acts as a sex pheromone. *Science* **296**, 138–141.
- Licht, T. (1989). Discriminating between hungry and satiated predators: the response of guppies (*Poecilia reticulata*) from high and low predation sites. *Ethology* **82**, 238–243.
- Liley, N. R. (1982). Chemical communication in fish. *Canadian Journal of Fisheries* **39**, 22–35.
- Lincoln, J., Coopersmith, R., Harris, E. W., Cotman, C. W., and Leon, M. (1988). NMDA receptor activation and early olfactory learning. *Developmental Brain Research* **39**, 309–312.
- Lindquist, S. B. and Bachman, M. D. (1980). Feeding behavior of the tiger salamander, *Ambystoma tigrinum*. *Herpetologica* **36**, 144–158.
- Lindquist, N., Hay, M. E., and Fenical, W. (1992). Defense of ascidians and their conspicuous larvae: adult vs. larval chemical defenses. *Ecological Monographs* **62**, 547–568.
- Lindroth, R. L. (1988). Adaptations of mammalian herbivores to plant chemical defenses. In *Chemical Mediation of Coevolution*, ed. K. C. Spencer, pp. 415–445. San Diego, CA: Academic Press.
- Lindroth, R. L. and Batzli, G. O. (1983). Detoxication of some naturally occuring phenolics by prairie voles: a rapid assay of glucuronidation metabolim. *Biochemical Systematics and Ecology* **11**, 405–409.
- (1984). Plant phenolics as chemical defenses: effects of natural phenolics on survival and growth of prairie voles (*Microtus ochrogaster*). *Journal of Chemical Ecology* **10**, 229–244.
- Lindroth, R. L., Batzli, G. O., and Avildsen, S. I. (1986). *Lespedeza* phenolics and *Penstemon* alkaloids: effects on digestion efficiencies and growth of voles. *Journal of Chemical Ecology* **12**, 713–728.
- Lindvall, T. and Svensson, L. T. (1974). Equal unpleasantness matching of malodorous substances in a community. *Journal of Applied Psychology* **59**, 264.
- Linn, C. E. Jr., Campbell, M. G., and Roelofs, W. L. (1987). Pheromone components and active spaces: what do moths smell and where do they smell it? *Science* 237, 650–651.
- Lipsitt, L. P., Engen, T., and Kaye, H. (1963). Developmental changes in the olfactory threshold of the neonate. *Child Development* **34**, 371–376.
- Löhner, L. (1926). Untersuchungen über die geruchsphysiologische Leistungsfähigkeit von Polzeihunden. *Archiv für die gesamte Physiologie* **121**, 84–94.
- Løkkeborg, S., Olla, B. L., Pearson, W. H., and Davis, M. W. (1995). Behavioral responses of sablefish, *Anoplopoma fimbria*, to bait odor. *Journal of Fisheries Biology* **46**, 142–155.
- Lomas, D. E. and Keverne, E. B. (1982). Role of vomeronasal organ and prolactin in the acceleration of puberty in female mice. *Journal of Reproduction and Fertility* **66**, 101–107.
- Lombardi, J. R. and Vandenbergh, J. G. (1977). Pheromonally induced sexual maturation in females: regulation by the social environment of the male. *Science* **196**, 545–546.

- Lombardi, J. R., Vandenbergh, J. G., and Whitsett, J. M. (1976). Androgen control of the sexual maturation pheromone in house mouse urine. *Biology of Reproduction* **15**, 179–186.
- Loop, M. S. and Scoville, S. A. (1972). Response of newborn *Eumeces inexpectatus* to preyobject extracts. *Herpetologia* **28**, 254–256.
- López, P. and Salvador, A. (1992). The role of chemosensory cues in discrimination of prey odors by the amphisbaenan *Blanus cinereus*. *Journal of Chemical Ecology* **18**, 87–93.
- Lorenz, K. (1963). Das sogenannte Böse: Zur Naturgeschichte der Aggression. Vienna: Borotha-Schoeler.
- Löyttiniemi, K. (1981). Nitrogen fertilization and nutrient contents in Scots pine in relation to browsing preference by moose (*Alces alces*). Folia Forestalia 478, 12–14.
- Löyttiniemi, K. and Hiltonen, R. (1978). Monoterpenes in Scots pine in relation to browsing preference by moose. (*Alces alces L.*) *Silvia Fennica* **12**, 85–87.
- Lucas, C. E. (1944). Excretions, ecology and evolution. *Nature* **153**, 378–379.
- Luschi, P. and Dall'Antonia, P. (1993). Anosmic pigeons orient from familiar sites by relying on the map-and-compass mechanism. *Animal Behaviour* **46**, 1195–1203.
- Lydell, K. and Doty, R. L. (1972). Male rat odor preferences for female urine as a function of sexual experience, urine age, and urine source. *Hormones and Behavior* **3**, 205–212.
- Lyman, B. J. and McDaniel, M. A. (1986). Effects of encoding strategy on long-term memory for odors. *Quarterly Journal of Experimental Psychology A: Human Experimental Psychology* **38**, 753–766.
- Mabry, T. J. and Gill, J. E. (1979). Sesquiterpene lactones and other terpenoids. In *Herbivores: Their Interaction with Secondary Plant Metabolites*, ed. G. A. Rosenthal and D. H. Janzen, pp. 501–537. New York: Academic Press.
- MacDonald, D., Kranz, K., and Aplin, R. D. (1984). Behavioral, anatomical and chemical aspects of scent marking amongst capybaras, *Hydrochoerus hydrochaeris* (Rodentia: Caviomorpha). *Journal of Zoology* (London) **202**, 341–360.
- MacDonald, K. B. (1980). Activity patterns in a captive wolf pack. Carnivore 31, 62-64.
- MacFarlane, A. (1975). Olfaction in the development of social preferences in the human neonate. In *Ciba Foundation Symposium*, No. 33: *Parent–Infant Interaction*, pp. 103–113. New York: Elsevier.
- MacGintie, G. E. (1939). The natural history of the blind goby, *Typhlogobius californiensis* Steindachner. *American Midland Naturalist* **21**, 489–505.
- MacLain, D. K. and Shure, D. J. (1985). Host plant toxins and unpalatability of *Neacoryphus bicruris* (Hemiptera: Lygaeidae). *Ecological Entomology* **10**, 291–298.
- Macrides, F., Bartke, A., and Dalterio, S. (1975). Strange females increase plasma testosterone levels in male mice. *Science* **189**, 1104–1106.
- Maderson, P. A. F. (1986). The tetrapod epidermis: a system protoadapted as a semiochemical source. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 13–25. New York: Plenum.
- Madison, D. M. (1977). Chemical communication in amphibians and reptiles, In *Chemical Signals in Vertebrates*, vol. 1, ed. D. Müller-Schwarze and M. M. Mozell, pp. 135–168. New York: Plenum.

- Madison, D. M., Sullivan, A. M., Maerz, J. C., McDarby, J. H., and Rohr, J. R. (2002). A complex, cross-taxon, chemical releaser of antipredator behavior in amphibians. *Journal of Chemical Ecology* **28**, 2271–2282.
- Madubunyi, L. C., Hassanali, A., Ouma, W., Nyarango, D., and Kabii, J. (1996). Chemoecological role of mammalian urine in host location by tsetse, *Glossina* spp. (Diptera: Glossinidae). *Journal of Chemical Ecology* **22**, 1187–1199.
- Magurran, A. E. (1986). Predator inspection behavior in minnow shoals: differences between populations and individuals. *Behavioral Ecology and Sociobiology* **19**, 267–273.
 - (1989). Acquired recognition of predator odour in the European minnow (*Phoxinus phoxinus*). Ethology **82**, 216–223.
- Mainardi, D., Marsan, M., and Pasqali, A. (1965). Causation of sexual preferences of the house mouse. The behavior of mice reared by parents whose odor was artificially altered. *Atti della Societa Italiana di Scienze Naturali e el Museo Civico de Storia Naturale di Milano* **104**, 325–338.
- Mair, R. G., Bouffard, J. A., Engen, T., and Morton, T. H. (1978). Olfactory, sensitivity during the menstrual cycle. *Sensory Processes* 2, 90–98.
- Malakoff, D. (1999). Following the scent of avian olfaction. Science 286, 704–705.
- Malone, N., Payne, C. E., Beynon, R. J., and Hurst, J. L. (2001). Social status, odour communication and mate choice in wild house mice. In *Chemical Signals in Vertebrates*, vol. 9, ed. Marchlewska-Koj, A., Lepri, J. J., and Müller-Schwarze. D., pp. 217–224. New York: Kluwer Academic/Plenum.
- Manning, C. J., Wakeland, E. K., and Potts, W. K. (1992). Communal nesting patterns in mice implicate MHC genes in kin recognition. *Nature* **360**, 581–583.
- Manteifel, Y. B. and Goncharova, N. V. (1992). Sensitivity and specificity of some L-amino acids perception in a European water turtle *Emys orbicularis*: chemotesting movements and avoidance reaction. *Comparative Biochemistry and Physiology* **102**, 527–531.
- Manteifel, Y., Goncharova, N., and Boyo, V. (1992). Chemotesting movements and chemosensory sensitivity to amino acids in the European pond turtle, *Emys orbicularis* L. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 397–401. New York: Plenum.
- Mapert, H. L., Cieslak, A., Alkan, O., *et al.* (1999). The golden hamster *aprodisin* gene. Structure, expression in parotid glands of female animals and comparison with a similar murine gene. *Journal of Biological Chemistry* **274**, 444–450.
- Marchisin, A. (1980). Predator-prey interactions between snake-eating snakes and pit vipers. Ph.D. Thesis, Rutgers University, Newark, NJ, USA.
- Marchlewska-Koj, A. (1990). Chemical interaction between adult female mice: the role of ovarian and adrenal hormones. In *Chemical Signals in Vertebrates*, vol. 5, ed. D. W. Macdonald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 209–216. New York: Oxford University Press.
- Marchlewska-Koj, A. and Kruczek, M. (1986). Female-induced delay of puberty in bank vole and European pine vole females. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 551–554 New York: Plenum Press.

- Marchlewska-Koj, A., Pochron, E. and Sliwowska, A. (1990). Salivary glands and preputial glands of males as source of estrus-stimulating pheromone in female mice. *Journal of Chemical Ecology* **16**, 2817–2822.
- Margolis, F. L., Kudrycki, K., Stein-Izsak, C., Grillo, M., and Akeson, R. (1993). From genotype to olfactory phenotype: the role of the Olf-1-binding site. *Ciba Foundation Symposia* **179**, 3–20.
- Marks, D. L., Swain, T., Goldstein, S., Richard, A., and Leighton, M. (1988). Chemical correlates of rhesus monkey food choice: the influence of hydrolyzable tannins. *Journal of Chemical Ecology* **14**, 213–235.
- Marlier, L., Schaal, B., Gaugler, C., and Messer, J. (2001). Olfaction in premature newborns: detection and discrimination abilities two months before gestational term. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 205–209. New York: Kluwer Academic/Plenum.
- Marples, N. M. (1993). Do wild birds use size to distinguish palatable and unpalatable prey types? *Animal Behaviour* **46**, 347–354.
- Marquis, R. J. and Batzli, G. O. (1989). Influence of chemical factors on palatability of forage to voles. *Journal of Mammalogy* **70**, 503–511.
- Marsden, A. M. and Bronson, F. A. (1964). Estrus synchrony in mice: alteration by exposure to male urine. *Science* **144**, 1469.
- Marshall, D. A. and Moulton, D. G. (1981). Olfactory sensitivity to α -ionone in humans and dogs. *Chemical Senses* **6**, 53–61.
- Martin, G. B., Oldham, C. M., Cognie, Y., and Pearce, D. T. (1986). The physiological responses of anovulatory ewes to the introduction of rams: a review. *Livestock Production Science* **15**, 219–247.
- Martin, I. G. and Beauchamp, G. K. (1982). Olfactory recognition of individuals by male cavies (*Cavia aperea*). *Journal of Chemical Ecology* **8**, 1241–1249.
- Martin, J. S. and Martin, M. M. (1983). Tannin essays in ecological studies. Precipitation of ribulose-1,5-biphosphate carboxylase/oxygenase by tannic acid, quebracho, and oak foliage extracts. *Journal of Chemical Ecology* **9**, 285–294.
- Martin, M. L., Price, E. O., Hallach, J. R., and Dally, M. R. (1987). Fostering lambs by odor transfer: the add-on experiment. *Journal of Animal Science* **64**, 1378–1383.
- Martof, B. S. (1962). Some observations on the role of olfaction among salient amphibia. *Physiological Zoology* **35**, 270.
- Maruniak, J. A., Silver, W. L., and Moulton, D. G. (1983). Olfactory receptors respond to blood-borne odorants. *Brain Research* **265**, 312–316.
- Maruniak, J. A., Wysocki, C. J., and Taylor, J. A. (1986). Mediation of male mouse urine marking and aggression by the vomeronasal organ. *Physiology and Behavior* **37**, 655–657.
- Maser, C., Trappe, J. M., and Nussbaum, R. A. (1978). Fungal-small mammal interrelationships with emphasis on Oregon coniferous forests. *Ecology* **59**, 799-809.
- Mason, J. R. (1989). Avoidance of methiocarb-poisoned apples by red-winged blackbirds. *Journal of Wildlife Management* **53**, 836–840.

- Mason, J. R. and Clark, L. (1986). Chemoreception and the selection of green plants as nest fumigants by starlings. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 369–384. New York: Plenum.
 - (1987). Behavioral assessment of olfactory and trigeminal responsiveness of starlings *Sturnus vulgaris* to nine anthranilates. *Chemical Senses* **12**, 679.
- Mason, J. R. and Stevens, D. A. (1981a). Discrimination and generalization among reagent grade odorants by tiger salamanders (*Ambystoma tigrinum*). *Physiology and Behavior* **27**, 647–653.
 - (1981b). Behavioral determinations of thresholds for *n*-butyl acetate and *n*-butyl alcohol in the tiger salamander (*Ambystoma tigrinum*). *Chemical Senses* **6**, 189–195.
- Mason, J. R., Rabin, M. D., and Stevens, D. A. (1982). Conditioned taste aversions: skin secretions used for defense by tiger salamanders (*Ambystoma tigrinum*). *Copeia*. 667–671.
- Mason, J. R., Greenspon, J. M. and Silver, W. L. (1987a). Capsaicin and its effects on olfaction and trigeminal chemoreception. *Acta Physiologica Hungarica* **69**, 469–479.
- Mason, J. R., Kamalesh, J. and K., Morton, T. H. (1987b). Generalization in olfactory detection of chemical cues containing carbonyl functions by tiger salamanders (*Ambystoma tigrinum*). 13, 1–18.
- Mason, J. R., Bean, N. J., and Galef, B. G., Jr. (1988). Attractiveness of carbon disulfide to wild Norway rats. *Proceedings of Vertebrate Pest Conference*, vol. 13, ed. A. C. Crabb and R. E. Marsh, pp. 95–97.
- Mason, R. T., Fales, H. M., Jones, T. H., *et al.* (1989a). Sex pheromones in snakes. *Science* **245**, 290–293.
- Mason, J. R., Bullard, R. W., Dolbeer, R. A., and Woronecki, P. P. (1989b). Red-winged blackbird (*Agelaius phoeniceus* L.) feeding responses to oil and anthocyanin levels in sunflower meal. *Crop Protection* **8**, 455–460.
- Mason, J. R., Adams, M. A., and Clark, L. (1989c). Anthranilate repellency to starlings: chemical correlates and sensory perception. *Journal of Wildlife Management*. **53**, 55–64.
- Mason, J. R., Bean, N. J. Shah, P. S., and Clark, L. (1991). Taxon-specific differences in responsiveness to capsaicin and several analogues: correlates between chemical structure and behavioral aversiveness. *Journal of Chemical Ecology* 17, 2539–2552.
- Mason, J. R., Epple, G., and Nolte, D. L. (1994). Semiochemicals and improvements in rodent control. In *Behavioral Aspects of Feeding: Basic and Applied Research in Mammals*, ed. B. G. Galef, M. Mainardi, and P. Valsecchi, pp. 327–345. Chur, Switzerland: Harwood Academic.
- Mason, R. T. (1992). Reptilian pheromones. In *Biology of the Reptilia*, vol. 18, *Hormones, Brain and Behavior*, ed. C. Gans and D. Crews, pp. 114–228. Chicago, IL: University of Chicago Press.
- Mason, R. T. and Greene, M. J. (2001). Invading pest species and the threat to biodiversity: pheromonal control of Guam brown tree snakes, *Boiga irregularis*. In *Chemical Signals in Vertebrates*, vol. 9, eds. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 361–368. New York: Kluwer Academic/Plenum.

- Mason, R. T. and Gutzke, W. H. N. (1990). Sex recognition in the leopard gecko, *Euble-pharis macularis* (Sauria: Gekkonidae). Possible mediation by skin-derived semiochemicals. *Journal of Chemical Ecology* **16**, 27–36.
- Massey, A. and Vandenbergh, J. G. (1980). Puberty delay by a urinary cue from female house mice in feral populations. *Science* **209**, 821–822.
- Matochik, J. A. (1988). Role of the main olfactory system in recognition between individual spiny mice. *Physiology and Behavior* **42**, 217–222.
- Matsunami, H. and Buck, L. B. (1997). A multigene family encoding a diverse array of putative pheromone receptors in mammals. *Cell* **90**, 775–784.
- May, R. M. (1972). Limit cycles in predator-prey communities. Science 177, 900-902.
- Mayer, J. J. and Brisbin, I. L., Jr. (1986). A note on the scent-marking behavior of two captive-reared feral boars. *Applied Animal Behaviour Science* **16**, 85–90.
- McAdoo, J. K., Evans, C. C., Roundy, B. A., Young, J. A., and Evans, R. A. (1983). Influence of heteromyid rodents on *Oryzopsis hymenoides* germination. *Journal of Range Management* **36**, 61–64.
- McArthur, C., Sanson, G. D., and Beal, A. M. (1995). Salivary proline-rich proteins in mammals: roles in oral homeostasis and counteracting dietary tannin. *Journal of Chemical Ecology* **21**, 663–691.
- McClenaghan, L. R. (1987). Lack of effect of 6-MBOA on reproduction in *Dipodomys merriami*. *Journal of Mammalogy* **68**, 150–152.
- McClintock, M. K. (1971). Menstrual synchrony and suppression. Nature 229, 244–245.
- McDonough, L. M., Brown, D. F., and Aller, W. C. (1989). Insect sex pheromones. Effect of temperature on evaporation rates of acetates from rubber septa. *Journal of Chemical Ecology* **15**, 779–790.
- McGlone, J. (2002). *Pig Production: Biological Principles and Applications*. Clifton Park, NJ: Delmar Learning.
- McGregor, J. H and Teska, W. R. 1989. Olfaction as an orientation mechanism in migrating *Ambystoma maculatum*. *Copeia* **1989**, 779–781.
- McKey, D. and Beckerman, S. (1993). Chemical ecology, plant evolution and traditional manioc cultivation systems. In *Man and the Biosphere Series*, vol. 13: *Tropical Forests People and Food*, ed. C. M. Hadlik, A. Hadlik, O. F. Linares, *et al.* pp. 321–338. Paris: UNESCO-MAB/Parthenon Publishing.
- McKey, D., Gartlan, J. S., Waterman, P. G., and Choo, G. M. (1981). Food selection by black colobus monkeys (*Colobus satanus*) in relation to plant chemistry. *Biological Journal of the Linnean Society* 16, 115–146.
- McKinnon, J. S. and Liley, N. R. (1987). Asymetric species specificity in response to female pheromone by males of two species of trichogaster (Pisces: Blontidae). *Canadian Journal of Zoology* **65**, 1129–1134.
- McLaughlin, S. K., Mckinnon, P. J., Robichon, A., Spickofsky, N., and Margolskee, R. F. (1993). Gustducin and transducin: a tale of two G proteins. *Molecular Basis of Smell and Taste Transduction* **179**, 186–200.
- McLean, S., Foley, W. J., Davies, N. W., et al. (1993). Metabolic fate of dietary terpenes from Eucalyptus radiata in common ringtail possom (Pseudocheirus peregrinus). Journal of Chemical Ecology 19, 1625–1643.

- McLennan, D. A. (2005). Changes in response to olfactory cues across the ovulatory cycle in brook sticklebacks, *Culaea inconstans*. *Animal Behaviour* **69**, 181–188.
- McMahon, T. E. and Tash, J. C. (1979). The use of chemosenses by threadfin shad, *Dorosoma* petenense to detect conspecifics, predators and food. *Journal of Fisheries Biology* **14**, 289–296.
- McNaughton, S. J. (1985). Interactive regulation of grass yield and chemical properties by defoliation, a salivary chemical, and inorganic nutrition. *Oecologia* **65**, 478–486.
- Mead, R. J., Oliver, A. J., King, D. R., and Hubach, P. H. (1985). The co-evolutionary role of fluoroacetate in plant–animal interactions in Australia. *Oikos* 44, 55–60.
- Mech, L. D. (1970). *The Wolf: The Ecology and Behavior of an Endangered Species.* Minneapolis, MI: University of Minnesota Press.
- Mehansho, H., Hagerman, A., Clements, S., et al. (1983). Modulation of proline-rich protein biosynthesis in rat parotid glands by sorghums with high tannin levels. *Proceedings of the National Academy of Sciences USA* **80**, 3948–3952.
- Mehansho, H., Butler, L. G., and Carlson, D. M. (1987). Dietary tannins and salivary proline-rich proteins: interactions, induction, and defense mechanism. *Annual Review of Nutrition* 7, 423–440.
- Meier, P. T. (1991). Response of adult woodchucks (*Marmota monax*) to oral-gland scents. *Journal of Mammalogy* **72**, 622–624.
- Meisami, E. and Wenzel, B. (1987). Is the northern fulmar's large olfactory bulb designed for high sensitivity in odor detection? *Chemical Senses* 12, 681–682.
- Melcer, T. and Chiszar, D. (1989). Striking prey creates a specific chemical search image in rattlesnakes. *Animal Behaviour* **37**, 477–486.
- Melchiors, M. A. and Leslie, C. A. (1985). Effectiveness of predator fecal odors as black-tailed deer repellents. *Journal of Wildlife Management* **49**, 358–362.
- Melrose, D. R., Reed, H. C. B., and Patterson, R. L. S. (1971). Androgen steroids associated with boar odour as an aid to the detection of oestrus in pig artificial insemination. *British Veterinary Journal* **127**, 497–502.
- Mennella, J. A. and Moltz, H. (1988). Infanticide in the male rat: the role of the vomeronasal organ. *Physiology and Behavior* **42**, 303–306.
 - (1989). Pheromonal emission by pregnant rats protects against infanticide by nulliparous conspecifics. *Physiological Behavior* **46**, 591–596.
- Meredith, M. (1983). Sensory physiology of pheromone communication. In *Pheromones and Reproduction in Mammals*, ed. J. G. Vandenbergh, pp. 199–252. New York: Academic Press.
- (1986). Vomeronasal organ removal before sexual experience impairs male hamster mating behavior. *Physiological Behavior* **36**, 737–743.
- Meredith, M. and O'Connell, R. J. (1979). Efferent control of stimulus access to the hamster vomeronasal organ. *Journal of Physiology* **286**, 301–316.
- Merkens, M., Harestad, A. S., and Sullivan, T. P. (1991). Cover and the efficacy of predator-based repellents for Townsend's vole, *Microtus townsendii*. *Journal of Chemical Ecology* 17, 401–412.

- Merkx, J., Slob, A. K., van der Werff, J. J., and ten Bosch, J. J. (1988). Vaginal bacterial flora partially determines sexual attractivity of female rats. *Physiology and Behavior* **44**, 147–149.
- Merritt, G. C., Goodrich, B. S., Hesterman, E. R., and Mykytowycz, R. (1982). Microflora and volatile fatty acids present in inguinal pouches of the wild rabbit, *Oryctolagus cuniculus*, in Australia. *Journal of Chemical Ecology* **8**, 1217–1226.
- Mester, A. F., Doty, R. L., Shapiro, A., and Frye, R. E. (1988). Influence of body tilt within the sagittal plane on odor identification performance. *Aviation, Space and Environmental Medicine* **59**, 734–737.
- Meyer, M. W. and Karasov, W. H. (1989). Antiherbivore chemistry of *Larrea tridentata*: effects on woodrat (*Neotoma lepida*) feeding and nutrition. *Ecology* **70**, 953–961.
- Meyers, C. W. and Daly, J. W. (1983). Dart-poison frogs. Scientific American 248, 120–133.
- Meyer, M. W. and Karasov, W. H. (1991). Chemical aspects of herbivory in arid and semiarid habitats. In *Plant Defenses Against Mammalian Herbivory*, ed. R.T. Palo and C. T. Robbins, pp. 167–187. Boca Raton, FL: CRC Press.
- Millar, J. G. and Haynes, K. F. (1988). *Chemical Ecology Methodology*. New York: Chapman and Hall.
 - (1998). Chemical Ecology Methodology. New York: Chapman & Hall.
- Mills, M. G. L., Gorman, M. L., and Mills, M. E. J. (1980). The scent marking behaviour of the brown hyaena *Hyaena brunnea*. *South African Journal of Zoology* **15**: 240–248.
- Milton, K. (1979). Factors influencing leaf choice by howler monkeys: a test of some hypotheses of food selection by generalist herbivores. *American Naturalist* **114**, 362–378.
- Minguez, E. (1997). Olfactory nest recognition by British storm-petrel chicks. *Animal Behaviour* **5**, 701–707.
- Mirza, R. S. and Chivers, D. P. (2001). Do chemical alarm signals enhance survival of aquatic vertebrates? An analysis of the current research paradigm. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 19–26. New York: Kluwer Academic/Plenum.
- Mitchell, D., Laycock, J. D., and Stevens, W. F. (1977). Motion sickness-induced pica in rats. *American Journal of Clinical Nutrition* **30**, 147–150.
- Moen, A. N. (1973). Wildlife Ecology. San Francisco, CA: W. H. Freeman.
- Molez-Verriere, N. (1988). Detection and capture of prey in two species of Galagidae, Primates. *Biology of Behavior* **13**, 30–42.
- Mombaerts, P. (1999). Seven-transmembrane proteins as odorant and chemosensory receptors. *Science* **286**, 707–711.
- Moncomble, A.-S., Coureaud, G., Quennedey, B., et al. (2005). The mammary pheromone of the rabbit: from where does it come? *Animal Behaviour* **69**, 29–38.
- Montagner, H. (1974). Communication non verbale et discrimination olfactive chez le jeune enfant: approche éthologique. In *L' Unite de l' Homme*, ed. E. Morin and M. Piatelli-Palmarini. Paris: Seuil.
- Montague, J. C., Pocock, D. C., and Wright, W. (1990). An examination of the animal browsing problem in Australian eucalypt and pine plantations. In *Proceedings of the*

- 14th Vertebrate Pest Conference, ed. L. R.Davis and R. E. Marsh, pp. 203–208. Davis, CA: University of California at Davis.
- Montgomery-St. Laurent, T., Fullenkamp, A. M., and Fischer, R. B. (1988). A role for the hamster's flank gland in heterosexual communication. *Physiology and Behavior* **44**, 759–762.
- Moodie, J. D. and Byers, J. A. (1989). The function of scent marking by males on female urine in pronghorns. *Journal of Mammalogy* **70**, 812–814.
- Moore, A. and Waring, C. P. (2001). The effects of a synthetic pyrethroid peticide on some aspects of reproduction in Atlantic salmon (*Salmo salar L.*). *Aquatic Toxicology* **52**, 1–12.
- Moore, B. D., Wallis, I. R., Palá-Paúl, J., et al. (2004). Antiherbivore chemistry of Eucalyptus: cues and deterrents for marsupial folivores. *Journal of Chemical Ecology* **30**, 1743–1769.
- Moore, C. L. (1984). Maternal contribution to the development of masculine sexual behavior in laboratory rats. *Developmental Psychobiology* **17**, 347–356.
- Moore, R. E. (1965). Olfactory discrimination as an isolation mechanism between *Peromyscus maniculatus* and *Peromyscus polionotus*. *American Midland Naturalist* **73**, 85–100.
- Moore, W. G. and Marchinton, R. L. (1974). Marking behavior and its social function in white-tailed deer. In *The Behavior of Ungulates and Its Relation to Management*, ed. V. Geist and F. Walter. pp. 447–456. Morges, Switzerland: International Union for the Conservation of Nature and Natural Resources.
- Mordue-Luntz, A. J., Ingvarsdöttir, A., Birkett, M. A., et al. (2004). The role of pheromones and kairomones in mate location of the salmon louse *Lepeophtheirus salmonis* (Crustacea; Caligidae). In *Proceedings of the Annual Meeting of International Society of Chemical Ecology*, July 2004, Ottawa, Canada.
- Mori, K., Nagao, H., and Yoshihara, Y. (1999). The olfactory bulb: coding and processing of odor molecule information. *Science* **286**, 711–715.
- Morin, P. P., Dodson, J. J., and Dore, F. Y. (1987). Laboratory identification of a sensitive period for olfactory imprinting in young Atlantic salmon. *American Zoologist* 27, 77A.
- Morris, M. P. and García-Rivera, J. (1955). The destruction of oxalates by the rumen contents of cows. *Journal of Dairy Science* **38**, 1169.
- Morrow, J. L. and McGlone, J. J. (1987). Preference of piglets for odors from sow feces. *Journal of Animal Science* **65**(Suppl. 1), 231.
 - (1988). Sensory systems and nipple attachment behavior in neonatal pigs. *Journal of Animal Science*. **66**(Suppl. 1), 243.
- Morrow-Tesch, J. and McGlone, J. J. (1990). Sources of maternal odors and the development of odor preferences in baby pigs. *Journal of Animal Science* **68**, 3563–3571.
- Morton, E. S. (1975). Ecological sources of selection on avian sounds. *American Naturalist* **109**, 17–34.
- Morton, J. F. (1970). Tentative correlations of plant usage and esophageal cancer zones. *Economic Botany* **24**, 217–226.
 - (1989). Tannin as a carcinogen in bush-tea: tea, maté, and khat. In *Chemistry and Significance of Condensed Tannins*, ed. R. W. Hemingway and J. J. Karchesy, pp. 403–416. New York: Plenum.

- Most, K. and Brückner, G. H. (1936). Über Voraussetzungen und den derzeitigen Stand der Nasenleistungen von Hunden. Zeitschrift für Hundeforschung 12, 9–30.
- Mott, M. (2004). Bees, giant African rats used to sniff land mines. *National Geographic News* 10 February 2004.
- Mucignat-Caretta, C., Caretta, A., and Cavaggioni, A. (1995). Acceleration of puberty onset in female mice by male urinary proteins. *Journal of Physiology* **486**, 517–522.
- Mucignat-Caretta, C., Cavaggioni, A., and Caretta, A. (2004). Male urinary chemosignals differentially affect aggressive behavior in male mice. *Journal of Chemical Ecology* **30**, 777–791.
- Müller-Schwarze, D. (1971). Pheromones in black-tailed deer (*Odocoileus heminonus columbianus*). *Animal Behaviour* **19**, 141–152.
 - (1972). Responses of young black-tailed deer to predator odors. *Journal of Mammalogy* **53**, 393–394.
 - (1979). Flehmen in the context of mammalian urine communication. In *Chemical Ecology: Odour Communication in Animals*, ed. F. J. Ritter, pp. 85–96. Amsterdam: Elsevier.
 - (1983). Experimental modulation of behavior of free-ranging mammals by semiochemicals. In *Chemical Signals in Vertebrates*, vol. 3, ed. D. Müller-Schwarze, and R. M. Silverstein, pp. 235–244. New York: Plenum.
 - (1987). Evolution of cervid olfactory communication. In Research Symposia of the National Zoological Park: Biology and Management of Cervidae, ed. C. M. Wemmer, pp. 331–344. Washington DC: Smithsonian Institution Press.
 - (1990). Leading them by their noses: animal and plant odours for managing vertebrates. In *Chemical Signals in Vertebrates*, vol. 5, ed. D. W. Macdonald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 585–598. Oxford: Oxford University Press.
 - (1992). Castoreum of beaver (*Castor canadensis*): function, chemistry and biological activity of its components. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty, and D. Müller-Schwarze, pp. 457–464. New York: Plenum.
 - (1998). Signal specialization and evolution in mammals. In *Chemical Signals in Vertebrates*, vol. 8, ed. R. E. Johnston, D. Müller-Schwarze, and P. W. Sorensen, pp. 1–14. New York: Plenum.
 - (2005). Thirty years on the odor trail: from the First to the Tenth International Symposium on Chemical Signals in Vertebrates. In Chemical Signals in Vertebrates, vol. 10, ed. R. T. Mason, M. P. LeMaster, & D. Müller-Schwarze, pp. 1–6. New York: Springer.
- Müller-Schwarze, D. and Giner, J. (2005). Cottontails and gopherweed: antifeedants from a spurge. In *Chemical Signals in Vertebrates*, vol. 10, ed. R. T. Mason, M. P. LeMaster, and D. Müller-Schwarze, pp. 409–412. New York: Springer.
- Müller-Schwarze, D. and Heckman, S. (1980). The social role of scent marking in beaver (*Castor canadensis*). *Journal of Chemical Ecology* **6**, 81–95.
- Müller-Schwarze, D. and Houlihan, D. (1991). Pheromonal activity of single castoreum constituents in beaver, *Castor canadensis*. *Journal of Chemical Ecology* **17**, 715–734.
- Müller-Schwarze, D. and Müller-Schwarze, C. (1971). Olfactory imprinting in a precocial mammal. *Nature* **229**, 55–56.

- (1973). Differential predation of South Polar skuas in an Adiplic penguin rookery. *Condor* **75**, 127–131.
- (1977). Interactions between South Polar skuas and Adie penguins. In *Adaptations within Antarctic Ecosystems*, ed. G. A. Llano, pp. 619-646. Washington, DC: Smithsonian Institution.
- (1985). Behavioral ecology of Labrador caribou. *National Geographic Society Research Reports* **21**, 321–326.
- Müller-Schwarze, D., Müller-Schwarze, C., and Franklin., W. F. (1972). Factors influencing scent marking in pronghorn (*Antilocapra americana*). Verhandlungen der Deutschen Zoologischen Gesellschaft **66**, 146–150.
- Müller-Schwarze, D., Müller-Schwarze, C., Singer, A. G., and Silverstein, R. M. (1974). Mammalian pheromone: identification of active compound in the subauricular scent of the male pronghorn. *Science* **183**: 860–862.
- Müller-Schwarze, D., Silverstein, R. M., Müller-Schwarze, C., Singer, A. G., and Volkman, N. J. (1976). Response to a mammalian pheromone and its geometric isomer. *Journal of Chemical Ecology* **2**, 389–398.
- Müller-Schwarze, D., Quay, W. B. & Brundin, A. (1977). The caudal gland of *Rangifer*: its behavioral role, histology and chemistry. *Journal of Chemical Ecology* 3, 591–601.
- Müller-Schwarze, D., Volkman, N. J., and Zemanek, K. (1977). Osmetrichia: specialized scent hairs in black-tailed deer. *Journal of Ultrastructure Research* **59**, 223–230.
- Müller-Schwarze, D., Ravid, U., Claesson, A., *et al.* (1978a). The "deer lactone": source, chiral properties, and responses by black-tailed deer. *Journal of Chemical Ecology* **4**, 247–256.
- Müller-Schwarze, D., Källquist, L., Mossing, T., Brundin, A., and Andersson, G. (1978b). Response of reindeer to interdigital secretion of conspecifics. *Journal of Chemical Ecology* **4**, 325–335.
- Müller-Schwarze, D., Altieri, R., and Porter, N. (1984). Alert odor from skin gland in deer. *Journal of Chemical Ecology* **10**, 1707–1729.
- Müller-Schwarze, D., Brashear, H., Kinnel, R., Hintz, K. A., and Skibo, C. (2001). Food processing by animals: do beavers leach tree bark to improve palatability? *Journal of Chemical Ecology* 27, 1011–1028.
- Muñoz, A. 2004. Chemo-orientation using conspecific chemical cues in the stripe-necked terrapin (*Mauremys leprosa*). *Journal of Chemical Ecology* **30**, 519–530.
- Murata, M., Miyagawa-Kohshima, K., Nakanishi, K., and Naya, Y. (1986). Characterization of compounds that induce symbiosis between sea anemone and anemone fish. *Science* **234**, 585–587.
- Murphy, B. P., Miller, K. V., and Marchinton, R. L. (1994). Sources of reproductive chemosignals in female white-tailed deer. *Journal of Mammalogy* **75**, 781–786.
- Murray, A. H., Iason, G. R., and Stewart, C. (1996). Effect of simple phenolic compounds of heather (*Calluna vulgaris*) on rumen microbial activity *in vitro*. *Journal of Chemical Ecology* 22, 1493–1504.
- Mykytowycz, R. (1965). Further observations on the territorial function and histology of the submandibular cutaneous (chin) glands in the rabbit, *Oryctolagus cuniculus* (L.). *Animal Behaviour* **13**, 400–412.

- Mykytowycz, R. (1968). Territorial marking by rabbits. Scientific American 218,116-126.
- Mykytowycz, R. and Ward, M. M. (1971). Some reactions of nestlings of the wild rabbit, *Oryctolagus caniculus* (L.), when exposed to natural rabbit odours. *Forma et Functio* 4, 137–148.
- Nathan, J. (1998). Balzac territory. New York Times, 7 June 1998, Section 2, p. 14.
- Nathanson, J. A. (1984). Caffeine and related methylxanthines: possible naturally occurring pesticides. *Science* **226**, 184–187.
- Natoli, E. (1985). Behavioral responses of urban feral cats to different types of urine marks. *Behaviour* **94**, 234–243.
- Natynczuk, S. E. and Macdonald, D. W. (1992). Scent communication in the rat. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 537–542. New York: Plenum.
- Negus, N. C., Berger, P. J., and Brown, B. W. (1986). Microtine population dynamics in a predictable environment. *Canadian Journal of Zoology* **64**, 785–792.
- Nelson, C. J., Seiber, J. N., and Brower, L. P. (1981). Seasonal and intraplant variation of cardenolide content in the California milkweed, *Asclepias eriocarpa*, and implications for plant defense. *Journal of Chemical Ecology* 7, 981–1010.
- Nelson, K. (1964). Behavior and morphology in the glandulocaudine fishes (Ostariophysi, Characidae). *University of California Publications in Zoology* **75**, 59–152.
- Nelson, R. J. and Shiber, J. R. (1990). Photoperiod affects reproductive responsiveness to 6-methoxy-2-benzoxazolinone in house mice. *Biology of Reproduction* **43**, 586–591.
- Neuhaus, W. (1953). Über die Riechschärfe des Hundes für Fettsäuren. Zeitschrift für vergleichende Physiologie **35**, 527–552.
 - (1955). Die Unterscheidung von Duftquantitäten bei Mensch und Hund nach Versuchen mit Buttersäure. Zeitschrift für vergleichende Physiologie 37, 234–252.
 - (1956a). Die Riechschwelle von Duftgemischen beim Hund und ihr Verhältnis zu den Schwellen unvermischter Duftstoffe. Zeitschrift für vergleichende Physiologie 38, 238–258.
 - (1956b). Die Unterscheidungsfähigkeit des Hundes für Duftgemische. Zeitschrift für vergleichende Physiologie **39**, 25–43.
- Neuhaus, W., and Müller, A. (1954). Das Verhältnis der Riechzellenzahl zur Riechschwelle beim Hund. *Naturwissenschaften* **41**, 237.
- Nevitt, G. A. (1991). Peripheral mechanisms of olfaction in two teleostean fishes. Ph.D. Thesis, DA 908505.
 - (1995). Antarctic procellariiform seabirds can smell krill. *Antarctic Journal of the USA* **29**, 168–169.
 - (1999). Foraging by seabirds on an olfactory landscape. American Scientist 87, 46-53.
- Nevitt, G. A., Viet, R. R., and Kareiva, P. (1995). Dimethyl sulphide as a foraging cue for Antarctic procellariiform seabirds. *Nature* **376**, 680–682.
- Nevo, E., Bodmer, M., and Heth, G. (1976). Olfactory discrimination as an isolating mechanism in speciating mole rats. *Experientia* **32**, 1511–1512.
- Newman, K. S. and Halpin, Z. T. (1988). Individual odours and mate recognition in the prairie vole, *Microtus ochrogaster*. *Animal Behaviour* **36**, 1779–1787.

- Newton, P. N. and Nishida, T. (1990). Possible buccal administration of herbal drugs by wild chimpanzees, *Pan troglodytes*. *Animal Behaviour* **39**, 798–801.
- Nicolaus, L. K. (1987). Conditioned aversions in a guild of egg predators: implications for aposematism and prey defense mimicry. *American Midland Naturalist* **117**: 405–419.
- Nicolaus, L. K. and Nellis, D. W. (1987). The first evaluation of the use of conditioned taste aversion to control predation by mongooses upon eggs. *Applied Animal Behaviour Science* 17, 329–346.
- Nicoletto, P. F. (1985). The relative roles of vision and olfaction in prey discrimination by the ground skink, *Scincella lateralis*. *Journal of Herpetology* **19**, 411–415.
- Ninomiya, K. and Kimura, T. (1986). Preference of female mice for odors from urine and preputial gland secretion of males. *Zoological Science* **3**, 1103.
 - (1988). Male odors that influence the preference of female mice: roles of urinary and preputial factors. *Physiology and Behavior* **44**, 791–796.
- Nishida, R. and Fukami, H. (1989). Ecological adaptation of an Aristolochiaceae-feeding swallowtail butterfly, *Atrophaneura alcinous*, to aristolochic acids. *Journal of Chemical Ecology* **15**, 2549–2564.
- Nitao, J. K. (1988). Artificial defloration and furanocoumarin induction in *Pastinaca sativa* (Umbelliferae). *Journal of Chemical Ecology* **14**, 1515.
- Nixon, A., Mallet, A. I., and Gomer, D. B. (1988). Simultaneous quantification of five odorous steroids, 16-androstenes, in the axillary hair of men. *Journal of Steroid Biochemistry* **29**, 505–510.
- Njemelä, P. and Danell, K. (1988). Comparison of moose browsing on Scots pine (*Pinus sylvestris*) and lodgepole pine (*P. contorta*). *Journal of Applied Ecology* **25**, 761–775.
- Nolte, D. L., Mason, J. R., and Clark, L. (1993). Avoidance of bird repellents by mice (*Mus musculus*). *Journal of Chemical Ecology* **19**, 427–432.
- Nolte, D. L., Mason, J. R., and Lewis, S. L. (1994a). Tolerance of bitter compounds by an herbivore, *Cavia porcellus*. *Journal of Chemical Ecology* **20**, 303–308.
- Nolte, D. L., Mason, J. R., Epple, G., Aronov, E., and Campbell, D. L. (1994b). Why are predator urines aversive to prey? *Journal of Chemical Ecology* **20**, 1505–1516.
- Nordeng, H. (1971). Is the local orientation of anadromous fishes determined by pheromones? *Nature* **233**, 411–413.
 - (1977). A pheromone hypothesis for homeward migration in anadromous salmonids. *Oikos* **28**, 155–159.
- Nordstrom, K. M., Belcher, A. M., Epple, G., et al. (1989). Skin surface microflora of the saddle-back tamarin monkey, *Saguinus fuscicollis*. *Journal of Chemical Ecology* **15**, 629–639.
- Novotny, M., Jemiolo, B., Harvey, S., Wiesler, D., and Marchlewska-Koj, A. (1986a). Adrenal-mediated endogenous metabolites inhibit puberty in female mice. *Science* **231**, 722–725.
- Novotny, M., Harvey, S., Jemiolo, B., and Alberts, A. (1986b). Synthetic pheromones that promote inter-male aggression in mice. *Proceedings of the National Academy of Sciences, USA* **82**, 2059–2061.

- Novotny, M. V., Ma, W., Wiesler, D., and Zidek, L. (1999). Positive identification of the puberty-accelerating pheromone of the house mouse: the volatile ligands associating with the major urinary protein. *Proceedings of the Royal Society of London Series B* **266**, 2017–2022.
- Nyby, J., Whitney, G., Schmitz, S., and Dizinno, G. (1978). Post pubertal experience establishes signal value of mammalian sex odor. *Behavioral Biology* **22**, 545–552.
- Nyström, P. and Abjörnsson, K. (2000). Effects of fish chemical cues on the interactions between tadpoles and crayfish. *Oikos* **88**, 181–190.
- O'Connell, R. J., Singer, A. G., Pfaffmann, C., and Agosta, W. C. (1979). Pheromones of hamster vaginal discharge: attraction to femtogram amounts of dimethyl disulfide and to mixtures of volatile compounds. *Journal of Chemical Ecology* **5**, 575–585.
- Ödberg, F. O. and Francis-Smith, K. (1977). Studies on the formation of ungrazed eliminative areas in fields used by horses. *Applied Animal Ethology* **3**, 27–34.
- O'Donnell, R. P., Ford, N. B., Shine, R., and Mason, R. T. (2004). Male red-sided garter snakes, *Thamnophis sirtalis parietalis*, determine female mating status from pheromone trails. *Animal Behaviour* **68**, 677–683.
- Ogurtsov, S. V. and Bastakov, V. A. (2001). Imprinting on native pond odour in the pool frog, *Rana lessonae* CAM. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 433–438. New York: Kluwer Academic/Plenum.
- Oh, H.K., Sakai, T., Jones, M. B., and Longhurst, W. M. (1967). Effect of various essential oils isolated from Douglas fir needles upon sheep and deer rumen microbial activity. *Applied Microbiology* **15**, 777–784.
- O'Hara, R. K. and Blaustein, A. R. (1985). *Rana cascadae* tadpoles aggregate with siblings: an experimental field study. *Oecologia* **67**, 44–51.
- Ohigashi, H., Huffman, M. A., Izutsu, D., *et al.* (1994). Toward the chemical ecology of medicinal plant use in chimpanzees: the case of *Vernonia amygdalina*, a plant used by wild chimpanzees possibly for parasite-related diseases. *Journal of Chemical Ecology* **20**, 541–553.
- Oli, M. K. and Dobson, F. S. (1999). Population cycles in small mammals: the role of age at sexual maturity. *Oikos* **86**, 557–565.
- Olivier, R. C. D. and Laurie, W. A. (1974). Habitat utilization by hippopotamus in the Mara River. *East African Wildlife* **12**, 249–271.
- Olsén, K. H. (1987). Chemoattraction of juvenile Arctic charr (*Salvelinus alpinus* L.) to water scented by conspecific intestinal content and urine. *Comparative Biochemistry and Physiology* 87, 641–644.
 - (1990). Further studies concerning chemoattraction among fry of Arctic charr (*Salvelinus alpinus* L.) to water conditioned by conspecifics. *Journal of Chemical Ecology* **16**, 2084–2090.
- Olsén, K. H. and Winberg, S. (1996). Learning and sibling odor preference in juvenile Artic char, *Salvelinus alpinus* (L.). *Journal of Chemical Ecology* **22**, 773–786.
- Olsén, K. H., Karlsson, L., and Helandes, A. (1986). Food search behavior in arctic charr, *Salvelinus alpinus* (L.) induced by food extracts and amino acids. *Journal of Chemical Ecology* 12, 1987–1998.

- Olsén, K. H., Bjerselius, R., Petersson, E., *et al.* (2000). Lack of species-specific primer effects of odours from female Atlantic salmon (*Salmo salar L.*) and brown trout (*Salmo trutta L.*). *Oikos* **88**, 213–220.
- Olsén, K. A., Johansson, A.-K., Bjerselius, R., Mayer, I., and Kindhal, H. (2002a). Mature Atlantic salmon (*Salmo salar L.*) male parr are attracted to ovulated female urine but not to ovarian fluid. *Journal of Chemical Ecology* **28**, 29–40.
- Olsén, K. H., Grahn, M., and Lohm, J. (2002b). Influence of MHC on sibling discrimination in arctic char, *Salvelinus alpinus* (L.). *Journal of Chemical Ecology* **28**, 783–795.
- Ophir, D., Guterman, A., and Gross-Isseroff, R. (1988). Changes in smell acuity induced by radiation exposure of the olfactory mucosa. *Archives of Otolaryngology and Head and Neck Surgery* **114**, 853–855.
- Or, K. and Ward, D. (2003). Three-way interactions between *Acacia*, large mammalian herbivores and bruchid beetles: a review. *African Journal of Ecology* **41**, 257–265.
- Ortmann, R. (1956). Über die Musterbildung von Duftdrüsen in der Sohlenhaut der weissen Hausmaus (Mus musculus albinus). Zeitschrift für Säugetierkunde 21, 138–141.
- Ouellet, J. P. and Ferron, J. (1988). Scent-marking behavior by woodchucks (*Marmota monax*). *Journal of Mammalogy* **69**, 365–368.
- Ovaska, K. (1988). Recognition of conspecific odors by the western red-backed salamander, *Plethodon vehiculum*. *Canadian Journal of Zoology* **66**, 1293–1296.
- Over, R., Cohen-Tannoudji, J., Dehnhard, M., Claus, R., and Signoret, J. P. (1990). Effect of pheromones from male goats on LH secretion in anoestrous ewes. *Physiology and Behavior* **48**, 665–668.
- Owadally, A. W. (1979). The dodo and the tambalacoque tree. Science 203,1363-1364.
- Owaga, M. L. A., Hassanali, A., and McDowell, P. G. (1988). The role of 4-cresol and 3-n-propylphenol in the attraction of tsetse flies to buffalo urine. *Insect Science and its Application* **9**, 95–100.
- Owaga, M. L. A. (1984). Preliminary observations on the efficacy of olfactory attractants derived from wild hosts of tsetse. *Insect Science and its Application* **5**, 87–90.
- Owre, O. T. and Northington, P. O. (1961). Indication of the sense of smell in the turkey vulture, *Cathartes aura* (Linnaeus) from feeding tests. *American Midland Naturalist* **66**, 200.
- Page, J. E., Balza, F., and Nishida, T. (1992). Biologically active diterpenes from *Aspilia mossambicensis*, a chimpanzee mecidinal plant. *Phytochemistry* **31**, 3437–3439.
- Pages, E. (1972). Comportement interindividual des Pangolins arboricoles du Gabon. *Biologia Gabonica* 1, 1–120.
- Palen, G. F. and Goddard, G. V. (1966). Catnip and estrus behavior in the cat. *Animal Behaviour* **14**, 372–377.
- Palo, R. T. (1984). Distribution of birch (*Betula* spp.), willow (*Salix* spp.) and poplar (*Populus* spp.) secondary metabolites and their potential role as chemical defense against herbivores. *Journal of Chemical Ecology* **10**, 499–520.
- Palo, R. T., Pehrson, A., and Knutsson, D. G. (1983). Can birch phenolics be of importance in the defense against browsing vertebrates? *Finnish Game Research* **41**, 75–80.

- Palo, R. T., Sunnerheim, K., and Theander, O. (1985). Seasonal variation of phenols, crude protein and cell wall content of birch (*Betula pendula* Roth.) in relation to ruminant in vitro digestibility. *Oecologia* **65**, 314–318.
- Pandey, S. D., and Pandey, S. C. (1986). Estrus suppression in wild mice: source of pheromonal cue. *Acta Physiologica Hungarica* **67**, 387–392.
- Papi, F. (1976). The olfactory navigation system of the pigeon. *Verhandlungen der Deutschen Zoologischen Gesellschaft* **69**, 184–205.
 - (1990). Olfactory navigation in birds. Experientia 46, 352-363.
- Papi, F. and Ioalè, P. (1988). Pigeon navigation: New experiments on interaction between olfactory and magnetic cues. *Comparative Biochemistry and Physiology*. *A, Comparative Physiology* **91**, 87–90.
- Papi, F., Fiore, L., Viaschi, V., and Benvenuti, S. (1972). Olfaction and homing in pigeons. *Monitore Zoologico Italiano* **6**, 85–95.
- Papi, F., Fiore, L., Fiaschi, V., and Benvenuti, S. (1973). An experiment for testing the hypothesis of olfactory navigation of homing pigeons. *Journal of Comparative Physiology* **83**, 93–102.
- Papi, F., Keeton, W. T., Brown, A. I., and Benvenuti, S. (1978). Do American and Italian pigeons rely on different homing mechanisms? *Journal of Comparative Physiology* **128**, 303–317.
- Paquet, P. C. and Fuller, W. A. (1990). Scent marking and territoriality in wolves of Riding Mountain National Park. In *Chemical Signals in Vertebrates*, vol. 5, ed. D. W. Macdonald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 394–400. Oxford: Oxford University Press.
- Park, D. and Propper, C. R. (2001). Repellent function of male pheromones in the redspotted newt. *Journal of Experimental Zoology* **289**, 404–408.
- Park, D., Hempleman, S. C., and Propper, C. R. (2001). Endosulfan exposure disrupts pheromonal systems in the red-spotted newt: a mechanism for subtle effects of environmental chemicals. *Environmental Health Perspectives* **109**, 669–673.
- Parker, G. H. (1911). The olfactory reactions of the common killifish, *Fundulus heteroclitus* (Linn.). *Journal of Experimental Zoology* **10**, 1–5.
- Parkes, A. S. and Bruce, H. M. (1961). Olfactory stimuli in mammalian reproduction. *Science* **134**, 1049–1054.
- Parsons, L. M. and Terman, C. R. (1978). Influence of vision and olfaction on the homing ability of the white-footed mouse (*Peromyscus leucopus noveboracensis*). *Journal of Mammalogy* **59**, 761–771.
- Paul, V. J. and Pennings, S. C. (1991). Diet-derived chemical defenses in the sea hare Stylocheirus longicauda (Quoy et Gaimard 1824). Journal of Experimental Marine Biology and Ecology 151, 227–243.
- Pawlik, J. R. (1993). Marine invertebrate chemical defenses. *Chemical Reviews* **93**, 1911–1922.
- Pawlik, J. R., McFall, G., and Zea, S. (2002). Does the odor from sponges of the genus *Ircinia* protect them from fish predators? *Journal of Chemical Ecology* **28**: 1103–1115.

- Pawson, M. G. (1977). Analysis of a natural chemical attractant for whiting *Merlangus merlangus* L. and cod *Gadus morhua* L. using a behavioral bioassay. *Comparative Biochemical Physiology* **56A**, 129–135.
- Payne, R. (2001). The tragedy of the live fish trade: Part 1. www.pbs.org/odyssey/voice/20010606.
- Pearce, D. T. and Oldham, C. M. (1988). Ovulation in the merino ewe in the breeding and anoestrous season. *Australian Journal of Biological Science* **41**, 23–26.
- Pearce, G. T. and Paterson, A. M. (1992). Physical contact with the boar is required for maximum stimulation of puberty in the gilt because it allows transfer of boar pheromone and not because it induces cortisol release. *Animal Reproduction Science* 27, 209–224.
- Pearl, I. A. and Darling, S. F. (1968). Studies on the hot water extractives of *Populus balsamifera* bark. *Phytochemistry* **7**, 1851–1853.
- Peckarsky, B. L. and Penton, M. A. (1988). Why do *Ephemerella* nymphs scorpion posture: a "ghost of predation past"? *Oikos* **53**, 185–193.
- Pedersen, P. E. and Blass, E. M. (1982). Prenatal and postnatal determinants of the first suckling episode in albino rats. *Developmental Psychobiology* **15**, 349–355.
- Pedersen, P. E., Stewart, W. B., Greer, C. A., and Shepherd, G. M. (1983). Evidence for olfactory function in utero: VNO probably monitors intrauterine environment. *Science* 221, 478–480.
- Pedersen, R. E. and Benson, T. E. (1986). Projection of septal organ receptor neurons to the main olfactory bulb in rats. *Journal of Comparative Neurology* **252**, 555–562.
- Peinetti, R., Pereyra, M., Kin, A., and Sosa, A. (1993). Effects of cattle ingestion on viability and germination rate of caldén (*Prosopis caldenia*) seeds. *Journal of Range Management* 46, 483–486.
- Pelletier, S. W. (1983). The nature and definition of an alkaloid. *Alkaloids, Chemical and Biological Perspectives* 1, 1–31.
- Pennycuick, P. R. and Cowan, R. (1990). Odour and food preferences of house mice, *Mus musculus*. *Australian Journal of Zoology* **38**, 241–247.
- Perret, M. and Schilling, A. (1987). Intermale sexual effect elicited by volatile urinary ether extract in *Microcebus murinus* (Prosimian, Primates). *Journal of Chemical Ecology* **13**, 495–507.
- Perrigo, G. and Bronson, F. H. (1983). Communication disparities between genetically diverging populations of deer mice. In *Chemical Signals in Vertebrates*, vol. 3, ed. D. Müller-Schwarze, and R. M. Silverstein, pp. 195–210. New York: Plenum.
- Perry, J. N., and Wall, C. (1984). Orientation of male pea moth, *Cydia nigricana*, to pheromone traps in a wheat crop. *Entomologia Experimentalis et Applicata* 37, 161–167.
- Pesaro, M., Balsamo, M., Gandolfi, G., and Tongiorgi, P. (1981). Discrimination among different kinds of water in juvenile eels, *Anguilla anguilla* (L.). *Monitore Zoologico Italiano* **15**, 183–191.
- Pesce, C. (1990). Stanching stench: Florida City a leader. USA Today, March 19, 1990.
- Peters, R. P. and Mech, L. D. (1975). Scent marking in wolves. *American Scientist* **63**, 628–637.

- Petit, C., Hossaert-McKey, M., Perret, P., Blondel, J., and Lambrechts, M. M. (2002). Blue tits use selected plants and olfaction to maintain an aromatic environment for nestlings. *Ecology Letters* **5**, 585.
- Petranka, J. W., Kats, L. B., and Sih, A. (1987). Predator–prey interactions among fish and larval amphibians: use of chemical cues to detect predatory fish. *Animal Behaviour* **35**, 420–425.
- Pettersson, L. B., Nilsson, P. A., and Bronmark, C. (2000). Predator recognition and defence strategies in crucian carp, *Carassius carassius*. *Oikos* 88, 200–212.
- Pfeiffer, W. (1978). Heterocyclic compounds as releasers of the fright reaction in the giant danio *Danio malabaricus* (Jerdon) (Cyprinidae, Ostariophysi, Pisces). *Journal of Chemical Ecology* **4**, 665–673.
- Pfeiffer, W. and Lamour, D. (1976). Die Wirkung von Schreckstoff auf die Herzfrequenz von *Phoxinus* (L.) (Cyprinidae, Ostariophysi, Pisces). *Revue Suisse Zoologique* 83, 861–873.
- Pfeiffer, W. and Lemke, J. (1973). Untersuchungen zur Isolierung und Identifizierung des Schreckstoffes aus der Haut der Elritze, *Phoxinus phoxinus* (L.) (Cyprinidae, Ostariophysi, Pisces). *Journal of Comparative Physiology* **82**, 407–410.
- Pfennig, D. W. (1997). Kinship and cannibalism. BioScience 47, 662-675.
- Pfister, J. A., Provenza, F. D., and Manners, G. D. (1990a). Ingestion of tall larkspur by cattle: separating effects of flavor from post-ingestive consequences. *Journal of Chemical Ecology* **16**, 1697–1705.
- Pfister, J. A., Müller-Schwarze, D., and Balph, D. F. (1990b). Effects of predator fecal odors on feed selection by sheep and cattle. *Journal of Chemical Ecology* **16**, 573–583.
- Pfister, J. A., Manners, G. D., Gardner, D. R., Price, K. W., and Ralphs, M. H. (1996). Influence of alkaloid concentration on acceptability of tall larkspur (*Delphinium spp.*) to cattle and sheep. *Journal of Chemical Ecology* **22**, 1147–1168.
- Pfister, J. A., Provenza, F. D., Manners, G. D., and Ralphs, M. H. (1997). Tall larkspur ingestion: can cattle regulate intake below toxic levels? *Journal of Chemical Ecology* **23**, 759–777.
- Phillips, J. A. and Alberts, A. C. (1992). Naive ophiophagus lizards recognize and avoid venomous snakes using chemical cues. *Journal of Chemical Ecology* **18**, 1775–1783.
- Pickrell, J. (2003). Cyanide on the side. http://sciencenow.sciencemag.org. 5/30/2003.
- Pigozzi, G. (1990). Latrine use and the function of territoriality in the European badger, *Meles meles*, in a Mediterranean coastal habitat. *Animal Behaviour* **39**, 1000–1002.
- Placyk, J. S. and Graves, B. M. (2002). Prey detection by vomeronasal chemoreception in a plethodontid salamander. *Journal of Chemical Ecology* **28**, 1017–1036.
- Poddar-Sarkar, M. (1996). The fixative lipid of tiger pheromone. *Journal of Lipid Mediators* and Cell Signalling **15**, 89–101.
- Poddar-Sarkar, M., Brahmachary, R. L., and Dutta, J. (1991). Short-chain fatty acids as putative pheromones in the marking fluid of the tiger. Indian Chemical Society 68, 255–256.

- Poduschka, W. and Firbas, W. (1968). Das Selbstbespeicheln des Igels, Erinaceus europaeus, Linne 1758, steht in Beziehung zur Funktion des Jacobson'schen Organes. Zeitschrift für Säugetierkunde 33, 160–172.
- Poindron, P., Levy, F., and Krehbiel, D. (1988). Genital, olfactory, and endocrine interactions in the development of maternal behavior in the parturient ewe. *Psychoneuroendocrinology* **3**, 99–125.
- Polak, E. H. and Provasi, J. (1992). Odor sensitivity to geosmin enantiomers. *Chemical Senses* 17, 23–26.
- Polkinghorne, C., Olson, J. M., Gallaher, D. G., and Sorensen, P. W. (2001). Larval sea lamprey release two unique bile acids to the water of a rate sufficient to produce detectable riverine pheromone plumes. *Fish Physiology and Biochemistry* **24**, 15–30.
- Pontet, A. and Schenk, F. (1988). Effects of conspecific odors on the activity and the ultrasonic vocalizations of the woodmouse, *Apodemus sylvaticus*, in a plus-maze during ontogeny. *Sciences et Techniques de L' Animal de Laboratoire* **13**, 105–109.
- Porter, R. H. (1986). Chemical signals and kin recognition in spiny mice (*Acomys cahirinus*). In *Chemical Signals in Vertebrates*. vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein, pp. 397–411. New York: Plenum.
- Porter, R. H. and Moore, J. D. (1981). Human kin recognition by olfactory cues. *Physiology and Behavior* **27**, 493–495.
- Porter, R. H., Cernoch, J. M., and McLaughlin, F. J. (1983). Maternal recognition of neonates through olfactory cues. *Physiology and Behavior* **30**, 151–154.
- Porter, R. H., Balogh, R. D., Cernoch, J. F., and Franchi, C. (1986). Recognition of kin through characteristic body odors. *Chemical Senses* 11, 389–395.
- Porter, R. H., McFadyen-Ketchum, S. A., and King, G. A. (1989). Underlying bases of recognition signatures in spiny mice, *Acomys cahirinus*. *Animal Behaviour* 37, 638–644.
- Porter, R. H., Makin, J. W., Davis, L. B., and Christensen, K. M. (1992). Breast-fed infants respond to olfactory cues from their own mother and unfamiliar lactating females. *Infant Behavior and Development* **15**, 85–93.
- Powers, J. B. and Winans, S. S. (1975). Vomeronasal organ: critical role in mediating sexual behavior of the male hamster. *Science* **187**, 961–963.
- Powers, J. B., Fields, R. B., and Winans, S. S. (1979). Olfactory and vomeronasal system participation in male hamsters' attraction to female vaginal secretion. *Physiology and Behavior* **22**, 77–84.
- Prakash, J. and Idris, M. (1982). Scent marking by the female of the Indian gerbil, *Tatera indica*, from two distinct desert habitats during oestrus. *Indian Journal of Experimental Biology* **20**, 915–916.
- Prasad, N. L. N. S. (1989). Territoriality in Indian Blackbuck antelope, *Antilope cervicapra* (Linnaeus). *Bombay Natural History Society Journal* **86**, 187–193.
- Preti G., Muetterties, E. L. Furman, J. M., Kennelly, J. J., and Johns, B. E. (1976). Volatile constituents of dog (*Canis familiaris*) and coyote (*Canis latrans*) anal sacs. *Journal of Chemical Ecology* **2**, 177–186.

- Preti, G., Cutler, W. B., Garcia, C. R., Huggins, G. R., and Lawley, H. J. (1986). Human axillary secretions influence women's menstrual cycles: the role of donor extract from women. *Hormones and Behavior* **20**, 474–482.
- Preti, G., Cutler, W. B., Christensen, C. M., *et al.* (1987). Human axillary extracts: analysis of compounds from samples which influence menstrual timing. *Journal of Chemical Ecology* **13**, 717–731.
- Preti, G., Zeng, X. N., Leyden, J. J., McGinley, K. J., and Spielman, A. I. (1992). Human axillary odors and their precursors. *Chemical Senses* 17, 685–686.
- Price, E. O., Dunn, G. C., Talbor, J. A., and Dally, M. R. (1984). Fostering lambs by odor transfer: the substitution experiment. *Journal of Animal Science* **59**, 301–307.
- Primor, N., Parness, J., and Zlotkin, E. (1978). Paradaxin: the toxic factor from the skin secretion of the flatfish *Paradachirus marmoratus* (Soleidae). In *Toxins: Animals, Plants and Microbial*, ed. P. Rosenberg. Oxford: Pergamon Press.
- Probst, B. and Lorenz, M. (1987). Increased scent marking in male Mongolian gerbils by urinary polypeptides of female conspecifics. *Journal of Chemical Ecology* **13**, 851–862.
- Provenza, F. D. (1995). Postingestive feedback as an elementary determinant of food preference and intake in ruminants. *Journal of Range Management* **48**, 2–17.
 - (2004). Linking herbivore experience, varied diets, and plant biochemical diversity. In *Proceedings of the Annual Meeting of International Society of Chemical Ecology*, July 2004, Ottawa, Canada.
- Provenza, F. D. and Lynch, J. J. (1994). How goats learn to distinguish between novel foods that differ in postingestive consequences. *Journal of Chemical Ecology* **20**, 609–624.
- Provenza, F. D., Kimball, B. A., and Villaba, J. J. (2000). Roles of odor, taste, and toxicity in the food preferences of lambs: implications for mimicry in plants. *Oikos* **88**, 424–432.
- $Prudente, A. (1963). \ Patologia geographica do cancer no Brasil. \textit{Boletin de Oncologia} \textbf{46}, 281.$
- Quadagno, D. M., Shubeita, H. E., Deck, J., and Francoeur, D. (1981). Influence of male social contacts, exercise, and all-female living conditions on the menstrual cycle. *Psychoneuroendocrinology* **6**, 239–244.
- Quay, W.B. (1970). Histology of the para-anal sebaceous glandular organs of the bat *Eonycteris spelaea* (Chiroptera: Pteropidae). *Anatomical Record* **166**, 189–197.
 - (1977). Structure and function of skin glands. In *Chemical Signals in Vertebrates*, vol. 1, ed. D. Müller-Schwarze, and M. M. Mozell, pp. 1–16. New York: Plenum.
- Quay, W. B. and Müller-Schwarze, D. (1970) Functional histology of integumentary glandular regions in black-tailed deer (Odocoileus heminonus columbianus). Journal of Mammalogy **51**, 675–694.
- Quinn, T. P. (1980). Locomotor responses of juvenile blind cave fish, *Astyanax jordani*, to the odors of conspecifics. *Behavioral and Neural Biology* **29**, 123–127.
- Quinn, T. P. and Busack, C. A. (1985). Chemosensory recognition of sibling in juvenile coho salmon (*Oncorhynchus kisutch*). *Animal Behaviour* **33**, 51–56.
- Quinn, V. S. and Graves, B. M. (1998). Home pond discrimination using chemical cues in *Chrysema picta*. *Journal of Herpetology* **32**, 457–461.

- Radwan, M. A. and Crouch, G. L. (1978). Selected chemical constituents and deer browsing preference of Douglas fir. *Journal of Chemical Ecology* **4**, 675–683.
- Rajan, R., Clement, J. P., and Bhalla, U. S. (2006). Rats smell in stereo. *Science* **311**, 666–670.
- Rajendren, G. and Dominic, C. J. (1986). Evaluation of involvement of accessory olfactory (vomeronasal) system in estrous cyclicity and mating in female mice. *Indian Journal of Experimental Biology* **24**, 573–577.
 - (1988). Effect of cyproterone acetate on the pregnancy-blocking ability of male mice and the possible chemical nature of the pheromone. *Journal of Reproduction and Fertility* **84**, 387–392.
- Ralphs, M. H., Olsen, J. D., Pfister, J. A., and Manners, G. D. (1988). Plant–animal interactions in larkspur poisoning in cattle. *Journal of Animal Science* **66**, 2334–2342.
- Randall, J. A. (1986). Preference for estrous female urine by male kangaroo rats (*Dipodomys spectabilis*). *Journal of Mammalogy* **67**, 736–739.
- Rasa, O. A. E. (1973). Marking behavior and its social significance in the African dwarf mongoose *Helogale undulata rufula*. *Zeitschrift für Tierpsychologie* **32**, 293–318.
- Rasmussen, L. E. L. (1988). Chemosensory responses in two species of elephants to constituents of temporal gland secretion and must hurine. *Journal of Chemical Ecology* **14**, 1687–1711.
- Rasmussen, L. E. L. and Greenwood, D. R. (2003). Frontalin: a chemical message of musth in Asian elephants (*Elephas maximus*). *Chemical Senses* **28**: 433–446.
- Rasmussen, L. E. L. and Hultgren, B. (1990). Gross and microscopic anatomy of the vomeronasal organ in the Asian elephant (*Elephas maximus*). In *Chemical Signals in Vertebrates*. vol. 5, eds. D. W. Mac Donald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 154–161. Oxford: Oxford University Press.
- Rasmussen, L. E. L. and Perrin, T. E. (1999). Physiological correlates of musth: lipid metabolites and chemical composition of exudates. *Physiology and Behavior* **67**, 539–549.
- Rasmussen, L. E. L. and Schulte, B. A. (1998). Chemical signals in the reproduction of Asian (*Elephas maximus*) and African (*Loxodonta africana*) elephants. *Animal Reproduction Science* **53**, 19–34.
- Rasmussen, L. E. L., Schmidt, M. J., Henneous, R., Groves, D., and Daves, G. D. (1982). Asian bull elephants: flehmen-like responses to extractable components in female elephant estrous urine. *Science* **217**, 159–162.
- Rasmussen, L. E., Buss, C. O., Hess, D. L., and Schmidt, M. J. (1984). Testosterone and dihydrotestosterone concentrations in elephant serum and temporal gland secretions. *Biology of Reproduction* **30**, 352–362.
- Rasmussen, L. E. L., Lee, T. D., Roelofs, W. L., Zhang, A., and Daves, G. D., Jr. (1996). Insect pheromone in elephants. *Nature* **379**:684.
- Rasmussen, L. E. L., Lee, T. D., Zhang, A., Roelofs, W. L., and Daves, G. D., Jr. (1997). Purification, identification, concentration and bioactivity of (*Z*)-7-dodecen-1-yl acetate: sex pheromone of the female Asian elephant, *Elephas maximus*. *Chemical Senses* **22**, 417–438.

- Raymer, J. D., Wiesler, D., Novotny, M., *et al.* (1984). Volatile constituents of wolf (*Canis lupus*) urine as related to gender and season. *Experientia* **40**, 707–709.
- Raymer, J., Wiesler, D., Novotny, M., *et al.* (1986). Chemical scent constituents in urine of wolf (*Canis lupus*) and their dependence on reproductive hormones. *Journal of Chemical Ecology* **12**, 297–314.
- Reardon, P. O., Leinweber, C. L., and Merrill, J. L. B. (1972). The effect of bovine saliva on grasses. *Journal of Animal Science* **34**, 897–98.
 - (1974). Responses of sideoats grama to animal saliva and thiamine. *Journal of Range Management* 27, 400–401.
- Reasner, D. S. (1987). Spatially selective alteration of the mitral cell layer: a critical review of the literature. *Chemical Senses* **12**, 365–379.
- Reasner, D. S. and Johnston, R. E. (1988). Acceleration of reproductive development in female Djungarian hamsters by adult males. *Chemical Senses* **13**, 729.
- Reed, J. P. (1969). Alarm substances and fright reaction in some fishes from the Southeastern United States. *Transactions of the American Fisheries Society* **98**, 664–668.
- Reger, R. L., Gerall, A. A., Wysocki, C. J., and Carter, C. S. (1987). LHRH neuronal system in the accessory olfactory bulb of the prairie vole, *Microtus ochrogaster*. In *Proceedings of the Annual Meeting of the Society for Neuroscience*, New Orleans.
- Regnier, F. E. and Goodwin, M. (1977). On the chemical and environmental modulation of pheromone release from vertebrate scent marks. In *Chemical Signals in Vertebrates*, vol. 1, ed. D. Müller-Schwarze, and M. M. Mozell, pp. 115–133. New York: Plenum.
- Rehnberg, B. G., and Schreck, C. B. (1986). The olfactory L-serine receptor in coho salmon: biochemical specificity and behavioral responses. *Journal of Comparative Physiology* **159**, 61–67.
- (1987). Chemosensory detection of predators by coho salmon, *Oncorhynchus kisutch*: behavioral reaction and the physiological stress responses. *Canadian Journal of Zoology* **65**, 481–485.
- Rehnberg, B. G., Smith, R. J. F., and Sloley, B. D. (1987). The reaction of pearl dace (Pisces, Cyprinidae) to alarm pheromone: time-course of behavior, brain amines and stress physiology. *Canadian Journal of Zoology* **65**, 2916–2921.
- Reichardt, P. B., Bryant, J. P., Clausen, T. P., and Wieland, G. D. (1985). Defense of winter-dormant Alaska paper birch against snowshoe hares. *Oecologia* **65**, 58–69.
- Reichardt, P. B., Bryant, J. P., Anderson, B. J., *et al.* (1990a). Germacrone defends Labrador tea from browsing by snowshoe hares. *Journal of Chemical Ecology* **16**, 1961–1970.
- Reichardt, P. B., Bryant, J. P., Mattes, B. R., *et al.* (1990b). Winter chemical defense of Alaskan balsam poplar against snowshoe hares. *Journal of Chemical Ecology* **16**, 1941–1959.
- Reidinger, R. F., Jr. and Mason, J. R. (1983). Exploitable characteristics of neophobia and food aversions for improvements in rodent and bird control. In *Proceedings of the 4th Symposium of the American Society for Testing and Materials: Vertebrate Pest Control and Management Materials*, ed. D. E. Kaukeinen, D. E., pp. 20–39, Washington, DC: American Society for Testing and Materials.

- Reis, P. J., Tunks, D. A., and Chapman, R. E. (1975). Effects of mimosine, a potential chemical deflecting agent, on wool growth and the skin of sheep. *Australian Journal of Biological Sciences* 1, 69–84.
- Renwick, J. H., Claring, W. B., Earthy, M. E., *et al.* (1984). Neural-tube defects produced in Syrian hamsters by potato glycoalkaloids. *Teratology* **30**, 371–381.
- Resink, J. W., van den Hurk, R., Peters, R. C., and van Oordt, P. G. W. J. (1987). Steroid glucuronides as sex attracting pheromones in the African catfish, *Clarias gariepinus*, *Proceedings of the 3rd International Symposium on Reproductive Physiology of Fish*, ed. D. R. Idler, L. W. Crim, and J. M. Walsh, p. 163. St. John's, Newfoundland: Memorial University Press.
- Resink, J. W., Schoonen, W. E. G. J., Albers, P. C., *et al.* (1989a). The chemical nature of sex attracting pheromones from the seminal vesicle of the African catfish, *Clarias gariepinus*. *Aquaculture* **83**, 137–151.
- Resink, J. W., Voorthuis, P. K., van den Hurk, R., Peters, R. C., and van Oordt, P. G. W. J. (1989b). Steroid glucuronides of the seminal vesicle as olfactory stimuli in African catfish, *Clarias gariepinus*. *Aquaculture* 83, 153–166.
- Resink, J. W., van den Berg, T. W. M., van den Hurk, R., Huisman, E. A., and van Oordt, P. G. W. J. (1989c). Induction of gonadotropin release and ovulation by pheromones in the African catfish, *Clarias gariepinus*. *Aquaculture* **83**, 167–177.
- Ressler, K. J., Sullivan, S. L., and Buck, L. B. (1993). A zonal organization of odorant receptor gene expression in the olfactory epithelium. *Cell*, **73**, 597–609.
 - (1994). Information coding in the olfactory system: evidence for a stereotyped and highly organized epitope map in the olfactory bulb. *Cell* **79** 1245–1255.
- Reynolds, J. and Keverne, E. B. (1979). The accessory olfactory system and its role in the pheromonally mediated suppression of oestrus in grouped mice. *Journal of Reproduction and Fertility* **57**, 31–35.
- Rhoades, D. F. (1985). Offensive–defensive interactions between herbivores and plants: their relevance in herbivore population dynamics and ecological theory. *American Naturalist* **125**, 205–238.
- Richardson, P. R. K. (1990). Scent marking and territoriality in the aardwolf. In *Chemical Signals in Vertebrates*. vol. 5, eds. D. W. MacDonald, D. Müller-Schwarze and S. E. Natynczuk, pp. 378–387. Oxford: Oxford University Press.
- Richardson, R. and Campbell, B. A. (1988). Effects of home nest odors on black–white preference in the developing rat: implications for developmental learning research. *Behavioral and Neural Biology* **50**, 361–366.
- Rick, C. M. and Bowman, R. I. (1961). Galapagos tomatoes and tortoises. *Evolution* **15**, 407–417.
- Rieger, J. F. and Jakob, E. H. (1988). The use of olfaction in food location by frugivorous bats. *Biotropica* **20**, 161–164.
- Rieser, J., Yonas, A., and Wikner, K. (1976). Radial localization of odors by newborns. *Child Development* 47, 856–859.
- Risser, J. M. and Slotnick, B. M. (1987). Nipple attachment and survival in neonatal olfactory bulbectomized rats. *Physiology and Behavior* **40**, 545–550.

- Rissman, E. F. and Johnston, R. E. (1986). Nutritional and social cues influence the onset of puberty in California voles. *Physiology and Behavior* **36**, 1–5.
- Rissman, E. F., Sheffield, S. D., Kretzmann, M. B., Fortune, J. E., and Johnston, R. E. (1984). Chemical cues from families delay puberty in male California voles. *Biology of Reproduction* **31**, 324–331.
- Ritter, F. J., Brüggemann, I. E. M., Gut, J., and Persoons, C. J. (1982). Recent pheromone research in the Netherlands on muskrats and some insects pests introduced from America into Europe: the muskrat, *Odatra zibethicus*, the American cockroach, *Periplaneta americana*, and the beet army worm, *Spodoptera exigua*. *American Chemical Society Symposium Series* **190**, 107–130.
- Rivard, G. and Klemm, W. R. (1989). Two body fluids containing bovine estrous pheromone(s). *Chemical Senses* **14**, 273–279.
 - (1990). Sample contact required for complete bull response to oestrous pheromone in cattle. In *Chemical Signals in Vertebrates*, vol. 5, ed. D W. Macdonald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 627–633. Oxford: Oxford University Press.
- Robbins, C. T., Hanley, T. A., Hagerman, A. E., *et al.* (1987). Role of tannins in defending plants against ruminants: reduction in protein availability. *Ecology* **68**, 98–107.
- Robbins, C. T., Hagerman, A. E., Austin, P. J., McArthur, C., and Hanley, T. A. (1991). Variation in mammalian physiological responses to a condensed tannin and its ecological implications. *Journal of Mammalogy* **72**, 480–486.
- Roberts, S. C. and Gosling, L. M. (2001). The economic consequences of advertising scent mark location on territories. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koi, J. J. Lepri, and D. Müller-Schwarze, pp. 11–17. New York: Plenum.
- Roberts, S. C. and Gosling, L. M. (2004). Manipulation of olfactory signaling and mate choice for conservation breeding: a case study of harvest mice. *Conservation Biology* **18**, 548–556.
- Roberts, S. C., Gosling, L. M., Thornton, E. A., and McClung, J. (2001). Scent-marking by male mice under the risk of predation. *Behavioral Ecology* **12**, 698–705.
- Robinson, I. (1990). The effect of mink odour on rabbits and small mammals. In *Chemical Signals in Vertebrates*, vol. 5, ed. D. W. Macdonald, D. Müller-Schwarze, and S. E. Natynczuk, pp. 566–572. New York: Oxford University Press.
- Robinson, T. (1979). The evolutionary ecology of alkaloids. In *Herbivores: Their Interaction with Secondary Plant Metabolites*, vol. 1, ed. G. A. Rosenthal, and M. R. Berenbaum, pp. 413–448. San Diego, CA: Academic Press.
- Rodriguez, E., Cavin, J. C., and West, J. E. (1982). The possible role of Amazonian psychoactive plants in the chemotherapy of parasitic worms: a hypothesis. *Journal of Ethnopharmacology* **6**, 303–309.
- Rodriguez, E., Aregullin, M., Nishida, T., *et al.* (1985). Thiarubrine A, a bioactive constituent of *Aspilia* (Asteraceae) consumed by wild chimpanzees. *Experientia* **41**, 419–420.
- Roeder, J. J. (1980). Marking behavior and olfactory recognition in genets (*Genetta genetta* L., Carnivora, Viverridae). *Behaviour* **72**, 200–210.

- Roessler, E. S. (1936). Viability of weed seeds after ingestion by California linnets. *Condor* **38**, 62–65.
- Rogers, J. G. and Beauchamp, G. K. (1976). Influence of stimuli from populations of *Peromyscus leucopus* on maturation of young. *Journal of Mammalogy* **57**, 320–330.
- Rollmann, S. M., Houck, L. D., and Feldhoff, R. C. (1999). Proteinaceous pheromone affecting female receptivity in a terrestrial salamander. *Science* **285**, 1907–1909.
 - (2000). Population variation in salamander courtship pheromones. *Journal of Chemical Ecology* **26**: 2713–2724.
 - (2003). Conspecific and heterospecific pheromone effects on female receptivity. *Animal Behaviour* **66**, 857–861.
- Romer, A. S. (1959). *Comparative Anatomy of Vertebrates*. [German edition.] Hamburg: Verlag Paul Parey.
- Rood, J. P. (1972). Ecological and behavioral comparisons of three genera of Argentine cavies. *Animal Behaviour Monographs* **5**, 1–83.
- Ropartz, P. (1968). Contribution à l'étude du déterminisme d'un effet de groupe chez les souris. *Comptes Rendus de l' Academie des Sciences Paris* **262**, 2070–2072.
- Roper, T. J., Shepherdson, D. J., and Davies, D. M. (1986). Scent marking with faeces and anal secretion in the European badger (*Meles meles*): seasonal and spatial characteristics of latrine use in relation to territoriality. *Behavior* **97**, 94–117.
- Rose, F. L. (1970). Tortoise chin gland fatty acid composition: behavioral significance. *Comparative Biochemistry and Physiology* **32**, 577–580.
- Rosell, F. and Bjørkøyli, T. (2002). A test of the dear enemy phenomenon in the Eurasian beaver. *Animal Behaviour* **63**, 1073–1078.
- Rosell, F. and Nolet, B. A. (1997). Factors affecting scent marking behavior in Eurasian beaver (*Castor fiber*). *Journal of Chemical Ecology* **23**, 673–689.
- Rosell, F. and Sun, L. (1999). Use of anal gland secretion to distinguish the two beaver species *Castor canadensis* and *C. fiber*. *Wildlife Biology* **5**, 119–124.
- Rosell, F., Bergan, F., and Parker, H. (1998). Scent marking in the Eurasian beaver (Castor fiber) as a means of territory defense. Journal of Chemical Ecology 24, 207–219.
- Rosenberg, T. (1995). The Haunted Land: Facing Europe's Ghosts After Communism. New York: Random House.
- Rosenthal. G. A. (1991). Nonprotein amino acids as protective allelochemicals. In *Herbivores: Their Interaction with Secondary Plant Metabolites*, vol. 1, ed. G. A. Rosenthal, and M. R. Berenbaum, pp. 1–34. San Diego, CA: Academic Press.
- Rosenthal, G. A. and Bell, E. A. (1979). Naturally occurring toxic nonprotein amino acids. In *Herbivores: Their Interaction with Secondary Plant Metabolites*, ed. G. A. Rosenthal, and D. H. Janzen, pp. 353–385. New York: Academic Press.
- Ross, P. and Crews, D. (1978). Stimuli influencing mating-behavior in garter snake, *Thamnophis radix*. *Behavioral Ecology* **4**, 133–142.
- Ross, C. W. and Detling, J. K. (1983). Investigations of trypsin inhibitors in leaves of four North American prairie grasses. *Journal of Chemical Ecology* **9**, 247–257.

- Rosser, A. E. and Keverne, E. B. (1985). The importance of central noradrenergic neurons in the formation of an olfactory memory in the prevention of pregnancy block. *Neuroscience* **15**, 1141–1147.
- Rothschild, M. (1965). The rabbit flea and hormones. Endeavour 24, 162–168.
- Rousi, M. and Häggman, J. (1984). Relationship between the total phenol content of Scots pine and browsing by the Arctic hare. *Silvae Genetica* **33**, 95–97.
- Rowsemitt, C. N. and O'Connor, A. J. (1989). Reproductive function in *Dipodomys ordii* stimulated by 6-methoxybenzoxazolinone. *Journal of Mammalogy* **70**, 805–809.
- Roy, J. and Bergeron, J. M. (1990a). Role of phenolics of coniferous trees as deterrents against debarking behavior of meadow voles (*Microtus pennsylvanicus*). *Journal of Chemical Ecology* **16**, 801–808.
 - (1990b). Branch-cutting behavior by the vole (*Microtus pennsylvanicus*): a mechanism to decrease toxicity in conifers. *Journal of Chemical Ecology* **16**, 735–741.
- Royce-Malmgren, C. H., and Watson, W. H., III. (1987). Modification of olfactory-related behavior in juvenile Atlantic salmon by changes in pH. *Journal of Chemical Ecology* **13**, 533–546.
- Rozenfeld, F. M. and Denoël, A. (1994). Chemical signals involved in spacing behavior of breeding female bank voles (*Clethrionomys glareolus* Schreber 1780, Microtidae, Rodentia). *Journal of Chemical Ecology* **20**, 803–813.
- Rozenfeld, F. M. and Rasmont, R. (1991). Odour cue recognition by dominant male bank voles, *Clethrionomys glareolus*. *Animal Behaviour* **41**, 839–850.
- Rozin, P. and Schiller, D. (1980). The nature and acquisition of a preference for chili pepper by humans. *Motivation and Emotion* **4**, 77–100.
- Rubin, B. D. and Katz, L. C. (1999). Optical imaging of odorant representation in the mammalian olfactory bulb. *Neuron* **23**, 499–511.
- Rubinoff, I. and Kropach, C. (1970). Differential reactions of Atlantic and Pacific predators to sea snakes. *Nature* **228**, 1288–1290.
- Russell, G. F. and Hills, J. I. (1971). Odor differences between enantiomeric isomers. *Science* **172**, 1043–1044.
- Russell, M. J. (1976). Human olfactory communication. Nature 260, 520-522.
 - (1983). Human olfactory communications. In *Chemical Signals in Vertebrates*, vol. 3, ed. D. Müller-Schwarze and R. M. Silverstein, pp. 259–273. New York: Plenum.
- Russell, M. J., Switz, G. M., and Thompson, K. (1980). Olfactory influences on the human menstrual cycle. *Pharmacology, Biochemistry and Behavior* **13**, 737–738.
- Russell, M. J., Mendelson, T., and Peeke, H. V. S. (1983). Mothers' identification of their infant's odors. *Ethology and Sociobiology* **4**, 29–31.
- Russell, M. J., Stone, G., and Russell, M. E. (1989). Human infants scent-mark their mothers. In *Proceedings of the Annual Meeting of the American Association for the Advancement of Science*, January 1989, San Francisco p. 209.
- Ryan, M. J., Fox, J. H., Wilczynski, R., and Rand, A. S. (1990). Sexual selection for sensory exploitation in the frog *Physalaemus pustulous*. *Nature* **343**, 66–67.

- Sachs, B. D. (1999). Airborne aphrodisiac odor from estrous rats: implications for pheromonal classification. In *Advances in Chemical Signals in Vertebrates*, vol. 8, ed. R. E. Johnston, D. Müller-Schwarze, and P. W. Sorensen, pp. 333–342. New York: Kluwer Academic-Plenum.
- Saglio, Ph., Fauconneau, B., and Blanc, J. M. (1990). Orientation of carp, *Cyprinus carpio* L., to free amino acids from Tubifex extract in an olfactometer. *Journal of Fisheries Biology* **37**, 887–898.
- Saito, T. R. (1986a). Induction of maternal behavior in sexually inexperienced male rats following removal of the vomeronasal organ. *Japanese Journal of Veterinary Science* **48**, 1029–1030.
 - (1986b). Role of the vomeronasal organ in retrieving behavior in lactating rats. *Zoological Science* **3**, 919–920.
- Salamon, M. (1995). Seasonal, sexual and dietary induced variations in the sternal scent secretion in the brushtail possum (*Trichosurus vulpecula*). In *Chemical Signals in Vertebrates*, vol. 7, ed. R. Apfelbach, D. Müller-Schwarze, K. Reutter, and E. Weiler, pp. 211–222. Oxford: Elsevier/Pergamon.
- Salmon, T. P. and Marsh, R. E. (1989). California ground-squirrel trapping influenced by anal-gland odors. *Journal of Mammalogy* **70**, 428–431.
- Sambraus, H. H. and Waring, G. H. (1975). Der Einfluss des Harns brünstiger Kühe auf die Geschlechtslust von Stieren. Zeitschrift für Säugetierkunde 40, 49–54.
- Sananes, C. B., Gaddy, J. R., and Campbell, B. A. (1988). Ontogeny of conditioned heart rate to an olfactory stimulus. *Developmental Psychobiology* **21**, 117–133.
- Sanchez-Criado, J. E. (1982). Involvement of the vomeronasal system in the reproductive physiology of the rat. In *Olfaction and Endocrine Regulation*, ed. W. Breipohl, pp. 209–217. London: IRL Press.
- Sanders, E. H., Gardner, P. D., Berger, P. J., and Negus, N. C. (1981). 6-Methoxy-benzoxazolinone: a plant derivative that stimulates reproduction in *Microtus montanus*. *Science* **214**, 67–69.
- San Diego Zoo (2002). Project: Giant panda Chemical communication in giant pandas. San Diego, CA; China. www.sandiegozoo.org/conservation/fieldproject. Accessed October 28, 2002.
- Sawyer, T. G., Marchinton, R. L., and Berisford, C. W. (1982). Scraping behavior in female white-tailed deer. *Journal of Mammalogy*. **63**, 696–697.
- Sawyer, T. G., Marchinton, R. L., and Miller, K. V. (1989). Response of female white-tailed deer to scrapes and antler rubs. *Journal of Mammalogy* **70**, 431–433.
- Schaal, B. (1988a). Natal discontinuity and chemosensory continuity: animal models and hypotheses for humans. *Année Biologique* **27**, 1–41.
 - (1988b). Olfaction in infants and children: developmental and functional perspectives. *Chemical Senses* **13**, 145–190.
- Schaal, B. and Orgeur, P. (1992). Olfaction *in utero*: can the rodent model be generalized? *Quarterly Journal of Experimental Psychology* **44B**, 245–278.
- Schaal, B., Montagner, H., Hertling, E., *et al.* (1980). Les stimulations olfactives dans les relations entre l'enfant et la mère. *Reproduction, Nutrition and Development* **20**, 843–858.

- Schaal, B., Orgeur, P., Lecanuet, J.-P., et al. (1991). Chimioréception nasale in utero: expériences préliminaires chez le foetus ovin. Comptes Rendus de l' Academie des Sciences, Paris 313 (Serie III), 319–325.
- Schaal, B., Marlier, L., and Soussignan, R. (1998). Olfactory function in the human fetus: evidence from selective neonatal responsiveness to the odor of amniotic fluid. *Behavioral Neuroscience* **112**, 1438–1449.
- Schaal, B., Marlier, L., and Soussignan, R. (2000). Human fetuses learn odours from their pregnant mother's diet. *Chemical Senses* **25**, 729–737.
- Schaal, B., Coureaud, G., Marlier, L., and Soussignan, R. (2001). Fetal olfactory cognition preadapts neonatal behavior in mammals. In *Chemical Signals in Vertebrates*, Vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri, and D. Müller-Schwarze, pp. 197–209. New York: Kluwer Academic/Plenum.
- Schaal, B., Coureaud, G., Langlois, D., *et al.* (2003). Chemical and behavioural characterization of the rabbit mammary pheromone. *Nature* **242**, 68–72.
- Schadler, M. H., Butterstein, G. M., Faulkner, *et al.* (1988). The plant metabolite 6-methoxybenzoxazolinone stimulates an increase in secretion of FSH and size of reproductive organs in *Microtus pinetorum*. *Biology of Reproduction* **38**, 817–820.
- Schaffer, V. (1940). *Die Hautdrüsenorgane der Säugetiere*. Vienna: Urban and Schwarzenberg. Schaller, G. B. (1972). *The Serengeti Lion*. Chicago, IL: the University of Chicago Press.
- Schank, J. C. (2001). Oestrus and birth synchrony in Norway rats, *Rattus norvegicus*. *Animal Behaviour* **62**, 409–415.
- Schecter, P. J. and Henkin, R. I. (1974). Abnormalities of taste and smell after head trauma. Journal of Neurology, Neurosurgery and Psychiatry 37, 802–810.
- Schellinck, H. M. and Brown, R. E. (2000). Selective depletion of bacteria alters but does not eliminate odors of individuality in *Rattus norvegicus*. *Physiology and Behavior* **70**, 261–270.
- Schilling, A., Serviere, J., Gendrot, G., and Perret, M. (1989). Functional activation of vomeronasal system by urine in a primate (*Microcebus murinus*): a 2-DG study. *Chemical Senses* 14, 224.
- Schisler, G. J. and Bergesen, E. P. (1996). Postrelease hooking mortality of rainbow trout caught on scented artificial baits. *North American Journal of Fish Management* **16**, 570–578.
- Schleidt, M. (1980). Personal odor and nonverbal communication. *Ethology and Sociobiology* 1, 225–231.
- Schleidt, M., Hold, B., and Attili, G. (1981). A cross-cultural study on the attitude towards personal odors. *Journal of Chemical Ecology* **7**, 19–31.
- Schmid, B. (1935). Über die Ermittlung des menschlichen und tierischen Individualgeruches durch den Hund. Zeitschrift für vergleichende Physiologie 22, 524–538.
- Schmidt, H. J. and Beauchamp, G. K. (1988). Adult-like odor preferences and aversions in three-year-old children. *Childhood Development* **59**, 1136–1143.
- Schmidt, U. (1975). Vergleichende Riechschwellenbestimmungen bei neotropischen Chiropteren (Desmodus rotundus, Artibeus literatus, Phyllostomus discolor). Zeitschrift fürSäugetierkunde 40, 269–296.

- Schmidt, U. and Greenhall, A. M. (1971). Untersuchungen zur geruchlichen Orientierung der Vampirfledermäuse (Desmodus rotundus). Zeitschrift für vergleichende Physiologie 74, 217–226.
- Schmidt, V. U., Schmidt, C., and Wysocki, C. J. (1986). Der Einfluss des Vomeronasalorgans auf das olfaktorisch geleitete Verhalten nestjunger Mäuse. *Zeitschrift für Säugetierkunde* **51**, 86–90.
- Schmidt-Koenig, K. (1987). Bird navigation: has olfactory orientation solved the problem? *Quarterly Review of Biology* **62**, 31–48.
- Scholl, J. P., Kelsey, R. G., and Shafizadeh, F. (1977). Involvement of volatile compounds of *Artemisia* in browse preference by mule deer. *Biochemical Systematics and Ecology* **5**, 291–295.
- Scholz, A. T., Horrall, R. M., Cooper, J. C., and Hasler, A. D. (1976). Imprinting to chemical cues: the basis for home stream selection in salmon. *Science* **192**, 1247–1249.
- Schulte, B. A., Müller-Schwarze, D., and Sun, L. (1995). Using anal gland secretion to determine sex in beaver. *Journal of Wildlife Management* **59**, 614–618.
- Schulte, B. A., Bagley, K., Correll, M., *et al.* (2005). Assessing chemical communication in elephants. In *Chemical Signals in Vertebrates*, vol. 10, eds. R. T. Mason, M. P. LeMasters, and D. Müller-Schwarze, pp. 140–151. New York: Springer.
- Schultz, T. H., Flath, R. A., Stern, D. J., et al. (1988). Coyote estrous urine volatiles. *Journal of Chemical Ecology* **14**, 701–712.
- Schultze-Westrum, T. G. (1965). Innerartliche Verständigung durch Düfte beim Gleitbeutler (*Petaurus breviceps papuanus*) Thomas (*Marsupialia Phalangeridae*). *Zeitschrift für vergleichende Physiologie* **50**, 151–220.
- Schwanzel-Fukuda, M. and Pfaff, D. (1989). Origin of luteinizing-hormone-releasing hormone neurons. *Nature* **338**, 161–164.
- Schwartz, C. C., Regelin, W. L., and Nagy, J. G. (1980). Deer preference for juniper forage and volatile oil treated foods. *Journal of Wildlife Management* 44, 114–120.
- Schwass, D. E. and Finley, J. W. (1985). Overview: the influence of nutrition on xenobiotic metabolism. *American Chemical Society Symposium Series* 277, 1–10.
- Schwende, F. J., Wiesler, D., Jorgenson, J. W., Carmack, M., and Novotny, M. (1986). Urinary volatile constituents of the house mouse, *Mus musculus*, and their endocrine dependency. *Journal of Chemical Ecology*, **12**, 277–296.
- Schwenk, K. (1994). Why snakes have forked tongues. Science 263, 1573-1577.
- Scott, A. P., Liley, N. R., and Vermeirssen, E. L. M. (1994). Urine of reproductively female rainbow trout, *Oncorhynchus mykiss* (Walbuam), contains a priming pheromone which enhances plasma levels of sex steroids and gonadotropin II in malts. *Journal of Fish Biology* 44, 131–148.
- Scott, G. R., Sloman, K. A., Rouleau, C., and Wood, C. M. (2003). Cadmium disrupts behavioural and physiological responses to alarm substance in juvenile rainbow trout (*Oncorhynchus mykiss*). *Journal of Experimental Biology* **206**, 1779–1790.
- Scott, N. L. and Rasmussen, L. E. L. (2005). Chemical communication of musth in captive male Asian elephants, *Elephas maximus*. In *Chemical Signals in Vertebrates*, vol. 10, eds. R. T. Mason, M. P. LeMaster, and D. Müller-Schwarze, pp. 118–127. New York: Springer.

- Scott, T. P. and Weldon, P. J. (1990). Chemoreception in the feeding behavior of adult American alligators, *Alligator mississippiensis*. *Animal Behaviour* **39**, 398–399.
- Scrivner, J. H., Howard, W. E., and Teranishi, R. (1984). Aldehyde volatiles for use as coyote attractants, In *Proceedings of the 11th Vertebrate Pest Conference*, ed. D. D. Clarke, pp. 157–160. Davis: University of California.
- Scudder, K. M., Stewart, N. J., and Smith, H. M. (1980). Response of neonate water snakes (*Nerodia sipedon sipedon*) to conspecific chemical cues. *Journal of Herpetology* **14**, 196–198.
- Scudder, K. M., Chiszar, D., and Smith, H. M. (1992). Strike-induced chemosensory searching and trailing behavior in neonatal rattlesnakes. *Animal Behaviour* **44**, 574–576.
- Segi, M. (1975). Tea-gruel as a possible factor for cancer for the esophagus. *Gann (Japanese Journal of Cancer Research)* **66**, 199–202.
- Seligman, P. J., Mathias, C. G. T., Omalley, M. A., *et al.* (1987). Phytophotodermatitis from celery among grocery store workers. *Archive of Dermatology* **123**, 1478–1482.
- Sengupta, S. (1981). Adaptive significance of the use of margosa leaves in nests of the house sparrow (*Passer domesticus*). *Emu* **81**, 114.
- Serizawa, S., Miyamichi, K., Nakatani, H., *et al.* (2003). Negative feedback regulation ensures the one receptor–one olfactory neuron rule in mouse. *Science* **302**, 2088–2094.
- Sever, D. M. (1988). Male *Rhyacotriton olympicus*, Dicamptodontidae: Urodela, has a unique cloacal vent gland. *Herpetologica* 44, 274–280.
- Shallenberger, R. J. (1975). Olfactory use in the wedge-tailed shearwater (*Puffinus pacificus*) on Manana Island, Hawaii. In *Olfaction and Taste*, vol. 5, ed. D. A. Denton and J. P. Coghlan, pp. 355–359. New York: Academic Press.
- Shappira, Z., Terkel, J., Egozi, J., Nyska, A., and Friedman, J. (1990). Reduction of rodent fertility by plant consumption. With particular reference to *Ziziphus spina-christi*. *Journal of Chemical Ecology* **16**, 2019–2026.
- Sheaffer, S. E. and Drobney, R. D. (1986). Effectiveness of lithium chloride induced taste aversions in reducing waterfowl nest predation. *Transactions of the Missouri Academy of Science* 20, 59–64.
- Sheffield, L. P., Law, J. M., and Burghardt, G. M. (1968). On the nature of chemical food sign stimuli for newborn garter snakes. *Communications in Behavioral Biology* 7–12.
- Shine, R., Langkilde, T. and Mason, R. T. (2003). Confusion within 'mating balls' of garter snakes: does misdirected courtship impose selection on male tactics? *Animal Behaviour* **66**, 1011–1017.
- Shivik, J. A. (1998). Brown tree snake response to visual and olfactory cues. *Journal of Wildlife Management* **62**, 105–111.
- Shivik, J. A., and Clark, L. (1997). Carrion seeking in brown tree snakes: importance of olfactory and visual cues. *Journal of Experimental Zoology* **279**, 549–553.
- Shkolnik, A. (1971). Diurnal activity in a small desert rodent. *International Journal of Biometeorology*, **15**, 115–120.

- Shutt, D. A. (1976). The effects of plant estrogens on animal reproduction. *Endeavour* **35**, 110–113.
- Siegel, M. A., Richardson, R., and Campbell, B. A. (1988). Effects of home nest stimuli on the emotional response of preweanling rats to an unfamiliar environment. *Psychobiology* **16**, 236–242.
- Siegel, S. (1979). The role of conditioning in drug tolerance and addiction. In *Psychopathology in Animals: Research and Clinical Implications*, ed. J. D. Keehn, pp. 143–168. New York: Academic Press.
- Signoret, J. A. (1975). Influence of the sexual receptivity of a teaser ewe on the mating preference in the ram. *Applied Animal Ethology* **1**, 229–232.
- Signoret, J. P., Cohen-Tannoudji, J., and Gonzalez, R. (1989). Olfactory aspects of sexual partner's effect on endocrine secretions in the domestic sheep. *Journal of Endocrinology* **123**(Suppl.), 15.
- Simon, G. S. and Madison, D. M. (1984). Individual recognition in salamanders: cloacal odors. *Animal Behaviour* **32**, 1017–1020.
- Sinclair, A. R. E., Jogia, M. K., and Andersen, R. J. (1988). Camphor from juvenile white spruce as an antifeedant for snowshoe hares. *Journal of Chemical Ecology* 14, 1505.
- Singer, A. G. and Macrides, F. (1990). Aphrodisin: pheromone or transducer? *Chemical Senses* **15**, 199–203.
- Singer, A. G., Agosta, W. C., O'Connell, R. J., *et al.*, (1976). Dimethyl disulfide: an attractant pheromone in hamster vaginal secretion. *Science* **191**, 948–950.
- Singer, A. G., Clancy, A. N., and Macrides, F. (1989). Conspecific and heterospecific proteins related to aphrodisin lack aphrodisiac activity in male hamsters. *Chemical Senses* 14, 565–575.
- Singer, A. G., Tsuchiya, H., Wellington, J. L., Beauchamp, G. K., and Yamazaki, K. (1993). Chemistry of odortypes in mice: fractionation and bioassay. *Journal of Chemical Ecology* **19**, 569–579.
- Singh, P. B., Brown, R. E., and Roser, B. (1987). MHC antigens in urine as olfactory cues. *Nature* **327**, 161–163.
- Sipos, M. L., Wysocki, C. J., Nygy, J. G., Wysocki, L., and Nemura, T. A. (1995). An ephemeral pheromone of female house mice: perception via the main and accessory systems. *Physiology and Behavior* **58**, 529–534.
- Skeen, J. T. and Thiessen, D. D. (1977). Scent of gerbil cuisine. *Physiology and Behavior* **19**, 11–14.
- Slotnick, B. H. and Schoonover, F. W. (1984). Redundancy: One-side bulbectomized rats have same absolute threshold and intensity difference threshold as intact rats. *Chemical Senses* **9**, 325–340.
- Smale, L., Pedersen, J. M., Block, M. L., and Zucker, I. (1990). Investigation of conspecific male odours by female prairie voles. *Animal Behaviour* **39**, 768–774.
- Smallwood, P. D. and Peters. W. D. (1986). Grey squirrel wood preferences: The effects of tannin and fat concentrations. *Ecology* **67**, 168–174.

- Smith, A. B. III, Belcher, A. M., Epple, G., Jurs, P. C., and Lavine, B. (1985). Computerized pattern recognition: a new technique for the analysis of chemical communication. *Science* **228**, 175–177.
- Smith, B. A. and Block, L. (1990). Preference of Mongolian gerbils for salivary cues: a developmental analysis. *Animal Behaviour* **39**, 512–521.
- Smith, D. V. and Margolskee, R. F. (2001). Making sense of taste. *Scientific American* **284**, 32–39.
- Smith, G. J. and Spear, N. E. (1978). Effects of the home environment on withholding behaviors and conditioning in infant and neonatal rats. *Science* **202**, 327–329.
 - (1981). Home environmental stimuli facilitate learning shock escape spatial discrimination in rats 7–11 days of age. *Behavioral and Neural Biology* **31**, 360–365.
- Smith, J. L. D., McDougall, C., and Michelle, D. (1989). Scent marking in free-ranging tigers, *Panthera tigris*. *Animal Behaviour* 37, 1–10.
- Smith, P. W., Parks, O. W., and Schwartz, D. P. (1984). Characterization of male goat odours: 6-trans-nonenal. *Journal of Dairy Science* **67**, 794–801.
- Smith, R. J. F. and Lawrence, B. J. (1989). Behavioral response of solitary fathead minnows, *Pimephalis promelas*, to alarm substance. *Journal of Chemical Ecology* **15**, 209–219.
- Smith, S. A. and Paselk, R. A. (1986). Olfactory sensitivity of the turkey vulture (*Cathartes aura*) to three carrion-associated odors *Auk* **103**, 586–592.
- Smith, T. E., Faulkes, C. G., and Abbott, D. H. (1997). Combined olfactory contact with the parent colony and direct contact with non breeding animals does not maintain suppression of ovulation in female naked mole-rats (*Heterocephalus glaber*). *Hormones and Behavior* **31**, 277–288.
- Smith, T. L. and Kardong, K. V. (2000). Absence of polarity perception by rattlesnakes of envenomated prey trails. *Journal of Herpetology* **34**, 621–624.
- Smith, W. P., Borden, D. L., and Endres, K. M. (1994). Scent-station visits as an index to abundance of raccoons: an experimental manipulation. *Journal of Mammalogy* **75**, 637–647.
- Smotherman, W. P. (1982). Odor aversion learning by the rat fetus. *Physiology and Behavior* **29**, 769–771.
 - (1987). Response of the rat fetus to olfactory stimulation presented *in utero*. *Society of Neuroscience* **13**, 8l.
- Smotherman, W.P. and Robinson, S. R. (1985). The rat fetus in its environment: behavioral adjustments to novel familiar, aversive and conditional stimuli presented in utero. *Behavioral Neuroscience* **99**, 521–530.
 - (1987). Prenatal expression of species-typical action patterns in the rat fetus (Rattus norvegicus). Journal of Comparative Psychology 101, 190–196.
 - (1988). Behavior of rat fetuses following chemical or tactile stimulation. *Behavioral Neuroscience* **102**, 24–34.
- Smotherman, W. P., Robinson, S. R., La Vallee, P. A. and Hennessy, M. B. (1987). Influences of the early olfactory environment on the survival, behavior and pituitary—adrenal activity of cesarean delivered preterm rat pups. *Developmental Psychobiology* **20**, 415–424.

- Snyder, G. K. and Peterson, D. T. (1979). Olfactory sensitivity in the black-billed magpie and in the pigeon. *Comparative Biochemistry and Physiology A* **62**, 921–925.
- Snyder, M. A. (1992). Selective herbivory by Abert's squirrel mediated by chemical variability in ponderosa pine. *Ecology* **73**, 1730–1741.
 - (1993). Interactions between Abert's squirrel and ponderosa pine: the relationship between selective herbivory and host plant fitness. *American Naturalist* **141**, 866–879.
 - (1998). Abert's squirrel (*Sciurus aberti*) in ponderosa pine (*Pinus ponderosa*) forests: directional selection, diversifying selection. In *Special Publication* 6: *Ecology and Evolutionary Biology of Tree Squirrels*, ed. M. A. Steele, J. F. Merritt, and D. A. Zegers, pp. 195–201. Martinsville, VA: Virginia Museum of Natural History.
- Soffie, M. and Lamberty, Y. (1988). Scopolamine effects on juvenile conspecific recognition in rats: possible interaction with olfactory sensitivity. *Behavioral Processes* 17, 181–190.
- Sola, C. (1995). Chemoattraction of upstream migrating glass eels, *Anguilla anguilla* to organic earthy and green odorants. *Environmental Biology of Fishes* **43**, 179–185.
- Sola, C. and Tosi L. (1993). Bile salts and taurine as chemical stimuli for glass eels *Anguilla* anguilla: a behavioral study. *Environmental Biology of Fishes* 37, 197–204.
- Soni, G. R. and Prakash, I. (1987). Effect of conspecific urine on behavior of soft-furred field-rat, *Rattus meltada*. *Behavioral Processes* **14**, 175–182.
- Sorensen, A. E. (1983). Taste aversion and frugivore preference. Oeologia 56, 117–120.
- Sorensen, P. W., and Stacey, N. E. (1990). Identified hormonal pheromones in the goldfish: the basis for a model of sex pheromones function in teleost fish. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 302–311. New York: Plenum.
 - (1999). Evolution and specialization of fish hormonal pheromones. In *Chemical Signals in Vertebrates*, vol. 8, ed. R. E. Johnston, D. Müller-Schwarze and P. W. Sorensen, pp. 15–47. New York: Kluwer Academic/Plenum.
- Sorensen, P. W., Hara, T. J., Stacey, N. E., and Goetz, F. W. (1988). F prostaglandins function as potent olfactory stimulants that comprise the postovulatory female sex pheromone in goldfish. *Biology of Reproduction* **39**, 1039–1050.
- Sorensen, P. W., Hara, T. J., Stacey, N. E., and Dulka, J. G., (1990a). Extreme olfactory specificity of male goldfish to the preovulatory steroidal pheromone 17α , 20β -dihydroxy-4-pregnen-3-one. *Journal of Comparative Physiology A* **166**, 373.
- Sorensen, P. W., Stacey, N. E., and Hara, T. J. (1990b). Acute olfactory sensitivity and specificity of mature male goldfish to water borne androgenic steroids: a class of inhibitory pheromones? *Chemical Senses* 15, 644.
- Sorensen, P. W., Irvine, I. A. S., Scott, A. P., and Stacey, N. E. (1992). Electrophysiological measures of olfactory sensitivity suggest that goldfish and other fish use species-specific mixtures of hormones and their metabolites as pheromones. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 357–354. New York: Plenum.
- Sosa, T., Chaves, N., Alias, J. C., Escudero, J. C., Henao, F., and Gutiérrez-Merino, C. (2004). Inhibition of mouth skeletal muscle relaxation by flavonoids of *Cistus ladanifer*

- L.: a plant defense mechanism against herbivores. *Journal of Chemical Ecology* **30**, 1087–1101.
- Spencer, P., Nunn, P. B., Hugon, J., *et al.* (1987). Guam amyotrophic lateral sclerosis–parkinsonism–dementia linked to a plant excitant neurotoxin. *Science* **237**, 517–522.
- Spickett, A. M., Kierans, J. E., Norval, R. A. I., and Clifford, C. M. (1981). *Ixodes matopi*, new species (Acarina: Ixodes): a tick found aggregating on preorbital gland scent marks of the klipspringer in Zimbabwe. *Onderstepoort Journal of Veterinary Research* 48, 23–30.
- Stabell, O. B. (1987). Intraspecific pheromone discrimination and substrate marking by Atlantic salmon parr. *Journal of Chemical Ecology* **13**, 1644.
- Stacey, N. E. and Hourston, A. S. (1982). Spawning and feeding behavior of captive Pacific herring, (*Clupea harengus pallasi*). *Canadian Journal of Fisheries* **39**, 489–498.
- Stager, K. E. (1964). The role of olfaction in food location by the turkey vulture (*Cathartes aura*). Los Angeles County Museum Contributions to Science No. 81.
- Stager, K. E. (1967). Avian olfaction. American Zoologist 7, 415–419.
- Stahl, E. (1888). Pflanzen und Schnecken. Biologische Studien über die Schutzmittel der Pflanzen gegen Schneckenfrass. Jenaische Zeitschrift für Medizin und Naturwissenschaft 22, 557–684.
- Stahlbaum, L. C. and Houpt, K. A. (1989). The role of the flehmen response in the behavioral repertoire of the stallion. *Physiology and Behavior* **45**, 1207–1214.
- Stattelman, A. J., Talbot, R. B., and Coulter, D. B. (1975). Olfactory thresholds of pigeons (*Columba livia*), quail (*Colinus virginianus*) and chickens (*Gallus domesticus*). *Comparative Biochemistry and Physiology A* **50**, 807–809.
- Staubli, U., Fraser, D., Faraday, R. and Lynch, G. (1987). Olfaction and the "data" memory system in rats. *Behavioral Neuroscience* **101**, 757–765.
- Stauffer, H. P. and Semlitch, R. D. (1993). Effects of visual, chemical and tactile cues of fish on the behavioral responses of tadpoles. *Animal Behaviour* **46**, 355–364.
- Steel, E. and Hutchinson, J. B. (1987). The aromatase inhibitor, 1,4,6-androstatriene-3,7-dione (ATD) blocks testosterone-induced olfactory behaviour in the hamster. *Physiology and Behavior* **39**, 141–145.
- Stehn, R. A. and Richmond, M. E. (1975). Male-induced pregnancy termination in the prairie vole, *Microtus ochrogaster*. *Science* **187**, 1211–1213.
- Stein, M., Ottenberg, P., and Roulet, N. (1958). A study of the development of olfactory preferences. *American Medical Association Archives of Neurology and Psychiatry* **80**, 264–266.
- Stern, K. and McClintock, M. K. (1998). Regulation of ovulation by human pheromones. *Nature* **392**, 177–179.
- Stevens, J. C., and Cain, W. S. (1985). Age-related deficiency in the perceived strength of six odorants. *Chemical Senses* **10**, 517–529.
- Stevens, J. C., Cain, W. S., and Burke, R. J. (1988). Variability of olfactory thresholds. *Chemical Senses* **13**, 643–654.
- Stewart, P. S., MacDonald, D. W., Newman, C., and Tattersall, F. H. (2002). Behavioral mechanisms of information transmision and reception by badgers, *Meles meles*, at latrines. *Animal Behaviour* **63**, 999–1007.

- Stickrod, G., Kimble, D. P., and Smotherman, W. P. (1982). In utero taste/odor aversion conditioning in the rat. *Physiology and Behavior* **28**, 5–7.
- Stoddart, D. M. (1976). Effects of weasels (*Mustela nivalis*) on trapped samples of their prey. *Oecologia* **22**, 439–441.
 - (1979). Specialized scent-releasing hair in the crested rat *Lophiomys imhausi*. *Journal of Zoology* **189**, 551–553.
 - (1982). Does trap odour influence estimation of population size of the short-tailed vole, *Microtus agrestis? Journal of Animal Ecology* **51**, 375–386.
 - (1983). The Ecology of Vertebrate Olfaction. London: Chapman & Hall.
- Stoddart, D. M. and Bradley, A. G. (1991). Measurement of short-term changes in heart rate and plasma concentration of cortisol and catecholamine in a small marsupial. *Journal of Chemical Ecology* **17**, 1333–1341.
- Stone, R. (1993). Guam: deadly disease dying out. Science 261, 424–426.
 - (2002). Toxicology. Fruit bats linked to mystery disease. Science 296, 24.
- Stonerook, M. J. and Harder, J. D. (1989). Male pheromones advance the age of first estrus in the gray, short-tailed opossum, *Monodelphis domestica*. *Biology of Reproduction* **40**(Suppl.1), 157.
- Stralendorff, F. V. (1987). Partial chemical characterization of urinary signaling pheromone in tree shrews (*Tupaia belangeri*). *Journal of Chemical Ecology* **13**, 655–679.
- Strier, K. B. and Ziegler, T. E. (1994). Insights into ovarian function in wild *Muriqui* monkeys (*Muriqui arachnoides*). *American Journal of Primatology* **32**, 31–40.
- Strom, G. H. (1976). Transport and diffusion of stack effluents. In *Air Pollution*, vol. 1, ed. A. C. Sku, pp. 401–503. New York: Academic Press.
- Strupp, B. J. and Levitsky, D. A. (1984). Social transmission of food preferences in adult hooded rat (*Rattus norvegicus*). *Journal of Comparative Psychology* **98**, 257–266.
- Sugiyama, T. (1983). Sex odour components in male goat. Estrous goats are interested in 4-ethyl fatty acids secreted by mature male goat. *Kagaku to Seibutsu* (*Chemistry and Biology, Japanese*) **21**, 420–430. [*Chemical Abstracts* 100, 18366s.]
- Sugiyama, T., Sasada, H., Masaki, F., and Yamashita, K. (1981). Unusual fatty acids with specific odor from mature male goats. *Agriculture and Biological Chemistry* **45**, 2655–2658.
- Sugiyama, T., Matsumura, H., Sasada, H., Masaki, J., and Yamashita, K. (1986). Characterization of fatty acids in the sebum of goats according to sex and age. *Agricultural and Biological Chemistry* **50**, 3049–3052.
- Sullivan, R. M. and Leon, M. (1986). Early olfactory learning induces an enhanced olfactory-bulb response in young rats. *Developmental Brain Research* 27, 278–282.
- Sullivan, R. M., Hofer, M. A., and Brake, S. C. (1986). Olfactory-guided orientation in neonatal rats is enhanced by a conditioned change in behavioral state. *Developmental Psychobiology* **19**, 615–624.
- Sullivan, T. P. (1986). Influence of wolverine (*Gulo gulo*) odor on feeding behavior of snow-shoe hares (*Lepus americanus*). *Journal of Mammalogy* **67**, 385–388.

- Sullivan, T. P. and Crump, D. R. (1984). Influence of mustelid scent gland compounds on suppression of feeding by snowshoe hares (*Lepus americanus*). *Journal of Chemical Ecology* **10**, 1809–1821.
 - (1986a). Avoidance responses of pocket gophers (*Thomomys talpoides*) to mustelid anal gland compounds, In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze, and R. M. Silverstein. pp. 519–531. New York: Plenum.
 - (1986b). Feeding responses of snowshoe hares (*Lepus americanus*) to volatile constituents of red fox (*Vulpes vulpes*) urine. *Journal of Chemical Ecology*. **12**, 729–739.
- Sullivan, T. P., Nordstrom, L. O., and Sullivan, D. S. (1985a). The use of predator odors as repellents to reduce feeding damage by herbivores. I. Snowshoe hares (*Lepus americanus*) *Journal of Chemical Ecology* **11**, 903–919.
 - (1985b). The use of predator odors as repellents to reduce feeding damage by herbivores. II. Black-tailed deer (*Odocoileus hemionus columbianus*). *Journal of Chemical Ecology* **11**, 921–935.
- Sullivan, T. P., Crump, D. R., and Sullivan, D. S. (1988). Use of predator odors as repellents to reduce feeding damage by herbivores. III. Montane and meadow voles (*Microtus montanus* and *Microtus pennsylvanicus*). *Journal of Chemical Ecology* **14**, 363–377.
- Sullivan, T. P., Crump, D. R., Wieser, H. and Dixon, E. A. (1990). Response of pocket gophers (*Thomomys talpoides*) to an operational application of synthetic semiochemicals of stoat (*Mustela erminea*). *Journal of Chemical Ecology* **16**, 941–949.
 - (1992). Influence of the plant antifeedant, pinosylvin, on suppression of feeding by snowshoe hares. *Journal of Chemical Ecology* **18**, 1573–1561.
- Sun, L. and Müller-Schwarze, D. (1997). Sibling recognition in the beaver: a field test for phenotype matching. *Animal Behaviour* **54**, 492–502.
- Sun, L., Xiao, B., and Dai, N. (1994). Scent marking behaviour in the male Chinese water deer. *Acta Theriologica* **39**, 177–184.
- Sunnerheim, K., Palo, R. T., Theander, O., and Knutsson, P. G. (1988). Chemical defense in birch. Platyphylloside: a phenol from *Betula pendula* inhibiting digestibility. *Journal of Chemical Ecology* **14**, 549.
- Sunnerheim-Sjöberg, K. (1992). (1*S*, 2*R*, 4*S*, 5*S*)-Angelicoidenol-2-O-*β*-D-glucopyranoside: a moose deterrent compound in Scots pine (*Pinus sylvestris* L.). *Journal of Chemical Ecology* **18**, 2025–2039.
- Sunnerheim-Sjöberg, K. and Hämäläinen, M. (1992). Multivariate study of moose browsing in relation to phenol patterns in pine needles. *Journal of Chemical Ecology* **18**, 659–672.
- Sutter, E. (1946). Das Abwehrverhalten nestjunger Wiedehopfe. Der Ornithologische Beobachter 43, 72–81.
- Sutton, O. G. (1953). Micrometeorology. New York: McGraw Hill.
- Sveinsson, T. and Hara, T. J. (1990). Olfactory receptors in Arctic char (*Salvelinus alpinus*) with high sensitivity and specificity for prostaglandin $F_{2\alpha}$. Chemical Senses **15**, 645–646.

- Svendsen., G. E. (1980). Patterns of scent-mounding in a population of beaver (*Castor canadensis*). *Journal of Chemical Ecology* **6**, 133–148.
- Svendsen, G. E. and Jollick, J. D. (1978). Bacterial contents of the anal and castor glands of beaver (*Castor canadensis*). *Journal of Chemical Ecology* **4**, 563–569.
- Svoboda, F. J. and Gullion, G. W. (1972). Preferential use of aspen by ruffed grouse in northern Minnesota. *Journal of Wildlife Management* **36**, 1166–1180.
- Swain, T. (1977). Secondary compounds as protective agents. *Annual Review of Plant Physiology* **28**, 479–501.
 - (1979). Phenolics in the environment. Recent Advances in Phytochemistry 12, 617–640.
- Swank, W. (1944). Germination of seeds after ingestion by ring-necked pheasants. *Journal of Wildlife Management* **8**, 223–231.
- Swihart, R. K., Pignatello, J. J., and Mattina, M. J. I. (1991). Aversive responses of white-tailed deer, *Odocoileus virginianus*, to predator urines. *Journal of Chemical Ecology* 17, 767–777.
- Tachibana, K., Sakaitanai, M., and Nakanishi, K. (1984). Pavoninins: shark-repelling ichthyotoxins from the defense secretion of the Pacific sole. *Science* **226**, 703–705.
- Tahvanainen, J., Helle, E., Julkunen-Tiitto, R., and Lavola, A. (1985). Phenolic compounds of willow bark as deterrents against feeding by mountain hares. *Oecologia* **65**, 319–323.
- Tang-Martinez, Z. (2001). The mechanisms of kin discrimination and the evolution of kin recognition in vertebrates: a critical re-evaluation. *Behavioral Processes* **13**, 21–40.
- Tang-Martinez, Z., Mueller, L. L., and Taylor, G. T. (1993). Individual odours and mating success in the golden hamster, *Mesocricetus auratus*. *Animal Behaviour* **45**, 1141–1151.
- Tattersall, D. B., Bak, S., Jones, P. R., *et al.* (2001). Resistance to an herbivore through engineered cyanogenic glucoside synthesis. *Science* **293**, 1826–1828.
- Tauber, G. (1992). A dubious battle to save the Kemp's Ridley sea turtle. *Science* **256**, 614–616.
- Teeter, J. (1980). Pheromone communication in sea lampreys (*Petromyzon marinus*): implications for population management. *Canadian Journal of Fisheries and Aquatic Science* 37, 2123–2132.
- Teicher, E. M. and Blass, M. H. (1980). Suckling. Science 210, 15-22.
- Teichmann, H. (1957). Das Riechvermögen des Aales (Anguilla anguilla L.). Naturwissenschaften 44, 242–246.
 - (1959). Concerning the power of the olfactory sense of the eel, *Anguilla anguilla* (L.). *Zeitschrift für vergleichende Physiologie* **42**, 206–254.
- Temeles, E. J. (1994). The role of neighbours in territorial systems: when are they 'dear enemies'? *Animal Behaviour* 47, 339–350.
- Temple, S. A. (1977). Plant-animal mutualism: coevolution with dodo leads to near extinction of plant (*Calvaria major*). *Science* **197**, 885–886.
- Terborgh, J. (1988). The big things that run the world: a sequel to E. O. Wilson. *Conservation Biology* **2**, 402–403.

- Terborgh J., Lopez, L., Nuñez P. *et al.* (2001). Ecological meltdown in predator-free forest fragments. *Science* **294**, 1923–1926.
- Terrick, T. D., Mumme, R. L. and Burghardt, G. M. (1995). Aposematic coloration enhances chemosensory recognition of noxious prey in the garter snake *Thamnophis radix*. *Animal Behaviour* **49**, 857–866.
- Terry, L. M. and Johanson, I. B. (1987). Olfactory influences on the ingestive behavior of infant rats. *Developmental Psychobiology* **20**, 313–332.
- Tester, A. L. (1963). Olfaction, gustation and the common chemical sense in sharks. *Sharks and Survival* **8**, 255–282.
- Tew, T. (1987). A comparison of small animal responses to clean and dirty traps. *Journal of Zoology (London)* **212**, 361–364.
- Tew, T. E., Todd, I. A., and Macdonald, D. W. (1994). Temporal changes in olfactory preferences in murid rodents revealed by live-trapping. *Journal of Mammalogy* **75**, 750–756.
- Thesen, A., Steen, J. B., and Døving, K. B. (1993). Behaviour of dogs during olfactory tracking. *Journal of Experimental Biology* **180**, 247–251.
- Thibodeaux, L. J. (1979). Chemodynamics, Environmental Movement of Chemicals in Air, Water, and Soil. New York: Wiley.
- Thieme, H. (1965). Die Phenylglykoside der Salicaceen: Untersuchungen über jahreszeitlich bedingte Veränderungen der Glykosidkonzentrationen, über die Abhängigheit des Glykosidgehalts von der Tageszeit und dem Alter den Pflanzenorgane. *Pharmazie* 20, 688–691.
- Thiessen D. D. and Cocke, R. (1986). Alarm chemosignals in a *Meriones unguiculatus*: preypredator interactions. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze and Silverstein, R. M. pp. 507–518. New York: Plenum.
- Thiessen, D. D., Clancy, A., and Goodwin, M. (1976). Harderian gland pheromone in the Mongolian gerbil *Meriones unguiculatus*. *Journal of Chemical Ecology* **2**, 231–238.
- Thoen, C., Bauwen, D., and Verheyen, R. F. (1986). Chemoreceptive and behavioral responses of the common lizard, *Lacerta vivipara*, to snake chemical deposits. *Animal Behaviour* **34**, 1805–1813.
- Thomas, D. W., Samson, C., and Bergeron, J. M. (1988). Metabolic costs associated with the ingestion of plant phenolics by *Microtus pennsylvaticus*. *Journal of Mammalogy* **69**, 512–515.
- Thomas, K. J. and Dominic, C. J. (1987a). Evaluation of the role of the stud male in preventing male-induced implantation failure (the Bruce effect) in laboratory mice. *Animal Behaviour* **35**, 1257–1260.
 - (1987b). Male-induced implantation failure (the Bruce effect) in laboratory mice: investigations on the olfactory memory of the newly inseminated female. *Archives of Biology* **98**, 263–272.
- Thomas, R. McG. (1996). Ernst Morch, anesthesiologist and heroic inventor, was 87 (obituary). *New York Times* January 18, p. B11.
- Tiedman, G. T., Oh, J. H., Oita, K., and Christoffers, G. W. (1976). Wildlife damage control II: partial identification of the active ingredients in big game repellents derived from

- fish and eggs. In Proceedings of the 172nd National Meeting of American Chemical Society San Francisco.
- Tipton, K. W., Floyd, E. H., Marshall, J. C., and McDevitt, J. B. (1970). Resistance of certain grain sorghum hybrids to bird damage in Louisiana. *Agronomy Journal* **62**, 211–213.
- Todd, N. (1962). The inheritance of the catnip response in the domestic cat. *Journal of Heredity* **53**, 54–56.
- Toftegaard, C. L., McMahon, K. L., Galloway, G. J., and Bradley, A. J. (2002). Processing of urinary pheromones in *Antechinus stuartii* (Marsupialia: Dasyuridae): Functional magnetic resonance imaging of the brain. *Journal of Mammalogy* **83**, 71–80.
- Tolhurst, B. E. and Vince, M. A. (1976). Sensitivity to odours in the embryo of the domestic fowl. *Animal Behaviour* **24**, 772–779.
- Tomasi, T. E. (1978). Function of venom in the short-tailed shrew, *Blarina brevicauda*. *Journal of Mammalogy* **60**, 751–759.
- Tomback, D. F. (1980). How nutcrackers find their seed stores. Condor 82, 10-19.
- Tosi, L. and Sola, C. (1993). Role of geosmin, a typical inland water odour, in guiding glass, eel *Anguilla anguilla* (L.) migration. *Ethology* **95**, 177–185.
- Tracy, C. R. and Dole, J. W. (1969). Orientation of displaced California toads, *Bufo boreas*, to their breeding sites. *Copeia* **1969**, 693–700.
- Traveset, A. (1998). Effects of seed passage through vertebrate frugivores' guts on germination: a review. *Perspectives in Plant Ecology, Evolution and Systematics* 1, 151–190.
- Tributsch, H. (1988). When the Snakes Awake: Animals and Earthquake Prediction. Cambridge, MA: MIT Press.
- Tristram, D. A. (1977). Intraspecific olfactory communication in the terrestrial salamander (*Plethodon cinereus*). *Copeia* 597–600.
- Trowbridge, B. J. (1983). Olfactory communication of the European otter *Lutra lutra* L. Ph.D. Thesis, University of Aberdeen, UK.
- Trumler, E. (1958). Beobachtungen an den Böhmzebras des Georg-von-Opel-Freigeheges für Tierforschung e.V., Kronberg im Taunus. J. Das Paarungsverhalten. Säugetierkundliche Mitteilungen 6, 1–18.
- Tsuchiya, H., Yamazaki, K., Beauchamp, G. K., and Singer, A. G. (1992). Chemical characterization of MHC-determined body odors. *Chemical Senses* 17, 593.
- Tucker, D. (1963). Physical variables in the olfactory stimulation process. *Journal of General Physiology* **46**, 453–489.
 - (1971). Non-olfactory responses from the nasal cavity: Jacobson's organ and the trigeminal system. In *Handbook of Sensory Physiology, vol.* 4, ed. L. M. Beidler, pp. 151–181. Berlin: Springer-Verlag.
- Twitty, V. C. (1966). Of Scientists and Salamanders. San Francisco, CA: W. H. Freeman.
- Umemura, K., Sugawara, K., and Ito, I. (1988). The source of the odor emitted from estrous cows. *Japanese Journal of Zootechnical Science* **59**, 779–786.
- Utaisincharoen, P., Mackessy, S, P., Miller, R. A., and Ju, A. T. (1993). Complete primary structure and biochemical properties of gilatoxin, a serine protease with kallikrein-like and angiotensin-degrading properties. *Journal of Biological Chemistry* **268**, 21975–21985.

- Valderrama, X., Robinson, J. G., Attygalle, A. B., Athula, B., and Eisner, T. (2000). Seasonal anointment with millipedes in a wild primate: a chemical defense against insects? *Journal of Chemical Ecology* **26**, 2781–2790.
- Vale, G. A. (1974). The responses of tsetse flies (Diptera: Glossinidae) to mobile and stationary baits. Bulletin of Entomological Research 64, 545–588.
 - (1980). Field studies of the responses of tsetse flies (Glossinidae) and other Diptera to carbon dioxide, acetone and other chemicals. *Bulletin of Entomological Research* **70**, 563–570.
 - (1981). An effect of host diet on the attraction of tsetse flies (Diptera: Glosinidae) to host odour. Bulletin of Entomological Research 71, 259–265.
- Vale, G. A. and Hall, D. R. (1985). The role of 1-octen-3-ol, acetone and carbon dioxide in the attraction of tsetse flies *Glossina* spp. (Diptera: Glossinidae) to ox odour. *Bulletin of Entomological Research* **75**, 209–217.
- Vale, G. A., Flint, S., and Hall, D. R. (1986). The field responses of tsetse flies, *Glossina* spp. (Diptera: Glossinidae), to odours of host residues. *Bulletin of Entomological Research* **76**, 685–693.
- Valentincic, T. and Caprio, J. (1992). Gustatory responses of channel catfish to amino acids. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 365–369 New York: Plenum.
- Van Damme, R. and Castilla, A. M. (1996). Chemosensory predator recognition in the lizard *Podarcis hispania*: effects of predation pressure relaxation. *Journal of Chemical Ecology* **22**, 13–22.
- Vandenbergh, J. G. (1969). Male odor accelerates female sexual maturation in mice. *Endocrinology* **84**, 658–660.
 - (1971). The influence of the social environment on sexual maturation in male mice. *Journal of Reproduction and Fertility* **24**, 383.
 - (1973). Effects of gonadal hormones on the flank gland of the golden hamster. *Hormone Research* **4**, 28–33.
 - (1987). Regulation of puberty and its consequences on population dynamics in mice. *American Zoologist* **27**, 891–898.
- Vandenbergh, J. G. and Hotchkiss, A. K. (2001). Interfetal communication and adult phenotype in mice. In *Chemical Signals in Vertebrates*, vol. 9, ed. A. Marchlewska-Koj, J. J. Lepri and D. Müller-Schwarze, pp. 183–187. New York: Kluwer Academic/Plenum.
- Vandenbergh, J. G., Whitsett, J. M., and Lombardi, J. L. (1975). Partial isolation of a pheromone accelerating puberty in female mice. *Journal of Reproduction and Fertility* 43, 515–523.
- Vandenbergh, J. G., Finlayson, J. S., Dobrogosz, W. J., Dills, S. S., and Kost, T. A. (1976). Chromatographic separation of puberty accelerating pheromone from male mouse urine. *Biology of Reproduction* **15**, 260–265.
- van den Berk, J. and Müller-Schwarze, D. (1984). Responses of wild muskrats (*Ondatra zibethicus* L.) to scented traps. *Journal of Chemical Ecology* **10**, 1411–1415.
- van den Hurk, R. V., Schoonen, W. G. E. J., van Zoelen, G. A., and Lambert, J. G. D. (1987). The biosynthesis of steroid glucuronides of the zebra fish *Brachydanio verio* and their

- pheromonal function as ovulation inducers. *General and Comparative Endocrinology* **68**, 179–188.
- van der Lee, S. and Boot, L. M. (1955). Spontaneous pseudopregnancy in mice. *Acta Physiologica Neerlandica* **4**, 442–446.
- Vander Wall, S. B. (1982). An experimental analysis of cache recovery in Clark's nutcracker. *Animal Behaviour* **30**, 84–94.
 - (1998). Foraging success of granivorous rodents: effects of variation in seed and soil water on olfaction. *Ecology* **79**, 223–241.
 - (2000). The influence of environmental conditions on cache recovery and cache pilferage by yellow pine chipmunks (*Tamias amoenus*) and deer mice (*Peromyscus maniculatus*). *Behavioral Ecology* **11**, 544–549.
 - (2003). How rodents smell buried seeds: a model based on the behavior of pesticides in soil. *Journal of Mammalogy* **84**, 1089–1099.
- Vander Wall, S. B., Beck, M. J., Briggs, R., *et al.* (2003). Interspecific variation in the olfactory abilities of granivorous rodents. *Journal of Mammalogy* **84**, 847–896.
- van Etten, C. H. and Tookey, H. L. (1979). Chemistry and biological effects of glucosinolates. In *Herbivores: Their Interactions with Secondary Plant Metabolites*, ed. G. A. Rosenthal and D. H. Janzen, pp. 471–500. New York: Elsevier.
- Van Hoven, W. (1991). Mortalities in kudu (*Tragelaphus strepsiceros*) populations related to chemical defence in trees. *Journal of African Zoology* **105**, 141–145.
- Vaughan, T. A. (1978). Mammalogy. Philadelphia, PA: Saunders College.
- Vaughan, T. A. and Czaplewski, N. J. (1985). Reproduction in Stephen's woodrat: the wages of folivory. *Journal of Mammalogy* **66**, 429–443.
- Vermeirssen, E. L. M., Scott, A. P., and Liley, N. R. (1997) Female rainbow trout urine contains a pheromone which causes rapid rise in plasma 17α,20β-dihydroxy-4-pregnen-3-one levels and milt amounts in males. *Journal of Fish Biology* **50**, 107–119.
- Vernet-Maury, E. (1980). Trimethylthiazoline in fox feces: a natural alarming substance for the rat. In *Proceedings of the VIIth International Symposium on Olfaction and Taste*, ed. H. van der Starre, p. 407. London: IRL Press.
- Vernet-Maury, E., Polak, E. H., and Demael, A. (1984). Structure–activity relationship of stress-inducing odorants in the rat. *Journal of Chemical Ecology* **10**, 1007–1018.
- Vickers, N. J. (2000). Mechanisms of animal navigation in odor plumes. *Biological Bulletin of the Marine Biological Laboratory* **198**, 203–212.
- Vieuille-Thomas, C. and Signoret, J. P. (1992). Pheromonal transmission of an aversive experience in domestic pig. *Journal of Chemical Ecology* **18**, 1551–1557.
- Viitala, J., Korpimäki, E., Palokangas, P., and Kolvula, M. (1995). Attraction of kestrels to vole scent marks visible in ultraviolet light. *Nature* **373**, 425–427.
- Vince, M. A. and Billing, A. E. (1986). Infancy in the sheep: the part played by sensory stimulation in bonding between ewe and lamb. In *Advances in Infancy Research*, vol. IV, ed. L. P. Lipsitt and C. Rovee-Collies, pp. 1–37. Norwood, NJ: Ablex.
- Vince, M. A. and Ward, T. M. (1984). The responsiveness of newly born Clun forest lambs to odour sources in the ewe. *Behavior* **89**, 117–127.

- Vince, M. A., Lynch, J. J., Mottershead, B. E., Green, G. C., and Elwin, R. L. (1987). Interactions between normal ewes and newly born lambs deprived of visual, olfactory and tactile sensory information. *Applied Animal Behaviour Science* **19**, 119–136.
- Vittoria, A. and Rendina, N. (1960). Fattori condizionanti la funzionalita tiaminica in piante superiori e cenni sugli effetti della bocca dei ruminanti sulle erbe pascolative. *Acta Medica Veterinaria* **6**, 379–403.
- Voigt, C. C. (2002). Individual variation in perfume blending in male greater sac-winged bats. *Animal Behaviour* **63**, 907–913.
- Voigt, C. C. and von Helversen, O. (1999). Storage and display of odor by male *Saccopterys bilieata* (Chiroptera; Emballonuridae). *Behavioral Ecology and Sociobiology* **47**, 29–40.
- vom Saal, F. S., Nagel, S. C., Palanza, P., Boechler, M., Parmigiani, S. and Welshons, W. V. (1995). Estrogenic pesticides: binding relative to estradiol in MCF-7 cells and effects of exposure during fetal life on subsequent territorial behavior in male mice. *Toxicology Letters* 77 343–350.
- von Frisch, K. (1941). Über einen Schreckstoff der Fischhaut and seine biologische Bedeutung. Zeitschrift für vergleichende Physiologie **29** 46–145.
- Wabnitz, P. A., Bowie, J. H., Tyler, M. J., Wallace, J. C., and Smith, B. P. (1999). Aquatic sex pheromone from a male tree frog. *Nature* **401**, 444–445.
- Waldman, B. (1981). Sibling recognition in toad tadpoles. The role of experience. Zeitschrift für Tierpsychologie **56**, 341–358.
- Waldvogel, J. A. (1987). Olfactory navigation in homing pigeons: are the current models atmospherically realistic? *Auk* **104**, 369–379.
- Waldvogel, J. A., Phillips, J. B., and Brown, A. I. (1988). Changes in the short-term deflector loft effect are linked to the sun compass of homing pigeons. *Animal Behaviour* **36**, 150–158.
- Wallace, P. (1977). Individual discrimination of humans by odor. *Physiology and Behavior* **19**, 577–579.
- Wallraff, H. G. (1988a). Olfactory deprivation in pigeons: examination of methods applied in homing experiments. *Comparative Biochemistry and Physiology A, Comparative Physiology* **89**, 621–630.
 - (1988b). Navigation by means of an olfactory map and a sun compass: the homing ability of pigeons. *Naturwissenschaften* **75**, 380–392.
 - (1989). Simulated navigation based on unreliable sources of information (Models on pigeon homing. Part 1). *Journal of Theoretical Biology* **137**, 1–19.
 - (1990). Navigation by homing pigeons. Ethology, Ecology and Evolution 2, 81.
 - (2004). Avian olfactory navigation: its empirical foundation and conceptual state. Animal Behaviour 67, 189–204.
- Wallraff, H. G. and Foà, A. (1981). Pigeon navigation: charcoal filter removes relevant information from environmental air. *Behavioral Ecology and Sociobiology* **9**, 67–77.
- Wallraff, H. G. and Neuman, M. F. (1990). Contribution of olfactory navigation and nonolfactory pilotage to pigeon homing. *Behavioral Ecology and Sociobiology* **25**, 293–302.
- Wallraff, H. G., Papi, F., Ioale, P., and Benvenuti, S. (1986). Magnetic fields affect pigeon navigation only while the birds can smell atmospheric odors. *Naturwissenschaften* **73**, 215–217.

- Walls, S. C., Mathis, A., Jaeger, R. G., and Gergits, W. F. (1989). Male salamanders with high-quality diets have faeces attractive to females. *Animal Behaviour* **38**, 546–548.
- Walsberg, G. E. (1975). Digestive adaptations of *Phainopepla nitens* associated with the eating of mistletoe berries. *Condor* 77, 169–174.
- Walther, F. R. (1978). Mapping the structure of the marking system of a territory of the Thomson's gazelle. *East African Wildlife Journal* **16**, 167–176.
- Wang, D., Chen, P., and Halpern, M. (1987). Further isolation and purification of the garter snake chemoattractant in earthworm wash. *Chemical Senses* **12**, 706.
- Wang, J. and Provenza, F. D. (1996). Food deprivation affects preference of sheep for foods varying in nutrients and a toxin. *Journal of Chemical Ecology* **22**, 2011–2021.
- Ward, D. and Young, T. P. (2002). Effects of large mammalian herbivores and ant symbionts on condensed tannins of *Acacia drepanolobium* in Kenya. *Journal of Chemical Ecology* **28**, 921–937.
- Ward, J. F., MacDonald, D. W., and Doncaster, C. P. (1997). Responses of foraging hedge-hogs to badger odour. *Animal Behaviour* **53**, 709–720.
- Ware, G. C. and Gosden, P. E. (1980). Anaerobic microflora of the anal sac of the red fox (*Vulpes vulpes*). *Journal of Chemical Ecology* **6**, 97–102.
- Warner, T. F. and Azen, E. A. (1988). Tannins, salivary proline-rich proteins and oesophageal cancer. *Medical Hypotheses* **26**, 99–102.
- Watkins, J. F., II, Gehlbach, F. R., and Kroll, J. C. (1969). Attractant–repellent secretions of blind snakes (*Leptotyphlops dulcis*) and their army ant prey (*Neivamyrmex nigrescens*). *Ecology* **50**, 1098–1102.
- Webb, J. K. and Shine, R. (1992). To find an ant: trail-following in Australian blindsnakes (Typhlopidae). *Animal Behaviour* **43**, 941–948.
- Webster, D. B. (1973). Audition, vision, and olfaction in kangaroo rat predator avoidance. *American Zoologist* **13**, 1346.
- Webster, D. R. and Weissburg, M. J. (2001). Chemosensory guidance cues in a turbulent chemical odor plume. *Limnology and Oceanography* **46**, 1034–1047.
- Weitzman, S. H. and Fink, S. V. (1985). Xenurobryconin phylogeny and putative pheromone pumps in glanduloaudine fishes. *Smithsonian Contributions to Zoology* **421**, 1–121.
- Wekesa, K. S. and Lepri, J. J. (1992). Removal of the vomeronasal organ impairs reproduction in male prairie voles. *Chemical Senses* 17, 718.
- Welch, B. L. and McArthur, E. D. (1981). Variation of monoterpenoid content among subspecies and accessions of *Artemisia tridentata* grown in a uniform garden. *Journal of Range Management* 34, 380–384.
- Welch, B. L., McArthur, E. D., and Davis, J. N. (1983). Mule deer preference and monoter-penoids (essential oils). *Journal of Range Management* **36**, 485–487.
- Welch, B. L., Pederson, J. C., and Rodriguez, R. L. (1989). Monoterpenoid content of sage grouse ingesta. *Journal of Chemical Ecology* **15**, 961–969.
- Weldon, P. J. (1982). Responses to ophiophagous snakes by snakes of the genus *Thamnophis. Copeia* **1982**, 788–794.
 - (2004). Defense anointing: extended chemical phenotype and unorthodox ecology. *Chemoecology* **14**, 1–4.

- Weldon, P. J. and Burghardt, G. M. (1979). The ophiophage defensive response in crotaline snakes: extension to new taxa. *Journal of Chemical Ecology* **5**, 141–151.
- Weldon, P. J. and Fagre, D. B. (1989). Responses by canids to scent gland secretions of the western diamondback rattlesnake (*Crotalus atrox*). *Journal of Chemical Ecology* **15**, 1589–1604.
- Weldon, P. J. and Schell, F. M. (1984). Responses by king snakes (*Lampropeltis getulus*) to chemicals from colubrid and crotaline snakes. *Journal of Chemical Ecology* **10**, 1509–1520.
- Weldon, P. J. and Williams J. A. (1988). Rathke's glands: pattern of secretion discharge and tests of antipredator activity. *American Zoologist* **28**, 162A.
- Weldon, P. J., Divita, F. M., and Middendorf, G. A., III (1987). Responses to snake odors by laboratory mice. *Behavioral Processes* **14**, 137–146.
- Weldon, P. J., Swenson, D. J., Olson, J. K., and Brinkmeier, W. G. (1990). The American alligator detects food chemicals in aquatic and terrestrial environments. *Ethology* **85**, 191–198.
- Weldon, P. J., Graham, D. P., and Mears, L. P. (1993). Carnivore fecal chemicals suppress feeding by alpine goats (*Capra hircus*). *Journal of Chemical Ecology* **19**, 2947–2952.
- Weldon, P. J., Aldrich, J. R., Klun, J. A., Oliver, J. E., and Debboun, M. (2003). Benzo-quinones from millipedes deter mosquitoes and elicit self-anointing in capuchin monkeys (*Cebus* spp.). *Naturwissenschaften* **90**, 301–304.
- Weller, L. and Weller, A. (1993). Human menstrual synchrony: a critical view. *Neuroscience* and Behavioral Review 17, 427–439.
- Wells, M. C. and Bekoff, M. (1981). An observational study of scent marking in coyotes, *Canis latrans. Animal Behaviour* **29**, 332–350.
- Welsh, R. G. and Müller-Schwarze, D. (1989). Experimental habitat scenting inhibits colonization by beaver, *Castor canadensis*. *Journal of Chemical Ecology* **15**, 887–893.
- Wenzel, B. M. (1968). Olfactory prowess of the kiwi. Nature 220, 1133-1134.
 - (1986). The ecological and evolutionary challenges of procellariiform olfaction. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze and R. M. Silverstein, pp. 357–368. New York: Plenum.
- Wenzel, B. M. and Sieck, M. H. (1966). Olfaction. *Annual Review of Physiology* 28, 381–432.
- Werner, D. I., Baker, E. M., Gonzalez, E. D. C., and Sosa, I. R. (1987). Kinship recognition and grouping in hatchling green iguanas. *Behavioral Ecology and Sociobiology* **21**, 83–90.
- Wheeler, J. W. (1977). Properties of compounds used as chemical signals. In *Chemical Signals in Vertebrates*, vol. 1, ed. D. Müller-Schwarze and R. M. Silverstein, pp. 61–70. New York: Plenum.
- White, P. J., Kreeger, T. J., Tester, J. R., and Seal, V. S. (1989). Anal-sac secretions deposited with feces by captive red foxes (*Vulpes vulpes*). *Journal of Mammalogy* **70**, 814–816.
- White, S. M., Flinders, J. T., and Welch, B. L. (1982). Preference of pygmy rabbits (*Brachylagus idahoensis*) for various populations of big sagebrush (*Artemisia tridentata*). *Journal of Range Management* **35**, 724–726.

- Whitten, W. K. (1958). Modification of the estrous cycle of the mouse by external stimuli associated with the male. Changes in the estrous cycle determined by vaginal smears. *Journal of Endocrinology* **17**, 307–313.
- Whitten, W. K., Wilson, M. C., Wilson, S. R., *et al.* (1980). Induction of marking behavior in wild red foxes (*Vulpes vulpes*) by synthetic urinary constituents. *Journal of Chemical Ecology* **6**, 49–55.
- Widowski, T. M., Ziegler, T. E., Elowson, H. M., and Snowdon, C. T. (1990). The role of males in the stimulation of reproductive function in female cotton-top tamarins, *Saguinus o. oedipus. Animal Behaviour* **40**, 731–741.
- Williams, B. L., Brodie, E. D., Jr., and Brodie E. D., III (2004). A resistant predator and its toxic prey: persistence of newt toxin leads to poisonous (not venomous) snakes. *Journal of Chemical Ecology* **30**, 1901–1919.
- Williams, J. R., Slotnick, B. M., Kirkpatrick, B. W., and Carter, S. B. (1992a). Olfactory bulb removal affects partner preference development and estrus induction in female prairie voles. *Physiology and Behavior* **52**, 635–639.
- Willliams, J. D., Holland, K. N., Jameson, D. M., and Bruening, R. C. (1992b). Amino acid profiles and liposomes: their role as chemosensory information carriers in the marine environment. *Journal of Chemical Ecology* **18**, 2107–2115.
- Williams, J. R. and Lenington, S. (1993). Factors modulating preferences of female house mice for males differing in t-complex genotype: role of t-complex genotype, genetic background, and estrous condition of females. *Behavior Genetics* 23, 51–58.
- Willis, C. M., Church, S. M., Guest, C. M., et al. (2004). Olfactory detection of human bladder cancer by dogs: proof of principle study. British Medical Journal 329, 712.
- Wilson, E. O. (1970). Chemical communication within animal species. In *Chemical Ecology*, ed. E. Sondheimer and J. B. Simeone, pp. 133–155. London: Academic Press.
- Wilson, E. O. and Bossert, W. H. (1963). Chemical communication among animals. *Recent Progress in Hormone Research* **19**, 673–716.
- Wilson, H. C. (1987) Female axillary secretions influence women's menstrual cycles: a critique. *Hormones and Behavior* **21**, 536–546.
- Wiltschko, R., and Wiltschko, W. (1987). Pigeon homing: olfactory experiments with inexperienced birds. *Naturwissenschaften* 74, 94–96.
 - (1992). The effect of temporary anosmia on orientation behavior. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 435–442. New York: Plenum.
- Wiltschko, W., Wiltschko, R. and Mathias, J. (1987a). The orientation behavior of anosmic pigeons in Frankfurt am Main, West Germany. *Animal Behaviour* **35**, 1324–1333.
- Wiltschko, W., Wiltschko, R., and Walcott, C. (1987b). Pigeon homing: different effects of olfactory deprivation in different countries. *Behavioral Ecology and Sociobiology* **21**, 333–342.
- Wiltschko, R., Wiltschko, W., and Kowalski, U. (1989). Pigeon homing: an unexpected effect of treatment with a local anesthetic on initial orientation. *Animal Behaviour* 37, 1050–1052.

- Winberg, S. and Olsén, H. (1992). The influence of rearing conditions on the sibling odour preference of juvenile Arctic charr, *Salvelinus alpinus* L. *Animal Behaviour* **44**, 157–164.
- Wines, M. (2004). For sniffing out land mines, a platoon of twitching noses. *New York Times*, 18 May 2004, pp. A1, A7.
- Wingate, L. P. (1956). Mambas strong smell. African Wildlife 10, 256–257.
- Wirsig, C. R. and Leonard, C. M. (1985). Terminal nerve damage affects hamster mating behavior. *Chemical Senses* **10**, 423.
- Witmer, L. M. (2001). Nostril position in dinosaurs and other vertebrates and its significance for nasal function. *Science* **293**, 850–854.
- Wolff, J. O. and Davis-Born, R. (1997). Response of gray-tailed voles to odours of a mustelid predator: a field test. *Oikos* **79**, 543–548.
- Wong, G. T. and Gannon K. S. (1996). Transduction of bitter and sweet taste by gustducin. *Nature* **381**, 796–800.
- Wood, W. F. (1990). New components in defense secretion of the striped skunk, *Mephitis mephitis*. *Journal of Chemical Ecology* **16**, 2057–2065.
 - (2004). Straight and branched-chain falty acids in preorbital glands of sika deer, *Cervus nippon. Journal of Chemical Ecology* **30**, 479–482.
- Wood, W. F., Morgan, G. C., and Miller, A. (1991). Volatile components in defensive spray of the spotted skunk, *Spilogale putorius*. *Journal of Chemical Ecology* 17, 1415–1420.
- Wood, W. F., Fisher, C. O., and Graham, G. A. (1993). Volatile components in defensive spray of the hog-nose skunk, *Conepatus mesoleucus*. *Journal of Chemical Ecology* **19**, 837–841.
- Woolhouse, A. D., Weston, R. J., and Hamilton, B. H. (1994). Analysis of secretions from scent-producing glands of brushtail possum (*Trichosurus vulpecula*). *Journal of Chemical Ecology* **20**, 239–253.
- Wrangham, R. W. and Nishida, T. (1983). *Aspilia* spp. leaves: a puzzle in the feeding behavior of wild chimpanzees. *Primates* **24**, 276–282.
- Wrangham, R. W. and Waterman, P. (1981). Feeding behavior of vervet monkeys on *Acacia tortilis* and *Acacia xanthoploea* with special reference to reproductive strategies and tannin production. *Journal of Animal Ecology* **50**, 715–731.
- Wright, J. and Weldon, P. J. (1990). Responses by domestic cats (*Felis catus*) to snake scent gland secretions. *Journal of Chemical Ecology* **16**, 2947–2953.
- Wuensch, K. L. (1982). Effect of scented traps on captures of Mus musculus and Peromyscus maniculatus. Journal of Mammalogy **63**, 314–315.
- Würdinger, I. (1979). Olfaction and feeding behavior in juvenile geese (Anser a. anser and Anser domesticus). Zeitschrift für Tierpsychologie **49**, 132–135.
- Würdinger, V. I. (1990). Die Reaktionen von Zebrafinken (*Taeniopygia guttata*) auf Düfte: eine Pilotstudie. *Die Vogelwarte* **35**, 359–367.
- Wyatt, T. D. (2003). *Pheromones and Animal Behaviour*. Cambridge, UK: Cambridge University Press.
- Wyatt, T. D., Phillips, D. G., and Grégoire, J.-C. (1993). Turbulence, trees and semiochemicals: windtunnel orientation of the predator, *Rhizophagus grandis*, to its barkbeetle prey, *Dendroctonus micans*. *Physiological Entomology* **18**, 204–210.

- Wysocki, C. J. (1989). Vomeronasal chemoreception: its role in reproductive fitness and physiology. In *Neural Control of Reproductive Function*, ed. J. Lakoski, B., Haber, R. Perez-Polo and D. Rossin, pp. 545–566. New York: Alan Liss.
- Wysocki, C. J., Beauchamp, G. K., and Erisman, S. (1980). Access of compounds to the vomeronasal organ in pine and meadow voles. *Proceedings of the Fourth Eastern Pine and Meadow Vole Symposium*, February 1980, Hendersonville, New York, pp. 20–23.
- Wysocki, C. J., Nyby, J., Whitney, G., Beauchamp, G., and Katz, Y. (1982). The vomeronasal organ: primary role in mouse chemosensory gender recognition. *Physiology and Behavior* **29**, 315–327.
- Wysocki, C. J., Katz, Y., and Bernhard, R. (1983). The male vomeronasal organ mediates female-induced testosterone surges. *Biology of Reproduction* **28**, 917–922.
- Wysocki, C. J., Beauchamp, G. K., Reidinger, R. R., and Wellington, J. L. (1985). Access of large and nonvolatile molecules to the vomeronasal organ of mammals during social and feeding behaviors. *Journal of Chemical Ecology* 11, 1147–1160.
- Wysocki, C. J., Bean, N. J., and Beauchamp, G. K. (1986). The mammalian vomeronasal system: its role in learning and social behaviors. In *Chemical Signals in Vertebrates*, vol. 4, ed. D. Duvall, D. Müller-Schwarze and R. M. Silverstein, pp. 471–485. New York: Plenum.
- Xiao-Nong Zeng, J., Leyden, J., Lawley, H. J., *et al.* (1991). Analysis of characteristic odors from human male axillae. *Journal of Chemical Ecology* 17, 1469–1492.
- Xu, Z., Stoddart, M., Ding, H., and Zhang, J. (1995). Self-anointing behavior in the rice-field rat, *Rattus rattoides*. *Journal of Mammalogy* **76**, 1238–1241.
- Yahr, P. (1977). Central control of scent marking. In *Chemical Signals in Vertebrates*, vol. 1, ed. D. Müller-Schwarze and M. M. Mozell, pp. 549–562. New York: Plenum Press.
- Yamamori, K., Nakamura, M., Matsui, T., and Hara, T. J. (1988). Gustatory responses to tetrodotoxin and saxitoxin in fish: a possible mechanism for avoiding marine toxins. *Canadian Journal of Fisheries and Aquatic Science* **45**, 2182–2186.
- Yamamoto, K., Kawai, Y., Hayashi, T., *et al.* (2000). Silefrin, a sodefrin-like pheromone in the abdominal gland of the sword-tailed newt, *Cynops ensicauda*. *FEBS Letters* **472**, 267–270.
- Yamazaki, K., Beauchamp, G. K., Matszaki, O., et al. (1986). Participation of the murine X and Y chromosomes in genetically determined chemosensory identity. Proceedings of the National Academy of Sciences, USA 83, 4438–4440.
- Yamazaki, K., Beauchamp, G. K., Kupniewski, D., *et al.* (1988). Familiar imprinting determines H-2 selective mating preferences. *Science* **240**, 1331–1332.
- Yamazaki, K., Beauchamp, G. K., Bard, J., and Boyse, E. A., (1990). Chemosensory identity and the Y chromosome. *Behavior and Genetics* **20**, 157–165.
- Yamazaki, K., Beauchamp, G. K., Imai, Y., Bard, J., and Boyse, E. A. (1992a). Expression of urinary H-2 odortypes by infant mice. *Chemical Senses* **17**, 723.
- Yamazaki, K., Beauchamp, G. K., Imai, Y., *et al.* (1992b). MHC control of odortypes in the mouse. In *Chemical Signals in Vertebrates*, vol. 6, ed. R. L. Doty and D. Müller-Schwarze, pp. 189–196. New York: Plenum.

- Yasumoto, T. and Murata, M. (1993). Marine toxins. Chemistry Review 93, 1897-1909.
- Yasumoto, T., Yasumara, D., Yotsu, M., *et al.* (1986). Bacterial production of tetrodotoxin and anhydrotetrodotoxin. *Agricultural and Biological Chemistry* **50**, 793–795.
- Yoerg, S. I. (1991). Social feeding reverses learned flavor aversions in spotted hyenas (*Crocuta crocuta*). *Journal of Comparative Psychology* **105**, 185–189.
- Yoshihara, Y., Nago, H., and Mori, K. (2001). Sniffing out odors with multiple dendrites. *Science* **291**, 835–837.
- Yousem, D. M., Maldjian, J. A., Siddiqui, F., et al. (1999). Gender effects on odor-stimulated functional magnetic resonance imaging. Brain Research 818, 480–487.
- Yun, S-S., Scott, A. P., and Li, W. (2003). Pheromones of the male sea lamprey, *Petromyzon marinus* L.: structural studies on a new compound, 3-keto allocholic acid, and 3-keto petromyzonol sulfate. *Steroids* **68**, 297–304.
- Zala, S. M. and Penn, D. J. (2004). Abnormal behaviors induced by chemical pollution: a review of the evidence and new challenges. *Animal Behaviour* **68**, 649–664.
- Zeng, C., Spielman, A. I., Vowels, B. R., et al. (1996a). A human axillary odorant is carried by apolipoprotein D. Proceedings of the National Academy of Sciences of the USA 93, 6626–6630.
- Zeng, X.-N., Leyden, J. J., Lawley, H. J. *et al.* (1991). Analysis of characteristic odors from human male axillae. *Journal of Chemical Ecology* **17**, 1469–1492.
- Zeng, X.-N., Leyden, J. J., Brand, J. G., *et al.* (1992). An investigation of human apocrine gland secretion for axillary odor precursors. *Journal of Chemical Ecology* **18**, 1039–1055.
- Zeng, X.-N., Leyden, J. J., Spielman, A. I., and Preti, G. (1996b). Analysis of characteristic human female axillary odors: qualitative comparison to males. *Journal of Chemical Ecology* **22**, 237–257.
- Zheng, W., Strobeck, C., and Stacey, N. E. (1997). The steroid pheromone $17\alpha,20\beta$ -dihydroxy-4-pregnene-3-one increases fertility and paternity in goldfish. *Journal of Experimental Biology* **200**, 2833–2840.
- Ziegler, T. E., Epple, G., Snowdon, C. T., *et al.* (1993). Detection of the chemical signals of ovulation in the cotton-top tamarin, *Saguinus oedipus*. *Animal Behaviour* **45**, 313–322.
- Zielinski, B., Arbuckle, W., Belanger, A., et al. (2003). Evidence for the release of sex pheromones by male round gobies (*Neogobius melanstomus*). Fish Physiology and Biochemistry **28**, 237–239.
- Zielinski, W. J. and Barthalmus, G. T. (1989). African clawed frog skin compounds: antipredatory effects on African and North American watersnakes. *Animal Behaviour* **38**, 1038–1086.
- Zimmerling, L. L. and Sullivan, T. P. (1994). Influence of mustelid semiochemicals on population dynamics of the deer mouse (*Peromyscus maniculatus*). *Journal of Chemical Ecology* **20**, 667–689.
- Zobel, A. M. and Brown, S. A. (1990). Dermatitis-inducing furanocoumarins on leaf surfaces of eight species of rutaceous and umbelliferous plants. *Journal of Chemical Ecology* **16**, 693–700.
- Zucker, W. V. (1983). Tannins: does structure determine function? An ecological perspective. *American Naturalist* **121**, 335–365.

INDEX

Aardvark (Orycteropus afer) 153	Air currents 6 Alarm
Aardwolf (<i>Proteles cristatus</i>) 23, 31, 153, 154, 159	-
Acacia	odors 191
	responses, fish 192 substance 192
fever tree (Acacia xanthophloea) 312	"alarm signals, damage-released" 192
umbrella thorn (A. tortilis) 312 whistling thorn (A. drepanolobium) 333	Albatross
Acanthochromis polyacanthus (see Damselfish) Accipenser baeri (see Sturgeon, Siberian)	black-browed (<i>Diomedea melanophris</i>) 74, 352 black-footed (<i>D. nigripes</i>) 114, 350
Accipenser gueldenstaedtii (see Sturgeon, Russian)	grey-headed (D. chrysostoma) 352
Accipenser stellatus (see Sturgeon, stellate)	wandering (D. exulans) 352
Acetate esters 26	Alcelaphus cokii (see Hartebeest, Coke's)
Acetone 372	Alces alces (see Moose)
6-Acetonylisoxanthopterin 194	Aldehydes in coyote lures 411
Acetophenone 190	Alder, green (Alnus crispa) 299, 300, 309
Achillea ligustica (see Yarrow, Ligurian)	induced defense 332
Achillea millefolium (see Yarrow)	Alert odors 191
Acidification of water, effects 392	Alfalfa (Medicago sativa) 278, 286, 307
Acomastylis [Geum] rossii (see Aven, alpine)	Algae (Shewanella sp.) 247
Acomys cahirinus (see Mouse, spiny)	Alkaloids,
Acomys russatus (see Mouse, golden spiny)	bioaccumulation 253
Acrocephalus schoenobaenus (see Warbler, sedge)	birds 50
Actinomycetes 66	poison dart frogs 49, 252
"Active signalers" 172	properties, occurrence 280
Active space 9, 33, 57	Alligator, American (Alligator mississipiensis) 349
Adaptations (herbivory)	Allocholic acid 66, 172
defensive 315	Allomarking 148
offensive 315	Almond, bitter (Prunus amygdalus) 291
Adrenocortical effects, mice 220	Alnus crispa (see Alder, green)
Aepyceros melampus (see Impala)	Alouatta belzebul (see Monkey, red-handed
Aeschna juncea (see Dragonfly)	howler)
Aeschna umbrosa (see Dragonfly)	Alouatta palliata (see Monkey, howler)
Aethia cristatella (see Auklet, crested)	Alpaca (Lama pacos) 140
Agaricus sp. (see Mushroom)	Amanita muscaria (see Mushroom, fly agaric)
Agelaius phoeniceus (see Blackbird, red-winged)	Amanita phalloides (see Mushroom, death cap)
Agnatha 171	Amazona amazonica (see Macaw, orange-winged)
Agouti (Dasyprocta leporina) 389	Ambystoma spp. (see Salamander)
Agrimonia sp. (see Agrimony)	Amines, biogenic 255
Agrimony (Agrimonia sp.) 266	Amino acids
Agropyron smithii (see Grass, western wheat)	as feeding deterrents 21
Ailuropoda melanoleuca (see Panda, giant)	barbel nerve response 339

non-protein 283	Anoplopoma fimbria (see Sablefish)
predator odors, fish 358	Anser anser (see Goose, greylag)
prey odors 339	Ant (Neivamyrmex sp.) 375
salmon fry feeding cues 342	Antarctica 60
y-Aminobutyric acid 245	Antechinus, brown (Antechinus stuartii) 103
2-amino-4', 5'-methoxyacetophenone 397	Antechinus stuartii (see Antechinus, brown)
Aminopropionitrile 283	Antifeedants
Ammonia 414	birds 394
Amniotic fluid 138	conspecific odors 407
dog 139	mammals 401
rat 139, 237	plant compounds 404
sheep 139, 409	selenium 406
species specificity 237	Antilocapra americana (see Pronghorn)
Amphibians 38, 49, 68, 98, 133, 145, 201	Antilope cervicapra (see Blackbuck)
behavior development 228	Antipredator ploy 196
defense 250, 259	Antipredator responses 18
kin recognition 129	snakes 229
odor thresholds 114	without predators 377
pre-mating isolation mechanisms 198	Ants as diet 253
prey odors 342	Anurans 68
priming pheromones 207	Apigenin 284
sex pheromones 176	Apis dorsata (see Honey bee)
territorial pheromones 152	Aplodontia rufa (see Mountain beaver)
VNO 96	Aplysinopsin 384
Amphibolurus ornatus (see Lizard)	Apocrine secretion binding proteins 26
Amphikuemin 384	Apodemus sylvaticus (see Mouse, wood)
Amphiprion ocellaris (see Fish, anemone)	Apolipoprotein D (apoD) 26
Amphiprion perideraion (see Fish, anemone)	Aposematic coloration 252
Amphisbaenan (Blanus cinereus) 348	Aposematism, olfactory 317
Amygdalin 281, 291	Apteryx sp. (see Kiwi)
Amyl acetate 408	Aquatic environment 15
Anadenanthera peregrina (see Yopo tree)	Arabidopsis thaliana 282
Anas platyrhynchos (see Mallard)	Arbacia punctulata (see Sea urchin)
Anchovy	Arborvitae, false (Hiba) (<i>Thujopsis dolabrata</i>) 275
inshore (nehu) (Stolephorus purpureus) 339,	Area odors 13
341	Area repellents 397
northern (Engraulis mordax) 339, 341	Arenicola marina (see Lugworm)
Androgenized fetuses 235	Arginine as attractant 374
Androgens in plants 288	Army worm (Spodophora exempta) 282
Andropogon gerardii (see Grass, big bluestem)	Aromatic plants in birds' nests 265, 378
4,16-androstadien-3-one 22, 27, 119	Arothron hispidus (see Fish, puffer,
Andropogon sp. (see Salsify)	white-spotted)
5α -androst-16-en-3 α -ol 48, 54, 55, 119, 187,	Artemia sp. (see Shrimp, brine)
211, 389, 406	Artemisia spp. (see Sagebrush)
5α-androst-16-en-3-one 27, 48, 54, 55, 118,	Arthole 310
119, 187, 211	Artibeus literatus (see Bats)
16-androstenes	Asarum caudatum (see Ginger, wild)
boar 32, 119	Asclepias spp. (see Milkweed)
pig fetuses 236	Aspen, quaking (Populus tremuloides) 299, 300, 304
Androstenol 48, 410 Androstenone 48, 410	induced defense 332
Angelfish (<i>Pterophyllum scalare</i>), priming 205	Aspilia (Aspilia mossambicensis) 381
Angelicoidenol 311	Asterropteryx semipunctatus (see Goby, starry fin)
Anguilla anguilla (see Eel) Anolis carolinensis (see Anole)	Astralagus bisulcatus (see Vetch, milkvetch, 2-grooved)
1110113 (4101111013) (300 1111010)	2-g100vcaj

picillata) 5,
oicillata) 5,
oicillata) 5,
oicillata) 5,
oicillata) 5,
, 3,
271
, 371
256, 375,
375
) 9, 17, 31,
) 9, 17, 31,
) 9, 17, 31,
) 9, 17, 31,
) 9, 17, 31,
) 9, 17, 31,
) 9, 17, 31,
) 9, 17, 31,
) 9, 17, 31,
) 9, 17, 31,
9, 17, 31,
201
201
201
201
201
201
201 7
201
201 7
201 7
201 7
201 7

Betula spp. (see Birch)	Blood 48
Bifidobacterium 53	Blood flukes (Trematoda) 373
Bile	schistosome (Schistosoma mansoni) 373
acids 66, 237	Boar (Sus scrofa) 32
salts 66	pheromone 116
Biotransformation of xenobiotics 322, 328	scent marking 148
acidemia 331	Bobcat (Felis rufus) 402
conjugation 330	Bobwhite, northern (See Quail, bobwhite)
degradation speed 331	Body bridging, rattlesnakes 364
microorganisms' role 332	Body odors 48, 49
oxidation 329	Body region odors 48
sodium loss 331	Boiga irregularis (see Snake, brown tree)
Birch (Betula spp.) 330	Boletus piperatus (see Mushroom, peppery
Alaska paper (B. neoalaskana) 337	bolete)
Arctic white (B. tortuosa) 301	Bombykol 117
canoe (paper B. papyrifera) 337	Bonasa umbellus (see Grouse, ruffed)
downy (B. pubescens) 310, 333, 334	Borneol 275
European white (<i>B. pendula</i>) 285, 301, 310,	Bornyl acetate 307
333, 334	
	Boundary layers 15
paper (B. resinifera) 299, 300 induced defense 332	Bouteloua curtipendula (see Grass, sideoats
	grama) Poutolous gracilis (see Crass, blue grama)
phenolics 285	Bouteloua gracilis (see Grass, blue grama)
Birds 42, 50, 71, 87, 153, 257	Brachylanio rerio (see Danio, zebra)
antifeedants 394	Brachylagus idahoensis (see Rabbit, pygmy)
aromatic plant material use 265	Brachyteles arachnoids (see Monkey, muriqui)
behavior development 231	Bracken, common (<i>Pteridium aquilinum</i>) 282
capsaicin	Branta canadensis (see Goose, Canada)
chemosensory foraging 349	Breath odor
contact repellents 396	humans 191, 365
defense of their prey 266	rats 197
detoxication of plant food 322	Broad tuning 91
embryo response to odors 231	Bruce effect 217
feeding on plants 304	Buckthorn (Rhamnus cathartica) 306, 384, 395
homing 71	Buffalo, African (Syncerus caffer) 372
methyl anthranilate	Bufo spp. (see Toad)
odor thresholds 114	Bullfinch (<i>Pyrrhula pyrrhula</i>) 305, 307
palatability 259	Bullhead catfish (Ictalurus nebulosus) 48, 49,
pheromone candidates 182	340
seed conditioning 387	Buried seeds 354, 356
seed passage 386	Busycon contrarium (see Whelk)
social odors 143	Butanoic acid 114
trigeminal nerve 107	(E)-2-Butene-1-thiol 24, 30, 32, 262
Birth control, herbs 287	(E)-2-Butenylmethyl disulfide
(+)-α-Bisabolol 300	(S)-(E)-2-Butenyl thioacetate 24, 262
Bison, North American (Bison bison) 383	2-sec-Butyl-4,5-dihydrothiazole 37, 149
Bitterbrush, antelope, Purshia tridentata 5	Butyrate esters, tamarin 168
Blackbird	Butyric acid
European (Turdus merula) 306, 384	children's tolerance 421
Red-winged (Agelaius phoeniceus) 305, 307,	dogs' threshold for 115
395	feeding stimulant, sheep 408
Blackbuck, Antilope cervicapra 4, 148	bat-pollinated flowers 389
Blarina brevicauda (see Shrew, short-tailed)	vampire bat attractant 371
Blarinatoxin 263	-
Blennius pavo (see Blenny)	Cactus lophophora (see Peyote)
Blenny (Blennius pavo) 38	Cadaverine 347

Caffeic acid 289	Carrot, wild (Daucus carota) 266
Caffeine 303	Carvacrol 275
Calidris canutus (see Knot)	(4R)-Carvone 22
Calidris maritima (see Sandpiper)	(4S)-Carvone 22
Caliosobruchus chinensis (see Beetle)	Cassava (see Manioc)
Callianassa affinis (see Shrimp, ghost)	Castor canadensis (see Beaver, North American)
Callinectus sapidus (see Crab, blue)	Castor fiber (see Beaver, Eurasian)
Calloselasma rhodostoma (see Pitviper)	Castration effect on odor detection 120
Calluna vulgaris (see Heather)	Catastomidae 175
Calvaria major (see Tambalacoque tree)	Cat, domestic 120, 137, 256, 257, 259
Calvatia gigantea (see Mushroom, giant puffball)	catmint 275
Camphene 310, 322	Catbird (Dumatella carolinensis) 386
Camphor 310, 322	Catfish 16, 91, 340
Campostoma anomalum (see Stone roller)	African (Glarias gariepinus) 206
L-Canavanine 283	channel (Ictalurus punctatus) 340, 342
Cancer, esophageal 288, 289	priming pheromones 206
Candida kruzei 53	sex attractant 173, 174
Canids 55, 154	Catha edulis (see Khat)
Canis familiaris (see Dog)	Catharacta maccormicki (see Skua, South Polar)
Canis latrans (see Coyote)	Cathartes spp. (see Vulture)
Canis lupus (see Wolf)	Catmint (Nepeta cataria) 275
Cannabis	Catostomus macrocheilus (see Sucker, largescale)
Cannabis indica 290	Cattle 18, 19, 48, 216, 257, 323, 324
C. sativa 290, 292	antifeedants 407
Cannibalism 120	calf odor in cross-fostering 408
Capra hircus (see Goat)	estrus synchronization 409
Capreolus capreolus (see Roedeer)	gland secretions 29
Capsaicin 21, 397, 398	oestrus odor 190
Capybara (Hydrochaeris hydrochaeris) 52, 57	perineal and neck glands 52
Carassius carassius (see Carp, crucian)	puberty acceleration 409
Carassius auratus (see Goldfish)	seed conditioning 386
Carbachol 413	sex attractants 185
Carbamate 412	tannins 311
Carbamylcholine chloride 413	toxic plants 280, 282
Carbon dioxide 372	tsetse fly 372
Carbon disulfide as food cue 197, 406	Cavia aperea (see Cavy)
Carbonyl sulfide 197, 406	Cavia sp. (see Guinea pig)
Carcinogens 288	Cavy (Cavia aperea) 134
Cardenolides 279	Cebus olivaceus (see Monkey, wedge-capped
Cardiac glycosides 264, 279	capuchin)
Carduelis cannabina (see Linnet)	Cedar, red (<i>Thuja plicata</i>) 402
Caretta caretta (see Turtle, loggerhead)	Centrocercus urophasianus (see Grouse, Sage)
Caribou (Rangifer tarandus) 81	Cephalostachyum ef. uiguieri (see Bamboo,
Caribou tarandus (see Reindeer)	cyanogenic)
Carica papaya (see Papaya)	Cercopithecus aethiops (see Monkey, vervet)
Carnivores 128, 184, 201	Cervus spp. (see Deer)
Carollia perspicillata (see Bats, short-tailed fruit)	Ceryle rudis rudis (see Kingfisher, pied,
Carp	palatability)
Cyprinus carpio 17, 18, 175	Chaconine 301
amino acids as food cues 340, 342	Chalcides ocellatus (see Lizard, ocellated skink)
priming, milt production 205	Char
crucian (Carassius carassius) 359	Arctic (Salvelinus alpinus = Salmo alpinus) 65
grass (Ctenopharyngodon idella) 341, 342, 385	66, 110, 111, 114, 130, 142, 227
Carpinus betulus (see Yoke-elm)	amino acids as attractants 341
Carrion odorants 114	predator odors 358
CHILICH CHUIGHING III	predator odoro JJU

brook (Salvelinus fontinalis) 111 lake (Salvelinus namaycush) 111	southern quahog (Mercenaria campechiensis) 340
Charicidae 20, 49	Claviceps sp. (see Ergot)
Chelodina longicollis (see Turtle, eastern	Clethrionomys spp. (see Vole)
	Clostridium 53
long-necked) Chelonia midas (see Turtle, green sea)	Clover 307
Chelonia midas (see Turtle, green sea)	"clover disease" 286
Chelydra serpentina (see Turtle, snapping)	
Chemical fish lures 392	subterranean (Trifolium subterraneum) 273, 286
Chemical imprinting 392	
Chemical search image 339	white (Trifolium repens) 286, 293
"Chemical spies" 172	Clupeus harengus pallasi (see Herring, Pacific)
"Chemical stranger" 49	Cobrotoxin 257
Chemoreception 4	Coca (Erythroxylon novogratense) 290, 292
and age 120	Cod, Atlantic (Gadus morhua) 86, 339,
gravity effects 15	341
hormonal influences 119	Coevolution
humidity effects 5	plants and herbivores 334
maturation 234, 236	predator–prey 268
radiation effects 14	Coffea arabica (see Coffee)
Chen caerulescens atlantica (see Goose, greater	Coffee (Coffee arabica) 302
Snow)	Coliforni hagtoria 52
Chicken, domestic (Gallus gallus domesticus) 73,	Coliform bacteria 53
116	Colohus virginianus (see Quail, bobwhite)
avoidance of salamanders 251	Colobus polykomos (see Monkey, black-and-white
Children 125	colobus)
odor preferences 239	Colosthetus sp. (see Frog)
Chimpanzee (<i>Pan troglodytes</i>), clay eating 326	Coluber constrictor (see Snake, racer)
Chinning	Commelina spp. (Dayflower) 381
rabbits 48	Competition 268
tree shrews 26	Competitive exclusion 268
Chin touching, salamanders 39	Conditioned flavor aversion 121, 405
Chipmunk	Conditioned taste aversion 260
Eastern (Tamias striatus) 5, 11	Conepatus mesoleucus (see Skunk, hog-nosed)
Siberian (Eutamias sibiricus asiaticus) 376, 377	Coniferyl benzoate 272, 287, 305
yellow pine (Tamias amoenus) 5, 357	Coniine 280
Cholestanol 181	Conium maculatum (see Hemlock)
Cholesterol 40, 149	Connochaetes spp. (see Wildebeest)
Christ's thorn (Ziziphus spina-christi) 287	Conservation 363, 396, 411
Chrysemys picta (see Turtle, painted)	Convallaria majalis (see Lily-of-the-valley)
Chrysemys (Pseudemys) scripta elegans (see Turtle,	Cottocomephorus grewingki (see Sculpin,
red-eared)	yellow-finned Baikal)
Chub	Coprine 281
hornyhead (Nocomis biguttatus) 175	Coprinus atramentarius (see Inky, alcohol)
speckled (Hybopsis aestivalis) 358, 359	Coragyps atratus (see Vulture, black)
Cichlasoma severum (see Cichlid, severum)	Coronella austriaca (see Snake, smooth)
Cichlids	Corvus corax (see Raven)
severum (Cichlasoma severum) 359	Cottontail, eastern (Sylvilagus floridanus)
tiger oscar (Astronotus ocellatus) 359	bioassay 313
Ciguatoxin 247	food choice 276
1,8-Cineole 307, 310, 322	interspecific responses 412
(E)-Cinnamaldehyde 412	Cottus carolinae (see Sculpin, banded)
Cistus ladanifer (see Rockrose)	Coturnix coturnix (see Quail, Japanese)
Citral 232	p-Coumaric acid 286
Clam	Coumestrol 286
pill (Musculium rosaceum) 343	Countermarking 165

Cowbird, brownheaded (Molothrus ater) 114,	Dace
116, 395	common (Leuciscus leuciscus) 194
Coyote (Canis latrans) 23, 37, 48, 402	finescale (<i>Phoxinus neogaeus</i>) 20, 359
chemical lures 411	pearl (Semotilus margarita) 194, 206
odor effect 368, 407	Dactylis glomerata (see Orchardgrass)
prey detection 355	Dactylopsila palpator (see Triok, long-fingered)
scent-rolling 256	Damselfish (Acanthochromis polyacanthus) 194
urine marking 31, 159, 167	Danaus plexippus (see Monarch butterfly)
Crab, blue (Callinectus sapidus) 340	Dandelion (<i>Taraxacum officinale</i>) 307
Crane, greater sandhill (<i>Grus canadensis</i>) 412	Danio
Crassostrea virginica (see Oyster)	giant (Danio malabricus) 194
Crataegus monogyna (see Hawthorn)	zebra (D. <i>rerio</i>) 173
Crayfish	Danio spp. (see Danio)
Oronectus rusticus 360	Daphnane 277
Procambarus clarki 363	Daphne (Daphne mezereum) 277, 360
Creeper, blue-bell (Sollya heterophylla) 306	Daphne mezereum (see Daphne)
Creosote bush (<i>Larrea tridentata</i>) 299	Daphnetoxin 277
p-Cresol 149	Daption capense (see Petrel, pintado)
Cricetomys gambianus (see African/Gambian giant	Darter, redline (Etheostoma ruflineatum) 343
pouched rat)	Darwin 238
Critical periods 241, 243	Dasyurus geoffroyii (see Marsupials,
Crocidura russula (see Shrew, greater	western quoll)
white-toothed)	Daucus carota (see Carrot, wild)
Crocodiles 41, 349	14-Deacetylnudicauline 280
Crocuta crocuta (see Hyena, spotted)	Deadnettle, purple (Lamium purpureum) 266
Cross-Appalachian tracer experiment	"Dear enemy" phenomenon 127
(ĈAPTEX) 79	Decahydroquinolines 252
Cross-fostering 140	(Z)-4-Decenal 182
Crotalus spp. (see Rattlesnake)	Dedicated motor patterns 83
Cryptomys damarensis (see Mole-rat,	Deer lactone 22, 37
Damaraland)	Deer 215
Ctenopharyngodon idella (see Carp, grass)	black-tailed (Odocoileus hemionus columbianus)
Culaea inconstans (see Stickleback, brook)	antifeedants 402
Cuy (Cavia sp.) 35	complex scent 49
3-Cyanoalanine 283	conditioned aversion 405
Cyanocitta cristata (see Jay, blue)	"deer lactone" 37
Cyanogenic glycosides 281, 298	feeding 315, 406
Cycads	flehmen 102, 185
queen sago (Cycas circinalis) 289	isomer discrimination 22
queen sago (Cycas rumphii) 289	maternal imprinting 243
Cycas circinalis (see Cycads, queen sago)	metatarsal odor 195, 264
Cycas rumphii (see Cycads, queen sago)	osmetrichia 56
Cycasin, biotransformation 332	predator odor responses 367, 368
1, 2-Cyclohexadione 300	rub-urination 57
Cyclohexanol 67	scent glands 44
Cyclopamine 288	tarsal organ 45
Cyclopia, sheep 288	terpenoid effects 285
Cycloposine 288	visual signals 34
Cyclosmates (fish) 16, 83	mule (Odocoileus hemionus)
p-Cymene 310, 330	feeding 310, 315, 406
Cynodon plectostachyus (see Grass, star) Cynops spp. (see Newt)	juniper defense 312 metatarsal gland 5
	proline-rich salivary proteins 323,
Cyprinidae 20, 175 Cypriniformes 20	324
Cyprinus carpio (see Carp)	tannin binding 324
Syprimus curpio (see Carp)	tallilli bilidilig 324

tarsal organ 45	Dimethyl disulfide
winter diet 310, 406	deer antifeedant 403
musk (Moschus sp.) 153	golden hamster 189
pampas (Ozotoceros bezoarticus) 298	rats 238
red (Cervus elaphus) 57	3,3-Dimethyl-1,2-dithiolane 403
antifeedants 275, 402, 403, 404	3,4-Dimethyl-1,2-dithiolane 184
wapiti (Cervus elaphus canadensis) 368, 399	5,5-Dimethyl-2-ethyltetrahydrofuran-2-ol
water (Hydropotes sp.) 159	210
white-tailed (Odocoileus virginianus)	2,5-Dimethyl-3-methylene-1,5-heptadine 306
female behavior 184	3,5-Dimethyl-2-octanone, wolf 32, 190
female odors 38	N,N-Dimethyl-2-phenylethylamine 351
food choice 276	2,5-Dimethylpyrazine 37, 213
metatarsal gland 5	Dimethyl sulfide 351
repellents 399, 402	Dimethyl tetrasulfide 403
rubs 150, 187	2,3-Dimethylthietane 184, 370, 403, 412
rutting behavior 150	Dimethyl trisulfide 403
scrapes 150, 187	Dinoflagellates (Gambierdiscus toxicus) 247
tarsal organ 45	Dioclea megacarpa (see Sea purse)
urine volatiles 37	Diomedea spp. (see Albatross)
visual display 34	Dipodomys spp. (see Kangaroo rat)
Dehydroepiandrostenone sulfate 226	Dipsosaurus dorsalis, (see Iguana, desert)
3, 4-Dehydro- <i>exo</i> -brevicomin 37, 149	Directional smelling 87, 122
Delphinium spp. (see Larkspur)	Discrimination
Dendroaspis sp. (see Snake, mamba)	population 142
Dendrobates spp. (see Frog, dart-poison)	species 142
Deoxycorticosterone 21	Dispersion
2-Deoxyglucose 242	coefficient 10
Desert animals 81	Gaussian plume model 10, 11
Desmognathus spp. (see Salamander)	time-average (Sutton) model 10, 11, 12
Desmothus rotundus (see Bats)	Display, "superiority" (muskox) 149
Dhurrin 281, 282, 291	Distichlis stricta (see Grass, salt)
Diadophis punctatus (see Snake, ring-necked)	"Disturbance signals" 192
2, 4-Diaminobutyric acid 283	Disulfides 23
2, 6-Diamino-4-oxodihydropteridine 194	Dodecanoic acid 27
Dicrostonyx torquatus (see Lemming, collared)	(Z)-7-Dodecen-1-yl acetate (elephant) 26, 33,
Didelphis virginiana (see Oppossum, Virginia)	186, 190
Diet	Dodecyl propionate 141
effects on odor or secretions 49, 50, 138,	Dodo (Raphus cucullatus) 385
253	Dodo tree (see Tambalacoque tree)
effect on predator odor 370	Dog (Canis familiaris)
quality signals, salamanders 176	air scenting 115, 414
Digestion inhibition 284	amniotic fluid 139
(see Foxglove)	anal sac compounds 23
Digitalis sp. 279	cow odor discrimination 190
Digitoxin 279	direction of track 417
Dihydroaplysinopsin 384	discriminating odors of twins 417
17α,20β-Dihydroxy-4-pregnen-3-one	disease odors 417
$(17\alpha, 20\beta P)$, goldfish 114, 174, 204	indole 38
Dihydrotestosterone 191	police 415, 417
2,5-Dihydro-2,4,5-trimethylthiazoline	response to odor mixtures 118
403	scent glands 49
Dik-dik (Madoqua kirki) 149	sensitivity to venom 257
Dimethyl anthranilate 394	tracking 5, 115, 414
(9E)-1,10-Dimethyl-9-decalol (geosmin) 66,	Dogfish 16
118	Dogrose (Rosa canina) 306
	,

Domestication 95	Esox niger (see Pickerel, chain)
Dominance area, deer 150	Estrus synchronization
Dopamine 194	cattle 409
Dorosoma petenense (see Shad, threadfin)	humans 224
Dragonfly	mice 214
Aeschna juncea 362	Ethanethiol 114
A. umbrosa 195	Etheostoma ruflineatum (see Darter, redline)
Dromedary, long-distance food detection 6	4-Ethyloctanoic acid 32, 190, 217
Drymarchon corais (see Snake, eastern indigo)	4-Ethyl-oct-2-enoic acid 190
Ducks, domestic 143	3-Ethyl phenol 372
Dumatella carolinensis (see Catbird)	4-Ethyl phenol 168, 190, 372
Dytiscus verticalis (see Beetle, diving)	Etiocholanolone glucuronide, gobi 174
Dyvicence (etc Decirc, art mg)	Eubacterium 53
Early experience, mammals 242	Eublepharis macularius (see Gecko, leopard)
Earthquake odors, animal response 418	Eucalyptus spp.
Earthworm (Lumbricus sp.) as prey 344	effect on gland secretion 50
Eavesdropping 374	herbivore metabolism 330
Ebéna (Virola theiodora) 292	herbivores, avoidance 306
Eel (Anguilla anguilla) 16, 66, 67, 113, 142,	in koala diet 297, 334
227	Eumeces spp. (see Lizard)
Eddies 8	Euphausia superba (see Krill, Antarctic)
Eddy diffusion 7	Eupomacentrus partitus (see Fish, bi-color
Eft, red (Notophthalmus viridens) 251	damsel)
Egg predation 412	Eurycea spp. (see Salamander)
Elaphe spp. (see Snake)	Eutamias sibiricus asiaticus (see Chipmunk,
Elasmobranchs 84	
	Siberian) Evalution of pheromone communication, 199
Elder (Sambucus nigra) 306 Electro-olfactogram (EOC) fish 175	Evolution of pheromone communication 199 Exaptation 179, 268
Electro-olfactogram (EOG), fish 175 Elephant 6, 26, 33, 52, 84, 185	Excretions 36
African (Loxodonta africana) 190, 311, 386	Extended phenotype 376
Asian (Elephas maximus) 32, 186, 190	Extended phenotype 370
Elephas maximus (see Elephant, Asian)	Falco tinnunculus (see Kestrel)
	Familiarization 125
Elk (see deer, red) Embryonic malformations 288	carnivores 125
	mice 125
Emetin dihydrochloride 413 Emodin 395	pronghorn 125
Emys orbicularis (see Turtle, European pond)	rodents 125
Enantiomers 22, 118	Fanaloka (Fossa fossa) 370
Endocrine disruptors 421	$(E,E)-\alpha$ -Farnesene 37, 149
	(E) - β -Farnesene 37, 149
Engraulis mordax (see Anchovy, northern) Enhydra lutris (see Otter, sea)	Fatty acids 27, 40, 54, 116, 226, 414
Environmental effects on odor thresholds 119	Feather lice (Phthiraptera), bird toxin
Enzymes, induced 330	effect 262
Ephemeral ponds, predation ecology 362	Feces 37, 152, 153
Epidermal growth factor 383	Feeding behavior 18
Equids 148	Feeding stimulants 406
Equus caballus (see Pony)	Felis concolor (see Mountain lion)
Ergot (Claviceps sp.) 290	Felis rufus (see Bobcat)
Erigeron sp. (see Flea bane)	Felis wiedi (see Margay)
Erinaceus europaeus (see Hedgehog)	Ferrets 29, 134, 367 Mustela fura 128, 184, 259
Eryobdella punctata (see Leech, red)	Mustela furo 128, 184, 259 chemical lures 411
Erythroxylon novogratense (see Coca) Escherichia coli 53, 237, 267	
	prey odor imprinting 244
Esox americanus (see Pickerel, redfin) Esox lucius (see Pike)	Ficus exasperata 381 Finch, zebra (Taeniopygia guttata) 306
LOUA IULIUS (SEE FIRE)	1'111C11, ZCD1a (14CHUDYYYU YUUUUU) 300

Fir, Douglas (Pseudotsuga menziesii) 402, 405, 406	rabbit (Spilopsyllus cuniculi) 376 Xenopsylla dipodilli 373
Fireflies (<i>Photinus</i> spp.) 315	Flea bane (Erigeron sp.) 266
Fish 17, 29, 49, 61, 84, 113, 130	Fleas (Siphonaptera) 373
acidification effects 392	Flehmen 101, 119, 184, 185, 186
anadromous 61	Fluoroacetate 335
anemone	Food
	caches
Amphiprion ocellaris 384	birds 354
Amphiprion perideraion 384	
behavior development 227	rodents 356
benefits of clay 328	flavor, chickens 396
blind cave 142	odors, seabirds 353
Astyanax mexicanus 142	preferences 303
Caecobarbus geertsii 142	preservatives, pika 382
Phreatichthys andruzzi 142	processing 319
box (Ostraction sp.) 250	Foraging, buried seeds 5
damsel, bi-color (Eupomacentrus partitus) 266	Forest
deep-sea (Gonostoma bathyphilus,	air flow 12
Gonostomatidae) 87	breeze 12
defense 246, 250	edges 12
feeding attractants 392	quiet zone 13
female pheromones 174	Formononetin 286, 287
kin recognition 129	Fossa fossa (see Fanaloka)
lures, chemical 392	Fox, red (Vulpes vulpes)
male pheromones 172	anal gland secretion as antifeedant 403
mosquito	bacteria in anal sacs 53
Gambusia affinis 230	as egg predator 413
Gambusia patruelis 358, 359, 363, 393	feces and urine 21, 160, 402
olfactory epithelium 86	odor effect on mammals 48, 378, 407
olfactory organ 85	response to artificial scent marks 168
pheromone hypothesis of migration 62	scent marks 159
priming pheromones 203	sex difference, fecal marking 167
puffer (Tetraodontidae) 247, 250	sulfide 30
white-spotted ("death") (Arothron	supracaudal gland 56
hispidus) 247	TMT 37
reef 266	urine marking and food 167
"scent marking" 151	Foxglove 279
seed conditioning 385	purple (Digitalis purpurea) 405
sequential imprinting 62	Freemartins 235
sex pheromones 172	Fregetta tropica (see Storm-petrel, black-bellied)
"sniffing" ("coughing") 83	Frog
sole	African clawed (Xenopus laevis) 253
peacock (Pardachirus pavoninus) 248,	American bull (<i>Rana catesbiana</i>) 255, 362
250	Australian turtle (Myobatrachus gouldii) 343
Red Sea Moses (P. marmoratus) 248, 250	cascade (Rana cascadae) 129
species discrimination 142	Colosthetus sp. 253
	dart-poison (Dendrobatidae) 49, 259
stream imprinting 62 tannic aid effects 328	Dendrobates auratus 253
toxic	Dendrobates pumilio 68
actively 248	Phyllobates aurotaenia 252
passively 247	Phyllobates bicolor 252
water pollution effects 393	Phyllobates terribilis 252
Flavonoids 273	European common brown (Rana temporaria),
Flea	VNO 97, 98, 362
Parapulex chephrenis 373	European edible (Rana esculenta) 68, 361

Frog (cont.)	Genet (Genetta genetta) 201
gray treefrog (Hyla chrysoscelis) 360, 361	Genetic modification, manioc 283
green (Rana clamitans) 68, 360, 362	Genistein 286, 287
magnificent tree (Litoria splendida), sex	Genotypic polymorphism, snakes 229
pheromone 40, 176, 177	Geochelone spp. (see Tortoise)
northern leopard (Rana pipiens) 343	Geophagy 324
pool (Rana lessonae) 29, 68, 361	humans 326
red-legged (Rana aurora) 363	Geosmin 66, 118
spotted chorus (<i>Pseudacris clarki</i>) 68	Gephyrotoxins 252
Strecker's chorus (<i>Pseudacris streckeri</i>) 68	Gerbil 53, 199
western chorus (<i>Pseudacris triseriata</i>) 68	Asia Minor (Meriones tristrami) 287
wood (Rana sylvatica) 362	Indian (Tatera indica) 161
Frontalin, elephant 22, 32	Mongolian (Meriones unguiculatus), 44
Fruits	diet-dependent odors 50, 51
flavor, birds 384	Harderian gland 48
odor 118	hormones and marking 166
	individual discrimination 134
plant metabolites 302	keeper substance 30
Fugu (puffer fish)	
Fugu pardalis 247	mate recognition 134
F. rubripes 247	osmetrichia 56
Extract porthern (Extractive decidie) 250, 251	paternal behavior 141
Fulmar, northern (Fulmarus glacialis) 350, 351	polypeptides 168
Functional groups 20	as prey 355
Furocoumarins 293, 294	scent marking 160, 183
Fusobacterium 53	spatial orientation 80
	Tatera spp. 5
Gadus morhua (see Cod, Atlantic)	Wagner's gerbil (Gerbillus dasyurus) 373
Galago	Gerbillus dasyurus (see Gerbil, Wagner's)
Allen's squirrel (Galago alleni) 357	Gerenuk (Litocranius walleri) 159, 160, 166
Demidoff's dwarf (Galagoides demidovii)	Germacrone 276, 312
357	Gerrhosaurus spp. (see Lizard)
Galagoides demidovii (see Galago, Demidoff's	Gila monster
dwart)	Hyloderma horridum 258, 259
Galidia elegans (see Mongoose, Madagascar	H. suspectum 258, 259
ring-tailed)	Gilatoxin 258
Gallic acid 272	Ginger, wild (Asarum caudatum) 405
Gallus gallus domesticus (see Chicken, domestic)	Gland
Gambierdiscus toxicus (see Dinoflagellates)	abdominal, salamanders 39, 176
Gambusia affinis (see Fish, mosquito)	anal
Gambusia patruelis (see Fish, mosquito)	beaver 31, 158, 167
Gases, effect on chemoreception 14	birds 42
Gastrolobium sp. 335	fox 160
Gastrophyne carolinensis (see Toad,	hog-nosed skunk 30
narrow-mouthed)	mongoose 29, 32
Gaultheria shallon (see Salal)	otter 122
Gazella spp. (see Gazelle)	spotted skunk 32
Gazelle	striped skunk 32
Dorcas (Gazella dorcas) 389	vole 51
seed conditioning 386	anal fin appendage, fish 38
Thomson (G. thomsoni) 159, 160	anogenital, meadow vole 33
Gecko 366	apocrine 43, 44, 191
leopard (Eublepharis macularius) 181	axilla, humans 27, 44, 191
western banded (Coleonyx variegatus)	axillary (Rathke's), turtles 41
181	caudal, fish 38
Genes, odorant receptor 93	caudal peduncle, fish 172

cheek, mongoose 29	rostral, frogs 177
chest, galago 28	sebaceous 43
chin	secretions, seasonal variation 27
rabbit 149	and reproductive state 27
tortoises 181	snout ("morilla"), capybara 52
cloacal	sternal, possum 51
birds 42	subauricular, pronghorn 30, 44, 45,
crocodiles 41	167
salamanders 39	subcaudal, badger 27
turtles 41	variation of secretion 27
cornual organ, goat 32	sublingual, shrew 263
crural venom, platypus 262	submaxillary, shrew 263
dorsal	supracaudal, red fox 56
crocodiles 41	tarsal, deer 44, 45
hyrax 35	temporal, elephant 32, 52
pronghorn 167	ventral, Mongolian gerbil 30, 44, 51
femoral, lizards 40, 52, 126	Glandular secretions 38
iguana 29, 52, 123, 126	Glarias gariepinus (see Catfish, African)
flank, golden hamster 52, 104, 147, 167	Glaucomys sabrinus (see Squirrel, flying)
foot, mouse 58	Glomerular convergence 93
granular, toads 254	Glomeruli 93
Harderian, rodents 48, 52	Glossina spp. (see Tsetse fly)
hedonic, woodland salamanders 39	Glucose 194
holocrine 44	Glutamate 245
infraorbital, muskox 149	Glycoalkaloids 301
inguinal, turtles 41	
. •	Goat (<i>Capra hircus</i>) 32, 323, 324 flehmen 185
interdigital, reindeer 196	
ischiadic, pronghorn 35 lateral, turtles 41	male odor compounds 190
	maternal imprinting 243 novel foods 317
mandibular, crocodiles 41 mental	odor "labeling" 244
salamanders 58, 176	predator odor effects 407
turtles 41	priming pheromones 219
mesorchial, fish 174	tannins 311
metacarpal, boar 148	toxic diet 297
metatarsal, Odocoileus 4, 44, 195	Goatfish, Hawaiian (Parupeneus porphyreus)
mucus, amphibians 38	339
musk, crocodiles 41	Gobius jozo (see Goby, black)
nasolabial, plethodontid salamanders	Goby
39	black (Gobius jozo) 173, 174
neck, cattle 52	blind (Othonops eos, formerly Typhlogobius
nucho-dorsal, snakes 41 oral, monitor lizards 376	californiensis) 172
•	starry fin (Asterropteryx semipunctatus) 194
parotoid	Goldenrod, rough (Solidago rugosa) 266, 26
frogs 177	Goldfish (<i>Carassius auratus</i>) 175, 192
toads 254	female pheromones 173, 174
pelvic, salamanders 39	odor threshold 114
perineal, cattle 52	as prey 343
posterolateral region, meadow vole 33	priming pheromones 203, 206
pre-anal, snakes 41	Gonostoma bathyphilus (see Fish, deep-sea)
preen (uropygial), birds 42, 182, 262	Gonostomatidae (see Fish, deep-sea)
preorbital, gazelle 160	Gopherus polyphemus (see Tortoise, Gopher)
preputial	Gourami
boar 148	blue (Trichogaster trichopterus) 198
rat 141	snakeskin (T. pectoralis) 198

Goose	Hamster
Canada (Branta canadensis) 305, 307, 395	Djungarian (Rhodopus sungorus) 211
greater snow (Chen caerulescens atlantica) 305,	golden (Mesocricetus auratus)
307	aphrodisin 105, 189
greylag (Anser anser) 231, 305	body region odors 136
Gopher, pocket (<i>Thomomys talpoides</i>) 397, 403	chemosensory systems 104
Grackle, boat-tailed (Quiscalus major) 395	dimethyl disulfide 189
Grandiflorenic acid 381	dominance 147
Grapholitha molesta (see Moth, oriental fruit)	flank gland 167, 188
Grass 281	hormone effects 52
big bluestem (Andropogon gerardii) 285	individual discrimination 128
foxtail bristle (Setaria italica) 306	long-lasting scent marks 170
grama	mating behavior, timing 183
blue (Bouteloua gracilis) 285	mounting pheromone 24, 189
sideoats (Bouteloua curtipendula) 383	over-marking 165
Indian rice (Oryzopsis hymenoides) 389	
	predator odor response 48
Kentucky blue (<i>Poa pratensis</i>) 305	priming 219
little bluestem (Schizachyrium scoparium =	scent marking rhythm 189
Andropogon scoparius) 285	spatial orientation 80
reed canary (Phalaris arundinacea) 294	tannin effects 285, 324
salt (Distichlis stricta) 286, 380	testosterone and olfactory
star (Cynodon plectostachyus) 282	performance 120
timothy (Phleum pratense) 293	vaginal secretion 104, 189
western wheat-(Agropyron smithii) 285	VNO and behavior 104, 106
Grasshopper (<i>Poekilocerus bufonius</i>) 280	Hapalemur aureus (see Lemur, golden bamboo)
Grasshoppers, saliva 383	Hardenbergia comptoniana (see Vine, lilac)
Grayling (Thymallus thymallus) 66	Hare
Grosbeak, black-headed (<i>Pheucticus</i>	
	European (Lepus europaeus)
melanocephalus) 268	feeding 285
Grouse	sodium loss 332
ruffed (Bonasa umbellus) 299, 301, 304,	mountain (Lepus timidus) 299
307	detoxication 332
sage (Centrocercus urophasianus) 310, 322	feeding 285, 299, 309
Growth rate, plants 333	phenols 312
Growth suppression, voles 285	snowshoe (Lepus americanus) 299, 300, 309
Grus canadensis (see Crane, greater sandhill)	antifeedants 403, 404, 405
Guam ALS-PDC (disease) 289	browsing and induced defense 332
Guinea pig (<i>Cavia</i> sp.) 35, 50, 51, 52	green alder defense 312
bitter substances 316	diffuse coevolution 336
VNO stimulation 104, 106	population cycles 336
Gull, glaucous-winged (Larus glaucescens)	"resistance centers" 336
266	Hartebeest, Coke's (Alcelaphus cokii), VNO 98
Gulo gulo (see Wolverine)	Hasler's fish orientation hypotheses 61
Guppies, tannin effect 328	Hawthorn (Crataegus monogyna) 306
Guppy, Trinidadian (Poecilia reticulata)	Heather (Calluna vulgaris) 296
alarm odor 192, 194	Hedera helix (see Ivy)
antipredator response 360	Hedgehog 377
Gymnocorymbus ternetzi (see Tetra, black)	African pygmy (four-toed) (Atelerix
Gypsy moth (Lymantria dispar) 117	pruneri) 267, 377
* *	Egyptian (desert) (Hemiechinus auritus) 267,
Habitat imprinting 393	377
Haementeria officinalis (see Leech, Mexican)	European (Erinaceus europaeus) 18, 55, 257,
Hair loss, livestock 283	259, 366, 377
Hallucinogens 290, 292	Heloderma heloderma horridus (see Lizard, Mexican
Halobaena caerulea (see Petrel, blue)	bearded)
11 DIAC	bourdou _j

Helogale undulata rufula (see Mongoose, African	estrus synchronization 224 fetus
dwarf)	
Hemicentetes semispinosus (see Tenrec, streaked)	odor learning 236
Hemiechinus auritus (see Hedgehog, Egyptian)	odor responses 235
Hemlock (Conium maculatum) 280	follicular phase odors 225 food aversions 419
Hemlock, eastern (Tsuga canadensis) 402	
Heptadecanoic acid 27	individual odors 136
2-Heptanone 213	individual recognition 239
(E)-4-Hepten-2-one 213	infants' "scent marking" 171
(E)-5-Hepten-2-one 213	kin recognition 132
Heron, black-crowned night (Nycticorax	menstrual phase odors 225
nycticorax) 255	movies with scent 422
Herpailurus yagouaroundi (see Jaguarundi)	odors in advertising 421
Herpestes spp. (see Mongoose)	odor thresholds 118
Herring, Pacific (Clupeus harengus pallasi) 172	palatability of birds 260
Heterocephalus glaber (see Mole-rat, naked)	postnatal odor responses 238
Heterodon nasicus (see Snake, hognose)	priming effects, search for 224
Heteromyid rodents 389	puberty 224
1,2-Hexadecanediol 216	sex odors 191, 419
Hippopotamus 57	track odors 415
Histrionicotoxins 252	trigeminal nerve 107, 108
Hoatzin (Opisthocomus hoazin)	Humboldt, Alexander von 60, 354
body odor 143, 262	Humidity 4, 30
fatty acid production 322	Hyaena brunnea (see Hyena, brown)
Homing	Hyaenids 55, 153
birds, to burrow 71	Hydrohates palacicus (see Phytrol Pritich storm)
pigeons 74	Hydrobates pelagicus (see Petrel, British storm)
Homobatrachotoxin, birds 261	Hydrochaeris hydrochaeris (see Capybara)
Homogentisic acid 50	Hydrogen sulfide 118, 420
Homo sapiens (see Humans) Honey badger (Ratel: Mellipora capensis), 252	Hydropotes sp. (see Deer, water)
Honey badger (Ratel; Mellivora capensis) 352 Honey bee (Apis dorsata) 352, 354	6-Hydroxycyclohexenone 300
Honey guide (Indicatoridae) 352	6-Hydroxy-6-methyl-3-heptanone 106, 210
greater (Indicator indicator) 352	11 <i>β</i> -Hydroxytestosterone 5-Hydroxytryptamine 194
least (Indicator exilis) 352	Hyena
scaly-throated (Indicator variegatus) 352	brown (Hyaena brunnea), 29, 45, 49
Honeysuckle, Tatarian (<i>Lonicera tatarica</i>) 386	scent marking 122, 153, 154, 159
Hoopoe (Upupa epops) 42, 144, 262	spotted (Crocuta crocuta) 154
Hormonal control of odors 52	scent marking 159
Hormone, luteinizing 215	social unlearning of aversion 414
Hormone-to-pheromone hypothesis 200	Hyla chrysoscelis (see Frog, grey treefrog)
Hornet (Vespa orientalis) 259, 260	Hyla crucifer (see Spring peeper)
Horse 184, 257, 293, 407	Hyloderma horridum (see Lizard, Mexican
Horsenettle, Carolina (Solanum carolinense) 302	beaded)
Human (Homo sapiens) 32, 44, 133	Hyloderma suspectum (see Gila monster)
androstenol in axillary secretion 226	Hypericum perforatum (see St. Johnswort)
aversion therapy 419	Hypomesus pretiosus (see Smelt, surf)
axillary odors	Hypothalamic dopamine system 222
effects 225	Hypoxanthine-3N-oxide 194
infants' response 239	Hyrax 35
body smell library 420	Procavia capensis syriaca 295
children's odor tolerance 421	1.5501100 capellosis 5,11000 250
clay eating 326	Ictalurus nebulosus (see Bullhead catfish)
degradation speed, xenobiotics 331	Ictalurus punctatus (see Catfish, Channel)
diet odors 418	Icterus spp. (see Oriole)

Ifrita, blue-capped (<i>Ifrita kowaldi</i>) 261	Kalahari 153
Iguana, desert (Dipsosaurus dorsalis) 41, 83	Kangaroo
femoral secretion 83, 84	Brush-tailed bettong (Bettongia
food finding 98	penicillata) 335
proteins 84	burrowing bettong (Bettongia lesueur) 335
VNO for food detection 97	eastern bettong (Bettongia gaimardi) 335
Iguana, green (Iguana iguana) 29, 52, 131	eastern grey (Macropus giganteus) 335
Iguana iguana (see Iguana)	grey (Macropus fuliginosus) 306, 313, 335
Ilex paraguensis (see Maté)	western grey (Macropus rufus) 335
Immunoglobulins 298	Kangaroo rat
Impala (Aepyceros melampus) 311, 312	banner-tailed (Dipodomys spectabilis) 183
Imprinting	Merriam's (D. merriami) 183, 369
filial 243	Ord's (D. ordii) 380, 381
maternal 243	panamint (D. panamintinas) 5
prey odors 244	and seed conditioning 388
Indicator spp. (see Honey guide)	Kaurenoic acid 381
Individual odors 27	Keeper substances 29, 30
Individual recognition 48	Kestrel (Falco tinnunculus) 354
Indolizidines 253	Khat (Catha edulis) 289, 292
Induced plant defense 332	Killifish (<i>Oryzias latipes</i>) 248
Infanticide 104, 132	Kin recognition 129
Information currents 17	frog, cascades, Rana cascadae 18,
Inky, alcohol (Coprinus atramentarius)	19
281	humans 132
Introduced predators, Australia 366	odor 81
α -Ionone 117	saliva 48
Ircinia sp. (see Sponge)	toad, American 18, 19
Isobutylamine 106	Kingfisher, pied (Ceryle rudis rudis),
2-Isobutyl-3-methoxypyrazine 67	palatability 260
Isoflavonoids 273	Kite, red (Milvus milvus) 412
Isomers 22	Kiwi (Apteryx sp.), prey detection 349
Δ^3 -Isopentenylmethyl sulfide, fox 30	Klipspringer (Oreotragus oreotragus) 375
Isopentenyl pyrophosphate 30	Knot (Calidris canutus) 352
Isopentyl acetate 390	Koala (Phascolarctus cinereus) 297, 334
Isopentyl sulfide, wolf 32, 190	Kob, Uganda (Kobus kob thomasi) 184
Isopropyl acetate 390	Kobus kob thomasi (see Kob, Uganda)
Isosmates 16, 83	Kobus leche kafuensis (see Lechwe, Kafue)
Isovaleric acid 168	Krill 17, 24
Iso-vanillin 22	Antarctic (Euphausia superba) 352
Isovelleral 276, 316	odor 81, 351
Isoxanthopterin 194	Kudu (Tragelaphus strepsiceros) 298, 311,
Ivy (Hedera helix) 306	312
Ixodes spp. (see Ticks)	
	Labeled lines 91
Jaguar (Panthera onca) 366, 369	Labrador tea (Ledum groenlandicum) 276,
Jaguarundi (Herpailurus yagouaroundi) 369	312
Jay	Labriformidin 279
blue (Cyanocitta cristata) 280, 320	Labriformin 279
gray (Perisorius canadensis) 266	Lacerta vivipara (see Lizard, common)
Jervine 288	Lactarius sp. (see Mushroom, lactarius)
Juniper (Juniperus sp.) 310, 319	γ-Lactones 149
alligator (Juniperus deppeana) 310	Lagodon rhomboides (see Pinfish)
Rocky Moutain (J. scopulorum) 310	Lagomorphs 309
Utah (J. osteosperma) 310	feeding inhibitors 309
Juniperus spp. (see Juniper)	population control 399

Lamium purpureum (see Deadnettle, purple)	Life history shifts 363
Lamprey 114	Lignins 274
Lampropeltis spp. (see Snake)	Lily-of-the-valley (Convallaria majalis) 283
Land mine detection 417	Limonene 289, 307, 396
Larkspur (Delphinium sp.) 317, 408	Linamarin 281, 291, 321
duncecap (Delphinium occidentale) 280	Linnet (Carduelis cannabina) 259
tall (D. barbeyi) 314	Lion 31, 37, 368, 403, 407
Larrea tridentata (see Creosote bush)	Lipids, iguana femoral secretion 123
Larus glaucescens (see Gull, glaucous-winged)	Lippia plicata 381
Laser-induced fluorescence 12	Lithium chloride 197, 412, 414
Lathyrus spp. (see Pea)	Litocranius walleri (see Gerenuk)
Latrine sites	Litoria splendida (see Frog, magnificent tree)
badger 164	Livestock 185, 291
hyena 154	Lizard
raccoon dog 164	Amphibolurus ornatus 52
Leaf miner (Perileucoptera coffeella) 303	Anole (Anolis carolinensis) 266
Learning	bearded dragon (Pogona vitticeps) 314
observational 319	black-lined plated (Gerrhosaurus
of odors	nigrolineatus) 181
mice 244	common (Lacerta vivipara) 181, 365
and olfactory bulb 244	response to predatory snake odor 365
rat 241	defense 258
Lechwe, Kafue (Kobus leche kafuensis) 184	defence of prey 266
Lecythis (Lecythis ollaria) 283	foraging 348
Ledum groenlandicum (see Labrador tea)	giant plated (Gerrhosaurus validus) 348
Leech	Iberian wall (Podarcis hispanica atrata) 378
medicinal (Hirudo medicinalis), feeding	Komodo monitor (<i>Varanus komodoensis</i>) 376
cues 374	lace monitor (<i>Varanus varius</i>) 376
Mexican (Haementeria officinalis), as snake	Mexican beaded (Heloderma heloderma
prey 347	horridus) 258
red (Erpobdella punctata), as snake prey 347	monitor (Varanus albigularis) 366
Legumes 283	savanna monitor (Varanus exanthematicus),
Lek species 184	preying 348
Lemming	skink
brown (Lemmus trimoucronatus = sibiricus) 128,	broad-headed (Eumeces laticeps) 143, 348
336	common five-lined (Eumeces fasciatus) 143
collared (Dicrostonyx torquatus) 336	ocellated (Chalcides ocellatus) 348
Lemmus trimoucronatus (= sibiricus; see Lemming,	southeastern five-lined (Eumeces
brown)	inexpectatus) 348
Lemur	VNO 97
black (Lemur macaco) 128, 144	western fence (Sceleporis sp.) 152
brown (L. fulvus) 128, 134, 144	Lobster 16, 17
golden bamboo (Hapalemur aureus) 282	Lonicera tatarica (see Honeysuckle, Tatarian)
mouse (Microcebus murinus)	Lophiomus imhausi (see Rat, African crested)
priming 219	Lophortyx californicus (see Quail, California)
VNO 101, 106	Lotaustralin 281, 291
ring-tailed (Lemur catta), VNO 98	Lotus corniculatus (see Trefoil, birdsfoot)
Lemur spp. (see Lemur)	Louse (Menacanthus sp.), birds 266
Lepidochelys kempi (see Turtle, Kemp's	Loxodonta africana (see Elephant, African)
ridley sea)	Lugworm (Arenicola marina) 339, 341
Lepomis spp. (see Sunfish)	Lumbricus sp. (see Earthworm)
Leptotyphlops dulcis (see Snakes, blind)	Lupines 280
Lepus spp. (see Hare)	Lupinine 280
Lethocerus americanus (see Waterbug, giant)	Luteinizing hormone (LH) 105, 218
Leuciscus leuciscus (see Dace, common)	Luteining hormone-releasing hormone 218

Lutra spp. (see Otter)	skin glands 43
Lycodonomorphus spp. (see Snake water)	species discrimination 144
Lycopersicum cheesmanii (see Tomato, Galapagos)	territorial marking 153
Lymantria dispar (see Gypsy moth)	VNO 98
1 ()1) /	Manduca sexta (see Tobacco hornworm)
Macaca sp. (see Monkey)	Manioc (Manihot esculenta), 282, 298
Macaw, orange-winged (Amazona	processing (detoxification) 321
amazonica) 325	Manihot esculenta (see Manioc)
Macronectes sp (see Petrel, giant)	Map
Macropisthodon (orange-necked keelback) 41	gradient 75
Macropus spp. (see Kangaroo)	mosaic 75
Madoqua kirki (see Dik-dik)	Margay (Felis wiedi) 369
Magpie, black-billed (<i>Pica pica</i>) 116, 354	Marine mammals 357
Main olfactory system 84, 89 Major histocompatibility complex (MHC)	Marking fluid, tiger 31
	Marmosets
odors 132	Callithrix sp. 166
fetal odor type 235	Cebuella sp. 166
individual odors 135	Marmota monax (see Woodchuck)
kin recognition 130	Marsupials 145, 306
mate choice 132, 187	greater glider (Petauroides volans) 330
mice 28	western quoll (Dasyurus geoffroyii) 366
odor learning 243	Marten, pine (Martes martes) 104
peptides 30	scent marking 126
VNO receptor activation 106	Martes martes (see Marten, pine)
Major urinary proteins (MUPs) 24, 25, 149,	Marula (Sclerocarya caffra) 386
218	Maté (Ilex paraguensis) 289
Malacosoma americanum (see Tent caterpillar,	Mate recognition 134
Eastern)	Maternal behavior 120
Mallard (Anas platyrhynchos) 182	female rats 141
Mammals 84, 122, 189	male rats 104
alarm odors 195	Mating balls, garter snakes 178
behavior development 231	Mating plug, garter snakes 180
capsaicin 396, 397	Mauremys leprosa (see Turtle, terrapin)
chemial repellents 399	6-MBOA (6-methoxybenzoxazolinone) 379
diet effects 50	381
diffuse coevolution 335	Medicagenic acid 278
dominance 145	Medicago sativa (see Alfalfa)
food choice 306	Meles meles (see Badger, European)
foraging 355	Mellivora capensis (see Badger, honey)
fungal spores 390	Menacanthus sp. (see Louse, birds)
hormone effects 52	Mephitis mephitis (see Skunk, striped)
kin recognition 129	4-Mercapto-4-methyl-2-pentanone 111
main olfactory system 89	3-Mercaptopropan-1-ol 403
methyl anthranilate 397	3-Mercaptothiopropan-1-ol 403
odor discrimination 126	Mercenaria campechiensis (see Clams, southern
odor thresholds 115	quahog)
orientation 80	Meriones tristrami (see Gerbil, Asia Minor)
parental odors 136	Meriones unguiculatus (see Gerbil, Mongolian)
predator odors 366	Mesocricetus auratus (see Hamster, golden)
priming pheromones 207	Metabolic cost, detoxification 296
recycling animal materials 267	Methacrolein 310
reproduction regulators 379	Methiocarb 396
scent marking evolution 201	Methionine 404
seed conditioning 389	2-Methoxyacetophenone 397
sex odors 183	6-Methoxybenzoxazolinone (see 6-MBOA)
JEA 04013 103	O INICIDO Y DELIZONAZUIIIUIIC (SEE UTIVI DE DE

2-Methoxy-3-isobutyl pyrazine 318	fathead (Pimephales promelas) 20, 175
2-Methoxy-3-methyl-1,4-benzoquinone 377	alarm substance 192
2-Methoxyphenol 372	male pheromone 172
(S)-8-Methoxypsoralen 289	predator recognition 360
β-N-Methylamino-L-alanine 289	as prey 344
Methyl anthranilate 89, 394, 397	Miroestrol 287
2-Methyl-1,4-benzoquinone 377	Misgurnus anguillicaudatus (see Weatherfish,
3-Methyl-1-butanethiol 24, 262	oriental)
(S)-3-Methylbutanylthioacetate 24, 262	Mite, fowl (Ornithonyssus sylviarum) 266
3-Methyl-3-butenyl methyl sulfide 403	Mole-rat
2-Methylbut-2-enal (rabbit mammary	blind (Spalax ehrenbergi) 144, 199
pheromone) 137	Damaraland (Cryptomys damarensis) 208
3-Methylbutylmethyl sulfide 403	naked (Heterocephalus glaber) 207
Methyl capsaicin 398	<i>Molothrus ater</i> (see Cowbird, brownheaded)
5-Methyl-2-furoic acid 50, 51	Monarch butterfly (Danaus plexippus) 268, 279
6-Methyl-5-hepten-2-one (salmon lice	Mongoose 257
attractant) 372	Aegyptian (Herpestes ichneumon) 32
(E)-3-Methyl-2-hexenoic acid 32, 54, 191	African dwarf (Helogale undulata rufula) 29,
(Z)-3-Methyl-2-hexenoic acid 191	134
3, 7-Di-O-methyl kaempferol 284	Indian (Herpestes auropunctatus) 18, 134,
Methyl ketones	413
garter snakes 178	Madagascar ring-tailed (Galidia elegans) 370
gecko 181	Monkey 388
Methyllycaconitine 280, 314	black-and-white colobus (Colobus
Methyl-3-methylbut-3-enyl sulfide 23	polykomos) 311
4-Methyl octanoic acid 217	howler (Alouatta palliata) 311
3-Methyl phenol 372	Macaca sp., degradation speed 331
4-Methyl phenol 190, 372	muriqui (Brachyteles arachnoids) 380
2-Methylquinoline 24	red-handed howler (Alouatta belzebul), clay
2-Methyl-2-thiazoline 232	eating 326
Micelles 17	rhesus (Macaca mulatta) 312
Microalgae (Phaeocystis pouchetti) 351	vervet (Cercopithecus aethiops) 312
Microbial odors 53, 373	wedge-capped capuchin (Cebus olivaceus) 267,
Microcebus murinus (see Lemur, mouse)	377
Microgale dobsoni (see Tenrec, long-tailed)	Monkeys, seed conditioning 386
Micropterus spp. (see Bass)	Monodelphis domestica (see Oppossum,
Microtus spp. (see Vole)	short-tailed)
Milk odor 137	Monoterpenes 404
Milkweed (Asclepiadaceae) 279	Montezuma Wildlife Refuge 400
Indian (Asclepias eriocarpa) 279	Moose (Alces alces) 310, 312, 333
scarlet (bloodflower) (Asclepias	Moray eels (Muraenidae) 248
curassavica) 280	Morone saxatilis (see Bass, striped)
Utah (Asclepias labriformis) 279	Morpholine 29, 61, 113, 114, 231
Millipede (Orthoporus dorsovittatus) 267	Moschus sp. (see Deer, musk)
Milvus milvus (see Kite, red)	Mosesins 248
Mimicry	Mosquitoes 377
Batesian 251, 395	Aedes aegypti 373
Müllerian 318	Anopheles gambiae 373
L-Mimosine 283	control 393
Mimuyi (stomach oil deposits) 153	Moth, oriental fruit (Grapholitha molesta) 26
Mink (Mustela vison) 370, 403, 413	Mountain beaver (Aplodontia rufa) 402, 405
Minnow	Mountain lion (Felis concolor) odor 368, 402, 407
European (<i>Phoxinus phoxinus</i> = $laevis$) 192	Mouse 199
alarm pheromone 79, 194, 206	aboriginal house (Mus spretus) 126
conditioned fright reaction 360	aztec (Peromyscus aztecus) 264, 266

Mouse (cont.) 199	urine mark and aggression 165
black-eared (Peromyscus melanotis) 264, 266	urine marking 183
California (Peromyscus californicus) 141	urine mark, "shelf life" 170
deer (Peromyscus maniculatus) 5, 202	urine volatiles 37
antipredator response changes 378	Whitten Effect 214
estrus urine attracting predators 374	spiny (Acomys cahirinus) 50, 123, 127,
finding buried seeds 356	129, 269
as non-target species, repellents 399	behavior development 237
plant toxins 282	flea attractants 373
Prairie (Peromyscus maniculatus bairdii) 401	golden (<i>Acomys russatus</i>), competition 269
pregnancy block 217	odor in competition 268
puberty delay 212, 213	urinary pheromones (MUPs) 24, 29, 30
seed digestion 302	white-footed (Peromyscus leucopus) 81, 129, 132
sexual priming of males 217	mate choice 188
as snake prey 346	pregnancy block 217
familiarization 125	puberty delay 213
foot glands 58	wood (Apodemus sylvaticus)
harvest	antipredator responses 366, 367, 370
Eurasian (Micromys minutus) 411	pups' odor preferences 242
Sumichrast's (Reithrodontomys	sex odor preferences 401
sumichrasti) 264, 266	trap response 401
house (Mus musculus) 201	see also VNO, rodents
biotransformation, degradation speed 331	Mud snails 16
carbon disulfide as attractant 197	Mugil cephalus (see Mullet, striped)
dominance 145	Mullet, striped (Mugil cephalus) 340
estrus suppression 214	Muntjac (Muntiacus reevesi) 149
estrus synchronization 214	Murina spp. (see Bats, tube-nosed)
evolution of communication 202	Muscarine 291
fecal volatiles 24	Muscimol 313
feeding 407	Musculium rosaceum (see Clam, pill)
female choice of male odor 166, 189	Mushroom 313
female urine affects testosterone in	Agaricus sp. 316
males 219	death cap (Amanita phalloides) 313
individual discrimination 134	fly agaric (Amanita muscaria) 290, 291, 313
mate choice 132, 187	giant puffball (Calvatia gigantea) 313
maturation 234	lactarius (Lactarius sp.) 316
6-MBOA effects 381	late fall-oyster (Panellus serotinus) 313
multicomponent pheromone 26	peppery bolete (Boletus piperatus) 316
odor discrimination 126	San Isidro (Psilocybe cubensis) 290, 291
over-marking 165	Teonancatl (Psilocybe mexicana) 292
predator odor response 367	vomiting Russula (Russula emetica) 316
pregnancy block 217	Mushrooms 276, 316
priming, growth 208	Muskox (Ovibos moschatus) 149
priming, sexual 216	Muskrat (Ondatra zibethica) 242, 400
proline-rich salivary proteins 323	Mus spp. (see Mouse)
puberty acceleration 210, 211	Mustela erminea (see Weasel, short-tailed)
pups' odor responses 236	Mustela furo (see Ferrets)
receptor zones 93	Mustela nivalis (see Weasel, least)
reproductive isolation 199	Mustela putorius (see Polecat)
scent changes over time 170	Mustela sibirica (see Weasel, Siberian)
scent marking 161	Mustela vison (see Mink)
smell and optimal outbreeding 131	Mustelid secretions, urine 402
spatial orientation 80	Mustelus griseus (see Shark, dog)
t-complex 187	β -Myrcene 307
toxins 283	Myrtaceae 306

NaCl as cue for leech 374	Norepinephrine 194
Najad, spiny (N <i>ajas marina</i>) 385	Nose-tapping, salamanders 39, 96
Natrix natrix (see Snake, grass)	Notechis scutulatus (see Snake, tiger)
Natrix spp. 41	Notonecta sp. (see Backswimmers)
Navigation	Notophthalmus viridens (see Eft, red)
defined 61	Notophthalmus viridescens (see Newt,
geographical 80	red-spotted)
landmark 80	Notophthalmus v. viridescens (see Newt, eastern
topographical 80	red-spotted)
Neem tree (Azadirachta indica) 265	Notropis coccogenis (see Shiner, striped)
Negaprion brevirostris (see Shark, lemon)	Notropis texanus (see Shiner, weed)
Neighbor effect, plants 313	Notropis venustus (see Shiner, blacktail)
Neisseria spp. 53	Nucifraga columbiana (see Nutcracker, Clark's)
Neivamyrmex sp. (see Ant)	Nutcracker, Clark's (Nucifraga columbiana) 354
Neodiprion sertifer (see Sawfly, European pine)	Nutrient-toxin titration hypothesis 302
Neophobia 317	Nyctereutes procyonides (see Raccoon dog)
Neotoma spp. (see Woodrat)	Nycticorax nycticorax (see Heron, black-crowned
Nepeta cataria (see Catmint)	night)
Nepetalactone 275	8)
Nereis sp. (see Sandworm)	Oak 407
Nerodia spp. (see Snake)	Emory (Quercus emoryi) 333
Nerium oleander (see Oleander)	red (Quercus rubra, subgenus Erythrobalanus),
Nerodia sipedon (see Snake, water)	acorn tannins 308
Nerve	Oceanites oceanicus (see Storm-petrel, Wilson's)
barbel 339	Oceanodroma leucorhoa (see Storm-petrel,
facialis 108	Leach's)
glosspharyngeal 85, 108	Ocellated skink (<i>Chalcides ocellatus</i>), VNO for
olfactory (fish) 85, 338	food detection 97
terminal 108	Ochotona princeps (see Pika)
trigeminal 107	Octadecanediol-1-2 216
prenatal function 234	Octanal 182
vagus 108	1-Octen-3-ol 318, 372
Neural pathways, centrifugal 95	Odocoileus spp. (see Deer)
Neurolathyrism 283	Odor
Neurotoxicity 289	alarm 191
Newt 16, 343	alert 191
Alpine (Triturus alpestris) 201	aversion
California (<i>Taricha torosa</i>) 259, 363	fetal rat 234
crested (<i>Triturus carnifex</i>) 201	gravity effect 15
eastern red-spotted (<i>Notophthalmus v.</i>	primary 317
viridescens) 69, 145	complexity 119
Italian (<i>Triturus italicus</i>) 201	fecal 138
red-bellied (Cynops pyrrhogaster) 39, 176	filial 139
red-spotted (Notophthalmus viridescens) 343,	gestalt 26
421	image 26
rough-skinned (<i>Taricha granulosa</i>) 250,	individual 133
363	memory 244
sword-tailed (Cynops ensicauda) 176	mosaic 26
western (<i>Taricha rivularis</i>) 69	"on-demand-odor" 33
Nightshade, American black (Solanum	pattern 26
americanum) 302	plumes 12
Nipple odors 236	preferences, children 239
Ngorongoro crater 154	profile 26
Nocomis biguttatus (see Chub, hornyhead)	and survival, rat pups 237
Nonanal 30	trails, snakes 228
INUITALIAI JU	11a113, 311aNC3 220

Oestrone-3-sulfate 185	Orthoporus dorsovittatus (see Millipede)
Oilbird (Steatornis caripensis) 144, 354	Orthopristis chrysopterus (see Pigfish)
Oleander (Nerium oleander) 279	Orycteropus afer (see Aardvark)
Oleandrin 279	Oryctolagus cuniculus (see Rabbit)
Oleic acid 30	Oryx, Arabian (Oryx leucoryx) 6
Olfaction-taste interactions 121	Oryzias latipes (see Killifish)
Olfaction-trigeminus interactions 122	Osmetrichia 55, 201
Olfaction–VNO interactions 121	Ostariophysi 20, 194
Olfactory aposematism 258	Ostraction sp. (see Fish, box)
Olfactory bulb	Othonops eos (formerly Typhlogobius californiensis
early learning 244	see Goby, blind)
noradrenergic pathways 139	Otter 122, 135
processing 93	Eurasian (<i>Lutra lutra</i>) 144, 160
size 87	spraints (fecal marks) 159, 167
spatial segregation of input 93	sea (Enhydra lutris) 266
	· · · · · · · · · · · · · · · · · · ·
Olfactory cortex, projections 94	Over-marking 165
Olfactory impairment, amalrara, 14	Ovibos moschatus (see Muskox)
Olfactory impairment, smokers 14	Ovul humanima (Speeding sunicularis) 265
Olfactory imprinting	Owl, burrowing (Speotyta cunicularia) 265
mammals 242, 408	Oxalic acid 284
reptiles 231	Oxalis acetosella (see Sorrel, wood)
Olfactory isolation mechanism 198	Oxalis pes-caprae (see Soursob)
Olfactory memory 120, 134, 218, 223	Oxylobium sp. 335
Olfactory receptor neurons 89, 91	Oxytocin 120, 139
specialist 110	Oyster (Crassostrea virginica) 340
Oncorhynchus kisutch (see Salmon, coho)	Ozone hole 293
Oncorhynchus mykiss (see Trout, rainbow)	Ozotoceros bezoarticus (see Deer, pampas)
Oncorhynchus nerka (see Salmon, sockeye)	
Ondatra zibethica (see Muskrat)	Pachyptila desolata (see Prion, Antarctic)
Opheodrys vernalis blanchardi) (see Snake, green)	Paco-vicuña 140
Opisthocomus hoazin (see Hoatzin)	Padre Island National Seashore 71
Opossum,	Pagodroma nivea (see Petrel, snow)
short-tailed (Monodelphis domestica) 211	Palatability
Virginia (Didelphis virginiana) 135, 257	amphibians 251
conditioned aversion 318	birds 260
as egg predator 413	Panda, giant (Ailuropoda melanoleuca) 48, 334
fungivory 291, 313	Pandalus borealis (see Shrimp, northern)
primary toxic mushroom avoidance 316	Panellus serotinus (see Mushroom, late
seed digestion 302	fall-oyster)
willow defense 312	Pangolin (Pangolinus arboricoles) 153
Orchardgrass (Dactylis glomerata) 123	Pangolinus arboricoles (see Pangolin)
Orcinol 296, 297	Panthera onca (see Jaguar)
Oreochromis spp. (see Tilapia)	Panthera t. tigris (see Tiger)
Oreotragus oreotragus (see Klipspringer)	Pan troglodytes (see Chimpanzee, clay eating)
Orientation	Papaya (Carica papaya) 297
definition 61	Papyriferic acid 300
guided 80	Parapulex chephrenis (see Flea)
Oriole	Parasites, host odors 371
black-backed (Icterus galbula abeilli) 268	Pardachirus marmoratus (see Fish, Red Sea Moses
northern (Icterus galbula) 268	sole)
Ornithonyssus sylviarum (see Mite, fowl)	Pardachirus pavoninus (see Fish, peacock sole)
Ornithorhynchus anatinus (see Platypus,	Pardaxins 248
duck-billed)	Parental behavior 136
Oronectus rusticus (see Crayfish)	Parrots, geophagy 324
Orthoaminoacetophenone 397	Parsnip (<i>Pastinaca</i> sp.) 307, 333
OTTOMINIOACCIOPHENONE JJ/	1 a10111 (1 a0111 and 5p.) 30/, 333

Pars-pro-toto 196, 264	pH and volatility 14
Parturition 120	Phascolarctus cinereus (see Koala)
Parupeneus porphyreus (see Goatfish, Hawaiian)	Phasianus colchicus (see Pheasant)
Parus caeruleus ogliastrae (see Tit, Corsica blue)	Pheasant (Phasianus colchicus) 387
Passer spp. (see Sparrow)	β -Phellandrene 308
Passiflora edulis (see Passion fruit)	Phenol 372
Passion fruit (Passiflora edulis) 297	Phenolic glycosides 301, 309
Pastinaca sp. (see Parsnip)	Phenolics 272
Pasting, brown hyena 29, 153	birch 285
Paternal behavior 141	Phenols 301, 372
Path integration 80	2-Phenoxyethanol (rabbit) 149
Paullinia yoco (see Yoco)	Phenylacetic acid 30, 51
Pavoninins 248	2-Phenylethanol 29, 62, 113, 231, 352
Pea	2-Phenylethylmethyl sulfide 403
flat (Lathyrus sylvestris) 283	Pheromaxein, pig 4, 54
perennial (L. latifolius) 283	Pheromone
sweet (L. odoratus) 283	aggregation, snakes 143
Peeper, spring (Hyla crucifer) 341	alarm, fish 359
Pelamis platurus (see Snake, yellow-bellied sea)	blend 26
Penaeus duoarum (see Shrimp, pink)	cloacal, snake 181
Penstemon digitalis 307	copulatory, garter snakes 180
2-pentanone 390	courtship, salamanders 176
(Z)-26-Pentatriaconten-2-one, garter	-to-hormone hypothesis 200
snakes 179	mammary ("nipple"), rabbit 136,
(Z)-2-Penten-1-yl acetate 37, 213	171
<i>n</i> -Pentyl acetate 37, 106, 213	mounting (hamster) 24
Peptides 177	multicomponent 26, 27
Peptostreptococcus sp. 53	functions 28
Perca flavescens (see Perch, yellow)	priming 409
Perca spp. (see Perch)	receptors, VNO 105
Perch	sex 171
common (Perca fluviatilis) 362	signaling 410
yellow (Perca flavescens) 192, 359	trailing, snakes 143
Perileucoptera coffeella (see Leaf miner)	transport 54
Perineal pouches, guinea pig 35	Pheucticus melanocephalus (see Grosbeak,
Perisorius canadensis (see Jay, gray)	black-headed)
Peromyscus spp. (see House)	Phleum pratense (see Grass, timothy)
Petauroides volans (see Marsupials, greater	Photinus spp. (see Fireflies)
glider)	Photophytodermatitis 296
Petaurus breviceps (see Sugar glider)	Photosensitizers 296
Petrel 24, 262	Photosensitization 294
Antarctic (Thalassoica antarctica) 153,	Phototoxic effects 295
352	Phoxinus neogaeus (see Dace, finescale)
blue (Halobaena caerulea) 74, 352	Phoxinus phoxinus (laevis) (see Minnow,
giant (Macronectes sp.) 351, 352	European)
Kerguelen (Pterodroma brevirostris) 352	Phyllobates spp. (see Frog, dart-poison)
pintado (Cape; Daption capense) 351, 352	Phyllostomus discolor (see Bats)
snow (Pagodroma nivea) 73, 153, 351	Phyllotreta nemorum (see Beetle, flea)
white-chinned (Procellaria aequinoctialis) 74,	Phytoestrogens 286
351, 352	Phytohemagglutinins 301
see also Storm-petrel	Pica pica (see Magpie, black-billed)
Petromyzonol sulfate 66, 172	Picea spp. (see Spruce)
Peyote (Cactus lophophora) 290, 292	Pickerel
Phaeocystis pouchetti (see Microalgae)	chain (Esox niger) 359
Phalaris arundinacea (see Grass reed canary)	redfin (Fsox americanus) 359

Pig	Plethodontids 96, 98
aggressive behavior 411	Plume models 10
alarm odor 195	Poa pratensis (see Grass, Kentucky blue)
boar odor effects on VNO 105	Podarcis hispanica atrata (see Lizard, Iberian wall)
caecal odor attractant 138	Poecilia reticulata (see Guppy, Trinidadian)
deoxycholic acid 236	Poekilocerus bufonius (see Grasshopper)
environmental odors 60	Pogona vitticeps (see Lizard, bearded dragon)
maternal pheromone 137	Polarity 22
nipple odor 236	and scent marks 14
pheromone transport 55	Polecat (Mustela putorius = Putorius putorius) 48,
priming pheromone 211	356
puberty acceleration 409	Pollution, air 420
sex pheromone 410	Polypeptides, gerbil 168
truffle detection 388	Pony (Equus caballus) 149
venom resistance 257	Poplar 299, 301
Pigeons, odor thresholds 87, 116	Poplar, balsam (Populus balsamifera) 299, 300
geographical differences in homing 77	301
homing 74	induced defense 332
odor cues in homing 78	see also Aspen quaking
Pigfish (Orthopristis chrysopterus) 340, 341	Population
Pika (Ochotona princeps), "hay piles" 319,	cycles 221, 332
382	density
Pike (Esox lucius) 192, 359, 360, 361	and gland activity 40
Pikeminnow (Ptychocheilus lucius) 393	and scent marking, beaver 166
Pimephales promelas (see Minnows, fathead)	differences, salamander secretions 177
Pine	estimates 412
American red (Pinus resinosa) 307, 319	"top down" regulation (Lake Guri) 298
Jeffrey (Pinus jeffreyi) 5, 357	toxic plant effects 297
lodgepole (Pinus contorta) 315, 405	Populus balsamifera (see Poplar, balsam)
Monterey (Pinus radiata) 404	Populus tremuloides (see Aspen, quaking)
ponderosa (Pinus ponderosa) 308	Possum
Scots (Pinus sylvestris) 309, 310, 378, 402	brush-tailed (Trichosurus vulpecula) 30, 50,
white (Pinus strobus) 307, 319	51, 125
α-Pinene 275, 308, 310, 322	conjugation 330
β-Pinene 308, 310, 396	Eucalyptus diet compounds 306
Pine oil 404	excretion 330
Pinfish (Lagodon rhomboides) 340, 341	pinene detoxification 275
Pinosylvin 273, 300, 309, 405	ring-tailed (Pseudocheirus peregrinus) 330
Pinosylvin methyl ether 273, 300, 309, 405	Postovulatory prostaglandin pheromone,
Pinus spp. (see Pine)	goldfish 204
2-Piperidone 23	Potato
Pithuranthus triradiatus 295	alkaloids
Pitohui 50	Pinta Boca (Solanum stenotonum) 326
hooded (Pitohui dichrous) 261	Solanum acaule 326
variable (P. kirhocephalus) 261	Solanum megistacrolobum 326
Pituophis melanoleucus (see Snake, pine)	secondary metabolites 300
Pitviper (Calloselasma rhodostoma) 49	Potentiation 318
Pitymys subterraneus (see Vole, European pine)	Predation risk 194
Plant secondary metabolites 271, 299	Predator control 411
Plasma cortisol 194	Predator inspection behavior 192, 359
Plasmodus rotundus (see Bat, vampire)	Pre-exposure effects 128
Platophylloside 285	Pre-familiarization, harvest mouse 411
Platypus, duck-billed (Ornithorhynchus	Pregnancy block
anatinus) 262	evolutionary significance 222
Plethodon spp. (see Salamanders, woodland)	post-implantation 217
Plethodontid receptivity factor (PRF) 177	pre-implantation 217
· · · · · · · · · · · · · · · · · · ·	

5 <i>β</i> -Pregnane-3 <i>α</i> ,17 <i>α</i> -diol-20-one-glucuronide,	salamander pheromone 176
catfish 174	tamarins 25
Pregnenolone 191	Proteles cristatus (see Aardwolf)
Prenatal olfaction 232	Proteus spp. 53
Preovulatory primer pheromone, goldfish	Protoadaptation 179
204	Prunasin 281, 297
Preputial secretion, mice 54, 188	Prunus amygdalus (see Almond, bitter)
Prey fish, predator odors 358	Pseudacris spp. (see Frog)
Prey odors 338	Pseudocheirus peregrinus (see Possum,
Primates	Ring-tailed)
concealed ovulation 186	Pseudomonas spp. 53, 267
hunting by smell 357	Pseudotsuga menziesii (see Fir, Douglas)
sex odors 186	Pseustes sulphureus (see Snake, puffing)
VNO 101, 134	Psilocybe spp. (see Mushroom)
Priming, growth and development 208	Ptarmigans 305
Priming pheromones	Pteridium aquilinum (see Bracken, common)
ecological significance 220	Pterodroma brevirostris (see Petrel, Kerguelen)
effect on life history 222	Pterophyllum scalare (see Angelfish)
fish 206	Ptychocheilus lucius (see Pikeminnow)
VNO role 216	Ptychocheilus oregonensis (see Squawfish,
Prion, Antarctic (Pachyptila desolata) 126	northern)
Proboscids 311	Puberty
Procambarus clarki (see Crayfish)	acceleration 210, 409
Procavia capensis syriaca (see Hyrax)	delay 212
Procellaria aequinoctialis (see Petrel,	humans 224
white-chinned)	Pueraria mirifica (see White kwao krua)
Procellariiforms 71, 87	Puffer fish (see Fugu)
foraging 350	Puffinus spp. (see Shearwater)
Procyon lotor (see Raccoon)	Pumiliotoxins 252
L-Proline 114, 283, 323	Purshia tridentata (see Bitterbush)
Proline-rich salivary proteins 323	Putrescine 347
Pronghorn (Antilocapra americana)	Pyrazine 26, 67, 318, 351, 352
familiarization 125	Pyridine-N-oxide 194
glands 44, 167	Pyrrhula pyrrhula (see Bullfinch)
isovaleric acid 168	Pyrrolizidines 253, 264
lipid secretion 45	•
male-specific glands 45	Quail
"marking trips" 161	bobwhite (Colinus virginianus) 116, 302
non-volatiles 30	California (Lophortyx californicus) 287
overmarking 148	Japanese (Coturnix coturnix) 287
scent mark longevity 31	Quebracho tree (Schinopsis lorentzii) 285, 305
visual signal 34	404
winter diet 310	Quercetin 284, 285
<i>n</i> -Propyl dithiolane 407	Quercus spp. (see Oak)
3-Propyl-1,2-dithiolane 397, 403	Quinol 297
3-Propylphenol 372	2-Quinolinemethanethiol 24, 262
4-Propylphenol 372	S-2-Quinolinemethyl thioacetate 24
2-Propylthietane 184, 397, 403, 412	Quinones in fruits 395
<i>n</i> -Propylthiolane 407	Quiscalus major (see Grackle, boat-tailed)
Prostaglandins 111, 114	guntum miger (ett Graeine, beat tantea)
Proteinase inhibitors 301	Rabbit (Oryctolagus cuniculus)
Proteins 24	antifeedant 403
elephant 26	chin-marking 48, 147, 159, 160
iguana 29, 123	over-marking 166
odorant binding 91	dominance odor 149
olfactory marker 92	flea 376

Rabbit (Oryctolagus cuniculus) (cont.)	prenatal olfaction 232
hormone-dependent dominance 166	priming and VNO 216
mammary ("nipple") pheromone 136, 237	proline-rich salivary proteins 323, 324
microorganisms 53	receptor zones 93
odor imprinting 243	response to milk odor 137
predator odor response 370	spatial orientation 80
population control 399	tannin processing 324
prenatal odor learning 232	taste aversion to salamanders 252
priming effects 212, 218	testosterone effect 52
pygmy (Brachylagus idahoensis) 315	urine effects on VNO 106
testosterone effect 52	urine volatiles 37
VNO 98, 237	rice (Rattus rattoides) 377
Raccoon (Procyon lotor) 413	roof (Rattus rattus) 407
Raccoon dog (Nyctereutes procyonides) 164	soft-furred (metad; Rattus meltada
Rajasthan desert 161	pallidior) 406
Ramphotyphlops nigrescens (see Snake, blind,	see also Kangaroo rat
Australian)	Rattlesnakes 83, 345
Rana aurora sp. (see Frog)	banded rock (Crotalus viridis viridis) 345, 346
Rangifer tarandus (see Caribou)	canebrake (Crotalus horridus atricaudatus) 364
Raphus cucullatus (see Dodo)	diamondback (Crotalus atrox) 256
Rat	northern Pacific (Crotalus viridis oregonus),
African crested (Lophiomus imhausi) 56	preying 345, 346
African/Gambian giant pouched (<i>Cricetomys</i>	prairie (<i>Crotalus viridis</i>) 228
gambianus) 417	behavior development 228, 230
hispid cotton (Sigmodon hispidus) 147	odor trails 228
Norway or laboratory (Rattus norvegicus)	prey striking and searching 345, 352
amniotic fluid 139, 237	predator odor response 365
antipredator response 37, 366, 367 bacteria 53	pygmy (Sistrurus miliarius) antipredator "body-bridging" 229, 364
bait shyness 406	strike-induced chemosensory searching 345
broad tuning 91	timber (<i>Crotalus horridus</i>) 364
castration effect on reception 120	behavior development 228, 230
caecal odor 50	response to prey odors 343
clay eating 326	twin-spotted (<i>Crotalus pricei</i>) 345
communication on food 197	venom immunity 257
cow odor discrimination 190	Rattus spp. (see Rat)
decoding "odor gestalt" 113	Raven (Corvus corax) 260, 412
fetal effect of alcohol 233	Recycled animal and plant materials 265
food odor communication 55	Reed canary grass (Phalaris arundinacea) 280
individual discrimination 134	Regional selectivity 21
individual odor variation 52, 53	Reindeer (Caribou tarandus) 81, 196
induced ovulation 215	Reithrodontomys sumichrasti (see Mouse,
maternal care and trigeminal nerve 108	Sumichrast's harvest)
maternal pheromone 241	Release modulators 29
maternal stimulation 242	Removal rate hypothesis 302
maturation of chemoreception 234	Renal effects 293
6-MBOA effects 381	Repellents
milk odor 137	conspecific odors 400
norepinephrine effect 120	contact 396
odorant thresholds 117	excretions 402
odor imprinting 243	non-target species, effects 399
odor memory 244	population effects 399
odors and pup survival 237	predator odors 397, 399, 407
ovulation and VNO 105	for predators 412
palatability of bird eggs and flesh 259	for rodents 403
prenatal odor conditioning 232, 233	Reproductive behavior 18

Reproductive cycle, behavior variation 189 Reproductive inhibition by plant	Sablefish (Anoplopoma fimbria) 114 Sagebrush 310, 315
compounds 286 Reptiles	big (Artemisia tridentata) 310, 315, 322 black (A. nova) 310
behavior development 228	Saguinus spp. (see Tamarin)
chemoreception 87 defense 256	Saiga antelope (Saiga tatarica) 57 Saiga tatarica (see Saiga antelope)
diet effects 49	
food choice 304	St. Johnswort (Hypericum perforatum) 294 Salal (Gaultheria shallon) 402
glands 40	Salamander 38, 68, 129, 259, 342
hormone effects 52	Allegheny mountain dusky (Desmognathus
odor detection 83	ochrophaeus) 207
odor thresholds 114	brook (Eurycea wilderea) 96, 361
orientation 69	long-toed (Ambystoma macrodactylum) 195
predator detection 363	northern two-lined (Eurycea bislineata) 360
priming pheromones 207	olympic torrent (<i>Rhyacotriton olympicus</i>) 39
reproductive isolation 198	pygmy (Desmognathus wrighti) 58
territorial displays 152	small-mouth (Ambystoma texanum) 360, 361,
VNO 97, 106	362
Reservoirs 54	spotted (Ambystoma maculatum) 12, 69,
anal sacs, carnivores 54	251
castor sacs 54	Texas blind (Typhlomolge rathbuni) 176
Retrieving pups, VNO effects 104	tiger (Ambystoma tigrinum) 114, 252, 343
Rhamnus cathartica (see Buckthorn)	woodland (<i>Plethodon</i> spp.) 39, 58, 96, 134
Rhinophrynus dorsalis (see Toad, Mexican	Jordan's (P. jordani) 176, 198, 207
burrowing)	red-backed (<i>P. cinereus</i>) 39, 133, 152, 176,
Rhizophagus grandis (see Beetle, predatory)	251, 343, 361
Rhododendron sp. 277	P. dorsalis 361
Rhodopus sungorus (see Hamster, Djungarian)	P. montanus 143, 240
Rhyacotriton olympicus 39	P. richmondi 361
Rivers	P. shermani 143, 240
black-water 274, 328	P. yonahlossee 143, 240
"white" 328	slimy (<i>P. glutinosus</i>) 198
Robin, American (<i>Turdus migratorius</i>) 302, 386	western red-backed (P. vehiculam) 145
Rockrose (Cistus ladanifer) 284	Salicaldehyde 300
Rodents	Salicylic acid 272
dominance 145	Saliva 32, 48, 383
familiar individuals 127	Salix spp. (see Willow)
food choice and plant metabolites 307	Salmon 61
male marking 183	Atlantic (Salmo salar)
mate choice 187	acidification effects 392
repellents 403	amino acids as feeding cues 342
territorial marking 157	behavior development 227
see also individual species	kin recognition 131
Roedeer (Capreolus capreolus) 402	olfactory imprinting 61
Rosa canina (see Dogrose)	orientation 16, 61, 65
Rosaceae 281	pesticide effect on priming 421
Rousettus aegyptiacus (see Bats, Egyptian fruit)	prey odor 342
Rub-urination 57	priming pheromone 205
"ruff-sniff", auklet 182	salmon lice attractant 371
Ruminants	sex attractant 175
microbes in biotransformation 332	Chinook 18, 393
phenolics effects 285	coho (Oncorhynchus kisutch) 65
Ruppia martima (see Tasselweed)	kin recognition 129, 130
Russula emetica (see Mushroom, vomiting	odor threshold 113, 114
Russula)	olfactory imprinting 61, 392

Salamander (cont.)	Scophthalmus maximus (see Turbot)
population odors 142	Schreckstoff 192, 194
predator odor avoidance 358	Scilliroside 404
sockeye (Oncorhynchus nerka) 392	Sculpin,
Salmon lice (Lepeophtheirus salmonis) 371	banded (Cottus carolinae) 343
Salmo salar (see Salmon)	yellow-finned Baikal (<i>Ćottocomephorus</i>
Salmo trutta (see Trout, brown)	grewingki), sex attractant 174
Salsify (Andropogon sp.) 307	Sea anemone
Saltgrass (see Grass, salt)	Radianthos kuekenthali 384
Salvelinus alpinus = Salmo alpinus (see Char,	Stoichactis kenti 384
Arctic)	Seabirds 17
Salvelinus fontinalis (see Trout, brook)	food odors 353
Samandarin 255	foraging 350
Sambucus nigra (see Elder)	see also individual species
Sandpiper (<i>Calidris maritima</i>) 352	Sea lamprey (Petromyzon marinus)
Sandrats 33	attractants 64, 66
Sandworm (Nereis sp.) as prey 344	bile acid 172
Saponins 277, 301	host odor 371
Sarcorhamphus papa (see Vulture, king)	sex attractant 171
Sawfly, European pine (Neodiprion sertifer) as	Sea purse (Dioclea megacarpa) 283
prey 355, 357	Seasonal variation behavior 188
Scent	
hair 55	plant chemical defenses 301
mark,	Sea squill (Urginea maritime) 404
aardwolf 31	Sea urchin (Arbacia punctulata) 340
age of 27, 170	Secondary plant metabolites, physiological
complex composition, beaver 168	effects 284
life expectancy 29, 32	Secretion
marking	flank gland,
beaver 157, 160	golden hamster 128
coyote 156	shrew 135
and feeding 166	viscosity 167
gazelle 160	Seeds 302
gerbil 160	conditioning 386, 388
gerenuk 160	dispersal, birds 384
hyena 153	passage through animals 384
mouse 161	birds 386
otter 160	fish 386
population density 161	mammals 386
pronghorn 161	reptiles 386
seasonal variation 168	Selenocystathionin 283
syndrome 161	Self-anointing 376, 377
tiger 160	canids 376
woodchuck 159	hedgehog 55
matching 126, 165	"selfish herd" 194
mound, beaver 122, 165	Self-medication 381
rolling 55	Semotilus margarita (see Dace, pearl)
Schistosoma mansoni (see Blood flukes,	Sensitive periods 241
schistosome)	Sensory drive 200
Sciurus spp. (see Squirrel)	Sensory exploitation 200
Schinopsis lorentzii (see Quebracho)	Septal organ of Masera 108
Schizachyrium scoparium (see Grass, little	Sesquiterpene 312
bluestem)	Sesquiterpene lactones 276, 382
Sclerocarya caffra (see Marula)	Sesquiterpene phytoalexins 301
Schreckstoff 192, 194	Setaria italica (see Grass, foxtail bristle)

Setting, effects of 17, 18	brine (Artemia sp.) 360
Sex pheromone, pig 4	ghost (Callianassa affinis) 172
3α -androstenol 4	northern (Pandalus borealis) 339, 341
5α -androstenone 4	pink (Penaeus duoarum) 340, 341
Shad, threadfin (Dorosoma petenense) 358, 360	Sideroxylon grandiflorum (see Tambalacoque tree)
Sharks	Sigmodon hispidus (see Rat, hispid cotton)
dog (Mustelus griseus) 248	Silefrin 176
hammerhead (Sphyrnidae) 84	Sistrurus miliarus (see Rattlesnakes, pygmy)
lemon (Negaprion brevirostris) 248	Skatole (methyl indole) 375, 414
white-tip reef (Triaenodon obesus) 248	Skink, blue-tongued (<i>Tilique scincoides</i>) 126
Shearwater	Skua, South Polar (Catharacta maccormicki) 260,
Buller's (Puffinus bulleri) 350	351
greater (<i>P. gravis</i>) 350	Skunk
Manx (<i>P. puffinus</i>) 114, 350	hog-nosed (Conepatus mesoleucus) 30
pink-footed (<i>P. creatopus</i>) 350	spotted (Spilogale putorius) 32
short-tailed (P. tenuirostris) 350	striped (Mephitis mephitis), 23, 32, 262
sooty (P. griseus) 350, 351	as egg predator 413 "Smelling of rain" 81
wedge-tailed (<i>P. pacificus</i>) 143	
Sheep	Smelt, surf (Hypomesus pretiosus) 339, 341
alkaloids 311, 312	Smoking 119
amniotic fluid 139	Snake 69 African water
antifeedants 402, 404, 407, 408	
attachment 138, 408	Lycodonomorphus laevissimus 253
clover effect on lambing 286	Lycodonomorphus rufulus 253
conditioned aversion 318	anal sacs 264
coping with toxins 302	avomic 69
cross-fostering, 140	black rat (Elaphe obsoleta obsoleta) 369
embryonic malformations 288	blind
feeding behavior 123	American (Typhlops pusillus) 375
feeding stimulants 408	Australian (Ramphotyphlops nigrescens) 375
flehmen 102, 185	Texas (Leptotyphlops dulcis) 256, 266, 375
induced ovulation 215	brown (Storeria dekayi) 228, 229
inguinal wax 138	brown tree, Boiga irregularis 18, 19, 180
lamb odor 244	chemical cues for control 347, 394
mating behavior 123	impact on fauna 346
parturition and odor response 245	prey odors 347
predator odor 37	bull (see pine snake)
prenatal odor responses 232	corn (Elaphe guttata) 69, 343
tannin processing 323, 324	defense 259
toxic plants 280, 284, 297, 314	eastern indigo (Drymarchon corais) 364
"She-males," snakes 178	European adder (Vipera berus) 178, 365
Shewanella sp. (see Algae)	feeding behavior development 229
Shiner	fox (Elaphe vulpina vulpina) 343
blacktail (Notropis venustus) 358, 359	garter 228, 365
striped (N. coccogenis) 343	aquatic (Thamnophis elegans aquaticus) 229
weed (N. texanus) 358, 359	avoidance toxic prey 255
Short-chain fatty acids 23	blackbelly (Thamnophis melanogaster), prey
Shrew	learning 347
common (Sorex araneus) 401	Canadian (Thamnophis sirtalis parietalis) 178
greater white-toothed (Crocidura russula) 37,	feeding behavior development 206, 229,
135	230
masked (Sorex cinereus) 355	food odor preferences 344
short-tailed (Blarina brevicauda) 251, 263,	multisensory preying 347
355, 366	Plains (Thamnophis radix) 69, 178, 181,
Shrimp	229, 362

0 1 ()	0.1.1.11
Snake (cont.)	Solubility 22
predator chemicals response 365	Songbirds 87
red-sided (Thamnophis sirtalis) 69, 98, 196,	Songthrush (Turdus philomelos) 306
198	Sorex spp. (see Shrew)
species identification 143	Sorghum spp. (see Sorghum)
toxin tolerance 251	Sorghum
VNO zone-to-zone projection 97	grain (Sorghum bicolor) 282, 289, 306
	sweet (Sorghum vulgare) 291
western terrestrial (Thamnophis	
elegans) 229, 230, 345, 362	Sorrel, wood (Oxalis acetosella) 284
gopher (see pine snake)	Soursob (Oxalis pes-caprae) 284
grass (Natrix natrix) 365	Spalax ehrenbergi (see Mole rat, naked)
gray rat (Elaphe obsoleta spiloides) 256	Sparrow
green (Opheodrys vernalis blanchardi) 229	house (Passer domesticus) 265, 306, 307
food odor 344	tree (P. montanus) 266
hognose (Heterodon nasicus) 365	Speotyta cunicularia (see Owl, burrowing)
king (Lampropeltis getulus) 229, 257, 364, 365	Spermophilus spp. (see Squirrel, ground)
prey odors 346	Sphyrnidae (see Shark, hammerhead)
mamba (<i>Dendroaspis</i> sp.) 256	Spices as bird repellents 396
milk (Lampropeltis triangulum) 256	Spilogale putorius (see Skunk, spotted)
orange-necked keelback (Macropisthodon	Spilopsyllus cuniculi (see Flea, rabbit)
sp.) 41	
	Splendipherin 40, 177
pine (bull or gopher; <i>Pituophis</i>	Spodophora exempta (see Army worm)
melanoleucus) 229, 365, 370	Sponge (Ircinia sp.) 265
response to mouse odor 344	Spraint (otter) 122
response to predator chemicals 365	Spring peeper (Hyla crucifer) 255
puffing (Pseustes sulphureus) 364	Spruce
racer (Coluber constrictor) 365	Norway (Picea abies) 307, 319
rat (Elaphe climacophora) 256, 376	Sitka (P. sitchensis) 405
ring-necked (Diadophis punctatus) 256, 361	white (P. glauca) 299, 300, 307, 319
smooth (Coronella austriaca) 365	Squalene 30, 178
southern water (Nerodia fasciata) 365	Squawfish, northern (Ptychocheilus
tiger (Notechis scutulatus) 258	oregonensis) 358
venom 49, 257	Squid 17
VNO 97	Squirrel
water (Nerodia [Natrix] sipedon) 228, 254, 343,	Abert's (tassel-eared) (Sciurus aberti) 308
360	flying (Glaucomys sabrinus) 313
yellow-bellied sea (Pelamis platurus) 257	gray (Sciurus carolinensis)
yellow-bellied water (Nerodia	acorn burying 320
erythrogaster) 255	acorn tannins 308
Snake plant (Turbina corymbosa) 292	mushroom eating 313
	,
Sniffing land mines 417	ground
Snow leopard (<i>Uncia uncia</i>) 368	Arctic (Spermophilus parryi plesius) 313
social odors, familial 125	Belding's (Spermophilus beldingi) 129
Sodefrin 39, 176	California (Spermophilus beecheyi) 370, 401
Sodium uptake, macaws 325	Columbian (Spermophilus columbianus) 127,
α-Solamargine 302	129
Solanine 280, 301, 327	Franklin's (Spermophilus franklinii) 413
Solanum americanum (see Nightshade, American	13-lined (Spermophilus tridecemlineatus) 129
black)	red (Tamiasciurus hudsonicus)
Solanum carolinense (see Horsenettle, Carolina)	mushroom preferences 316
Solanum spp. 302	toxic mushrooms 313
Solanum stenotonum (see Potato, Pinta Boca)	Staphylococcus spp. 53
α -Solasonine 302	S. aureus 53, 54
Solidago rugosa (see Goldenrod, rough)	Starling (Sturnus vulgaris) 265, 378, 394
Sollya heterophylla (see Creeper, blue-bell)	Steatornis sp. (see Oilbird)
	- '

Sternotherus odoratus (see Turtle, stinkpot)	Syncerus caffer (see Buffalo, African)
Steroidal bufogenins 255	Synomones 383
Steroids,	
beetles 21	Tachycineta bicolor (see Swallow, tree)
human axilla 27	Tadarida brasiliensis mexicana (see Bat, Mexican
odoriferous 20, 54	Taeniopygia guttata (see Finch, zebra)
pregnolones 21	Tamaliscus korrigum (see Topi)
Sterols 40	Tamarin,
Stickleback, brook (Culaea inconstans) 173	cotton-top (Saguinus o. oedipus) 29, 48, 212
Stigmastanol 181	scent marking 220
Stigmasterol 181	red-bellied (Saguinus labiatus) 369
Stilbenes 273	saddleback (Saguinus fuscicollis) 25, 26, 53
Stimulus generalization 111	144, 168
Stingrays (Dasiatidae) 250	Tambalacoque (Dodo) tree (Sideroxylon
Stoat (see Weasel, short-tailed)	grandiflorum, formerly Calvaria
Stolephorus purpureus (see Anchovy, inshore)	major) 385
Stomach oil, petrels 262, 264	Tamias spp. (see Chipmunk)
Stonefish (Scorpaenidae) 249	Tamiasciurus hudsonicus (see Squirrel, red)
Synanceja trachynis 249	Tannins
Synaneichthyes verrucosus 249	carcinogens 288
Stone roller (Campostoma anomalum) 343	condensed (catechin) 274, 275
Storeria dekayi (see Snake, brown)	description 273
Storm-petrel	digestion effects 285
black-bellied (Fregetta tropica) 351	hydrolyzable 274, 275
British storm (Hydrobates pelagicus) 73	juniper 319
Leach's (Oceanodroma leucorhoa) 350, 351	sorghum 306
Wilson's (Oceanites oceanicus) 71, 350, 351	South African plants 311
Streptococcus spp. 53	and tortoises 304
S. aurealis 267	Taraxacum officinale (see Dandelion)
S. faecalis 53	Taricha spp. (see Newt)
Stress, rat pups 139	Tasselweed, beaked (Ruppia maritima) 385
Strike-induced chemosensory searching,	Taste 108
rattlesnakes 83	aversion 316
Sturgeon	conditioned 317, 260
Russian (Accipenser gueldenstaedtii) 114	bitter 316
Siberian (A. baeri) 114	bud 109
stellate (A. stellatus) 114	experience in utero 233
Sturnus vulgaris (see Starling)	fish 108
Sucker, largescale (Catostomus macrocheilus) 358	papillae 109
Suckling, chemical cues 237	rat fetus 233
Sugar glider (Petaurus breviceps) 125, 145	receptor cells 109
"Suint", ram 215	Tatera indica (see Gerbil, Indian)
Sulfur compounds 21, 37, 370, 399, 403,	Taurine 66
405, 414	Taxiphyllin 281
Sunfish 21	Taxus cuspidata (see Yew, Japanese)
bluegill (Lepomis macrochirus) 359	t-complex, mice 187, 210
green (Lepomis cyanellus) 360, 362	Temperature effects 3, 55
Sus scrofa (see Boar)	Tenrec
Swallow, tree (Tachycineta bicolor) 114, 116	long-tailed (Microgale dobsoni) 370
"Swimmer's itch" 374	streaked (Hemicentetes semispinosus) 370
Swordtail (Xiphophorus helleri) 359	tailless (Tenrec ecaudatus) 370
Sylvia communis (see Whitethroat)	Tenrec ecaudatus (see Tenec, tailless)
Sylvilagus floridanus (see Cottontail)	Tent caterpillar, Eastern (Malacosoma
Synanceja trachynis (see Stonefish)	americanum) 291
Synaneichthyes verrucosus (see Stonefish)	Teratogens 288

Terpene alcohols 404 Terpenoids 274, 319 α -Terpineol 322 Territorial marking, badger 27	Fowler's (B. woodhousei fowleri) 68, 255 glands 253 Great Plains (B. cognatus) 362
Testosterone 105	Mexican (B. valliceps) 68 Mexican burrowing (Rhinophrynus
fish 174	dorsalis) 343
Testudo hermanni (see Tortoise, Greek land)	narrow-mouthed (Gastrophyne
Tetra, black (Gymnocorymbus ternetzi) 206	carolinensis) 255
Δ ⁹ -Tetrahydrocannabinol 290	natterjack (Bufo calamita) 342
2,4,6,10-Tetramethyl-1-undecanoic acid 32 Tetraphyllin B 297	southwestern (B. microsaphus) 362 toxins used by hedgehogs 377
Tetrodotoxin 110, 114, 247, 250	western (B. boreas) 68, 195, 343, 363
Thalarctos maritimus (see Bear, polar)	Tobacco hornworm (Manduca sexta) 303
Thalassoica antarctica (see Petrel, Antarctic)	Tomato, Galapagos (Lycopersicum cheesmanii var.
Thalassoma bifasciatum (see Wrasse,	minor) 385
blue-headed)	Tongue-flicking, reptiles 97
Thamnophis spp. (see Snake, garter)	Topi (Tamaliscus korrigum), VNO 98
Thiamine 383	Topography and airflow 12
Thiazoles 67	Tortoise 69, 181
Thioacetates 23, 32	Galapagos (Geochelone elephantopus) 385
Thiols 23, 32, 114	Giant (Geochelone gigantea) 304
Thomomys talpoides (see Gopher, pocket)	Gopherus sp. 181
Thresholds	Gopher (Gopherus polyphemus) 4
birds 116	Greek land (Testudo hermanni) 69
odor detection 113	Toxins, antiparasite 262
odor recognition 113	Tragelaphus strepsiceros (see Kudu)
β -Thujaplicine 275	Trail odors 196
β -Thujone 322	6-Transnonenal 190
Thujopsine 275	Tree shrew
Thuja plicata (see Cedar, red)	Tupaia belangeri 26
Thujopsis dolabrata (see Arborvitae, false)	Tupaia glis 166, 167
Thunnus albacares (see Tuna, yellowfin)	Trefoil, birdsfoot (Lotus corniculatus) 291
Thymallus thymallus (see Grayling)	Triaenodon obesus (see Sharks, white-tip reef)
Ticks	Trichogaster spp. (see Gourami)
effect of bird chemicals 262	Trichosurus vulpecula (see Possum, brush-tailed)
Ixodes matopi 375	Trifolium spp. (see Clover)
Ixodes mentzii 375	Trigeminal irritants 394, 397
Tiger (Panthera t. tigris) 31, 38, 366, 407	7α ,12 α ,24-Trihydroxy- 5α -cholan-3-one-24-
scent marking 159, 160	sulfate 172
Tilapia	2,4,6-Trihydroxydihydrochalcone 300
oreochromis sp. 385	Trimethylamine 23, 114, 352
redbelly (<i>Tilapia zilii</i>) 341	2,3,5-Trimethylphenylmethyl carbamate 412
spotted (Tilapia mariae) 360	3,4,5- Trimethylphenylmethyl carbamate 412
Tilapia spp. (see Tilapia)	Trimethylthiazoline (TMT) 37, 369, 403, 407
Tilique scincoides (see Skink, blue-tongued)	Triok, long-fingered (Dactylopsila palpator) 263
Tit, Corsica blue (Parus caeruleus ogliastrae) 267	(Z)-24-Tritriaconten-2-one, garter snakes 178
Toad	Triturus spp. (see Newt)
American (Bufo americanus)	Trout
chemical defense 255, 341	brook (Salvelinus fontinalis) 65
kin discrimination 129, 131, 228	host for sea lamprey 371
setting, effect 19	brown (Salmo trutta) 175
cane (giant) (B. marinus) 256, 343	host for sea lamprey 371
common European (B. bufo) 362	priming 205
alarm substance 195	rainbow (Oncorhynchus mykiss)
bufotalin 255	amino acids as food cues 340

effect on prey 362	Urginea maritima (see Sea quill)
kin recognition 129, 131	Urine 37
odor sensitivity 114	guinea pig 51
pheromone specificity 111	wolf 32
prey odors 342	Urine balls, sandrats 33
priming pheromone 205, 206	Urodeles 69
specialist receptors 110	Ursus americanus (see Bear, black)
water pollution effects 393	Ursus arctos (see Bear, brown)
Truffle, Périgord (Tuber melanosporum) 389	
Tryptophan 194	Vanillin 22
Tsetse fly (Glossina spp.) 372	Vanillyl acetamide 398
T-shirt test 132, 191, 239	Varanus albigularis (see Lizard, monitor)
Tsuga canadensis (see Hemlock, eastern)	Varanus exanthematicus (see Lizard, savanna
Tuber melanosporum (see Truffle, Périgord)	monitor)
Tubifex, amino acids 340	Varanus komodoensis (see Lizard, Komodo
Tuna 16	monitor)
yellowfin (Thunnus albacares) 339, 341	Varanus varius (see Lizards, lace monitor)
Tupaia spp. (see Tree shrew)	Vegetation and air flow 12
Turbina corymbosa (see Snake plant)	Veratryl acetamide 398
Turbot (Scophthalmus maximus) 371	Veratryl amine 397, 398
Turbulence 7	Vernodalin 382
environment, properties 16	Vernonia sp. 276
flow 6	V. amygdalina 382
water 15	Vernonioside B ₁ 382
Turdus merula (see Blackbird, European)	Vespa orientalis (see Hornet)
Turdus migratorius (see Robin, American)	Vespertilionidae (see Bats)
Turdus philomelos (see Songthrush)	Vetch
Turtle 69	common (Vivia sativa) 283
chemosensory foraging 348	milkvetch, two-grooved (Astralagus
eastern long-necked (Chelodina longicollis) 70	bisulcatus) 318
European pond (Emys orbicularis) 348	Vicuña (Vicugna vicugna) 140, 159
green sea (Chelonia midas) 231	Vine, lilac (Hardenbergia comptoniana) 313
imprinting 71	4-Vinylguaiacol 286
Kemp's ridley sea (Lepidochelys kempi) 71,	4-Vinylphenol 286
258, 393	Vipera berus (see Snake, European adder)
loggerhead (Caretta caretta) 258, 348	Virola theiodora (see Ebéna)
odor thresholds 114	Viscous environment, properties 16
painted (Chrysemys picta) 70	Visual signals 33
red-eared (Chrysemys [Pseudemys] scripta	Vivia sativa (see Vetch)
elegans) 98	Volatiles 22
sea, egg losses 413	mixtures 26
secretions 258	over vegetation 13
snapping (Chelydra serpentina) 231, 255,	Volatility 22
360	Vole (Microtus spp.) 404
stinkpot (Sternotherus odoratus) 258	bank (Clethrionomys glareolus) 147, 150, 199
terrapin (Mauremys leprosa) 181	213, 401
Typhlomolge rathbuni (see Salamander, Texas	antipredator responses 366, 367, 370
blind)	trap response 401, 406
Typhlops pusillus (see Snake, blind, American)	boreal redback (Clethrionomys gapperi) 399
,	California (Microtus californicus) 212, 298,
Ultrasonic vocalizations 104, 121	401
Ultraviolet light 354	California red-backed (Clethrionomys
Uncia uncia (see Snow leopard)	occidentalis) 313
Ungulates 150, 184, 187	cycles 297
Upupa epops (see Hoopoe)	European common (Microtus arvalis) 131

	,
Vole (Microtus spp.) (cont.)	VNO (Vomeronasal organ)
European field (short-tailed) (M. agrestis) 217,	in amphibians 96
219, 315, 354, 366, 367, 401	and behavior 103, 104, 106
European pine (Pitymys subterraneus) 213	in mammals 98, 101, 105, 134, 237
gray-tailed, Microtus canicaudus 18, 132, 367	and luteinizing hormone 219
Levant (M. guentheri) 211	and mating 104, 105
long-tail (M. longicaudus) 399	and maturation 234
meadow (M. pennsylvanicus)	and pheromones 216
alkaloids 309, 312	in primates 101, 134
antifeedants 403	rodents 101, 105, 106, 130, 216, 219
body region odors 49	pump 99, 100
conifer diet 307	receptors
"dear enemy" phenomenon 127	V ₁ 103, 105
diet-dependent odor 50, 51	
familiar individuals 127	V ₂ 105
	reptiles 97
food palatability 307	system 96, 98
food processing 319	Vulnerability score, birds 260
metabolic rate 296	Vulpes vulpes (see Fox, red)
as non-target species, repellents 399	Vulture
odor response, timing 189	black (Coragyps atratus) 350
phenolics effects 284	greater yellow-headed (Cathartes
predator avoidance 366, 367	melambrotus) 350
pregnancy block 217	king (Sarcorhamphus papa) 350
priming 211	
renal lesions 293	lesser yellow-headed (Cathartes
	burrovianus) 350
saponins 309	turkey (Cathartes aura) 115, 116, 349
seasonal differences 188	
signal longevity 33	Wallabia bicolor (see Wallaby, swamp)
tannin processing 324	Wallaby, swamp (<i>Wallabia bicolor</i>),
vomeronasal organ (see VNO)	antifeedants 402
montane (M. montanus) 285, 379, 381, 403	Wapiti (see Deer, red)
Orkney (M. arvalis orcadensis) 378, 402	Warbler, sedge (Acrocephalus schoenobaenus)
pine (M. pinetorum) 101, 106	259
prairie (M. ochrogaster)	Water-air interface 17
food palatability 307	Water buffalo, African 6
individual odors 134	Waterbug, giant (Lethocerus americanus) 195
induced estrus 211, 215	Waterfowl, egg predators 413
kin recognition 131	Wax esters, beaver 31
luteinizing hormone release 219	
	Waxwings 89
main olfactory system 96	cedar (Bombycilla cedrorum) 114, 116
mate recognition 134	Weasel
mating and VNO 104, 105	least (Mustela nivalis) 366, 374
phenolics effects 284, 285	short-tailed (stoat; M. erminea) 135, 170,
pregnancy block 217, 218	366, 374, 397, 403
priming, growth 208	Siberian (M <i>ustela sibirica</i>) 377
priming, sexual 216	see also Ferrets
puberty delay 213	Weatherfish, oriental (Misgurnus
tannin effects 285, 324	anguillicaudatus) 21
VNO 106	Whales
root (Microtus oeconomus), induced	baleen 81, 357
enzymes 330	toothed 357
Townsend's (M. townsendii) 18, 379, 381	Whelk (Busycon contrarium) 340
antifeedants 404	White kwao krua (<i>Pueraria mirifica</i>) 287
repellents 399	
woodland (M. pinetorum) 380, 381	Whitethroat (Sylvia communis) 259
woodiand (w. pinciorani) 300, 301	Whitten effect 214

Wildebeest (Connochaetes sp.) 81 common (C. taurinus) 99 Wildfires and plant defense 337 Willow 299, 301, 309 Arctic (Salix arctica) 305 S. fragilis 301 S. purpurea 301 Wolf (Canis lupus) body region odors 48 odor effect on prey 402 prey detection 355 scent marking 159 territorial marking 154 testosterone effect 53 urine compounds 32, 190 urine marking 154, 160, 167 Wolverine (Gulo gulo) 403 Woodchuck (Marmota monax) 127, 147, 159 Woodrat desert (Neotoma lepida) 299

Stephens' (N. stephensi) 319

Wrasse, blue-headed (Thalassoma bifasciatum) 265, 266

Xenopus laevis (see Frog, African clawed) Xenopsylla dipodilli (see Flea) Xiphophorus helleri (see Swordtail)

Yarrow (Achillea millefolium) 266 Ligurian (Achillea ligustica) 267 Yew, Japanese (Taxus cuspidata) 402 Yoco (Paullinia yoco) 303 Yoke-elm (Carpinus betulus) 407 Yopo tree (Anadenanthera peregrina) 292

Zebra 6
Zebrafish (*Danio* [formerly *Brachydanio*] *rerio*) 91, 93
male sex pheromone 174
primer pheromone 206
Ziziphus spina-christi (see Christ's thorn)
Zone-to-zone projection in olfactory bulb 93